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BHAS 42 CELL TRANSFORMATION ASSAY VALIDATION STUDY REPORT

Series on Testing and Assessment

No. 208

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OECD Environment, Health and Safety Publications

Series on Testing and Assessment

No. 208

BHAS 42 CELL TRANSFORMATION ASSAY VALIDATION STUDY REPORT

IOMC

INTER-ORGANIZATION PROGRAMME FOR THE SOUND MANAGEMENT OF CHEMICALS

A cooperative agreement among **FAO, ILO, UNDP, UNEP, UNIDO, UNITAR, WHO, World Bank and OECD**

Environment Directorate
ORGANISATION FOR ECONOMIC CO-OPERATION AND DEVELOPMENT
Paris 2014

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**OECD Environment Directorate,
Environment, Health and Safety Division
2 rue André-Pascal
75775 Paris Cedex 16
France**

Fax: (33-1) 44 30 61 80

E-mail: ehscont@oecd.org

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FOREWORD

The project for developing a validation report and a Test Guideline for the Bhas 42 Cell Transformation Assay (CTA) was proposed by Japan and included in the Test Guideline work plan in 2010, after completion of international validation studies. The validation report was subsequently developed and peer-reviewed by the EU Reference Laboratory for Alternatives to Animal Testing (EURL ECVAM) Scientific Advisory Committee (ESAC). The ESAC peer review served as a basis for the development of the EURL ECVAM Recommendation on the Cell Transformation Assay based on the Bhas 42 cell line, publicly available on the EURL ECVAM website.

The validation report of the Bhas CTA was circulated for comments to the Working group of the National Coordinators for the Test Guidelines Programme (WNT) in October 2013. It was discussed by the expert group on cell transformation assays in a meeting held in Paris in January 2014 and subsequently endorsed by the WNT at its 26th meeting in April 2014. The Joint Meeting of the Chemicals Committee and the Working Party on Chemicals, Pesticides and Biotechnology agreed to its declassification on 7th July, 2014.

This document is published under the responsibility of the Joint Meeting of the Chemicals Committee and the Working Party on Chemicals, Pesticides and Biotechnology.

30 July, 2012

Bhas 42 Cell Transformation Assay

Validation Study Report

Validation Management Team

Makoto Hayashi	Biosafety Research Center, Foods, Drugs and Pesticides (Iwata, Shizuoka, Japan)
Hajime Kojima	Japanese Center for the Validation of Alternative Methods, National Institute of Health Sciences (Setagaya, Tokyo, Japan)
Raffaella Corvi	European Centre for the Validation of Alternative Methods (Ispra, Italy)
William Stokes	Interagency Coordinating Committee on the Validation of Alternative Methods, the National Toxicology Program Interagency Center for the Evaluation of Alternative Toxicological Methods, National Institute of Environmental Health Sciences (Research Triangle Park, North Carolina, USA)
Abigail Jacobs	Food and Drug Administration (Rockville, Maryland, USA)
Takeshi Morita	National Institute of Health Sciences (Setagaya, Tokyo, Japan)
Leonard Schechtman	Innovative Toxicology Consulting LLC (Lake Worth, Florida, USA)
Masaya Suzuki	Biosafety Research Center, Foods, Drugs and Pesticides (Iwata, Shizuoka, Japan)

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List of Abbreviations

BSRC	Biosafety Research Center, Foods, Drugs and Pesticides
CAS no.	Chemical Abstract Service registry number
CTA	Cell Transformation Assay
CV	Crystal violet
DF5F	Dulbecco's modified Eagle's medium/Ham's F12 supplemented with 100 units/mL of penicillin, 100 µg/mL of streptomycin and 5% FBS
DMSO	Dimethyl Sulfoxide
DRP	Detailed review paper
DRP 31	Detailed Review Paper on Cell Transformation Assays for Detection of Chemical Carcinogens, OECD Environment, Health and Safety Publications, Series on Testing and Assessment, No. 31 [OECD, 2007]
ECVAM	European Centre for the Validation of Alternative Methods
FBS	Fetal Bovine Serum
FDSC	Food and Drug Safety Center
Harlan	Harlan Cytotest Cell Research GmbH
HRI	Hadano Research Institute
IARC	International Agency for Research on Cancer
IC ₅₀	50% inhibitory concentration
IC ₉₀	90% inhibitory concentration
ICCVAM	Interagency Coordinating Committee on the Validation of Alternative Methods
ID	Identification
JaCVAM	Japanese Centre for the Validation of Alternative Methods
JBRC	Japan Bioassay Research Center
M10F	Eagle's minimum essential medium supplemented with 100 units/mL of penicillin, 100 µg/mL of streptomycin and 10% fetal bovine serum
MCA	3-Methylcholanthrene
Mitsubishi	Mitsubishi Chemical Medience
MNNG	<i>N</i> -methyl- <i>N'</i> -nitro- <i>N</i> -nitrosoguanidine
NEDO	New Energy and Industrial Technology Development Organization
NIHS	National Institute of Health Sciences
NOEL	No-effect level
NTP	National Toxicology Program
OECD	Organization for Economic Co-operation and Development
SHE	Syrian Hamster Embryo
TPA	12- <i>O</i> -tetradecanoylphorbol-13-acetate

1 **Modular approach**

2
3 The European Centre for the Validation of Alternative Methods (ECVAM) developed a
4 modular approach to the validation of alternative methods, where the various
5 information requirements for peer-review and as generated during the validation
6 process are broken down into 7 independent modules. In this report, module 1 (test
7 definition) is described in section 1, Rationale for the proposed test, section 2,
8 Organization of the study, sections 3.1.1 and 4.1.1, Study design (on the 6-well method
9 and on the 96-well method, respectively) and sections 3.2 and 4.2, Protocol, and briefly
10 mentioned in section 5, General discussion. After the presentation of assay results in
11 sections 3.3 and 4.3, modules 2-5 (within-laboratory reproducibility, transferability,
12 between-laboratory reproducibility and predictive capacity) are assessed in sections 3.4
13 and 4.4, Discussion and section 5, General discussion. Modules 6 and 7 (applicability
14 domain and performance standards) are not addressed in this report.

15

1 Preface

2
3 The Organisation for Economic Co-operation and Development (OECD) has published a
4 detailed review paper (DRP) on the cell transformation assays (CTAs) [OECD, 2007],
5 where Syrian Hamster Embryo (SHE) and BALB/c 3T3 CTAs are dealt with as assays
6 having potential to detect non-genotoxic as well as genotoxic carcinogens. Meanwhile, a
7 new CTA using the Bhas 42 cells which were clonally derived from the BALB/c 3T3 cells
8 transfected with *v-Ha-ras* gene was developed at about the same time as the
9 compilation of the review. Hadano Research Institute (HRI), Food and Drug Safety
10 Center (FDSC) undertook a research project financed by the New Energy and Industrial
11 Technology Development Organization (NEDO, Japan) to refine, evaluate and validate
12 the Bhas 42 CTA. HRI applied the assay to various chemicals under the project, and the
13 accumulated data showed that the assay was promising to predict chemical
14 carcinogenicity. The study presented in this report was executed to validate the Bhas 42
15 CTA in a formal inter-laboratory validation study. Two protocol variants of the Bhas 42
16 CTA were considered in this study: the 6-well plate and the 96-well plate protocols. The
17 ultimate goal of this study is to develop an OECD Test Guideline for the Bhas 42 CTA.
18 The study was supervised by the Validation Management Team (VMT) established by
19 the Japanese Centre for the Validation of Alternative Methods (JaCVAM). This report
20 includes a short introduction on the context and background of the study, the
21 presentation of the results generated in the validation study and the conclusions by the
22 VMT.
23

1 Summary

2
3 Carcinogenicity is a key element in the hazard and risk assessment of chemicals for
4 human health and has traditionally been assessed using the chronic carcinogenicity
5 bioassay in rodents. Rodent carcinogenicity tests are costly in terms of time, money, and
6 the large number of animals sacrificed. With the aims of reducing the number of rodent
7 bioassays conducted, as well as the associated time and costs, while maintaining
8 efficient discrimination of carcinogens from non-carcinogens, several *in vitro*
9 alternatives have been developed. Among them, *in vitro* genotoxicity tests have been
10 routinely employed to provide adjunct data with which to screen chemicals for
11 carcinogenicity. However, direct DNA damage is but one of mechanisms involved in
12 chemical carcinogenesis, since some carcinogens are non-genotoxic. *In vitro* cell
13 transformation mimics multistage carcinogenesis in experimental animals, and cell
14 transformation assays (CTAs) can detect both non-genotoxic as well as genotoxic
15 carcinogens. The conventional CTAs have been carried out using primarily Syrian
16 hamster embryo (SHE) cells, BALB/c 3T3 cells and C3H/10T1/2 cells. The Bhas 42 cells
17 were established from the BALB/c 3T3 cells by the transfection of v-Ha-*ras* gene more
18 than twenty years ago and postulated to be an initiated cell population in the two-stage
19 carcinogenesis theory. Using the Bhas 42 cells, a sensitive short-term CTA was
20 developed which is capable of detecting both tumor-initiating and tumor-promoting
21 activities of chemical carcinogens.

22
23 The evolution of the Bhas 42 CTA has spawned two methods. One is a 6-well method
24 where the assay is carried out using 6-well micro-plates and the other is a 96-well
25 method where 96-well micro-plates are used. The Bhas 42 CTA was developed, refined
26 and applied to various chemicals using 6-well micro-plates. Later the 96-well method
27 was added as a modification that has the potential to be utilized for high throughput
28 automated applications. The protocol of each of the 6-well and 96-well methods consists
29 of two assay components, the initiation assay and the promotion assay, to detect the
30 tumor-initiating activity and the tumor-promoting activity of chemicals, respectively.
31 The assay procedures are fundamentally the same between the 6-well and 96-well
32 methods. The cells are apportioned relative to each well-size (6-well micro-plate or
33 96-well micro-plates), cultured and treated with a test chemical for the same duration
34 and the production of transformed foci is observed.

35
36 Hadano Research Institute (HRI), Food and Drug Safety Center (FDSC) undertook a
37 New Energy and Industrial Technology Development Organization (NEDO) Project in
38 Japan. Among the purposes of this large project was to refine, evaluate and validate the
39 Bhas 42 CTA. HRI finished three validation studies, pre-validation and validation
40 studies on the 6-well method and a validation study on the 96-well method. The report
41 of the pre-validation study on the 6-well method (available elsewhere) is excluded from
42 this report since a validation management team (VMT) was not organized for that study.
43 Nevertheless those study results are compared with those of other validation studies in
44 section 5, General discussion. This validation study report provides details on the latter
45 two validation studies (6-well method and 96-well method) that were supervised by the
46 CTA Advisory Committee and VMT.

47
48 The validation study performed on the 6-well method involved six laboratories from
49 three countries. Twelve coded chemicals were examined in total and each chemical was
50 tested by three laboratories. In the initiation assay, concordant results were obtained by
51 three laboratories for eight out of ten chemicals and in the promotion assay, concordant

1 results were achieved for ten of twelve chemicals. The 6-well method discriminated all
2 six carcinogens and two tumor-promoters from four non-carcinogens. Thus, the present
3 study demonstrated that the Bhas 42 CTA using 6-well micro-plates is transferable
4 between laboratories, reproducible between laboratories and applicable to the
5 prediction of chemical carcinogenicity. In addition, by comparison of the present results
6 with intra-laboratory data previously published, within-laboratory reproducibility in
7 the 6-well method was also confirmed.

8
9 The validation study performed on the 96-well method involved four participating
10 laboratories. The study was conducted in a stepwise manner (pre-validation phase,
11 validation phase I and validation phase II). 3-Methylcholanthrene (MCA) and
12 12-*O*-tetradecanoylphorbol-13-acetate (TPA) were examined in the pre-validation phase,
13 and coded chemicals were tested in the validation phases I and II. A total of 25
14 chemicals were tested including duplicate chemicals between phases. In the
15 pre-validation phase, the respective results of MCA and TPA were quite similar among
16 all four laboratories. In validation phase I, seven coded chemicals were examined and
17 each chemical was tested by the four laboratories. In the initiation assay, concordant
18 results were obtained by four laboratories for six chemicals. In the promotion assay,
19 concordant results among all four laboratories were achieved for four chemicals and for
20 the remaining three chemicals concordant results were obtained by three laboratories.
21 In validation phase II, 16 coded chemicals were examined and each chemical was tested
22 by two laboratories. The results were concordant between the two respective
23 laboratories for all 16 chemicals. MCA and TPA were tested under coded conditions in
24 phase I, and the results were quite similar to those in the pre-validation phase.
25 Benzo[*a*]pyrene was examined in phases I and II and the judgments were identical
26 between both phases in every laboratory. The transformation frequency of each positive
27 control was statistically significantly different from that of corresponding negative
28 control and the cluster of transformation frequency in positive controls was quite
29 separate from that in negative controls. Overall, 10 out of 12 carcinogens and
30 tumor-promoters were positive and 8 out of 9 non-carcinogens were negative in the
31 96-well method of Bhas 42 CTA. These results proved that the 96-well method is robust,
32 transferable between laboratories, reproducible both within and between laboratories,
33 and applicable to the prediction of chemical carcinogenicity.

34
35 The 6-well method and the 96-well method produced the same results for 15 out of the
36 17 chemicals duplicated between the validation study on the 6-well method and that of
37 the 96-well method.

38
39 The VMT reviewed the results of the validation studies on the 6-well method and the
40 96-well method.

41

1 Rationale for the proposed test

3 1.1 Introduction

5 The assessment of carcinogenicity is one of the most important issues in the regulation
6 of chemicals. Carcinogenicity is traditionally assessed using chronic carcinogenicity
7 bioassays performed in rodents. Standard carcinogenicity tests are performed using
8 hundreds of mice and rats and incur large expenditures in terms of cost and time,
9 usually 2 years. The new European chemical legislation (REACH, Registration,
10 Evaluation, Authorization and Restriction of Chemicals), which limits animal use for
11 safety assessment, has recently been enacted. Situations such as this, other regulatory
12 global requirements, and public pressures necessitate the use of validated cost-effective
13 *in vitro* alternative test methods that can reduce/replace the use of animals for the
14 assessment of chemical carcinogenicity. Genotoxicity tests (*in vivo* and *in vitro*) have
15 come to serve as adjuncts to routine *in vivo* carcinogenicity tests, both for screening
16 purposes and prediction purposes and to help resolve mechanism(s) of action. However,
17 a significant number of carcinogens are considered to be non-genotoxic [Hernández *et*
18 *al.*, 2009]. In order to avoid this limitation by using standard genotoxicity tests, it is
19 desirable that a more encompassing reliable *in vitro* test other than the genotoxicity
20 assays be included in the battery of short-term assays for the prediction of chemical
21 carcinogenicity. An *in vitro* cell transformation assay (CTA) is such a unique system.
22 The CTA, of which there are several, is an assay measuring the phenotypic conversion
23 from normal to malignant characteristics in cultured mammalian cells exposed to test
24 chemicals. Among the available CTAs, focus formation assays, generally using BALB/c
25 3T3 cells or C3H 10T1/2 cells, and colony forming assays, most often using Syrian
26 hamster embryo (SHE) cells, have been acknowledged as the principal *in vitro*
27 screening transformation methods for chemical carcinogens [Kakunaga and Yamasaki,
28 1985; Montesano *et al.*, 1986]. However, these CTAs have not yet been fully embraced by
29 regulatory authorities for routine use in the regulation of potential chemical
30 carcinogens [IARC/NCI/EPA Working Group, 1985; OECD, 2007].

31
32 Although a two-stage theory of chemical carcinogenesis has been hypothesized
33 [Berenblum I, 1941b], at present, it is considered to involve multiple stages. However,
34 the two-stage model of carcinogenesis can serve as a suitable, though abridged,
35 representation of multi-stage carcinogenesis. The two-stage carcinogenesis model been
36 described as being comprised of an initiation stage and a promotion stage. In the
37 initiation stage, a target cell is damaged with a genotoxic carcinogen (tumor-initiator),
38 the damage is fixed as a mutation in the genome, and the fixed mutation is never
39 reversed and is retained in the genome. Some of those mutations render the target cells
40 predisposed for carcinogenesis although they are not yet cancerous. The process to this
41 point is the initiation stage and such cells are termed initiated cells. In the promotion
42 stage, the initiated cell is stimulated with a non-genotoxic carcinogen such as TPA to
43 proliferate and proceed to tumor formation. The carcinogen that promotes the initiated
44 cell toward tumor formation is termed a tumor-promoter. Thus, the tumor-promoter is
45 non-genotoxic and does not produce tumors by itself, but promotes carcinogenesis in
46 initiated cells. This two-stage carcinogenesis theory serves to explain the phenomenon
47 in mouse skin that neither treatment with a sub-threshold dose of a genotoxic
48 carcinogen, such as benzo[a]pyrene, alone, nor treatment with a tumor-promoter, such
49 as croton oil, alone produced tumors but that a sequential treatment with a
50 sub-threshold dose of a genotoxic carcinogen and followed by treatment with a
51 tumor-promoter produced tumors [Berenblum, 1941a]. The two-stage carcinogenesis

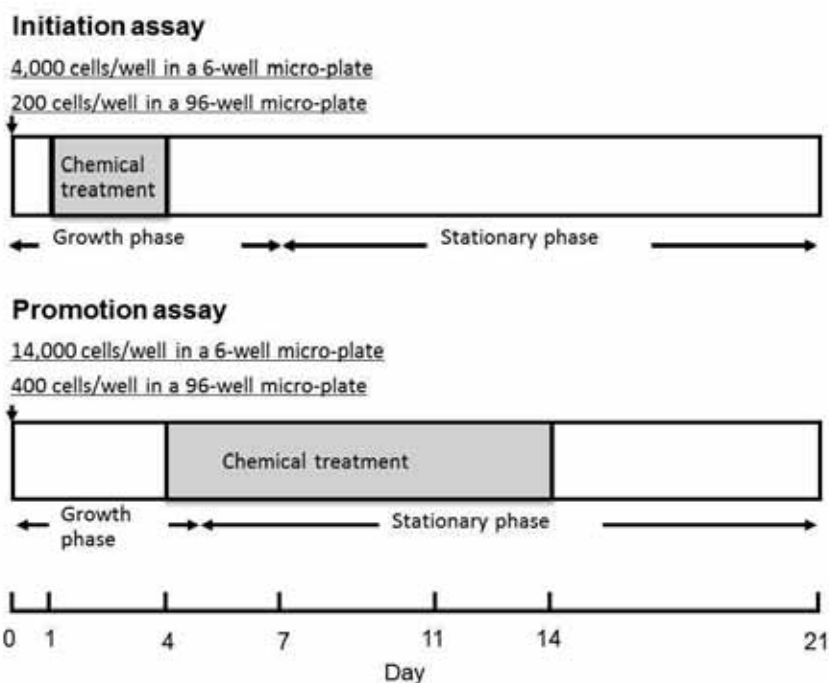
1 progression has been reported in other organs and in other experimental animals as
2 well as in mouse skin [Beebe *et al.*, 1995; Nishiwaki-Matsushima *et al.*, 1991; Reddy *et*
3 *al.*, 1977; Sakai, 2008]. Many non-genotoxic carcinogens are considered to be
4 tumor-promoters, since somatic cells can be initiated by a small amount of
5 environmental and/or endogenous genotoxic carcinogens. Meanwhile, tumor-promoters
6 are different from receptor-mediated non-genotoxic carcinogens which are able to cause
7 tumors without initiation by a genotoxic carcinogen.

8
9 Cell transformation *in vitro* is regarded as a model system for carcinogenesis *in vivo*
10 [Smets, 1980; Barrett *et al.*, 1986; Landolph, 1985]. Initiation and promotion stages
11 have been described in cell transformation just like the two-stages in mouse skin
12 carcinogenesis [Mondal *et al.*, 1976; Hirakawa *et al.*, 1982]. A sub-threshold dose of a
13 genotoxic carcinogen (tumor-initiator) alone does not cause cell transformation in the
14 cultures nor does any tumor-promoter alone. When the cells are treated with a
15 sub-threshold dose of a genotoxic carcinogen and then with a tumor-promoter, however,
16 transformation is induced in the target cells. Therefore, two-stage CTAs that mimic a
17 two-stage carcinogenesis test *in vivo* can be performed using BALB/c 3T3 and C3H
18 10T1/2 cells. In the two-stage CTAs, the target cells are initiated with a sub-threshold
19 dose of a known genotoxic carcinogen prior to treatment with a test chemical. In this
20 way, many non-genotoxic carcinogens can be detected as promoters of cell
21 transformation [Kuroki and Sasaki, 1985; Sakai, 2008; Sakai *et al.*, 2002]. Generally,
22 genotoxic carcinogens can be detected as transforming agents by conventional
23 (one-stage) CTAs and as initiators by two-stage CTAs [IARC/NCI/EPA Working Group,
24 1985; Sakai and Sato, 1989; Tsuchiya *et al.*, 1999].

25
26 The Bhas 42 CTA is a sensitive short-term system that has reduced associated cost and
27 labor compared with those associated with the conventional BALB/c 3T3 and
28 C3H10T1/2 CTAs [Sasaki *et al.*, 2012]. Bhas 42 cells were developed from BALB/c 3T3
29 cells through the transfection with a plasmid pBR322 containing Ha-MuSV-DNA, clone
30 H1 (v-Ha-*ras*) [Dhar *et al.*, 1982; Ellis *et al.*, 1980; Sasaki, K. *et al.*, 1988]. It has been
31 confirmed that the cell population still retains v-Ha-*ras* gene (presently at passage 18)
32 and m-RNA of the viral *ras* gene is expressed in the cultures as well [Tanabe *et al.*, in
33 preparation]. The Bhas 42 cells are transformed by known tumor-promoters, including
34 12-*O*-tetradecanoylphorbol-13-acetate (TPA), okadaic acid and lithocholic acid, without
35 initiating treatment with a known tumor-initiator such as 3-methylcholanthrene (MCA)
36 [Omori *et al.*, 2004], and are presumed to be initiated toward transformation by the
37 introduced *ras* sequence [Sasaki *et al.*, 1990]. Using the Bhas 42 cells, Omori *et al.*
38 developed a short-term CTA to identify tumor-promoters [Omori *et al.*, 2004]. The Bhas
39 42 CTA does not require pretreatment with an initiator to detect tumor-promoting
40 activity, and the assay period is 3 weeks instead of 6 weeks, as is the case with the
41 BALB/c 3T3 two-stage CTA [Sakai, 2008]. Because of the increased sensitivity of cells to
42 transformation, the assay scales such as the dish size, the dish number, the medium
43 volume, the serum concentration in the cultures and the number of medium exchanges
44 have been diminished, resulting in a reduction in the cost and labor expended. Asada *et*
45 *al.* [2005] modified the Bhas 42 CTA such that it was capable of detecting
46 tumor-initiating activity as well as tumor-promoting activity of chemicals. The current
47 protocol consists of two assay components, the initiation assay and the promotion assay,
48 to detect the tumor-initiating activity and the tumor-promoting activity of chemicals,
49 respectively. In the initiation assay, the cells are inoculated at a low density and treated
50 with a test chemical in the beginning of the assay period so that the target cells can
51 undergo cell division several times before reaching contact inhibition of growth at
52 confluence, and thus DNA damage is fixed as mutations in the genes. In the promotion

1 assay, the cells are seeded more densely than in the initiation assay, and the treatment
2 with a test chemical is started at sub-confluence and continued beyond confluence, and
3 thus longer than in the initiation assay.

4
5 HRI undertook a research project financed by NEDO to characterize, evaluate, refine
6 and validate the Bhas 42 CTA. Under the project, the Bhas 42 CTA has been applied to
7 98 chemicals including carcinogens and non-carcinogens. The results have been
8 published in Mutation Research [Sakai et al., 2010], the PDF file of the article is
9 presented as Annex 16. The results confirm that the performance of the assay for the
10 prediction of chemical carcinogenicity is superior or equivalent to that of conventional
11 genotoxicity assays and that the assay is capable of detecting Ames-negative and
12 Ames-discordant carcinogens in addition to Ames-positive carcinogens.



16
17
18 Fig. 1. Illustration of the Bhas 42 cell transformation assay methodology.

19
20
21
22 The above-mentioned studies on the Bhas 42 CTA, i.e. development, improvement and
23 application to 98 chemicals, were performed using 6-well micro-plates (6-well method).
24 Meanwhile, the Bhas 42 CTA using 96-well micro-plates (96-well method) has been
25 developed in anticipation of its potential utility for high throughput automated
26 applications. The assay procedures are fundamentally the same between the 6-well and
27 96-well methods as shown in Fig. 1. The cells are seeded in proportion to the respective
28 well-sizes, 6-well and 96-well micro-plates, cultured and treated with a test chemical on
29 the very same schedule and the induction of transformed foci is observed.
30 Quantification of transformation frequency, however, is different, i.e., in the 6-well
31 method, the number of transformed foci in every well of a given plate is recorded, and in
32 the 96-well method, the number of wells in a given plate having transformed foci is

1 recorded. Accordingly, the statistical analysis of each is different between the two
2 methods. An analysis by multiple comparison using the Dunnett test is performed in
3 the 6-well method and the chi-square test with Bonferroni adjustment is carried out in
4 the 96-well method.

5
6 Three validation studies on the Bhas 42 CTA, a pre-validation study of the 6-well
7 method, a validation study of the 6-well method and a validation study of the 96-well
8 method, were carried out under the NEDO project.

9
10 The pre-validation study of 6-well method involved six laboratories and nine coded
11 chemicals were examined. Each chemical was tested by three laboratories. The
12 pre-validation study results have been published previously [Tanaka *et al.*, 2009], and
13 are excluded from this report since a VMT was not organized for the pre-validation
14 study. But its results are compared with those of other validation studies in section 5,
15 Discussion.

16
17 The validation study of the 6-well method involved six laboratories from three countries,
18 USA, Germany and Japan, and twelve coded chemicals were tested. Each chemical was
19 assayed by three laboratories. This study is included in this validation report. The
20 summary of results has been published by the Executive Team and the participating
21 laboratories [Sakai *et al.*, 2011].

22
23 The validation study of 96-well method was proceeded in a stepwise manner
24 (pre-validation phase, validation phase I and validation phase II) and included four
25 laboratories in Japan. MCA and TPA were examined in the pre-validation phase, and
26 coded chemicals were tested in validation phases I and II. A total of 25 chemicals were
27 tested including duplicate chemicals between phases. Each chemical was assayed by
28 four laboratories in the pre-validation phase and validation phase I studies, and by two
29 laboratories in the validation phase II studies.

30
31 The Bhas 42 cells were established by Sasaki *et al.* in the National Institute of Health
32 Sciences, Japan [Sasaki, *et al.*, 1988]. The cells were propagated and at present are
33 stored at HRI, FDSC (Hadano, Japan) and Japanese Collection of Research
34 Bioresources (JCRB) Cell Bank, National Institute of Biomedical Innovation (Osaka,
35 Japan). Those cells have been confirmed to be free from mycoplasma and adequate for
36 transformation assays. They are distributed to worldwide laboratories through Health
37 Science Research Resources Bank (HSRRB, Osaka, Japan)
38 [http://cellbank.nibio.go.jp/cellbank_e.html].

39
40 The current report, which was prepared by HRI with the support of the VMT, presents
41 the outcome of the validation studies on the 6-well method and the 96-well method in
42 the Bhas 42 CTA under the NEDO Project.

43 44 45 46 **1.2 Objective and goals**

47
48 The aim of these studies was to scientifically validate the Bhas 42 CTA in formal
49 inter-laboratory studies in order to assess the within- and between-laboratory
50 reproducibility of the assay, its transferability and its predictive capacity. The ultimate
51 goal is to demonstrate its utility for adoption as an OECD Test Guideline that could be

1 used as a component of a test battery for the prediction of chemical carcinogenicity. In
2 this respect, the Bhas 42 CTA should supplement conventional genotoxicity tests with
3 the ability of detecting non-genotoxic carcinogens including tumor-promoters. The
4 present objective was to assess the transferability, reliability (reproducibility within and
5 between laboratories) and relevance (predictive capacity) of this test system with a set
6 of coded chemicals whose carcinogenicities are known *in vivo*. It is recommended that
7 both of the 6-well method and the 96-well method should be integrated in the OECD
8 test guideline so as to call attention to the availability of both variations and their
9 different potential applications (i.e. manual procedures or automated procedures).

13 1.3 Intended use

15 The Bhas 42 CTA is expected to have its greatest impact as a component of a battery
16 with genotoxicity assays for the prediction of chemical carcinogenicity. No single
17 genotoxicity assay is capable of predicting or assessing all chemicals for their
18 carcinogenic potential. DNA damage is not the only mechanism involved in
19 carcinogenesis. Some carcinogens are not genotoxic [Hernández *et al.*, 2009].
20 Non-genotoxic events, which are at least initially independent of direct DNA damage,
21 can play a critical role in chemical carcinogenesis. The strength of the Bhas 42 CTA is
22 that it (a) can detect both genotoxic and non-genotoxic carcinogens [Asada *et al.*, 2005],
23 (b) is sensitive to tumor-promoters [Omori *et al.*, 2004, 2005], (c) discriminates
24 tumor-initiating activity and tumor-promoting activity of carcinogens [Sakai, 2008].
25 When applied to mycotoxins derived from *Fusarium* and related substances, it was
26 expected that the Bhas 42 CTA would show a good correlation with an *in vivo* two-stage
27 carcinogenesis test using experimental animals for estimation of the tumor-promoting
28 activity [Sakai *et al.*, 2007]. In the in-house assay of 98 chemicals, the Bhas 42 CTA
29 could identify a considerable number of carcinogens which are negative or discordant in
30 the Ames assay [Sakai *et al.*, 2010].

34 1.4 Current use

36 The Bhas 42 cells have been used for academic studies in Japan. The role of cytokines in
37 tumor-promotion and the mechanisms of stomach cancer caused by *Helicobacter pylori*
38 have been investigated utilizing the Bhas 42 cells as well as BALB/c 3T3 cells, though
39 the protocols are different from those proposed in the present validation report [Komori
40 *et al.*, 1993; Suganuma *et al.*, 1999, 2001, 2005]. The Bhas 42 CTA has been applied by
41 researchers in Kanagawa Prefectural Institute of Public Health (Japan) and National
42 Institute of Health Sciences (Japan) to abietic acid and dehydroabietic acid, which are
43 the components of resin from a needle-leaf tree and possible contaminants in paper and
44 paperboard for food contact use, and those resin components have been shown to have
45 positive activity in the promotion assay [Ohmori and Kawamura, 2009]. In-house use of
46 the Bhas 42 CTA by industry for safety assessment of chemicals, pharmaceuticals,
47 cosmetics, pesticides and biocides is limited, since, at present, such companies tend to
48 employ the conventional genotoxicity assays that are accepted by regulatory authorities.
49 It is expected that the utilization of Bhas 42 CTA will be expanded as a consequence of
50 the development of an OECD Test Guideline.

1
2 **1.5 Recent research**
3

4 The Bhas 42 CTA was assessed in-house against 98 chemicals in order to characterize it
5 and evaluate its performance for the detection of chemical carcinogens [Sakai *et al.*,
6 2010]. When the assay results were compared with the existing genotoxicity data, the
7 Bhas 42 CTA could detect a considerable number of the carcinogens that are negative or
8 discordant in the Ames test, and most of those Ames-negative and Ames-discordant
9 carcinogens were detected by the promotion assay in the Bhas 42 CTA. This fact
10 suggested that the Bhas 42 cells behaved as initiated cells in CTA. The performance
11 indices were calculated from the assay results of 52 carcinogens and 37 non-carcinogens
12 (The other nine chemicals were of unknown carcinogenicity). The concordance was 78%,
13 sensitivity 73%, specificity 84%, positive predictivity 86%, negative predictivity 69%,
14 false negatives 27% and false positives 16%. Of these values, the concordance, specificity,
15 negative predictivity and false positives were superior to those of conventional
16 genotoxicity tests and the other performance indices were equivalent to those of
17 conventional genotoxicity tests. From these results, the authors concluded that the
18 overall accuracy of prediction of chemical carcinogenicity would be improved by
19 introducing the Bhas 42 CTA into the standard battery of *in vitro* assays.
20
21
22

23 **1.6 Published data on inter-laboratory reproducibility**
24

25 No formal between-laboratory trial had been previously conducted to assess the
26 transferability and reproducibility of the Bhas 42 CTA and its applicability to the
27 prediction of chemical carcinogenicity. However, there were two inter-laboratory
28 collaborative studies conducted prior to the validation studies presented in the current
29 report. The first study, which involved 14 participating laboratories, was conducted on
30 the promotion assay by the Non-genotoxic Carcinogen Study (NGCS) Group in the
31 Environmental Mutagen Society of Japan [Omori *et al.*, 2005]. After confirmation that
32 these laboratories could obtain positive results with two tumor-promoters, TPA and
33 lithocholic acid, 12 coded chemicals were assayed. Each chemical was tested in four
34 laboratories. For eight chemicals, all four laboratories obtained consistent results, and
35 for two of the other four chemicals, one of the four laboratories showed inconsistent
36 results. It should be noted that only the promotion assay of Bhas 42 CTA had been
37 developed at this stage. The second study was carried out by HRI as the lead laboratory,
38 after the Bhas 42 CTA protocol was modified so as to detect tumor-initiating activity in
39 addition to tumor-promoting activity. This second study was conducted as a part of the
40 NEDO Project and is referred to as “a pre-validation study of the 6-well method” in this
41 report. The pre-validation study has been published [Tanaka *et al.*, 2009], and the
42 results are compared with those of the validation studies on the 6-well method and on
43 the 96-well method in section 5 General discussion.
44
45
46

47 **1.7 Patents**
48

49 The test method and the cells employed in this study are in the public domain and have
50 not been patented.
51

2. Organization of the study

2.1 Executive team

The current validation study was carried out by HRI as a part of the project entrusted from NEDO.

Executive Team

Noriho Tanaka (Director), HRI, FDSC (Kanagawa, Japan)

Makoto Umeda, HRI, FDSC

Ayako Sakai, HRI, FDSC

Assistant

Shoko Arai, HRI, FDSC (until December 31, 2010)

2.2 Validation management team

Validation management team

Chairman Makoto Hayashi (Biosafety Research Center, Foods, Drugs and Pesticides (BSRC), Shizuoka, Japan)

Representative of JaCVAM Hajime Kojima (Japanese Center for the Validation of Alternative Methods (JaCVAM), National Institute of Health Sciences (NIHS), Tokyo, Japan)

Representative of ECVAM Raffaella Corvi (European Centre for the Validation of Alternative Methods (ECVAM), Ispra, Italy)

Representative of ICCVAM William Strokes (Interagency Coordinating Committee on the Validation of Alternative Methods (ICCVAM), the National Toxicology Program Interagency Center for the Evaluation of Alternative Toxicological Methods, National Institute of Environmental Health Sciences, North Carolina, USA)

Representative of ICCVAM (Provisional, 2008) Abigail Jacobs (Food and Drug Administration, Maryland, USA)

Expert Takeshi Morita (NIHS)

Expert Leonard Schechtman (Innovative Toxicology Consulting LLC, Florida, USA)

Statistician Masaya Suzuki (BSRC) until July 31, 2010

*Advisory Committee**

Chairman Makoto Hayashi (NIHS; currently BSRC)

Expert Raffaella Corvi (ECVAM)

Representative JaCVAM Hajime Kojima (JaCVAM, NIHS)

Expert Takeshi Morita, (NIHS)

Statistician Sebastian Hoffmann, (ECVAM; currently seh consulting + services, Germany)

* This Committee was active in the beginning of the validation study on Bhas 42 CTA using 6-well micro-plates and reorganized to eventually become the formal VMT on 2008.10.29.

2.3 Participating laboratories

The validation study of the 6-well method included six laboratories, four in Japan, one from the USA and one from Germany; the validation study of the 96-well method included four Japanese laboratories. Those laboratories are listed below, and their data are shown in Tables and Figures under lab identification (ID) codes presented in Table 1. HRI, Mitsubishi Chemical Medience (Mitsubishi), BSRC and Japan Bioassay Research Center (JBRC) had considerable expertise in the Bhas 42 CTA, since they had participated in the pre-validation study on the Bhas 42 CTA (6-well method) [Tanaka et al., 2009]. Harlan Cytotest Cell Research (Harlan) had a little experience and BioReliance had limited knowledge of the Bhas 42 system, although they had taken part in the ECVAM validation study on the BALB/c 3T3 and SHE CTAs. The latter two laboratories were included in order to involve “naive” laboratories experienced in CTAs but not specifically in the Bhas 42 CTA.

6-well method validation study

- Laboratory I: HRI (Kiyoshi Sasaki, Dai Muramatsu, Kumiko Hayashi, Nobuko Endou, Sachiko Kuroda; Kanagawa, Japan)
 Laboratory II: Mitsubishi (Hideki Hirose, Nana Ishii; Tokyo, Japan)
 Laboratory III: BSRC (Fukutaro Mizuhashi, Sawako Kasamoto, Miho Nagai; Shizuoka, Japan)
 Laboratory IV: JBRC (Masumi Asakura; Kanagawa, Japan)
 Laboratory V: BioReliance Corporation (Kamala Pant, Shannon W. Bruce, Jamie E. Sly; Rockville, Maryland, Japan)
 Laboratory VI: Harlan (Albrecht Poth, Susanne Bohnenberger, Thorsten Kunkelmann; Rossdorf, Germany)

96-well method validation study

- Laboratory 1: BSRC (Fukutaro Mizuhashi, Sawako Kasamoto, Miho Nagai, Maiko Takai; Shizuoka, Japan)
 Laboratory 2: JBRC (Masumi Asakura; Kanagawa, Japan)
 Laboratory 3: HRI (Kiyoshi Sasaki, Dai Muramatsu, Nobuko Endou; Kanagawa, Japan)
 Laboratory 4: Mitsubishi (Nobuhiko Tashiro, Nana Ishii; Tokyo, Japan)

Table 1
 Participating laboratories, the validation studies to which they contributed, and their identification (ID) codes in respective studies

Laboratory	6-Well method	96-Well method		
		Pre-validation	Phase I	Phase II
HRI	Lab I	Lab 3	Lab 3	Lab 3
Mitsubishi	Lab II	Lab 4	Lab 4	--*
BSRC	Lab III	Lab 1	Lab 1	Lab 1
JBRC	Lab IV	Lab 2	Lab 2	Lab 2
BioReliance	Lab V	--	--	--
Harlan	Lab VI	--	--	--

* Not participated

1
2
3 **2.4 Quality assurance schemes employed by the testing laboratories**
4

5 The present study was conducted under conditions that closely approximated Good
6 Laboratory Practice (GLP) by all laboratories.
7

8
9
10 **2.5 Meetings held**
11

12 27/08/2007 (HRI)

13 Workshop for technical training of the Bhas 42 CTA using 6-well micro-plates (6-well
14 method)*

15 Albrecht Poth: RCC (Harlan)

16 Kamala Pant: BioReliance

17 Kiyoshi Sasaki: HRI.

18 Subjects: explanation of the Bhas 42 CTA and its protocol, technical training and
19 judgment of transformed foci.

20 * Since the other participating laboratories had experience in the 6-well method, they
21 did not take part in this workshop.
22

23 10-12/01/2008 (HRI)

24 Meeting of Advisory Committee

25 Makoto Hayashi: NIHS

26 Raffaella Corvi: ECVAM, Italy

27 Hajime Kojima: JaCVAM, NIHS

28 Takeshi Morita: NIHS

29 Sebastian Hoffmann: ECVAM, Italy

30 Plus participating laboratories

31 Subjects: protocol for the 6-well method and consultation on statistical analysis of the
32 data from the 6-well method validation study.
33

34 14/03/2008 (HRI)

35 Meeting on appropriate statistics for the Bhas 42 CTAs using 6-well micro-plates and
36 96-well micro-plates

37 Sebastian Hoffmann (ECVAM)

38 Kiyoshi Sasaki (HRI)

39 Ayako Sakai (HRI)

40 Kumiko Hayashi (HRI)

41 Subject: consultation on statistical methods suitable for data analysis in the 6-well
42 method and the 96-well method.
43

44 18/04/2008 (HRI)

45 Meeting on the validation study of Bhas 42 CTA using 96-well micro-plates (96-well
46 method)

47 Hajime Kojima (JaCVAM)

48 Noriho Tanaka (HRI)

49 Ayako Sakai (HRI)

50 Subject: providing a framework for the validation study on the 96-well method
51

1 15/10/2008 (Kyoto Univ.)
2 Meeting on appropriate statistics for the 96-well method
3 Takashi Omori (Kyoto Univ.)
4 Shoko Arai (HRI)
5 Subjects: preparation of data sheet for the 96-well method validation study and
6 construction of a database for the study
7
8 29-30/10/2008 (HRI)
9 1st VMT Meeting
10 VMT members: Makoto Hayashi, Raffaella Corvi, Hajime Kojima, Takeshi Morita, Abby
11 Jacob
12 Adviser: Takashi Omori (Kyoto Univ. 29 October)
13 Participating laboratories: HRI, Mitsubishi, BSRC, JBRC
14 Subjects: study plan, protocol and test chemicals for the validation study of 96-well
15 method.
16
17 18/11/2008 (HRI)
18 Workshop for technical inter-laboratory transfer of 96-well method protocol
19 Participating laboratories: HRI, Mitsubishi, BSRC, JBRC
20 Subjects: explanation of protocol and technical training
21
22 24/12/2008 (BSRC)
23 Meeting on statistics
24 Takashi Omori (Kyoto Univ.)
25 Masaya Suzuki (BSRC)
26 Shoko Arai (HRI)
27 Subject: decision on the method of statistical analysis for the data in the validation
28 study of 96-well method
29
30 17/01/2009 (Kyoto Univ.)
31 Meeting on statistics
32 Takashi Omori (Kyoto Univ.)
33 Masaya Suzuki (BSRC)
34 Shoko Arai (HRI)
35 Subject: analysis of the results in pre-validation phase of the 96-well method validation
36 study
37
38 26/01/2009 (HRI)
39 Joint meeting of Japanese VMT and participating laboratories about the results of
40 pre-validation phase
41 VMT members: Makoto Hayashi and Hajime Kojima
42 Statistics team: Masaya Suzuki and Shoko Arai
43 Participating Laboratories: HRI, Mitsubishi, BSRC, JBRC
44 Subject: evaluation of results in the pre-validation phase, harmonization of focus
45 identification and counting among laboratories, adoption of assay acceptance criteria*
46 and planning of phase I in the validation study of 96-well method
47 * In accordance with the discussion at this joint meeting, the protocol for the 96-well
48 method was revised (Ver. 3, 2009.1.26)
49
50 15/10/2009 (HRI)
51 Joint meeting of Japanese VMT and participating laboratories regarding the results of
52 phase I

1 VMT members: Makoto Hayashi and Takeshi Morita
2 Statistics team: Masaya Suzuki and Shoko Arai
3 Participating Laboratories: HRI, Mitsubishi, BSRC, JBRC
4 Subject: preliminary evaluation of the results in the phase I validation study of 96-well
5 method
6
7 9-10/11/2009 (NEDO Shiroganedai Training Center)
8 2nd VMT Meeting
9 VMT members: Makoto Hayashi, Hajime Kojima, Raffaella Corvi, William Stokes,
10 Leonard Schechtman, Takeshi Morita:
11 Participating laboratories: HRI, Mitsubishi, BSRC, JBRC
12 Subjects: evaluation of the phase I results and planning of phase II in the 96-well
13 method validation study.
14
15 6-7/10/2010 (NEDO Kawasaki Head Office)
16 3rd VMT Meeting
17 VMT members: Makoto Hayashi, William Stokes, Raffaella Corvi, Hajime Kojima,
18 Leonard Schechtman, Takeshi Morita
19 Participating laboratories: HRI, BSRC, JBRC
20 Subjects: evaluation of the results in the 96-well method phase II validation study and
21 approval of the 96-well method; evaluation of the results in the 6-well method
22 validation study and approval of the 6-well method
23
24 15/9/2011 (Keisuikan, Sinmachi-Campus, Doshisha Univ., Kyoto)
25 Meeting for international Bhas 42 CTA validation study
26 VMT member: Makoto Hayashi, Hajime Kojima, Raffaella Corvi, Takeshi Morita and
27 Leonard Schechtman
28 The other participants: Noriho Tanaka, Makoto Umeda, Ayako Sakai, Kiyoshi Sasaki
29 and Koji Yamakage
30 Subjects: Future schedule on the Bhas 42 CTA to develop OECD Test Guideline: VMT
31 members and the other participants agreed that the Bhas 42 CTA validation study
32 report should undergo peer review by the ESAC through the ECVAM process.
33

3 Validation Study on the 6-well method

The summary of this validation study has been published by the executive team and the participating laboratories [Sakai *et al.*, 2011].

3.1 Study design and chemicals tested

3.1.1 Study design

In the validation study of 6-well method, six laboratories participated. Twelve coded chemicals were tested. Each chemical was assayed by three laboratories so that the inter-laboratory reproducibility could be assessed. The intra-laboratory reproducibility was retrospectively assessed by comparing data between this validation study and pre-validation study reported by Tanaka *et al.* [2009] and also between this validation study and in-house HRI studies [Sakai *et al.*, 2010]. The transferability of the test protocol was determined by comparing the data from HRI, which had developed the current protocol, with those from the other laboratories in this validation study.

A workshop on the technical training necessary for the proper conduct of the Bhas 42 CTA using 6-well micro-plates was held by HRI on 27 August, 2007 before starting the validation study of the 6-well method. The study directors of Harlan and BioReliance participated in the workshop to learn the protocol, the assay procedures and judgment criteria for identifying transformed foci. Since the other participating laboratories had experience in the pre-validation study of 6-well method [Tanaka *et al.*, 2009], they did not take part in this workshop.

3.1.2 Chemicals tested and negative and positive controls

The chemicals were selected so that 2/3 were carcinogens including non-genotoxic ones and 1/3 were non-carcinogens (Tables 2 and 3). Ideally, it would have been good to include chemicals with low potency. However, it is difficult to assess precise potency based on available data. It could be argued that the process of dilution of test chemicals resulting in reduced concentrations may approximate potency reduction so as to mimic the testing of low potency chemicals. The chemical properties and classes of 12 test chemicals and two positive controls are presented in Annex 14. The test chemicals, their Chemical Abstract Service registry numbers (CAS no.), their code names for the validation study and the laboratories that took charge of each chemical are shown in Table 2. Each of the participating laboratories independently determined the appropriate solvent (either water or DMSO) for the test chemicals with the exception of dibenz[*a,h*]anthracene and mezerein. Solvent determination was based upon the results of preliminary solubility testing. The test chemical dose ranges for the transformation assays were selected according to dose setting procedures using the cell growth assays. For the two chemical exceptions, the solvent, dimethyl sulfoxide (DMSO) was pre-selected and the maximum concentrations in culture medium of each were likewise pre-assigned (≤ 10 $\mu\text{g/mL}$ and ≤ 0.01 $\mu\text{g/mL}$, respectively). These amounts were predetermined due to the expense and limited availability of each chemical.

MCA and TPA were chosen as positive controls for the initiation assay and the

1 promotion assay, respectively, because they induce a strong positive response in the
 2 Bhas 42 CTA, have been used as positive controls since the development of Bhas 42 CTA
 3 [Omori *et al.*, 2004; Asada *et al.*, 2005], and are common positive controls in the BALB/c
 4 3T3 and C3H10T1/2 CTA two-stage systems. The historical data of these chemicals as
 5 positive controls were available at the lead laboratory, HRI. The doses of MCA and TPA
 6 (1 µg/mL and 0.05 µg/mL, respectively) to be used as positive controls were determined
 7 based on the HRI historical data.

8
 9
 10
 11 Table 2
 12 Test chemicals, code names and laboratory allocation in the 6-well method validation
 13 study.

14 Chemical	15 CAS no. ^a	16 Carcinogenicity ^b	17 Code	18 Allocation to lab					
				19 I	20 II	21 III	22 IV	23 V	24 VI
25 2-Acetylaminofluorene	53-96-3	+	A	○			○		○
26 Cadmium chloride	10108-64-2	+	F	○	○	○			
27 Dibenz[<i>a,h</i>]anthracene*	53-70-3	+	C	○	○			○	
28 Lithocholic acid	434-13-9	TP ^c	E	○			○		○
29 Methapyrilene HCl	135-23-9	+,TP	H	○	○		○		
30 Mezerein*	34807-41-5	TP	G	○		○	○		
31 Sodium arsenite	7784-46-5	+	D	○		○			○
32 <i>o</i> -Toluidine HCl	636-21-5	+	B	○				○	○
33 Anthracene	120-12-7	-	I	○	○				○
34 L-Ascorbic acid	50-81-7	-	L	○	○		○		
35 Caffeine	58-08-2	-	K	○				○	○
36 D-Mannitol	69-65-8	-	J	○		○	○		
37 3-Methylcholanthrene	56-49-5	+	Positive control for initiation assay						
38 TPA ^d	16561-29-8	+	Positive control for promotion assay						

39 ^a Chemical abstract service registry number.

40 ^b +, carcinogen; -, non-carcinogen; TP, tumor-promoter

41 ^c Tumor-promoter

42 ^d 12-*O*-tetradecanoylphorbol-13-acetate.

43 * The solvent (DMSO) and the highest concentrations in the culture medium
 44 (dibenz[*a,h*]anthracene, 10 µg/mL; mezerein, 0.01 µg/mL) were designated in advance.

45
 46 Data regarding genotoxic and carcinogenic activity, including tumor-promoting activity,
 47 of the test chemicals were garnered from (a) the Detailed Review Paper on Cell
 48 Transformation Assays for Detection of Chemical Carcinogens, OECD Environment,
 49 Health and Safety Publications, Series on Testing and Assessment, No. 31 (DRP 31)
 50 [OECD, 2007], (b) a review by Kirkland *et al.* [2005], and (c) other original publications
 51 [Reddy *et al.*, 1975; Lijinsky *et al.*, 1992]. In DRP 31, the reference data to *in vivo*
 52 carcinogenicity are from well-regarded sources, such as IARC, Gold & Zeiger [1997],
 53 NTP databases. The carcinogenicity and genotoxicity of the test chemicals are presented
 54 in Table 3.

55 The chemicals tested were purchased and coded by the HRI chemical repository
 56 supervisor, who was independent of the participating laboratories and the executive
 57 team, and distributed by him to the chemical repository officer of each individual
 58 participating laboratory. The suppliers, catalog numbers and lot numbers of the test
 59 chemicals are presented in Annex 1 (Table 54).

1
2
3 Table 3
4 Genotoxicity and carcinogenicity of the test chemicals in the 6-well method validation
5 study.

Chemicals	Genotoxicity ^a				IARC class	Carcinogenicity ^b
	<i>In vitro</i>			<i>In vivo</i>		
	Ames ^c	ML ^d	CA ^e	MN ^f		
2-Acetylaminofluorene	+	+	+	+		+
Cadmium chloride	-	+	+/-	-	1	+
Dibenz[a,h]anthracene	+	+	+		2A	+
Lithocholic acid	-	+/-	-			TP ^g
Methapyrilene HCl	-	+/-	+			+, TP ^h
Mezerein	-					TP
Sodium arsenite	-	+		+	1	+
<i>o</i> -Toluidine HCl	+/-					+
Anthracene	+/-	+	-		3	-
L-Ascorbic acid	+w/-		-	+		-
Caffeine	-		+	+	3	-
D-Mannitol	-	-	-	-		-

21 ^a Source: DRP 31 [OECD, 2007] and a review by Kirkland *et al.* [2005]: +, positive; -,
22 negative; +/-, positive and negative results in DRP 31; +w, weakly positive.

23 ^b Source: DRP 31 [OECD, 2007] and original papers: +, carcinogen; -, non-carcinogen;
24 TP, tumor-promoter

25 ^c Ames test

26 ^d Mouse lymphoma test

27 ^e Chromosomal aberration test

28 ^f Micronucleus test

29 ^g Reddy *et al.*, [1975].

30 ^h Lijinsky *et al.*, [1992].

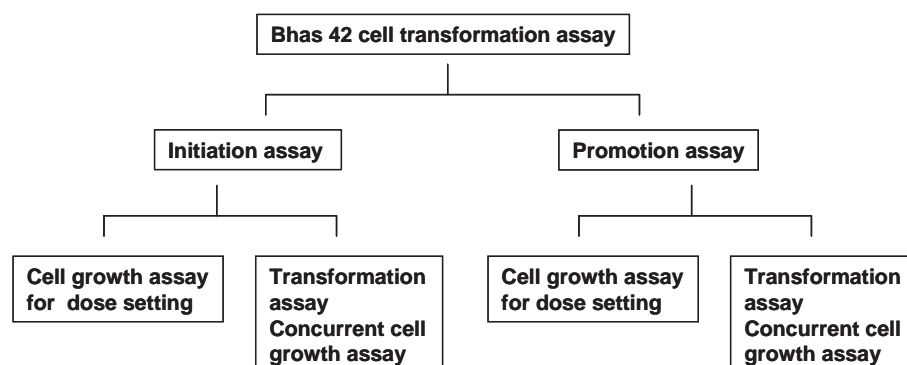
35 3.1.3 Timeline

37 27/08/2007	Workshop for technical transfer of Bhas 42 CTA using 6-well micro-plates.
39 1/10/2007	Start of the 6-well method validation study
40 1-26/10/2007	Discussion, questions and answers about the protocol exchanged between HRI and participating laboratories (via e-mail)
42 26/10/2007	Determination of the protocol for the 6-well method validation study
44 1/11/2007	Shipment of test chemicals to Japanese laboratories
45 12/11/2007	Shipment of test chemicals to overseas laboratories
46 10-12/02/2008	Meeting of Advisory Committee
47 13/10/2009	Last submission of study results: Data submission from Lab 6 was delayed. The other laboratories reported all the data including re-test by October, 2008.
50 28/10/2009	Decoding of the test chemicals.
51 6-7/10/2010	Evaluation and approval of the study results by VMT (in the 3rd VMT Meeting)

1 15/9/2011 Meeting for the international Bhas 42 CTA validation study:
2 Future schedule on the Bhas 42 CTA to develop OECD/TG: VMT
3 members and the other participants agreed that the Bhas 42 CTA
4 validation study report should undergo peer review by the ESAC
5 through the ECVAM process.
6
7
8
9

10 3.2 Protocol

11
12 The transformation assay was performed according to the procedures as described
13 previously [Sakai *et al.*, 2010]. The detailed test protocol used in the 6-well method
14 validation study is presented in Annex 2. Fig. 2 shows a schematic representation of the
15 Bhas 42 CTA protocol. The protocol consists of two assay components, the initiation
16 assay to detect tumor-initiating activity and the promotion assay to detect
17 tumor-promoting activity. The cell growth assays were carried out prior to the
18 transformation assays to determine the doses applicable to the cell transformation tests.
19 Cell growth assays were also performed concurrently with every transformation assay
20 (every initiation and promotion assay) to estimate the effect of each treatment on the
21 cell growth and survival.
22
23
24
25



37
38
39 Fig. 2. Schematic of the Bhas 42 cell transformation assay protocol.
40
41
42
43

44 3.2.1 Preparation

45 46 3.2.1.1 Cells

47
48 The Bhas 42 cells from the same cell pool at passage 17 were distributed by HRI to the
49 participating laboratories. We confirmed that Bhas 42 cells at passage 18 retained the
50 transfected v-Ha-*ras* gene and expressed the introduced mRNA at a level similar to that
51 of the c-Ha-*ras* gene.

1
2 **3.2.1.2 Materials**

3
4 Eagle's minimum essential medium supplemented with 100 units/mL of penicillin, 100
5 µg/mL of streptomycin and 10% fetal bovine serum (FBS) (M10F) was used for cell
6 expansion (for mother cultures). Dulbecco's modified Eagle's medium/Ham's F12
7 supplemented with 100 units/mL of penicillin, 100 µg/mL of streptomycin and 5% FBS
8 (DF5F) was used for the transformation assays and cell growth assays.

9
10 The batch of FBS used throughout this validation study by all participating laboratories
11 was selected by HRI, having been prescreened to produce a minimal number of
12 transformed foci in the Bhas 42 cells treated with the solvent control, DMSO, and a
13 substantially elevated number of transformed foci in the cells treated with the positive
14 control, MCA (1 µg/mL) or TPA (0.05 µg/mL).

15
16 The all materials and reagents used are shown in the test protocol presented in Annex 2,
17 and their suppliers, catalog numbers and lot numbers are also listed.

18
19 **3.2.1.3 Cell stocks**

20
21 The distributed Bhas 42 cells were thawed, expanded and cryopreserved in aliquots so
22 as to generate a large batch of that cell pool in the respective laboratories for use in the
23 transformation assays. The cells were cultivated in M10F in a humidified 5% CO₂
24 incubator at 37°C and when the cultures reached about 70% confluence they were
25 subcultured using 0.25% trypsin in order to maintain a sub-confluent state. Within two
26 passages after thawing, the cultured cells were suspended at 5 x 10⁵ cells/mL in fresh
27 M10F containing 5% DMSO, frozen in 0.5 mL volumes at -80 °C and stored in liquid
28 nitrogen. The transformation assays were always started from this frozen cell stock.
29 The cell growth assay were not necessary performed using cells from this frozen stock,
30 however, properly passaged cells were always used in such the cell growth assays,
31 which were conducted for the determination of test chemical concentrations.

32
33 **3.2.1.4 Chemicals**

34
35 Test chemicals were dissolved or suspended in DMSO or sterile distilled water. The
36 vehicle of choice was decided by each laboratory depending on its respective solubility
37 test. The vehicles used and their final concentrations in the medium are presented in
38 the tables of assay results (Tables 4-15, below).

39
40 MCA and TPA were dissolved in DMSO and served as positive controls for the initiation
41 assay and the promotion assay, respectively. The same lots of each were purchased by
42 each of the participating laboratories.

43
44
45 **3.2.2 Initiation assay**

46
47 **3.2.2.1 Cell growth assay**

48
49 A cell suspension at 2,000 cells/mL in DF5F was distributed into each well of 6-well
50 micro-plates in a 2 mL volume (4,000 cells/well, day 0). Three wells were prepared for
51 each treatment group. At 24 h after seeding, the culture medium was replaced with
52 medium containing a test chemical at various concentrations or vehicle alone, or a test

1 chemical dissolved in DMSO or sterile distilled water was added directly to the culture
2 medium in the well without medium exchange. On day 4, the medium containing the
3 test chemical was replaced with the fresh DF5F. On day 7, the cells were fixed with 10%
4 formalin or methanol and stained with a 0.1% crystal violet (CV) solution in 5% ethanol.
5 CV was extracted from the stained cells with a solution containing 0.02 mol/L
6 hydrochloric acid in 50% ethanol. The optical density of the extract was measured at a
7 wavelength between 540 and 570 nm. The relative cell growth of cultures treated with a
8 chemical was calculated as follows:

9
10
$$\text{relative cell growth} = [(At - Ab)/(Ac - Ab)] \times 100$$

11
12 where At was the absorbance of CV extract from a well with the chemically-treated cells,
13 Ac was the absorbance of CV extract from a well with the solvent-treated cells and Ab
14 was the absorbance of CV extract from a well with medium only.

15 16 **3.2.2.2 Dose-setting for the transformation assay**

17
18 Five or more concentrations were selected based on the results of cell growth assays, so
19 that the concentrations used covered a range from little or no toxicity to a toxicity level
20 that resulted in less than 20% survival compared to the control cultures. In practice, the
21 concentrations generally employed included one dose below the no-effect level (NOEL),
22 two doses between the NOEL and the 50% inhibitory concentration (IC₅₀) and two doses
23 between the IC₅₀ and the 90% inhibitory concentration (IC₉₀).
24

25 The highest concentration of test chemical in the medium was 5 mg/mL in this study.

26 27 **3.2.2.3 Transformation assay to examine initiating activity**

28
29 One tube of the frozen Bhas 42 cells was rapidly thawed and grown in M10F up to about
30 70% confluence and then subcultured in DF5F to reach about 70% confluence again.
31 Thereafter, cells were cultured in DF5F. The cells were trypsinized and suspended at a
32 density of 2,000 cells/mL and seeded into each well of 6-well micro-plates in 2 mL
33 volumes (4,000 cells/well, day 0). Nine wells were prepared per concentration, of which
34 six wells were reserved for the transformation assay and three were reserved for the
35 concurrent cell growth assay. At 24 h after seeding, the cells were treated by the
36 addition to the cultures of a test chemical solution or the vehicle alone, or by complete
37 replacement of the medium containing either the test chemical or the solvent vehicle.
38 The treatment in the initiation phase was continued for 72 h. Following the exposure
39 period, all treatment media were removed and the cells were refed with medium
40 without the test chemical (day 4) and subsequently cultured in the normal medium
41 until day 21, receiving medium exchanges on day 7, day 11 and day 14. The cells were
42 then fixed with methanol and stained with Giemsa's solution. Each assay also included
43 MCA (1 µg/mL) as the positive control.
44

45 46 **3.2.3 Promotion assay**

47 48 **3.2.3.1 Cell growth assay**

49
50 Two milliliter of a cell suspension at 7,000 cells/mL was seeded into each well (14,000
51 cells/well, day 0), and on day 4, the culture medium was replaced with a medium
52 containing a test chemical. On day 7, the cells were fixed and stained with CV, and the

1 relative cell growth of cultures treated with a chemical was calculated according to the
2 procedures as described in 3.2.2.1.

3 4 **3.2.3.2 Dose-setting for the transformation assay**

5
6 Five or more concentrations were selected based on the results of cell growth assays.

7
8 For the chemicals that exhibited marked growth enhancement, the test concentrations
9 were selected to cover a range from little effect on cell growth to growth enhancement.
10 In practice, the concentrations generally employed included one dose below the NOEL,
11 three doses resulting in growth enhancement, and one more higher dose that produces
12 weak growth inhibition. For the chemicals that did not induce marked growth
13 enhancement, the test concentrations were selected to range from a dose two or three
14 levels lower than the no-effect concentration to that showing a survival less than 50%.
15 In practice, the concentrations generally employed included two doses below NOEL, two
16 doses between NOEL and IC₅₀, and one dose above IC₅₀.

17
18 For a chemical that caused a sharp decline of cell growth within a narrow concentration
19 range, one or two additional doses outside the established concentration range were set
20 up as a precaution against the fluctuation of cell response among experiments.

21
22 The highest concentration of test chemical in the medium was 5 mg/mL in this study.

23 24 **3.2.3.3 Transformation assay to examine promoting activity**

25
26 The promotion assay was carried out in the same manner as the initiation assay
27 (3.2.2.3) except for the following steps. The cells were seeded at a density of 7,000
28 cells/mL (14,000 cells/well, day 0) and cultured for 4 days without a medium exchange.
29 On day 4, day 7, and day 11, the culture medium was replaced with fresh medium
30 containing a test chemical or vehicle alone and the treatment in the promotion phase
31 was continued until day 14 (for a total of 10 days). The cells were then cultured in the
32 normal medium without the test chemical for one week until day 21. Each assay also
33 included TPA (0.05 µg/mL) as the positive control.

34 35 36 **3.2.4 Focus count and statistical analysis**

37
38 The transformed foci were judged on the basis of the morphological characteristics: (a)
39 more than 100 cells, (b) spindle-shaped cells whose morphology was distinctly different
40 from the contact-inhibited monolayer cells, (c) deep basophilic staining, (d) random
41 orientation of cells, especially at the edge of foci, (e) dense multilayering of cells, and (f)
42 invasive growth into the monolayer of surrounding contact-inhibited cells. The number
43 of transformed foci in every well was recorded and a statistical analysis for the increase
44 in the number of the transformed foci produced by a test chemical was performed by
45 multiple comparison using the one-sided Dunnett test (p<0.05). The statistical
46 significance of positive controls was evaluated by the t-test or Aspin-Welch test
47 depending on the result of the F-test for homoscedasticity (homogeneity of variance).
48 "Toxic" was recorded for wells in which the cells were not confluent at the end of the
49 transformation assay because of cytotoxicity resulting from chemical treatment.

50
51 A Photo Catalog used for judging the morphology of foci in the Bhas 42 CTA is attached
52 (Annex 3).

1
2
3 **3.2.5 Judgment**
4

5 The results in the Bhas 42 CTAs were judged positive when there existed two or more
6 sequential doses that induced statistically significant increases in the number of
7 transformed foci, and negative when there was no dose showing a statistically
8 significant increase of foci. When the statistically significant increase was at only one
9 dose, the assay result was regarded as equivocal, and then the initiation or promotion
10 assay together with the concomitant cell growth assay was repeated including the
11 positive dose in the first assay. The chemical was judged to be positive if a statistically
12 significant increase in the number of transformed foci resulted at one or more
13 concentrations in the second assay.
14

15
16 **3.2.6 Assay acceptance criteria**
17

18 In the 6-well method validation study, assay acceptance criteria were not described in
19 the protocol. Instead of acceptance criteria, the following determining factors, which are
20 described in protocol section D. Judgment, were observed: “When low transformation
21 frequency in the positive control or high transformation frequency in the negative
22 control is obtained, the experiment should be repeated using optimized test conditions.
23 When biologically suspicious results are obtained, a re-experiment may be requested
24 after discussion.”
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29 **3.3 Results**
30

31 This international validation study of the Bhas 42 CTA using 6-well micro-plates was
32 carried out on twelve coded chemicals in total for the initiation and promotion assays.
33 Each chemical was examined by three laboratories. The data for the negative and
34 positive controls obtained in individual laboratories are collected and illustrated in
35 Figures 3 and 4. The results of each individual test chemical in the Bhas 42 CTAs,
36 including the respective concurrent cell growth assays are presented in Tables 4 - 15
37 and in Figures 5 - 16. The raw data submitted by individual laboratories are presented
38 in Annex 4.
39

40
41 **3.3.1 Quality assurance aspects**
42

43 All the data were sent to Noriho Tanaka and Ayako Sakai. Since at that stage there was
44 no acceptance criteria in the protocol, all the data submitted from the participating
45 laboratories were accepted, and are presented and discussed in this validation study
46 report. In certain instances, specific experiments were repeated by a particular
47 laboratory. The data from both the first and repeated experiments are presented for
48 each chemical, including the reason why the experiment was repeated. The statistical
49 analysis was carried out by A. Sakai.
50
51

3.3.2 Negative and positive controls

The numbers of transformed foci produced in the negative (DMSO) and positive controls are presented in Tables 4-15 as part of the assay results derived for each test chemical. In Fig. 3, those control values are plotted pairwise, assay by assay, and the pairs are clustered by a given laboratory. The number of transformed foci was statistically analyzed by t-test or Aspin-Welch test and there were significant differences between positive controls and the corresponding negative controls in all the initiation and promotion assays ($p < 0.05$), although the absolute number of foci induced by the positive control varied between laboratories and sometimes within laboratories. In Fig 4, the numbers of transformed foci in the negative and positive controls are averaged laboratory by laboratory.

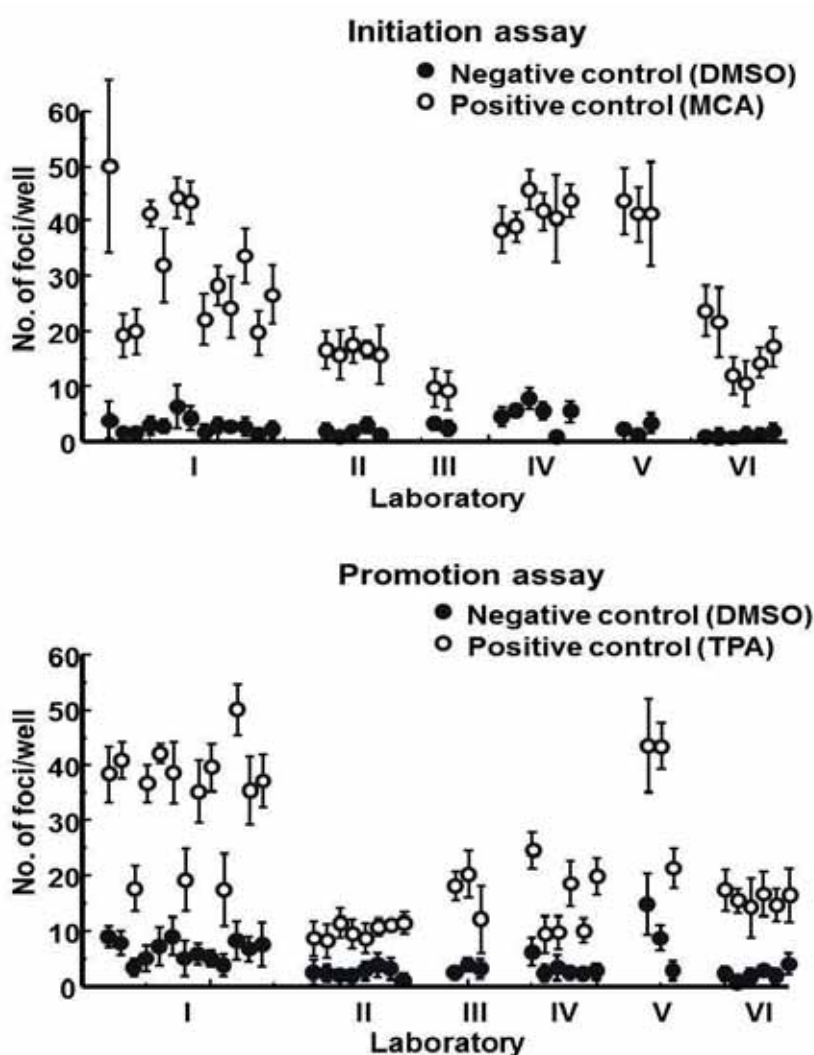
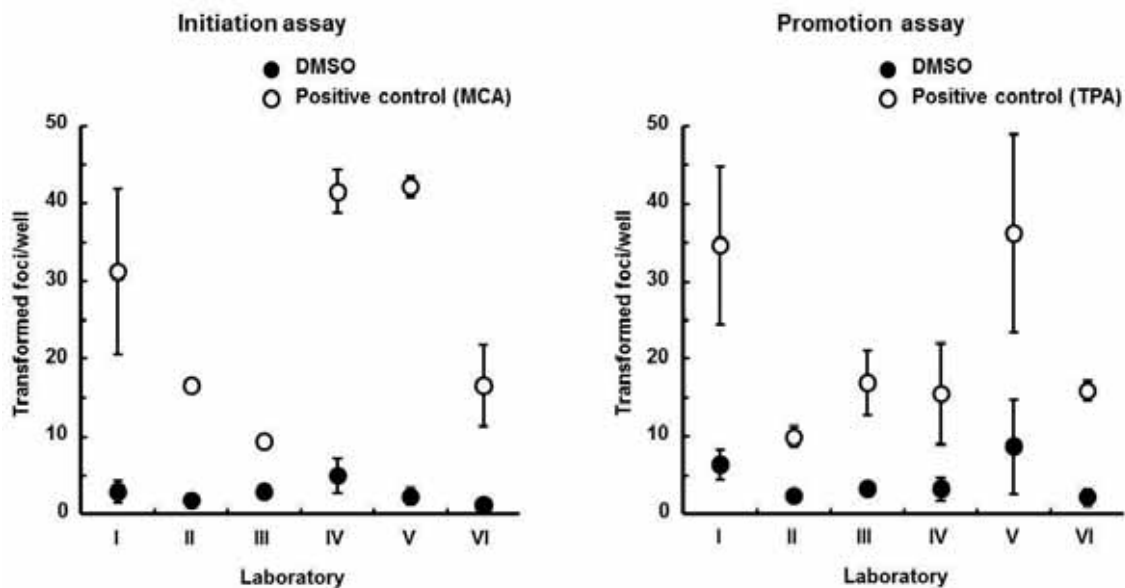


Fig. 3. Number of transformed foci per well in the negative and positive controls of the Bhas 42 cell transformation assay in the 6-well method validation study. Negative and positive controls are paired assay by assay and the pairs are clustered by a given laboratory. Bars indicate standard deviation.

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Fig. 4 Average number of the transformed foci per well produced in negative and positive controls of the Bhas 42 cell transformation assay in individual laboratories in the 6-well method validation study. Bars indicate standard deviation.

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3.3.3 2-Acetylaminofluorene

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The results for 2-acetylaminofluorene are shown in Table 4 and Fig. 5. All three laboratories judged 2-acetylaminofluorene as positive both in the initiation assay and in the promotion assay since it induced statistically significant increases in the number of transformed foci at two or more sequential concentrations in both assays in all laboratories.

1 Table 4.
 2 Results of transformation assay on 2-acetylaminofluorene in the 6-well method
 3 validation study

4 a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab IV		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.1% DMSO)	100	3.8 \pm 3.4	100	4.5 \pm 1.6		
0 (0.5% DMSO)					100	1.0 \pm 0.6
10	118	4.5 \pm 2.3	101	5.5 \pm 2.3		
12					103	2.7 \pm 2.1
14					86	5.7 \pm 2.5 *
15	130	7.0 \pm 1.7	104	5.8 \pm 3.1		
16					87	9.2 \pm 3.5 *
18					60	6.3 \pm 4.1 *
20	135	10.0 \pm 3.7 *	94	8.7 \pm 2.6 *	45	4.2 \pm 2.3
25	91	14.5 \pm 1.9 *	41	12.5 \pm 1.9 *	20	1.8 \pm 1.5
30	46	14.3 \pm 1.6 *	26	12.3 \pm 2.2 *	20	Toxic
35			18	14.2 \pm 2.9 *		
40			17	13.5 \pm 3.7 *	9	Toxic
50	22	8.8 \pm 5.7 *				
100	19	5.2 \pm 2.4				
300	14	7.8 \pm 3.5				
MCA 1 ^d (0.1% DMSO)	82	50.0 \pm 15.6 †	72	38.5 \pm 4.1 †		
MCA 1 (0.5% DMSO)					53	23.7 \pm 4.6 †

25 ^a % of cell growth compared to that of the solvent control.

26 ^b Average number of transformed foci/well \pm SD.

27 ^c Solvent control: final solvent concentration in the working culture media in parentheses

28 ^d Positive control in the initiation assay: final solvent concentration in the working culture media in
 29 parentheses.

30 * $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

31 † $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

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b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab IV		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.1% DMSO)	100	9.0 \pm 2.0	100	6.2 \pm 2.6		
0 (0.5% DMSO)					100	2.2 \pm 1.3
0.03	103	5.7 \pm 1.0				
0.1	103	4.2 \pm 1.9				
0.3	104	6.3 \pm 2.3				
1	101	9.2 \pm 3.1				
2.5					98	6.0 \pm 3.6
3	102	6.8 \pm 2.6				
5			100	9.5 \pm 1.9 *	79	10.2 \pm 2.9 *
10	85	13.0 \pm 3.6 *	98	10.8 \pm 3.2 *	88	15.5 \pm 2.1 *
15	83	19.5 \pm 3.9 *	89	12.0 \pm 2.1 *	58	11.7 \pm 4.5
20	71	19.3 \pm 3.4 *	85	9.7 \pm 1.6 *	61	5.3 \pm 3.8
25			78	10.0 \pm 2.5 *	49	Toxic
30			72	12.7 \pm 1.6 *		
35			78	18.2 \pm 1.7 *		
40			76	1.3 \pm 1.0		
50					47	Toxic
TPA 0.05 ^d (0.1% DMSO)	125	38.5 \pm 5.0 †	219	24.7 \pm 3.3 †		
TPA 0.05 (0.5% DMSO)					104	17.5 \pm 3.7 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

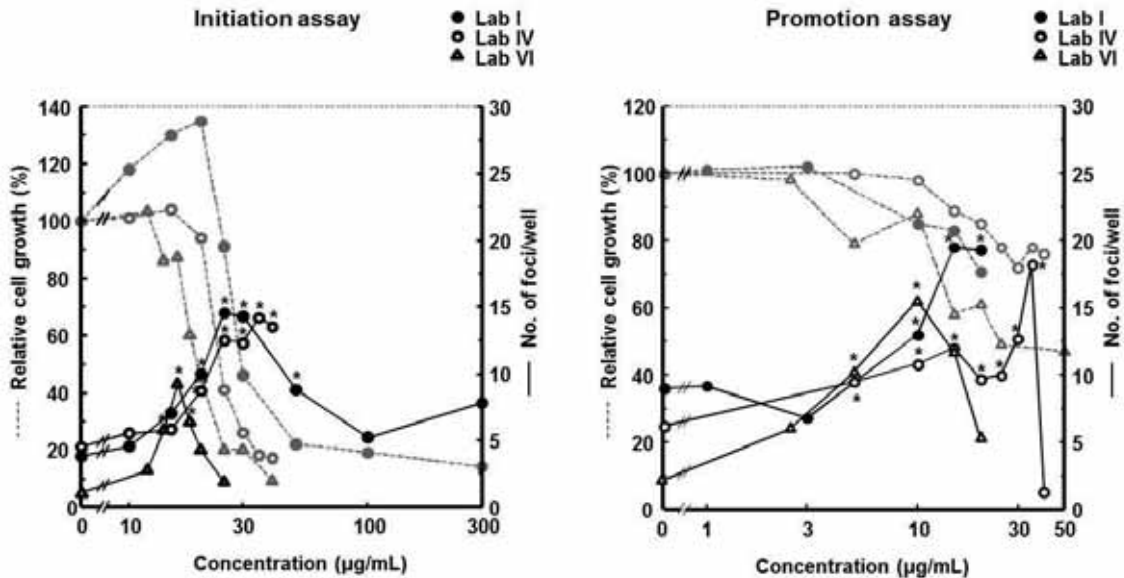


Fig. 5. Graphic view of the results of transformation assay and concurrent cell growth assay on 2-acetylaminofluorene in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

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2 **3.3.4 Cadmium chloride**
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4 The results for cadmium chloride are presented in Table 5 and Fig. 6. The chemical gave
5 negative results in the initiation assays as determined by all three laboratories.
6

7 The promotion assay was repeated by all three laboratories. The reason for the
8 repetition was the cell death caused by cytotoxicity at high concentrations of the test
9 chemical. All three laboratories performed the second assay in a lower concentration
10 range. In the first run of the promotion assay, Lab I obtained statistically significant
11 increases in the number of transformed foci at the lowest two concentrations, 0.4 and
12 0.6 µg/mL, but the chemical was toxic at the higher five doses. In the second run of the
13 assay, the positive results were confirmed at concentrations ≤0.6 µg/mL. In Lab II, the
14 seven highest of eight doses tested were completely cytotoxic in the first run of the assay.
15 However, in the second run of the assay, which employed lower doses, the number of
16 transformed foci at one dose (the highest dose tested, which was also the lowest dose
17 tested in the first run) was statistically significantly increased. Lab II again repeated
18 the assay for a third time and obtained a significant increase at a single dose again.
19 Thus, based upon the response criteria, cadmium chloride was judged to be positive in
20 the Lab II. Since Lab III set the test concentrations over a wide range, cell killing was
21 caused at only two doses in the first promotion assay. Meanwhile, Lab III obtained a
22 statistically significant increase in transformed foci at a single dose in that first assay.
23 In the second assay that was carried out using a lower and narrower concentration
24 range, there were the significant increases of foci at 3 consecutive doses. Therefore,
25 cadmium chloride was judged positive by Lab III.
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2 Table 5.
3 Results of transformation assay on cadmium chloride in the 6-well method validation
4 study

5 a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab III	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (5% Water)	100	4.8 \pm 2.5	100	1.5 \pm 1.4	100	3.0 \pm 1.7
0.1			111	1.7 \pm 1.4		
0.2			104	0.8 \pm 1.2		
0.4			96	2.0 \pm 1.4		
0.402					110	1.7 \pm 1.0
0.482					105	1.3 \pm 1.4
0.579					106	1.7 \pm 1.6
0.6			93	1.0 \pm 1.1		
0.694					95	1.3 \pm 1.5
0.8	88	2.7 \pm 0.8	72	0.7 \pm 1.2		
0.833					72	1.3 \pm 1.2
1	82	3.3 \pm 0.5	41	0.7 \pm 0.8	55	1.3 \pm 1.0
1.2	80	2.3 \pm 1.8	13	0.5 \pm 1.2	7	0.3 \pm 0.5
1.4	44	1.3 \pm 1.2				
1.44					5	1.0 \pm 0.6
1.5			4	0.2 \pm 0.4		
1.6	21	0.7 \pm 0.5				
1.73					-1	0.3 \pm 0.8
1.8	6	0.8 \pm 1.2				
2	3	0.3 \pm 0.5				
0 ^d (0.1% DMSO)	100	4.3 \pm 2.1		1.0 \pm 0.9		
MCA 1 ^e (0.1% DMSO)	74	43.5 \pm 3.8 †		15.8 \pm 4.4 †	14 ^f	9.2 \pm 2.0 ‡

30 ^a % of cell growth compared to that of the solvent control.

31 ^b Average number of transformed foci/well \pm SD.

32 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

33 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
34 media in parentheses.

35 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
36 parentheses.

37 ^f % of cell growth compared to that of 5% water.

38 † $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

39 ‡ $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. 5% water.

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b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I, 1st run		Lab I, 2nd run		Lab II, 1st run		Lab II, 2nd run		Lab II, 3rd run		Lab III, 1st run		Lab III, 2nd run	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well	CG	Foci/well	CG	Foci/well	CG	Foci/well	CG	Foci/well
0 ^c (5% Water)	100	5.3 \pm 2.3	100	2.5 \pm 0.8	100	2.6 \pm 1.1	100	1.3 \pm 0.8	100	1.3 \pm 1.5	100	2.0 \pm 0.6	100	1.2 \pm 0.8
0.001							92	1.0 \pm 0.6						
0.01							100	0.3 \pm 0.5						
0.05							97	1.7 \pm 1.4						
0.0625											97	2.2 \pm 0.8	98	1.2 \pm 1.6
0.1			101	2.0 \pm 0.9	102	0.8 \pm 1.3					99	2.3 \pm 0.8	102	1.3 \pm 0.5
0.125														
0.2			104	4.2 \pm 1.0	116	2.8 \pm 1.9					108	4.0 \pm 1.8 *	108	3.7 \pm 1.6
0.25														
0.275														
0.3														
0.35														
0.4														
0.45	114	21.8 \pm 4.4 *												
0.5														
0.6														
0.8														
1														
1.2														
1.4														
1.6														
2														
0 ^d (0.1% DMSO)	100	9.0 \pm 3.4	100	5.0 \pm 3.3	100	2.3 \pm 1.6	100	2.0 \pm 0.9	100	1.8 \pm 0.4	100	15.7 \pm 4.5 †	100	3.8 \pm 1.3
TPA 0.05 ^e (0.1% DMSO)	125	38.7 \pm 5.6 †	132	19.3 \pm 5.6 †	133	8.3 \pm 3.1 †	134	11.5 \pm 2.7 †	134	9.5 \pm 2.7 †	155 ^f	15.7 \pm 4.5 †	166	20.3 \pm 4.2 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

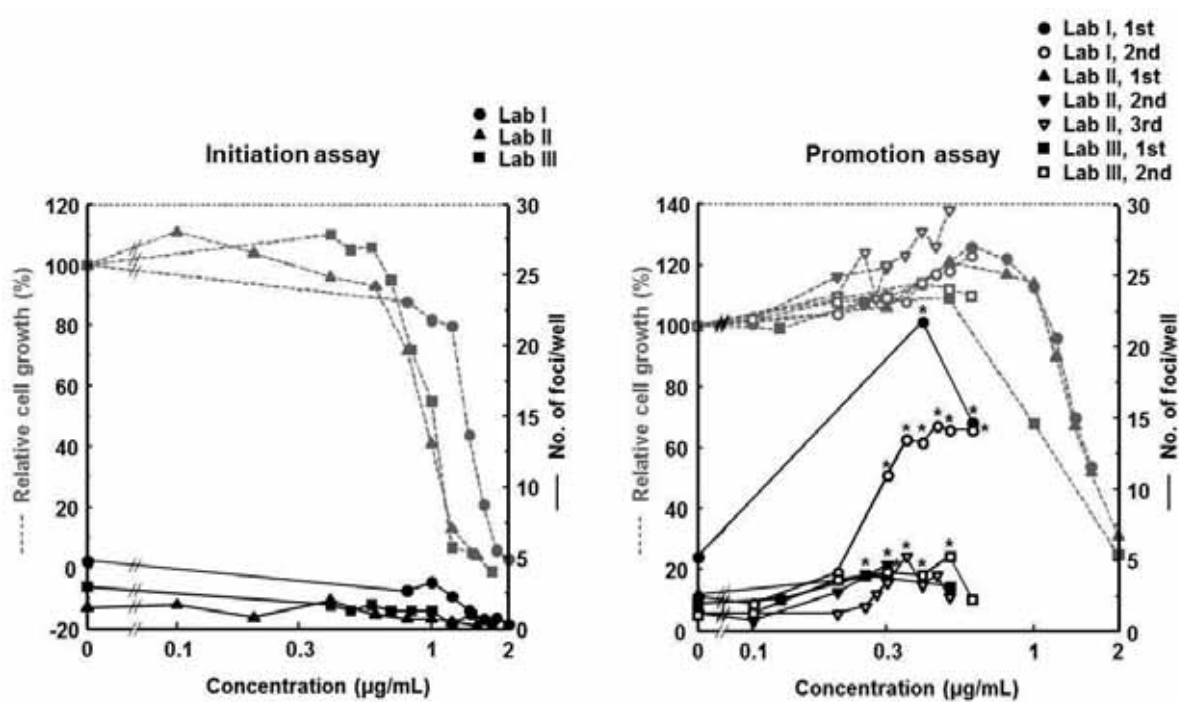
^f % of cell growth compared to that of 5% water.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

‡ $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. 5% water.

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Fig. 6. Graphic view of the results of transformation assay and concurrent cell growth assay on cadmium chloride in the 6-well method validation study.
* p<0.05; one-sided Dunnett test.

3.3.5 Dibenz[*a,h*]anthracene

The results for dibenz[*a,h*]anthracene are presented in Table 6 and Fig. 7. All three laboratories reported that dibenz[*a,h*]anthracene was clearly positive in the initiation assay, but negative in the promotion assay.

Table 6.

Results of transformation assay on dibenz[*a,h*]anthracene in the 6-well method validation study

a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab V	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.1% DMSO)	100	1.5 \pm 1.2			100	2.8 \pm 1.9
0 (0.2% DMSO)			100	0.8 \pm 0.8		
0.0033					89	8.0 \pm 1.4
0.01	113	6.0 \pm 2.5	99	2.5 \pm 1.5	73	13.3 \pm 3.1 *
0.03	110	8.7 \pm 2.9 *	80	6.2 \pm 2.4 *		
0.033					37	22.5 \pm 3.0 *
0.04			71	7.3 \pm 3.3 *		
0.05			65	8.3 \pm 2.9 *		
0.1	84	15.0 \pm 4.2 *	53	11.2 \pm 3.9 *	11	18.5 \pm 4.1 *
0.3	63	24.2 \pm 3.9 *				
0.33					3	22.7 \pm 7.1 *
1	58	24.2 \pm 3.3 *	30	12.7 \pm 2.9 *	4	16.3 \pm 4.2 *
3	54	17.8 \pm 4.5 *				
3.3					7	17.5 \pm 4.8 *
7	57	17.2 \pm 3.5 *				
10	58	17.5 \pm 3.5 *	38	9.3 \pm 2.4 *	1	17.3 \pm 7.8 *
0 ^d (0.1% DMSO)			100	2.0 \pm 1.3	100	1.2 \pm 0.4
MCA 1 ^e (0.1% DMSO)	79	20.0 \pm 4.1 †	53	16.7 \pm 3.4 †	23	41.3 \pm 4.9 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab V	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.1% DMSO)	100	3.3 \pm 1.6			100	8.7 \pm 2.3
0 (0.2% DMSO)			100	2.8 \pm 1.5		
0.001			81	1.5 \pm 0.5	83	9.0 \pm 2.1
0.0033					94	11.8 \pm 3.3
0.005			82	2.5 \pm 1.5		
0.01	83	2.2 \pm 2.1			70	11.2 \pm 3.1
0.03	80	2.3 \pm 2.0				
0.033					70	11.5 \pm 4.4
0.05			76	2.7 \pm 1.9		
0.1	74	2.7 \pm 1.2			67	10.5 \pm 3.0
0.2			69	1.5 \pm 1.0		
0.3	65	0.8 \pm 1.6				
0.33					59	5.0 \pm 4.1
1	66	1.0 \pm 1.3	71	0.2 \pm 0.4	71	2.3 \pm 2.4
3	66	1.7 \pm 0.8				
3.3					68	3.2 \pm 1.2
7	71	0.7 \pm 1.0				
10	66	1.0 \pm 0.9	74	1.0 \pm 1.3		
0 ^d (0.1% DMSO)			100	2.5 \pm 2.1		
TPA 0.05 ^e (0.1% DMSO)	118	17.7 \pm 4.2 †	128	8.8 \pm 3.2 †	136	43.5 \pm 4.1 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

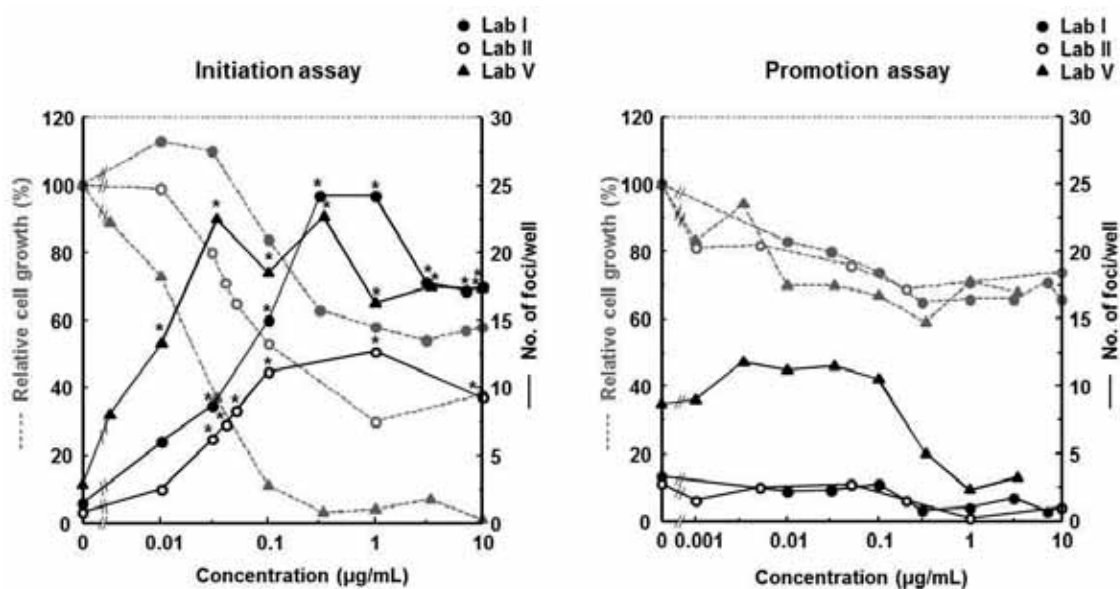


Fig. 7. Graphic view of the results of transformation assay and concurrent cell growth assay on dibenz[*a,h*]anthracene in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

3.3.6 Lithocholic acid

The results obtained with lithocholic acid are shown in Table 7 and Fig. 8. The data from all laboratories revealed that lithocholic acid was negative in the initiation assay and positive in the promotion assay.

Table 7.
Results of transformation assay on lithocholic acid in the 6-well method validation study

a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab IV		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (DMSO 0.1%)	100	6.3 \pm 3.9	100	5.7 \pm 1.0		
0 (DMSO 0.5%)					100	1.3 \pm 1.4
7			92	5.7 \pm 2.3		
10	105	5.5 \pm 1.9	94	4.5 \pm 3.3		
14			96	5.7 \pm 2.4		
15	106	4.5 \pm 2.3				
17.5	108	2.0 \pm 1.4				
20	104	3.7 \pm 2.3	85	3.7 \pm 1.9		
22					75	1.8 \pm 1.5
22.5	107	2.7 \pm 0.5				
25	89	4.3 \pm 2.5				
27.5	56	5.5 \pm 3.7				
28			112	3.2 \pm 1.2	10	0.2 \pm 0.4
28.25					9	0.5 \pm 0.5
28.5					7	0.8 \pm 0.8
28.75					8	0.5 \pm 0.8
29					11	0.0 \pm 0.0
29.25					9	Toxic
29.5					8	Toxic
29.75					8	Toxic
30	11	3.0 \pm 1.4			9	Toxic
40			3	4.2 \pm 0.8		
56			0	Toxic		
80			1	Toxic		
MCA 1 ^d (0.1% DMSO)	73	44.2 \pm 3.7 †	57	39.0 \pm 2.6 †		
MCA 1 (0.5% DMSO)					56	10.5 \pm 4.2 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab IV		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (DMSO 0.1%)	100	7.2 \pm 3.5	100	2.3 \pm 1.6		
0 (DMSO 0.5%)					100	2.7 \pm 0.8
2.5	93	9.3 \pm 2.6				
5	89	15.7 \pm 5.1 *				
10	80	27.3 \pm 4.2 *			83	19.7 \pm 5.2 *
12			86	6.3 \pm 4.1 *		
15	72	44.8 \pm 9.8 *				
18			86	13.5 \pm 2.1 *		
20	75	45.7 \pm 5.8 *			83	33.2 \pm 4.9 *
25	86	12.2 \pm 2.4	89	14.0 \pm 1.8 *		
30	16	Toxic			106	Toxic
35			84	Toxic	50	Toxic
40					38	Toxic
42.5					19	Toxic
45					14	Toxic
50			5	Toxic	11	Toxic
TPA 0.05 ^d (0.1% DMSO)	124	42.3 \pm 1.8 †	136	9.5 \pm 3.4 †		
TPA 0.05 (0.5% DMSO)					145	16.8 \pm 4.1 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

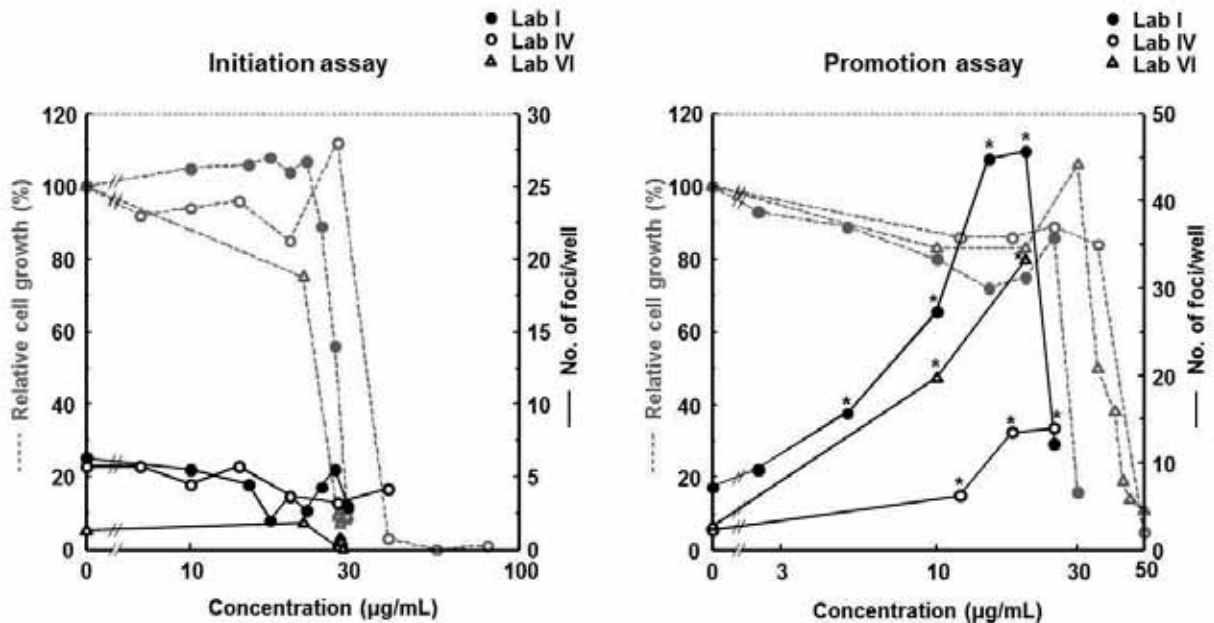


Fig. 8. Graphic view of the results of transformation assay and concurrent cell growth assay on lithocholic acid in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

3.3.7 Methapyrilene hydrochloride

The results for methapyrilene hydrochloride are shown in Table 8 and Fig. 9. All three laboratories demonstrated methapyrilene hydrochloride to be negative in the initiation assay and positive in the promotion assay.

Table 8.

Results of transformation assay on methapyrilene hydrochloride in the 6-well method validation study

a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (5% water)	100	2.7 \pm 1.6	100	2.0 \pm 1.4	100	6.8 \pm 1.8
53					120	6.2 \pm 1.2
75					149	7.2 \pm 3.1
100	109	3.3 \pm 1.5				
110					124	6.2 \pm 1.5
150	86	2.8 \pm 1.0			129	6.5 \pm 2.6
200	71	3.2 \pm 2.1	45	1.8 \pm 0.8		
210					132	5.8 \pm 1.5
220			23	1.0 \pm 0.6		
240			13	1.7 \pm 1.2		
250	56	3.0 \pm 1.9				
260			5	0.8 \pm 0.8		
280			3	1.3 \pm 0.8		
300	32	2.0 \pm 1.4	1	0.5 \pm 0.8	64	4.8 \pm 1.3
350	16	1.0 \pm 0.9				
400	6	1.7 \pm 1.0				
450	4	1.3 \pm 1.5				
0 ^d (0.1% DMSO)	100	3.0 \pm 1.4	100	1.8 \pm 1.2	100	5.5 \pm 1.5
MCA 1 ^e (0.1% DMSO)	45	28.3 \pm 3.6 †	54	17.5 \pm 3.1 †	70	41.8 \pm 3.4 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (5% Water)	100	5.7 \pm 2.3	100	2.3 \pm 1.8	100	2.7 \pm 1.2
5			104	3.0 \pm 1.5	111	6.7 \pm 1.5
10	121	10.5 \pm 2.3	116	4.5 \pm 2.6	116	9.2 \pm 5.2 *
20					126	16.0 \pm 1.4 *
25	130	16.2 \pm 2.6 *				
40					140	29.7 \pm 4.8 *
50	151	35.7 \pm 5.6 *	159	18.0 \pm 4.8 *		
75	168	46.0 \pm 8.0 *				
80					199	52.3 \pm 7.3 *
100	184	33.8 \pm 4.5 *	190	24.7 \pm 6.2 *		
160					217	34.5 \pm 2.3 *
200	164	5.3 \pm 1.9	161	1.0 \pm 1.3		
300	139	0.7 \pm 0.8				
400	114	Toxic				
500			72	Toxic		
0 ^d (0.1% DMSO)	100	5.0 \pm 1.5		2.7 \pm 1.6		2.5 \pm 0.8
TPA 0.05 ^e (0.1% DMSO)	126	39.7 \pm 4.3 †		8.7 \pm 2.8 †		18.7 \pm 4.2 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

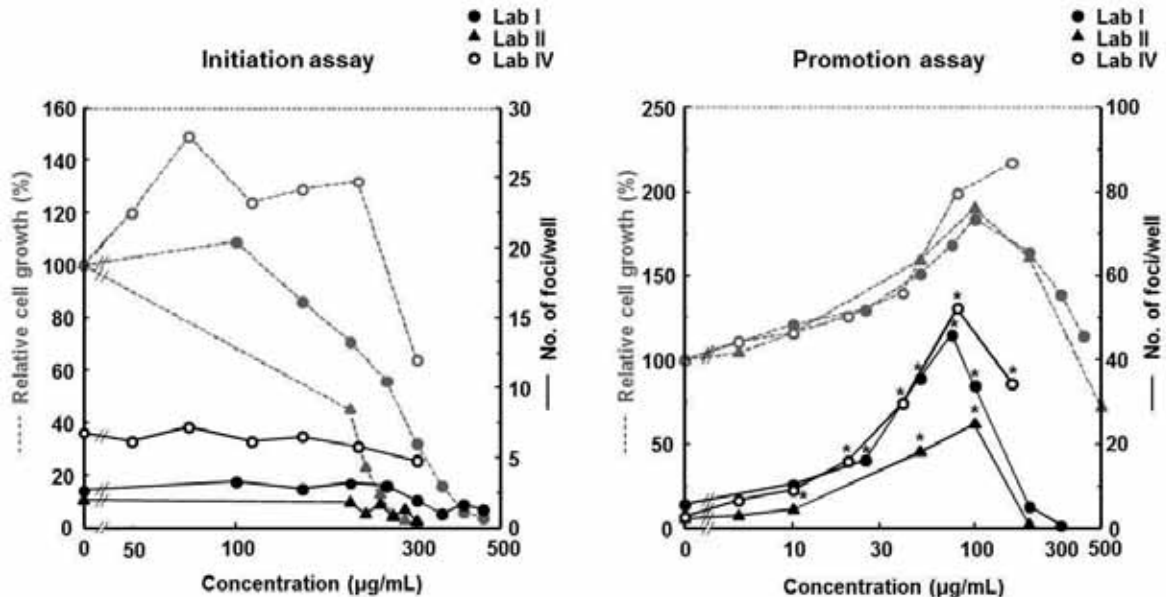


Fig. 9. Graphic view of the results of transformation assay and concurrent cell growth assay on methapyrilene hydrochloride in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

3.3.8 Mezerein

Table 9 and Fig. 10 show the results for mezerein. All three laboratories reported clearly positive results in the promotion assay. The chemical induced marked and dose-dependent increases in the number of transformed foci relative to the solvent control in all cases.

In the initiation assay performed by Lab I, mezerein did not induce statistically significant increases in the number of transformed foci at any concentration up to 0.01 µg/mL, which was the maximum concentration used in the present study. In the initiation assays performed by Lab III and Lab IV, a statistically significant increase in the number of transformed foci was observed only at the maximum (preset) concentration used. The results of the initiation assay by both laboratories remained inconclusive since neither laboratory repeated the assay at higher concentrations (due to limited availability) in an attempt to verify the initial results. Rather, they considered that the promotion assay had established that the chemical was clearly positive in the Bhas 42 CTA and that it was not necessary to further differentiate the initiation vs. promotion properties of the compound for the purpose of this exercise.

Table 9.

Results of transformation assay on mezerein in the 6-well method validation study

a) Initiation assay

Concentration (µg/mL)	Lab I		Lab III		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.1% DMSO)	100	1.7 ± 1.5	100	2.5 ± 1.4	100	7.8 ± 2.0
0.000010			104	1.8 ± 1.0		
0.000039			102	2.3 ± 0.8		
0.0001	94	1.7 ± 1.6				
0.000156			110	1.5 ± 1.5		
0.0003	94	1.5 ± 1.0				
0.0005	93	2.2 ± 0.8				
0.000625			113	1.7 ± 1.2		
0.00063					94	7.7 ± 1.5
0.001	94	1.5 ± 1.4				
0.0013					109	7.8 ± 3.2
0.0025			140	3.0 ± 0.6	130	7.3 ± 2.3
0.003	111	1.7 ± 0.8				
0.005	141	1.3 ± 1.2			195	10.2 ± 2.6
0.01	150	3.2 ± 1.9	202	13.3 ± 2.9 *	208	13.2 ± 1.7 *
MCA 1^d (0.1% DMSO)	56	22.2 ± 4.7 †	28	9.2 ± 3.5 †	71	45.7 ± 3.6 †

^a % of cell growth compared to that of that solvent control.

^b Average number of transformed foci/well ± SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* p<0.05; one-sided Dunnett test, vs. corresponding solvent control.

† p<0.05; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab III		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.1% DMSO)	100	5.7 \pm 1.9	100	3.2 \pm 1.8	100	3.3 \pm 2.3
0.000005			100	2.2 \pm 2.1		
0.000020			104	4.5 \pm 1.5		
0.000026					100	5.3 \pm 2.1
0.000064					102	4.3 \pm 2.0
0.000078			107	9.2 \pm 2.6		
0.0001	103	17.5 \pm 5.4 *				
0.00016					113	8.2 \pm 1.6 *
0.0003	121	37.2 \pm 4.2 *				
0.000313			143	31.8 \pm 6.0 *		
0.0004					139	18.3 \pm 2.2 *
0.0005	142	46.5 \pm 4.4 *				
0.001	184	62.7 \pm 2.1 *			208	40.2 \pm 4.1 *
0.00125			257	46.5 \pm 7.4 *		
0.0025					236	75.2 \pm 5.4 *
0.003	181	Toxic				
0.005	183	Toxic	206	37.2 \pm 7.9 *		
0.01	193	Toxic				
TPA 0.05 ^d (0.1% DMSO)	124	35.3 \pm 5.6 †	143	12.2 \pm 6.1 †	142	9.8 \pm 3.1 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

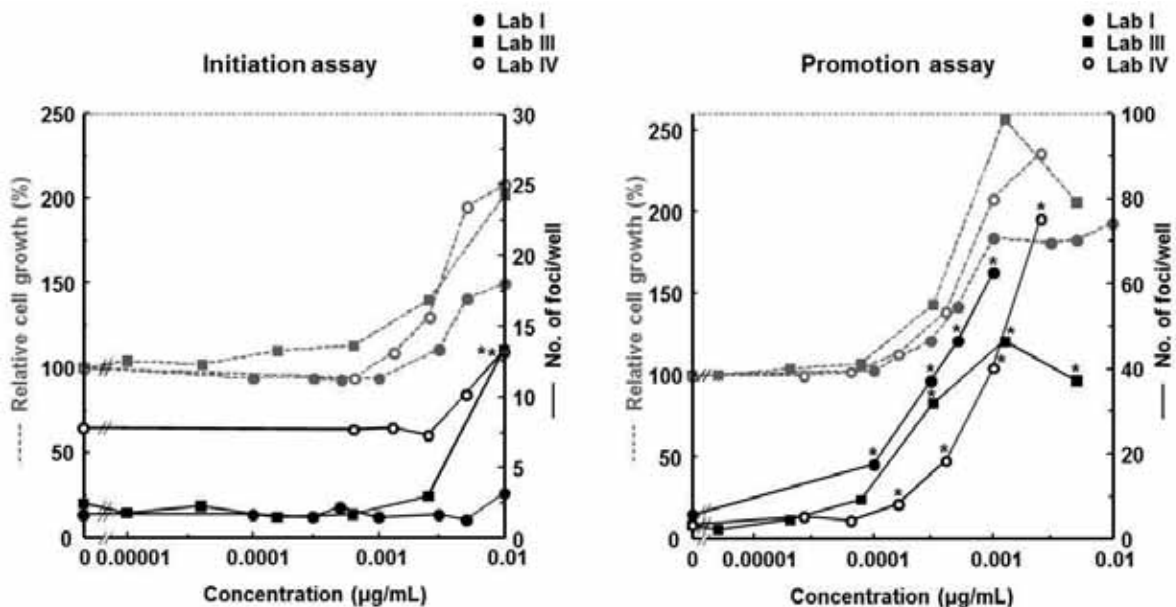


Fig. 10. Graphic view of the results of transformation assay and concurrent cell growth assay on mezezein in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

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2 **3.3.9 Sodium arsenite**
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4 Table 10 and Fig. 11 show the assay results for sodium arsenite. In the first initiation
5 assay of sodium arsenite by Lab I, there was a statistically significant increase in the
6 number of transformed foci at a single dose of 0.8 µg/mL. In accordance with the
7 protocol, Lab I repeated the initiation assay, but there was no significant increase of
8 transformed foci in the second assay. Thus, sodium arsenite was judged as negative in
9 the initiation assay in Lab I. Since Lab III first obtained statistically significant
10 increases of transformed foci at two non-sequential doses, 0.455 and 0.769 µg/mL, they
11 performed a second initiation assay and again obtained a significant increase at 0.769
12 µg/mL. Consequently, sodium arsenite was judged to be positive in the initiation assay
13 by Lab III. Lab VI carried out initiation assay only once and observed no increase of
14 transformed foci. However, sodium arsenite did not produce any cytotoxicity at the
15 tested concentrations in the concurrent cell growth assay. In view of its high solubility
16 in the solvent (water) and its lack of cytotoxicity at the lower concentration initially
17 used, sodium arsenite should have been retested by Lab VI at higher concentrations in
18 order to achieve cytotoxicity. Therefore, the initiation assay conducted on sodium
19 arsenite by Lab VI was judged to be incomplete because of inadequate dosing.
20

21 In the promotion assay on sodium arsenite, all three laboratories reported positive
22 results. Despite having obtained statistically significant increases of transformed foci at
23 two sequential doses in the first assay, however, Lab III repeated the promotion assay.
24 The compound was considered positive and the repeat was not necessary. Lab III's
25 decision was the result of having lost cultures at the two highest concentrations in the
26 transformation assay because of cytotoxicity, although the cell growth rates at
27 respective concentrations were 77% and 26% of control in the concurrent cell growth
28 assay. The second assay was voluntarily carried out in a lower and narrower
29 concentration range and confirmed the positive results obtained in the first assay.
30
31

Table 10.

Results of transformation assay on sodium arsenite in the 6-well method validation study

a) Initiation assay		Lab I, 1st run		Lab I, 2nd run		Lab III, 1st run		Lab III, 2nd run		Lab VI	
Concentration ($\mu\text{g}/\text{mL}$)	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well	CG	Foci/well	CG	Foci/well	CG
0 ^c (0.5% DMSO)	100	2.0 \pm 2.0	100	2.8 \pm 1.3	100	1.5 \pm 0.5	100	1.5 \pm 1.8	100	0.8 \pm 1.0	100
0 (5% Water)											
0.01											99
0.025											99
0.05											96
0.075											108
0.1	113	1.7 \pm 1.2									101
0.11											100
0.12											96
0.123			103	0.7 \pm 0.5							
0.13											104
0.14											100
0.15											98
0.159			96	1.5 \pm 0.8							
0.2	114	2.3 \pm 1.8									
0.207			107	1.3 \pm 0.5	103	2.7 \pm 2.0					
0.269			101	2.2 \pm 1.6	108	1.8 \pm 1.5					
0.3			99	2.3 \pm 1.5							
0.35	120	3.3 \pm 1.6	104	2.3 \pm 1.4	110	2.8 \pm 1.3	102	3.0 \pm 2.5			
0.4											
0.455			103	1.7 \pm 2.3	103	3.3 \pm 1.6 *	84	2.5 \pm 2.2			
0.5											
0.592	97	3.0 \pm 1.3	103	1.0 \pm 1.1	91	1.3 \pm 1.0	61	2.5 \pm 2.3			
0.6			106	1.7 \pm 1.4							
0.7											
0.769			74	3.5 \pm 1.0 *	35	6.3 \pm 2.7 *					
0.8	49	5.0 \pm 3.0 *	96	1.2 \pm 1.5							
0.9			96	1.2 \pm 1.2							
1	16	2.5 \pm 1.9	35	2.2 \pm 1.3	6	4.7 \pm 3.8					
2	3	0.8 \pm 1.0									
0 ^d (0.1% DMSO)	100	3.0 \pm 1.7	100	2.8 \pm 1.3	100	3.3 \pm 1.2					
MCA 1 ^e (0.1% DMSO)	60	41.3 \pm 2.3 †	47	32.0 \pm 6.7 †	22 ^f	15.0 \pm 3.3 ‡	21	9.7 \pm 3.6 †			
MCA 1 (0.5% DMSO)									79	12.0 \pm 3.5 †	

^a % of cell growth compared to that of the solvent control.^b Average number of transformed foci/well \pm SD.^c Solvent control: final solvent concentration in the working culture media in parentheses.^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.^f % of cell growth compared to that of 5% water.

* p<0.05; one-sided Dunnett test, vs. corresponding solvent control.

† p<0.05; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

‡ p<0.05; one-sided t-test or Aspin-Welch test, vs. 5% water.

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b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab III, 1st run		Lab III, 2nd run		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well	CG	Foci/well
0 ^c (0.5% DMSO)								
0 (5% Water)	100	4.5 \pm 1.8	100	2.5 \pm 1.4	100	0.7 \pm 0.8		
0.0158			107	2.5 \pm 2.1				
0.025					104	2.5 \pm 1.4	109	6.5 \pm 1.9 *
0.05	94	6.3 \pm 2.6	106	5.2 \pm 1.2 *	118	4.7 \pm 1.6 *	103	7.3 \pm 1.8 *
0.1	101	13.0 \pm 2.2 *			104	2.8 \pm 1.5 *	105	11.8 \pm 2.9 *
0.15					102	3.7 \pm 1.2 *		
0.158			109	6.2 \pm 2.6 *				
0.2					112	3.5 \pm 2.3 *	101	13.7 \pm 2.7 *
0.25					125	1.7 \pm 1.0		
0.3	99	9.0 \pm 0.9 *			127	0.2 \pm 0.4	96	1.5 \pm 0.8
0.4							96	0.7 \pm 0.8
0.5			97	0.3 \pm 0.8			95	0.0 \pm 0.0
0.7	86	1.7 \pm 0.8						
0.75							80	Toxic
1	83	Toxic						
1.3	85	Toxic						
1.58			77	Toxic				
1.7	78	Toxic						
5			26	Toxic				
0 ^d (0.1% DMSO)	100	5.0 \pm 2.4			100	2.5 \pm 1.0		
TPA 0.05 ^e (0.1% DMSO)	128	36.7 \pm 3.3 †	133 ^f	13.7 \pm 6.8 ‡	198	18.2 \pm 2.7 †		
TPA 0.05 (0.5% DMSO)							109	14.3 \pm 5.4 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

^f % of cell growth compared to that of 5% water.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

‡ $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. 5% water.

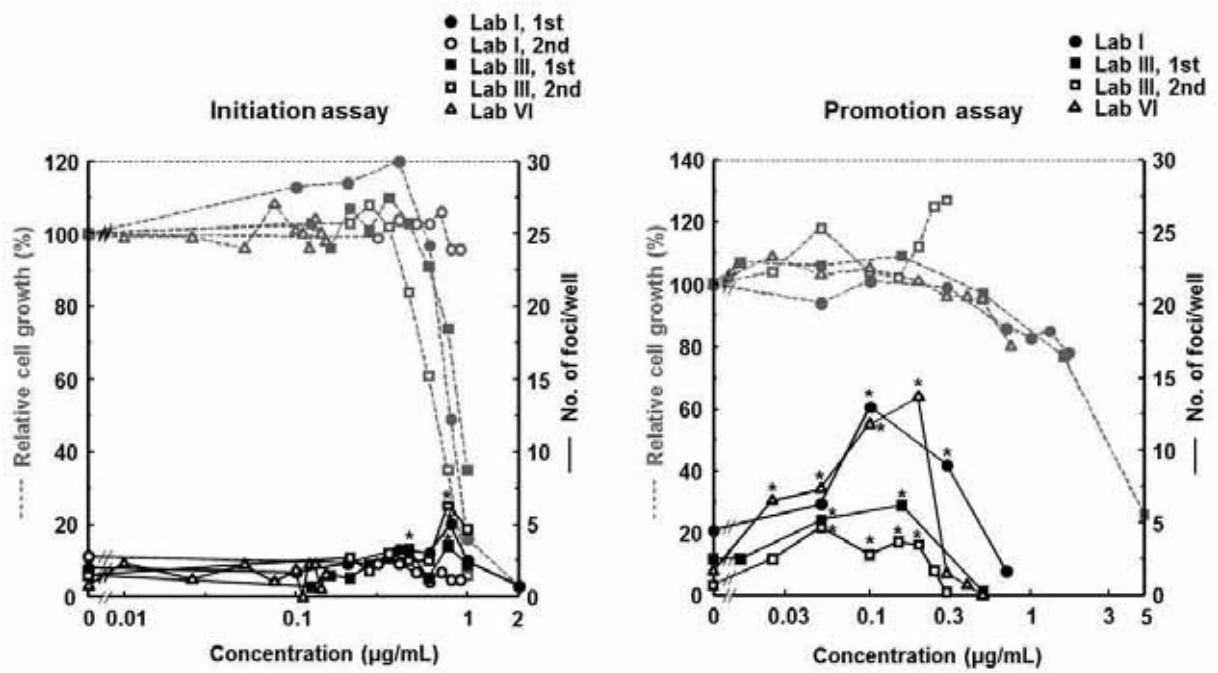


Fig. 11. Graphic view of the results of transformation assay and concurrent cell growth assay on sodium arsenite in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

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3.3.10 *o*-Toluidine hydrochloride

Table 11 and Fig. 12 represent the results of *o*-toluidine hydrochloride. Two laboratories, Lab V and Lab VI, gave positive results both in the initiation assay and in the promotion assay. The other laboratory, Lab I, gave negative results in both assays, although acceptable responses were obtained with the positive controls.

Table 11.
Results of transformation assay on *o*-toluidine hydrochloride in the 6-well method validation study

a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab V		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.5% DMSO)					100	1.0 \pm 1.5
0 (5% Water)	100	1.8 \pm 1.3	100	2.0 \pm 2.4		
100	98	2.8 \pm 1.8	92	2.5 \pm 2.5		
150					123	2.0 \pm 1.8
250			79	6.8 \pm 3.2 *		
300	77	2.5 \pm 1.5			89	1.7 \pm 1.2
450					109	1.8 \pm 1.3
500	61	3.2 \pm 2.5	50	7.0 \pm 2.0 *		
600					73	4.5 \pm 2.7 *
700	29	2.8 \pm 1.8				
750					64	3.7 \pm 1.0 *
1000	18	1.8 \pm 1.6	0	9.5 \pm 4.0 *	29	2.0 \pm 1.1
1250					13	0.3 \pm 0.5
1500					9	0.3 \pm 0.8
2000			-6	Toxic		
3000	3	Toxic	-4	Toxic		
5000	3	Toxic				
0 ^d (0.1% DMSO)	100	1.7 \pm 0.8	100	2.3 \pm 0.8		
MCA 1 ^e (0.1% DMSO)	53	19.3 \pm 3.9 †	33	43.7 \pm 6.0 †		
MCA 1 (0.5% DMSO)					40	21.7 \pm 6.3 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab V		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.5% DMSO)					100	0.7 \pm 0.8
0 (5% Water)	100	4.5 \pm 1.6	100	16.3 \pm 4.1		
10	94	5.5 \pm 2.0	94	27.0 \pm 4.6 *		
12.5					128	0.5 \pm 0.5
25					65	2.5 \pm 2.6
30	87	4.3 \pm 1.5	92	31.0 \pm 1.7 *		
100	83	3.7 \pm 1.6	75	26.7 \pm 7.4 *		
250			55	27.8 \pm 4.6 *	91	6.2 \pm 2.8 *
300	65	4.8 \pm 1.7				
500			42	28.0 \pm 6.4 *	48	8.0 \pm 3.5 *
700	50	Toxic				
1000	13	Toxic	33	6.0 \pm 4.3	68	0.5 \pm 0.8
1500					28	Toxic
2000					21	Toxic
0 ^d (0.1% DMSO)	100	7.8 \pm 2.2	100	14.8 \pm 5.5		
TPA 0.05 ^e (0.1% DMSO)	127	41.0 \pm 3.3 †	134	43.7 \pm 8.4 †		
TPA 0.05 (0.5% DMSO)					104	15.5 \pm 2.2 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

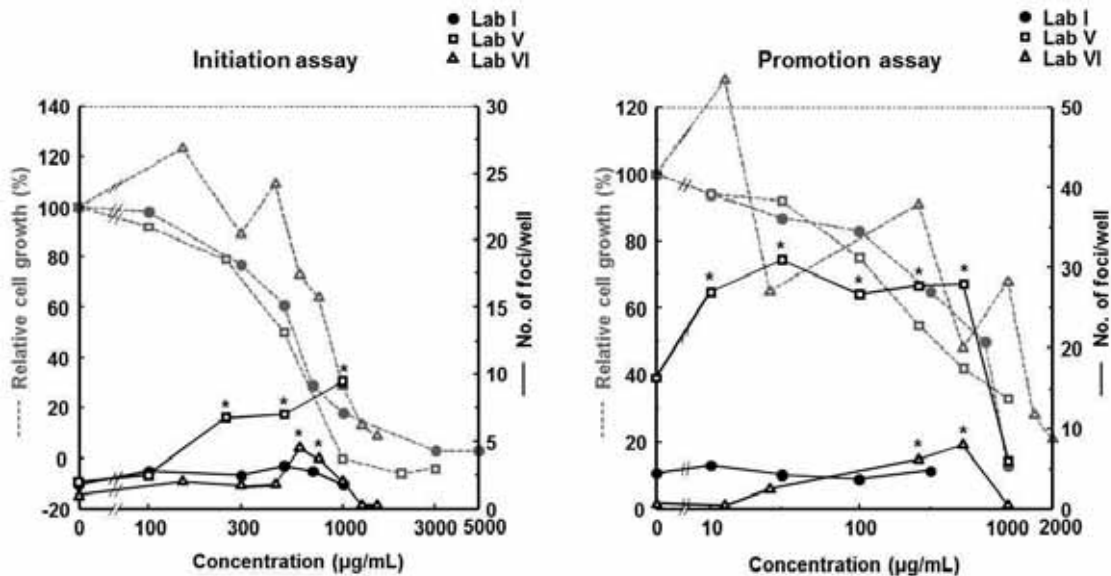


Fig. 12. Graphic view of the results of transformation assay and concurrent cell growth assay on σ -toluidine hydrochloride in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

3.3.11 Anthracene

The results for anthracene are exhibited in Table 12 and Fig. 13. All three laboratories obtained negative results in the initiation assay. In the promotion assay, the results were negative in two laboratories, Lab I and Lab II, but positive in Lab VI.

Inhibition of cell growth or focus formation by cytotoxicity was not observed at the concentrations tested in both initiation and promotion assays in all three laboratories. However, anthracene is poorly soluble in DMSO and other usable organic solvents and insoluble in water. The assays by three laboratories were conducted up to the limit of solubility in DMSO or beyond by using a chemical suspension in the vehicle. Despite this, the dosing employed in these assays was considered to be sufficient.

Table 12.
Results of transformation assay on anthracene in the 6-well method validation study

a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.5% DMSO)	100	4.2 \pm 2.6	100	2.5 \pm 0.8	100	1.3 \pm 1.2
1.25					95	1.5 \pm 1.0
1.56	107	5.8 \pm 2.7	99	2.0 \pm 0.6		
2.5					86	2.2 \pm 1.2
3.13	102	5.5 \pm 2.9	116	2.3 \pm 1.5		
5					95	2.5 \pm 1.6
6.25	109	4.0 \pm 1.9	112	3.5 \pm 2.6		
10					93	1.3 \pm 0.8
12.5	106	2.8 \pm 2.2	105	2.7 \pm 1.6		
20					100	2.5 \pm 1.0
25	110	5.7 \pm 2.1	110	1.8 \pm 1.3		
50	107	6.3 \pm 2.2				
0 ^d (0.1% DMSO)	100	2.7 \pm 0.8	100	3.0 \pm 1.5		
MCA 1 ^e (0.1% DMSO)	48	24.3 \pm 5.6 †	62	16.8 \pm 1.5 †		
MCA 1 (0.5% DMSO)					59	14.3 \pm 2.6 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.5% DMSO)	100	2.3 \pm 1.2	100	2.3 \pm 1.0	100	1.8 \pm 1.3
1.25					104	5.5 \pm 2.3
1.56	96	2.7 \pm 2.0	106	2.7 \pm 0.8		
2.5					97	6.7 \pm 2.3 *
3.13	94	2.0 \pm 0.9	102	3.8 \pm 2.8		
5					105	6.7 \pm 4.4 *
6.25	97	2.7 \pm 0.8	108	4.7 \pm 1.5		
10					96	9.8 \pm 4.4 *
12.5	98	2.8 \pm 0.8	93	3.5 \pm 2.5		
20					97	10.2 \pm 3.5 *
25	98	3.0 \pm 1.4	96	2.7 \pm 1.2		
50	96	1.3 \pm 0.8				
0 ^d (0.1% DMSO)	100	3.7 \pm 2.0	100	3.7 \pm 2.2		
TPA 0.05 ^e (0.1% DMSO)	131	17.5 \pm 6.5 †	137	10.8 \pm 1.6 †		
TPA 0.05 (0.5% DMSO)					136	14.7 \pm 2.9 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided Dunnett test, vs. corresponding solvent control.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

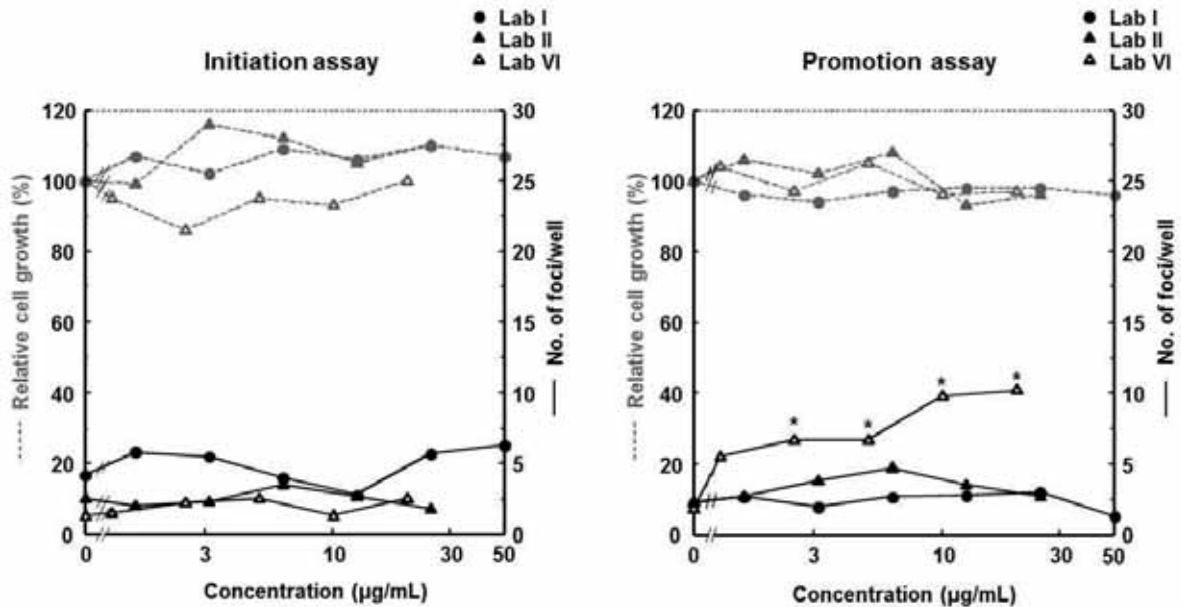


Fig. 13. Graphic view of the results of transformation assay and concurrent cell growth assay on anthracene in the 6-well method validation study.

* $p < 0.05$; one-sided Dunnett test.

3.3.12 L-Ascorbic acid

Table 13 and Fig. 14 show the results for L-ascorbic acid. All three laboratories reported L-ascorbic acid to be negative in both the initiation and promotion assays. Lab II voluntarily repeated the promotion assay. In the first promotion assay, Lab II observed that the transformed focus formation was reduced in the cultures treated with ascorbic acid and the number of foci diminished to 0 at the highest three concentrations. Despite this reduction in transformed foci, obvious general cytotoxicity was not observed. In the second promotion assay using a lower concentration range, Lab II confirmed that L-ascorbic acid decreased the number of transformed foci as compared to the negative control. Ascorbic acid also inhibited transformed focus formation in the promotion assays performed by the other two laboratories. Sodium ascorbate has been reported to inhibit (a) spontaneous focus formation, (b) MCA-induced focus formation, and (c) MCA-initiated and phorbol ester-promoted focus formation in the two-stage BALB/c 3T3 CTA [Sivak and Tu, 1980; Tsuchiya *et al.*, 2000].

Table 13. Results of transformation assay on L-ascorbic acid in the 6-well method validation study

a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (5% Water)	100	2.7 \pm 2.2	100	2.0 \pm 1.9	100	4.2 \pm 1.5
10			108	2.2 \pm 1.0		
25	91	1.7 \pm 1.2				
50	87	1.7 \pm 1.5				
75					96	4.7 \pm 2.0
100	78	2.7 \pm 1.0				
110					101	4.8 \pm 1.8
150	63	2.0 \pm 1.1	85	2.2 \pm 1.5	95	4.8 \pm 1.0
200	46	2.5 \pm 1.4				
210					91	3.7 \pm 1.6
250	25	2.8 \pm 0.8				
300	14	2.3 \pm 1.9	73	1.0 \pm 0.6	73	3.2 \pm 1.2
340			52	1.3 \pm 1.0		
380			32	0.2 \pm 0.4		
400	4	2.5 \pm 1.4	6	0.5 \pm 0.8		
420			0	0.2 \pm 0.4	5	2.8 \pm 2.0
600					1	Toxic
0 ^d (0.1% DMSO)	100	2.3 \pm 1.5	100	1.3 \pm 1.0		5.5 \pm 1.9
MCA 1 ^e (0.1% DMSO)	48	26.7 \pm 5.3 †	55	15.8 \pm 5.3 †		43.7 \pm 3.0 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab II, 1st run		Lab II, 2nd run		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well	CG	Foci/well
0 ^c (5% Water)	100	4.5 \pm 1.6	100	1.8 \pm 1.2	100	1.0 \pm 1.1	100	2.2 \pm 0.8
0.1					99	0.5 \pm 0.5		
1					103	0.3 \pm 0.5		
10					95	0.2 \pm 0.4		
50	92	1.2 \pm 1.2			102	0.0 \pm 0.0		
100	96	1.5 \pm 1.0			82	0.0 \pm 0.0		
110							104	0.5 \pm 0.5
150			101	0.2 \pm 0.4	92	0.0 \pm 0.0	100	0.7 \pm 0.5
200	92	1.5 \pm 0.8						
210							101	1.5 \pm 1.0
300	91	0.8 \pm 0.4	95	0.2 \pm 0.4			107	0.8 \pm 1.0
400	90	2.2 \pm 2.0						
420							111	1.7 \pm 1.2
500	89	Toxic						
600	86	Toxic	103	0.5 \pm 0.8			39	Toxic
700			102	0.3 \pm 0.5				
800	41	Toxic	92	0.0 \pm 0.0				
900			78	0.0 \pm 0.0				
1500			36	0.0 \pm 0.0				
0 ^d (0.1% DMSO)	100	7.5 \pm 4.0	100	3.2 \pm 1.9	100	1.0 \pm 1.3		2.7 \pm 1.4
TPA 0.05 ^e (0.1% DMSO)	123	37.2 \pm 4.7 \dagger	144	11.2 \pm 0.8 \dagger	156	11.5 \pm 1.9 \dagger		20.0 \pm 3.3 \dagger

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

\dagger $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

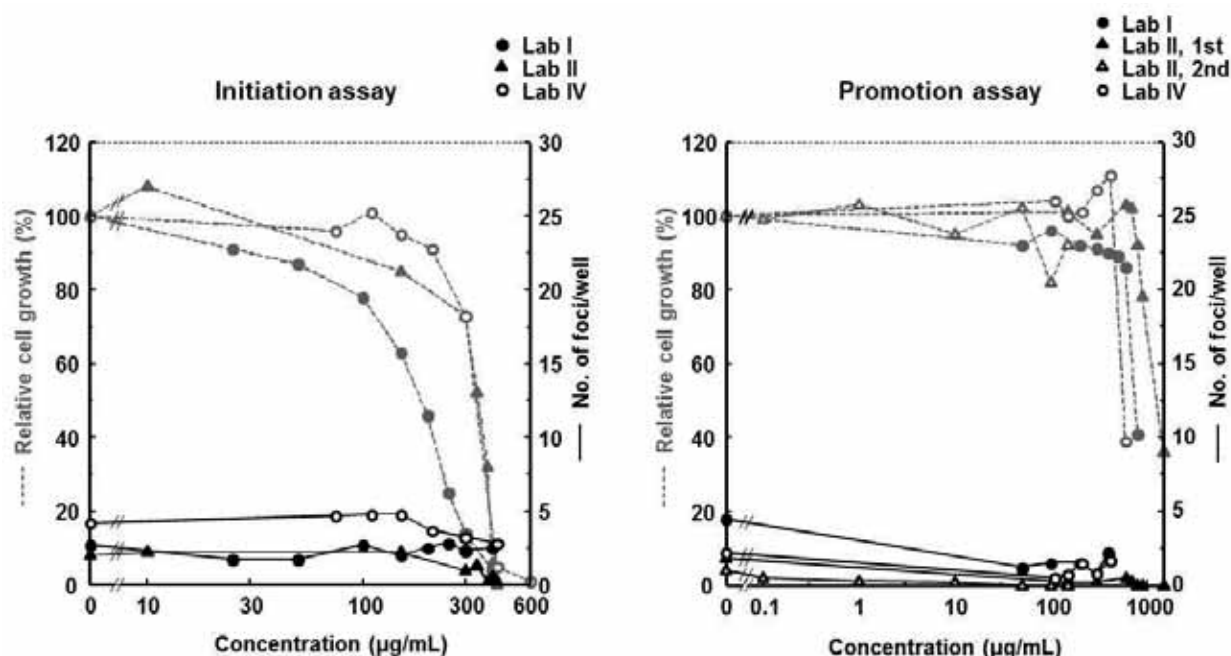


Fig. 14. Graphic view of the results of transformation assay and concurrent cell growth assay on L-ascorbic acid in the 6-well method validation study.

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2 **3.3.13 Caffeine**
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4 The results for caffeine are shown in Table 14 and Fig. 15. Two laboratories (Lab I and
5 Lab V) presented convincing results that caffeine was negative in the initiation assay.
6 Likewise, in the initiation assay reported by the other laboratory (Lab VI), no
7 statistically significant increase in the number of transformed foci was reported,
8 although cytotoxicity was also not induced at any of the concentrations of caffeine in the
9 concurrent cell growth assay. Therefore, it was concluded that the initiation assay by
10 Lab VI was incomplete because of inadequate dosing. Caffeine was judged negative by
11 all three laboratories in the promotion assay.
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15 Table 14.

16 Results of transformation assay on caffeine in the 6-well method validation study

17 **a) Initiation assay**

Concentration ($\mu\text{g/mL}$)	Lab I		Lab V		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.5% DMSO)					100	1.8 \pm 1.5
0 (5% Water)	100	2.3 \pm 1.6	100	2.7 \pm 1.2		
6.25					87	2.2 \pm 1.2
12.5					89	1.7 \pm 0.8
25					97	0.5 \pm 0.8
50					86	2.5 \pm 1.8
100	114	1.8 \pm 1.2	104	3.0 \pm 2.5	90	4.0 \pm 3.0
200			100	4.7 \pm 2.1		
300	92	1.7 \pm 1.0	76	5.2 \pm 3.3		
400	51	2.0 \pm 1.3	38	5.8 \pm 2.3		
500	23	1.7 \pm 1.0	18	3.7 \pm 2.3		
600	13	2.5 \pm 1.9				
700	11	1.0 \pm 1.3				
800	11	1.8 \pm 1.5				
0 ^d (0.1% DMSO)	100	1.3 \pm 1.2	100	3.3 \pm 1.8		
MCA 1 ^e (0.1% DMSO)	47	19.7 \pm 4.0 †	54	41.3 \pm 9.4 †		
MCA 1 (0.5% DMSO)					56	17.2 \pm 3.5 †

37 ^a % of cell growth compared to that of the solvent control.

38 ^b Average number of transformed foci/well \pm SD.

39 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

40 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
41 media in parentheses.

42 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
43 parentheses.

44 † $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.
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b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab V		Lab VI	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (0.5% DMSO)					100	4.0 \pm 1.8
0 (5% Water)	100	5.0 \pm 1.9	100	4.2 \pm 2.3		
6.25					88	3.3 \pm 1.2
10	93	2.0 \pm 1.4	80	1.5 \pm 1.0		
12.5					94	2.5 \pm 1.4
25	87	4.0 \pm 2.4			85	4.3 \pm 1.4
30			73	2.2 \pm 2.1		
50	82	3.5 \pm 1.0			88	1.5 \pm 1.0
100	70	1.5 \pm 0.8	35	2.5 \pm 1.6	78	1.8 \pm 1.5
200	57	0.2 \pm 0.4	33	2.3 \pm 1.0		
300	49	0.0 \pm 0.0	26	1.7 \pm 1.6		
500	39	0.2 \pm 0.4	27	0.5 \pm 0.5		
0 ^d (0.1% DMSO)	100	6.7 \pm 2.3	100	2.7 \pm 1.8		
TPA 0.05 ^e (0.1% DMSO)	121	35.5 \pm 6.1 †	106	21.5 \pm 3.5 †		
TPA 0.05 (0.5% DMSO)					141	16.5 \pm 4.8 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

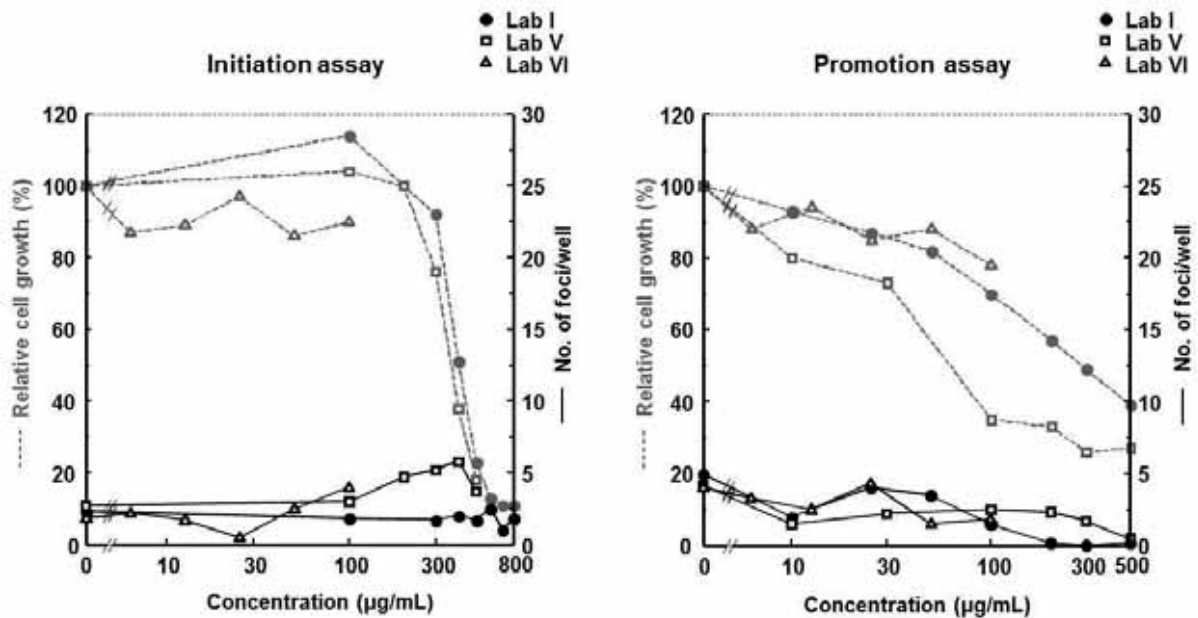


Fig. 15. Graphic view of the results of transformation assay and concurrent cell growth assay on caffeine in the 6-well method validation study.

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3 **3.3.14 D-Mannitol**
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5 Table 15 and Fig. 16 represent the results for D-mannitol. Both in the initiation assay
6 and in the promotion assay, D-mannitol was consistently negative in the three
7 laboratories up to the maximum concentration (5 mg/mL) tested.
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11 Table 15.
12 Results of transformation assay on D-mannitol in the 6-well method validation study

13 a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab III		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (5% Water)	100	2.7 \pm 1.0	100	2.7 \pm 1.4	100	1.2 \pm 1.3
15.8			103	1.0 \pm 0.9		
50			98	1.5 \pm 0.8		
100	107	1.5 \pm 1.0				
158			102	2.7 \pm 1.6		
300	105	2.0 \pm 2.4				
500			99	1.3 \pm 1.2		
1000	111	0.8 \pm 1.2				
1300					108	0.8 \pm 1.0
1580			110	0.8 \pm 0.8		
1800					111	1.8 \pm 0.8
2500	106	3.2 \pm 0.8			114	1.2 \pm 0.8
3500					109	1.8 \pm 1.2
5000	109	2.5 \pm 1.0	102	1.8 \pm 1.8	107	1.5 \pm 1.6
0 ^d (0.1% DMSO)	100	2.7 \pm 1.6				1.0 \pm 0.9
MCA 1^e (0.1% DMSO)	64	33.7 \pm 5.0 †	33^f	14.0 \pm 4.4 ‡		40.5 \pm 7.9 †

32 ^a % of cell growth compared to that of the solvent control.

33 ^b Average number of transformed foci/well \pm SD.

34 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

35 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
36 media in parentheses.

37 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
38 parentheses.

39 ^f % of cell growth compared to that of 5% water.

40 † p<0.05; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

41 ‡ p<0.05; one-sided t-test or Aspin-Welch test, vs. 5% water.
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b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab I		Lab III		Lab IV	
	CG ^a	Foci/well ^b	CG	Foci/well	CG	Foci/well
0 ^c (5% Water)	100	5.8 \pm 2.1	100	2.8 \pm 1.5	100	1.5 \pm 1.0
15.8			101	3.5 \pm 1.6		
50			93	3.0 \pm 1.4		
100	101	6.3 \pm 4.5				
158			102	2.2 \pm 1.9		
300	101	7.2 \pm 2.0				
500			101	2.8 \pm 2.3		
880					103	1.7 \pm 0.5
1000	102	9.2 \pm 3.2				
1300					107	1.5 \pm 0.8
1580			104	1.7 \pm 0.5		
1800					99	1.8 \pm 1.7
2500	103	5.2 \pm 2.6			110	1.7 \pm 1.9
3500					101	1.3 \pm 0.8
5000	103	3.7 \pm 2.0	106	4.3 \pm 1.5	99	1.2 \pm 1.2
0 ^d (0.1% DMSO)	100	8.2 \pm 3.6				2.3 \pm 1.0
TPA 0.05 ^e (0.1% DMSO)	134	50.2 \pm 4.6 †	157 ^f	8.7 \pm 3.6 ‡		10.2 \pm 2.1 †

^a % of cell growth compared to that of the solvent control.

^b Average number of transformed foci/well \pm SD.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

^f % of cell growth compared to that of 5% water.

† $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. corresponding solvent control.

‡ $p < 0.05$; one-sided t-test or Aspin-Welch test, vs. 5% water.

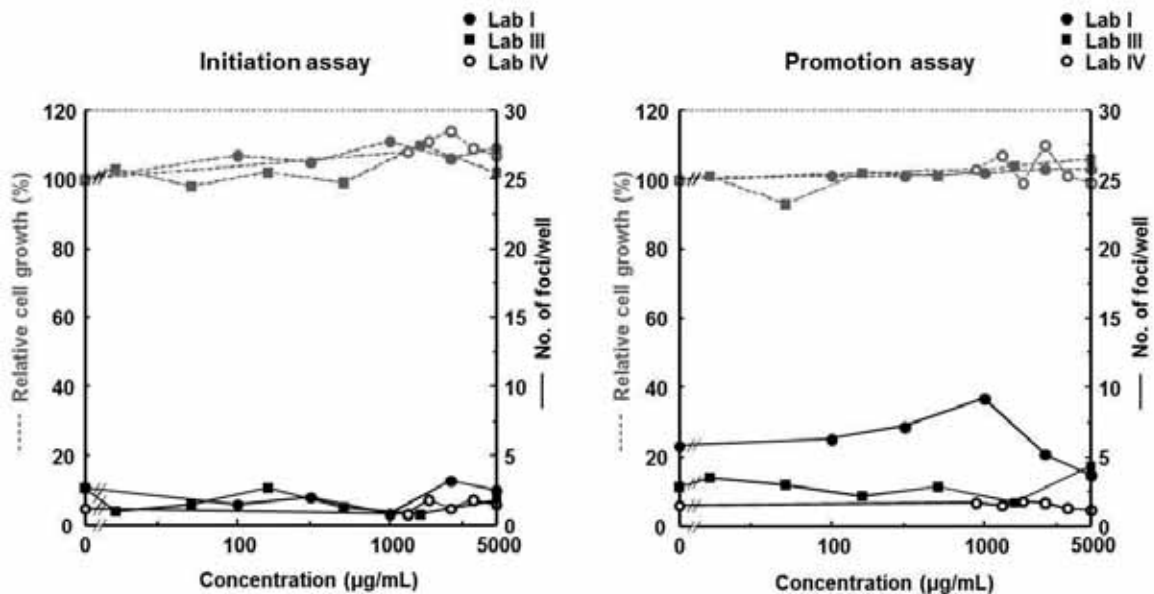


Fig. 16. Graphic view of the results of transformation assay and concurrent cell growth assay on D-mannitol in the 6-well method validation study.

3.4 Conclusion

The positive and negative calls for the test chemicals based on the results in the 6-well method validation study are summarized in Table 16. The positive or negative call for each chemical was individually determined based upon the Bhas 42 CTA results derived by each laboratory, into which the judgments in the initiation and promotion assays are integrated. Thus, a test chemical is positive in the Bhas 42 CTA, if it is positive in either of initiation and promotion assays. The overall judgment is determined by majority rule for the combined results obtained by all the participating laboratories.

3.4.1 Module 2 — Within-laboratory reproducibility

In this validation study, every test chemical was assayed including the negative control (DMSO) and each positive control (MCA in the initiation assay and TPA in the promotion assay). The numbers of transformed foci per well produced in the negative and positive controls are collected from all assays in the 6-well method validation study, clustered by individual laboratories and plotted in Fig. 3 for the initiation assay and the promotion assay. The average number of transformed foci per well in each laboratory is presented in Fig. 4. The ranges of the number of transformed foci per well produced in the negative control were narrow except for Lab V, which was one of two naive laboratories. The numbers of transformed foci in all the positive controls were statistically significantly increased in all the laboratories compared with those in the corresponding negative controls, although the number was considerably varied in Lab I (initiation and promotion assays) and Lab V (promotion assay). Thus, the VMT considered that the within-laboratory reproducibility for the negative and positive controls was satisfactory in all laboratories.

To estimate the within-laboratory reproducibility, the results obtained by Lab I were compared with those from a previously conducted Lab I in-house study (Table 17) [Sakai *et al.*, 2010]. With the exception of σ -toluidine, all assay results between the two studies were concordant. Therefore, the within-laboratory reproducibility for the Bhas 42 CTA was 92% (11/12) in Lab I.

Lithocholic acid, methapyrilene hydrochloride, mezerein and anthracene had been examined in the 6-well method pre-validation study [Tanaka *et al.*, 2009] prior to this validation program. We then compared the results obtained by the same laboratories in the pre-validation study and the validation study. In Table 18, Lab A and Lab IV, Lab F and Lab I, and Lab C and Lab II were the same laboratories. Their results between pre-validation and validation studies were determined to be highly comparable. Their calls in the transformation assay likewise agreed. The chemicals methapyrilene and mezerein, which showed potent promoting activity in the Bhas 42 CTA, caused weak but statistically significant increases in the number of transformed foci at their high concentrations in single instances in several initiation assays.

3.4.2 Module 3 — Transferability

In general, the Bhas 42 CTA can be performed in a laboratory that has experience in routine cell culture techniques. General cell culture laboratory equipment and instruments are sufficient to perform the proposed test method. All supplies and

1 reagents are readily available commercially. Like the other CTAs, however, the Bhas 42
2 CTA requires staff training beyond that of general cell culture techniques in order to
3 distinguish transformed foci from non-transformed foci. A training day was held in
4 January, 2007 at HRI for the pre-validation study of the 6-well method, which preceded
5 the validation study of the 6-well method (this validation study) and all the Japanese
6 laboratories that participated in the pre-validation study of the 6-well method, the
7 validation study of the 6-well method and the validation study of the 96-well method
8 were involved. Thus, Japanese laboratories had been trained in the procedures used for
9 the 6-well method Bhas 42 CTA and in the identification of Bhas 42 transformed foci.
10 Consequently, those laboratories were able to gain experience in the Bhas 42 CTA prior
11 to the conduct of the validation study of the 6-well method (this validation study). On
12 the other hand, American and German laboratories, Lab V and Lab VI, respectively,
13 which had been naive to the Bhas 42 CTA, received one day training at HRI for the
14 validation study of the 6-well method (this validation study) in August, 2007. There,
15 they were trained for the procedures of 6-well method Bhas 42 CTA and the
16 identification of Bhas 42 transformed foci. They started the validation study of the
17 6-well method (this validation study) after preliminary efforts using MCA, TPA and
18 DMSO (solvent control) in their own laboratories.

19
20 In the Bhas 42 CTA, foci are differentiated into two types, i.e., transformed foci and
21 non-transformed foci, instead of type III foci, type II foci and type I foci, which have
22 been used as descriptors in the BALB/c 3T3 CTA and the C3H101/2 CTA. A photo
23 catalog of non-transformed foci and different morphological variants of transformed foci
24 has been compiled to assist in the discrimination of transformed and non-transformed
25 foci. The photo catalog is attached to this report as Annex 3.

26
27 In Table 16, Lab I is the lead laboratory and Lab V and Lab VI are naïve laboratories.
28 Lab II, Lab III and Lab IV had some previous experience with the Bhas 42 CTA before
29 this validation study. To evaluate its inter-laboratory transferability, Bhas 42 CTA
30 results were compared between the lead laboratory and the naïve laboratories and
31 between the lead laboratory and the laboratories having Bhas 42 CTA experience. The
32 latter laboratories obtained results similar to those of the lead laboratory; furthermore,
33 the judgment calls for transformation were completely concordant. The naïve
34 laboratories, however, reported different results for *o*-toluidine hydrochloride and
35 anthracene. This experience suggests that the proper conduct of the Bhas 42 CTA
36 requires adequate training and practice to accurately discriminate and quantify
37 transformed foci and non-transformed foci.

38 39 40 **3.4.3 Module 4 – Between-laboratory reproducibility**

41
42 As shown in Figures 3 and 4, the numbers of transformed foci per well in the negative
43 controls for the initiation assay and the promotion assay were consistently low and less
44 than 10 in all the laboratories with an exception of negative control for the promotion
45 assay in Lab V, which was a naïve laboratory. Although the values in the positive
46 controls varied between laboratories, they were always statistically significantly
47 different from those in the corresponding negative controls in all the laboratories. Thus,
48 the VMT considered that the between-laboratory reproducibility was satisfactory for the
49 negative and positive controls.

50
51 In addition to the reproducibility findings resulting from the studies reported for
52 Module 2 and Module 3, above, the inter-laboratory reproducibility of the Bhas 42 CTA

was determined for the test chemicals listed in Table 16, each of which was distributed to three laboratories. In the initiation assay, concurring conclusions were reached by the three laboratories testing the same respective compounds with 8 out of 10 chemicals. Thus, the between-laboratory reproducibility for the initiation assay was 80% in this validation study. Since the tests of mezerein and caffeine were considered incomplete in two of three laboratories for mezerein and one of three laboratories for caffeine, those two chemicals were excluded from this calculation. In the promotion assay, concurrent conclusions were reached with 10 of 12 chemicals, and therefore between-laboratory reproducibility for the promotion assay was 83%. The judgments in the transformation assay were concordant between laboratories except for *o*-toluidine hydrochloride and anthracene, and consequently the between-laboratory reproducibility of Bhas 42 CTA was 83% (10/12). It is possible that the differences in results obtained were a consequence of those chemicals being examined by the naive laboratories, Lab V and Lab VI. Although mezerein was designated as equivocal (based upon the statistically significant increase in the number of transformed foci at only one dose) in the initiation assays conducted in two of the laboratories, the positive results obtained in the promotion assay were considered to have established mezerein as clearly positive in the Bhas 42 CTA. Likewise, despite the results for sodium arsenite in the initiation assay, which were split between negative and weakly positive, sodium arsenite was clearly positive in the promotion assay and thus considered positive in the Bhas 42 CTA in the three laboratories.

Between-laboratory reproducibility can be summarized as follows:

<u>Between-laboratory reproducibility</u>	
<u>Assay</u>	<u>%</u>
Initiation assay	80 (8/10)
Promotion assay	83 (10/12)
<u>Transformation assay (Bhas 42 CTA)</u>	<u>83 (10/12)</u>

3.4.4 Module 5 — Predictive capacity

The results demonstrated that the predictive capacity of Bhas 42 CTA is exceedingly high. When we compared *in vivo* carcinogenicity results and the overall judgments (judgment by majority rule among laboratories), all carcinogens including tumor-promoters were positive and all non-carcinogens were negative in the Bhas 42 CTA (Table 16). A 2x2 contingency table of the results obtained from 12 tested chemicals is presented below:

2x2 Contingency table of the results in the 6-well method Bhas 42 CTA validation study

		<i>In vivo</i> carcinogenicity		Total
		Carcinogen	Non-carcinogen	
Bhas 42 cell transformation assay	+	8	0	8
	-	0	4	4
Total		8	4	12

1 Table 16.

2 Judgments of the result for 12 test chemicals in the validation study of 6-well method.

3	4	5	6	Laboratory						Overall	Carcinogenicity
				I	II	III	IV	V	VI		
7	2-Acetylaminofluorene	Initiation	+			+		+	+	+	
8		Promotion	+			+		+	+		
9		Transformation ^b	+			+		+	+		
10	Cadmium chloride	Initiation	-	-	-				-	+	
11		Promotion	+	+	+				+		
12		Transformation	+	+	+				+		
13	Dibenz[<i>a,h</i>]anthracene	Initiation	+	+			+		+	+	
14		Promotion	-	-			-		-		
15		Transformation	+	+			+		+		
16	Lithocholic acid	Initiation	-			-		-	-	TP ^c	
17		Promotion	+			+		+	+		
18		Transformation	+			+		+	+		
19	Methapyrilene HCl	Initiation	-	-		-			-	+, TP	
20		Promotion	+	+		+			+		
21		Transformation	+	+		+			+		
22	Mezerein	Initiation	-		± ^d	±				TP	
23		Promotion	+		+	+			+		
24		Transformation	+		+	+			+		
25	Sodium arsenite	Initiation	-		+			i ^e		+	
26		Promotion	+		+			+	+		
27		Transformation	+		+			+	+		
28	<i>o</i> -Toluidine HCl	Initiation	-				+	+	+	+	
29		Promotion	-				+	+	+		
30		Transformation	-				+	+	+		
31	Anthracene	Initiation	-	-				-	-	-	
32		Promotion	-	-				+	-		
33		Transformation	-	-				+	-		
34	L-Ascorbic acid	Initiation	-	-		-			-	-	
35		Promotion	-	-		-			-		
36		Transformation	-	-		-			-		
37	Caffeine	Initiation	-				-	i	-	-	
38		Promotion	-				-	-	-		
39		Transformation	-				-	-	-		
40	D-Mannitol	Initiation	-		-	-			-	-	
41		Promotion	-		-	-			-		
42		Transformation	-		-	-			-		

41 ^a Judgment by majority rule.42 ^b Judgment in the Bhas 42 CTA, into which judgments in the initiation and promotion assays are integrated.43 ^c Tumor-promoter.44 ^d Equivocal: there was a statistically significant increase in transformation frequency at only one concentration.45 ^e Incomplete: unable to judge the results due to inadequate dosing.

46

1 Table 17.
 2 Within-laboratory reproducibility of the 6-well method: Comparison of Lab I results for
 3 the 6-well method validation study and the in-house study.

4	Compound	Assay	Validation study	In-house study ^a
5	2-Acetylaminofluorene	Initiation	+	+
6		Promotion	+	+
7		Transformation	+	+
8	Cadmium chloride	Initiation	-	-
9		Promotion	+	+
10		Transformation	+	+
11	Dibenz[<i>a,h</i>]anthracene	Initiation	+	+
12		Promotion	-	-
13		Transformation	+	+
14	Lithocholic acid	Initiation	-	-
15		Promotion	+	+
16		Transformation	+	+
17	Methapyrilene HCl	Initiation	-	-
18		Promotion	+	+
19		Transformation	+	+
20	Mezerein	Initiation	-	-
21		Promotion	+	+
22		Transformation	+	+
23	Sodium arsenite	Initiation	-	± ^b
24		Promotion	+	+
25		Transformation	+	+
26	<i>o</i> -Toluidine (HCl) ^c	Initiation	-	-
27		Promotion	-	+
28		Transformation	-	+
29	Anthracene	Initiation	-	-
30		Promotion	-	-
31		Transformation	-	-
32	L-Ascorbic acid	Initiation	-	-
33		Promotion	-	-
34		Transformation	-	-
35	Caffeine	Initiation	-	-
36		Promotion	-	-
37		Transformation	-	-
38	D-Mannitol	Initiation	-	-
39		Promotion	-	-
40		Transformation	-	-

41 ^a Results of Lab I in-house study [Sakai *et al.*, 2010].

42 ^b Equivocal: there was a statistically significant increase in transformation frequency at only one
 43 concentration.

44 ^c Hydrochloride was tested in the 6-well method validation study, but free *o*-toluidine was used in the
 45 in-house study.

46

1 Table 18

2 Within-laboratory reproducibility of the 6-well method: Comparison of results of 4
 3 chemicals that were duplicated between the pre-validation and validation studies of the
 4 **6-well method**

5 Compound	6 Assay	7 <u>Pre-validation^a</u>			8 <u>Validation^b</u>			9 Carcinogenicity <i>in vivo</i>
		10 <u>Laboratory^c</u>			11 <u>Laboratory</u>			
		12 A	13 C	14 F	15 I (F)	16 II (C)	17 IV (A)	
18 Lithocholic acid	19 Initiation	-	-	-	-	-	TP ^d	
	20 Promotion	+	+	+	+	+		
	21 Transformation ^e	+	+	+	+	+		
22 Methapyrilene HCl	23 Initiation			+	-		+ , TP	
	24 Promotion			+	+			
	25 Transformation			+	+			
26 Mezerein	27 Initiation	-	-	-		± ^f	TP	
	28 Promotion	+	+	+	+	+		
	29 Transformation	+	+	+	+	+		
30 Anthracene	31 Initiation		-	-	-	-	-	
	32 Promotion		-	-	-	-		
	33 Transformation		-	-	-	-		

34 ^a The results in Lab A, Lab C and Lab F were obtained from the report on the pre-validation study of
 35 the 6-well method [Tanaka et al., 2009].

36 ^b The results of Lab I, Lab II and Lab IV were obtained from Table 16 in the present validation report.

37 ^c Lab A and Lab IV are the same laboratory; Lab C and Lab II are the same laboratory; Lab F and Lab
 38 I are the same laboratory.

39 ^d Tumor-promoter.

40 ^e Judgment in the Bhas 42 CTA, into which judgments in the initiation and promotion assays are
 41 integrated.

42 ^f Equivocal: there was a statistically significant increase in transformation frequency at only one
 43 concentration.

34 3.4.5 Negative and positive controls

35 Although the number of foci induced by the positive control varied between laboratories
 36 and sometimes within a given laboratory (Fig. 3), there was a statistically significant
 37 difference between the positive control and the corresponding negative control in every
 38 initiation and promotion assay. Taken together with the results reported for Modules
 39 2-5, these data indicate that the Bhas 42 CTA 6-well method is both feasible and
 40 reliable.

41 3.4.6 Vehicle

42 In this validation study, the choice of solvent/vehicle (DMSO or distilled water) was left
 43 to each laboratory, considering the conditions of actual assay practice. Consequently,
 44 different solvents were chosen among the laboratories for 3 chemicals, sodium arsenite,
 45 *o*-toluidine hydrochloride and caffeine. Despite this difference, the judgment of
 46 transforming activity did not appear to be affected for these chemicals (Tables 10, 11
 47 and 14). Furthermore, there is no information in literature that these chemicals are

1 unstable in either solvent.
2
3

4 **3.4.7 Chemical treatment procedures in the initiation assay** 5

6 In the initiation assay the cells were treated with test chemical by the direct addition of
7 chemical solution to the culture medium or by the replacement of existing medium with
8 fresh medium containing the chemical, although in the promotion assay the chemical
9 treatment was always carried out by periodic exchanges of medium containing test
10 chemical, as described in section 3.2.2, Initiation assay in the text and in the protocol for
11 the 6-well method validation study in Annex 2. The choice of treatment procedure in the
12 initiation assay was left to each laboratory. Only Lab IV out of 6 participating
13 laboratories treated the cells by the medium exchange method. There was no difference
14 in the judgment of results obtained with the two treatment procedures in the initiation
15 assay for the particular 6 chemicals, 2-acetylaminofluorene, lithocholic acid,
16 methapyrilene hydrochloride, mezerein, L-ascorbic acid and D-mannitol (Table 16).
17

4 Validation Study on the 96-well method

4.1 Study design and chemicals tested

4.1.1 Study design

Four laboratories participated and the study proceeded stepwise (pre-validation phase, validation phase I and validation phase II). A total of 25 chemicals were tested including duplication between phases, i.e., chemicals tested in (a) both the pre-validation phase and validation phase I, or (b) both validation phase I and validation phase II. Such chemicals were recorded as having been tested twice. Before starting the validation study, a workshop for the technical transfer of the 96-well method was held by HRI on 18 November, 2008. All participating laboratories took part in the workshop to learn the protocol, the assay procedures and judgment criteria for identifying transformed foci (i.e., how to count wells with transformed foci).

In the pre-validation study, MCA and TPA were examined to confirm the transferability of the test protocol. The chemical names were accessible and the concentrations tested were given in this phase: 0.03, 0.1, 0.3 and 1 µg/mL of MCA, and 0.025, 0.05, 0.1 and 0.2 µg/mL of TPA. Based on the transformation results obtained for the negative and positive controls in the pre-validation study, assay acceptance criteria were established.

In the validation phase I study, seven coded chemicals were examined and all of the chemicals were tested by all four laboratories to assess the inter-laboratory reproducibility of the 96-well method (Tables 19 and 21). MCA and TPA were included among the coded chemicals to evaluate intra-laboratory reproducibility by comparing the results obtained between the pre-validation phase and the validation phase I. The selected chemicals except TPA were the same as those tested in the ECVAM-initiative pre-validation study of BALB/c 3T3 CTA so that the results could be compared between the two CTAs.

In the validation phase II study, 16 coded chemicals were examined. Lab 4 did not participate in the phase II study. Each chemical was tested by two laboratories to confirm the inter-laboratory reproducibility of the 96-well method (Table 20). In order to assess the predictivity of the Bhas 42 CTA for chemical carcinogenicity, the chemicals selected included both tumor-promoters and non-carcinogens, with the latter comprising half of the total chemicals (Table 22). Ideally, it would have been good to include chemicals with low potency. However, it is difficult to assess precise potency based on available data. It could be argued that the process of dilution of test chemicals resulting in reduced concentrations may approximate potency reduction so as to mimic the testing of low potency chemicals. Benzo[a]pyrene was included among the coded test chemicals to ensure within-laboratory reproducibility between phases I and II. The chemicals which were used in the 6-well method validation study were also included among the coded test chemicals to confirm that the 6-well method and the 96-well method were equivalent in the transformation response.

The chemical properties and classes of test chemicals for the 96-well method validation study are presented Annex 15.

4.1.2 Chemicals tested and negative and positive controls

The chemicals selected, their CAS number, their distribution among laboratories and chemical codes for the validation study are listed for phase I and II studies in Tables 19 and 20, respectively. In the phase I study, the solvent (DMSO) for MCA, 2-acetylaminofluorene, benzo[*a*]pyrene and TPA was pre-selected by the VMT and their highest concentrations were also pre-assigned to be $\leq 100 \mu\text{g/mL}$, $\leq 100 \mu\text{g/mL}$, $\leq 100 \mu\text{g/mL}$ and $\leq 1 \mu\text{g/mL}$, respectively, in the culture medium. In the phase II study, the approximate water solubility of each coded chemical was suggested and the top doses of benzo[*a*]pyrene, dibenz[*a,h*]anthracene, mezerein and phorbol were pre-assigned to be $\leq 10 \mu\text{g/mL}$, $\leq 10 \mu\text{g/mL}$, $\leq 0.1 \mu\text{g/mL}$ and $\leq 5 \mu\text{g/mL}$, respectively, in the medium. These amounts were predetermined because these chemicals were expensive and their availability was limited. For the other chemicals, each individual participating laboratory determined the dose ranges for the transformation assays according to dose setting procedures using the cell growth assays.

MCA and TPA were chosen as positive controls for the initiation assay and the promotion assay, respectively.

Table 19

Test chemicals, CAS numbers and code names with allocation to the laboratories in phase I of 96 well method validation study.

Chemical	CAS no ^a	Carcinogenicity ^b	Code			
			Allocation to lab			
			1	2	3	4
3-Methylcholanthrene*	56-49-5	+	A-01	C-03	B-06	D-02
Benzo[<i>a</i>]pyrene*	50-32-8	+	A-02	C-04	B-07	D-01
2-Acetylaminofluorene*	53-96-3	+	A-03	C-07	B-01	D-04
TPA ^c *	16561-29-8	+,TP	A-04	C-05	B-03	D-07
<i>o</i> -Toluidine	95-53-4	+	A-05	C-06	B-02	D-03
Anthracene	120-12-7	-	A-06	C-01	B-04	D-05
Phenanthrene	85-01-8	-	A-07	C-02	B-05	D-06
3-Methylcholanthrene	56-49-5	+	Positive control for Initiation assay			
TPA ^c	16561-29-8	+	Positive control for promotion assay			

^a Chemical Abstract Service registry number.

^b +, carcinogen; -, non-carcinogen; TP, tumor-promoter

^c 12-*O*-tetradecanoylphorbol-13-acetate.

*The solvent (DMSO) and the highest concentrations (3-methylcholanthrene, 100 $\mu\text{g/mL}$; benzo[*a*]pyrene, 100 $\mu\text{g/mL}$; 2-acetylaminofluorene, 100 $\mu\text{g/mL}$; TPA, 1 $\mu\text{g/mL}$) were designated by VMT in advance.

1
2 Table 20
3 Test chemicals, CAS numbers and code names with allocation to the laboratories in
4 phase II of 96-well method validation study.

Chemical	CAS no. ^a	Carcinogenicity ^b	Code		
			Allocation to lab		
			1	2	3
<i>N</i> -Methyl- <i>N</i> '-nitro- <i>N</i> -nitrosoguanidine	70-25-7	+	B121		A101
Benzo[<i>a</i>]pyrene*	50-32-8	+	B122		A102
Dibenz[<i>a,h</i>]anthracene*	53-70-3	+		C131	A103
Sodium arsenite	7784-46-5	+	B123		A104
Cadmium chloride	10108-64-2	+	B124		A105
Methapyrilene HCl	135-23-9	+, TP		C132	A106
Mezerein*	34807-41-5	TP		C133	A107
Lithocholic acid	434-13-9	TP		C134	A108
Pyrene	129-00-0	-		C135	A109
Caprolactam	105-60-2	-	B125		A110
Ampicillin sodium	69-52-3	-	B126		A111
L-Ascorbic acid	50-81-7	-	B127		A112
D-Mannitol	69-65-8	-		C136	A113
Caffeine	58-08-2	-		C137	A114
Phorbol*	17673-25-5		B128		A115
Eugenol	97-53-0	-		C138	A116
3-Methylcholanthrene	56-49-5	+	Positive control		
TPA ^c	16561-29-8	+	Positive control		

26 ^a Chemical Abstract Service registry number.

27 ^b +, carcinogen; -, non-carcinogen; TP, tumor-promoter

28 ^c 12-*O*-tetradecanoylphorbol-13-acetate.

29 * The highest concentrations in the final culture medium (benzo[*a*]pyrene, 10 µg/mL;
30 dibenz[*a,h*]anthracene, 10 µg/mL; mezerein, 0.1 µg/mL; phorbol, 5 µg/mL) were suggested by VMT in
31 advance.

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36 The carcinogenicity, including tumor-promoting activity, and genotoxicity of test
37 chemicals in phases I and II are presented in Tables 21 and 22, respectively. Those
38 activities of test chemicals were based on the review paper, DRP 31[OECD, 2007] and a
39 review by Kirkland *et al.*, [2005], except for the carcinogenicity of methapyrilene
40 hydrochloride, lithocholic acid and phorbol. Methapyrilene was included in the list of
41 carcinogens in DRP 31 and reported to be a tumor-promoter by Lijinsky *et al.* [1992].
42 The tumor-promoting activity of lithocholic acid was reported by Reddy *et al.* [1975].
43 Carcinogenicity (leukaemogenic action) of phorbol has been reported to be positive in
44 SWR mice [Berenblum and Lonai, 1970], and positive or negative depending on the
45 mouse strains [Armuth, 1976], and its tumor-promoting activity in mouse skin is also
46 positive in a sensitive strain, STS mice, but negative in other strains, Charles River CD1
47 mice and Sencar mice [Baird and Boutwell, 1971; Slaga *et al.*, 1976 and 1980].
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1
2 Table 21
3 Genotoxicity and carcinogenicity of the test chemicals for phase I of 96-well method
4 validation study.

Chemicals	Genotoxicity ^a				IARC class	Carcinogenicity ^b
	<i>In vitro</i>			<i>In vivo</i>		
	Ames ^c	ML ^d	CA ^e	MN ^f		
3-Methylcholanthrene	+	+	+			+
Benzo[a]pyrene	+	+	+	+	2A	+
2-Acetylaminofluorene	+	+	+	+		+
TPA ^g	–		+			+, TP
<i>o</i> -Toluidine	+/-	+/-	+	–	2A	+
Anthracene	+w/-	+	–		3	–
Phenanthrene	+w/-		–		3	–

15 ^a Source: DRP 31 [OECD, 2007] and a review by Kirkland *et al.* [2005]: +, positive; –, negative; +/-,
16 positive and negative results in DRP 31; +w, weakly positive.

17 ^b Source: DRP 31 [OECD, 2007]: +, carcinogen; –, non-carcinogen; TP, tumor-promoter.

18 ^c Ames test.

19 ^d Mouse lymphoma test.

20 ^e Chromosomal aberration test.

21 ^f Micronucleus test.

22 ^g 12-*O*-Tetradecanoylphorbol-13-acetate

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1
2 Table 22
3 Genotoxicity and carcinogenicity of the test chemicals for phase II of 96-well method
4 validation study.

Chemicals	Genotoxicity ^a				IARC class	Carcinogenicity ^b
	<i>In vitro</i>			<i>In vivo</i>		
	Ames ^c	ML ^d	CA ^e	MN ^f		
MNNG ^g	+	+	+	+		+
Benzo[<i>a</i>]pyrene	+	+	+	+	2A	+
Dibenz[<i>a,h</i>]anthracene	+	+	+		2A	+
Sodium arsenite	–	+		+	1	+
Cadmium chloride	–	+	+/-	–	1	+
Methapyrilene HCl	–	+/-	+			+ ,TP ^h
Mezerein	–					TP
Lithocholic acid	–	+/-	–			TP ⁱ
Pyrene	–	+	–		3	–
Caprolactam	–	–	–	–	4	–
Ampicillin sodium	–	–	–		3	–
L-Ascorbic acid	+w/-		–	+		–
D-Mannitol	–	–	–	–		–
Caffeine	–		+	+	3	–
Phorbol						*
<u>Eugenol</u>	–	+	+	+/-	3	–

24 ^a Source: DRP 31 [OECD, 2007] and a review by Kirkland *et al.* [2005]: +, positive; –, negative; +/-, positive and negative results in DRP 31; +w, weakly positive.

25
26 ^b Source: DRP 31 [OECD, 2007] except for lithocholic acid and phorbol: +, carcinogen; –, non-carcinogen; TP, tumor-promoter

27 ^c Ames test

28 ^d Mouse lymphoma test

29 ^e Chromosomal aberration test

30 ^f Micronucleus test

31 ^g *N*-Methyl-*N*'-nitro-*N*'-nitrosoguanidine

32 ^h Lijinsky *et al.*, [1992]

33 ⁱ Reddy *et al.*, [1975]

34 * Carcinogenicity in mice (leukaemogenic action) is positive or negative depending on the strains
35 [Berenblum and Lonai, 1970; Armuth, 1976]. Tumor-promoting activity in mouse skin is positive in a
36 sensitive strain but negative in other strains [Baird and Boutwell, 1971; Slaga *et al.*, 1976 and 1980].
37 These strains are mentioned in the text.
38

39
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41
42 In the pre-validation study, the chemicals tested (MCA and TPA) were selected by VMT
43 and distributed by the lead laboratory, HRI, to the other participating laboratories. In
44 phases I and II, the test chemicals were selected by VMT, and purchased, coded and
45 distributed by JaCVAM, which is institutionally and geographically independent of the
46 participating laboratories. The chemicals were distributed to the chemical repository
47 officer of each individual participating laboratory. The participating laboratories were
48 not informed of the chemical names or the chemical selection criteria used in phases I
49 and II. The suppliers, catalog numbers and lot numbers of the test chemicals are
50 presented in Tables 55 and 56 of Annex 5.

51 52 53 4.1.3 Time line

54
55 18/04/2008 Preparation of a framework for the validation study on the 96-well
56 method.

1	29-30/10/2008	1st VMT meeting: explanation of study plan, discussion on and
2		determination of the protocol for pre-validation study (Ver. 2), test
3		chemical selection for the pre-validation study to optimize the protocol
4		(MCA and TPA), schedule of the pre-validation phase; chemical
5		selection by VMT for the phase I validation study.
6	12/11/2008	Start of the pre-validation study. (MCA, TPA, Bhas 42 cells and FBS
7		were shipped by HRI).
8	18/11/2008	Workshop for technical transfer of the 96-well method according to the
9		protocol Ver. 2.
10	24/12/2008	Decision on the method of statistical analysis for the data.
11	13/1/2009	Deadline for data submission in the pre-validation study.
12	17/1/2009	Statistical analysis of the results in the pre-validation study by the
13		statistical team.
14	26/1/2009	Joint meeting of Japanese VMT and participating laboratories:
15		evaluation of the results in the pre-validation study, exchange of
16		information on the technical issues, harmonization of focus counting
17		among laboratories, decision regarding assay acceptance criteria*,
18		and planning of phase I study.
19		* As a result of the discussion in this joint meeting the protocol of
20		96-well method was amended for the phase I validation study (Ver. 3).
21	29/1/2009	Start of the phase I validation study. (The test chemicals were shipped
22		by JaCVAM to participating laboratories.)
23	16/9/2009	Deadline for data submission in the phase I validation study
24	15/10/2009	Joint meeting of Japanese VMT and participating laboratories: report
25		of the results in the phase I validation study, decoding of test
26		chemicals.
27	9-10/11/2009	2nd VMT Meeting: evaluation of the results in the phase I validation
28		study, and planning of the phase II study.
29	6/1/2010	Start of the phase II validation study. (Test chemicals were shipped by
30		JaCVAM to participating laboratories.)
31	14/7/2010	Deadline for data submission in the phase II study.
32	6-7/10/2010	3rd VMT Meeting: report and evaluation of the results in the phase II
33		validation study, approval of the 96-well method, decoding test
34		chemicals.
35	15/9/2011	Meeting for the international Bhas 42 CTA validation study: Future
36		schedule on the Bhas 42 CTA to develop OECD/TG: VMT members
37		and the other participants agreed that the Bhas 42 CTA validation
38		study report should undergo peer review by the ESAC through the
39		ECVAM process.
40		
41		
42		

4.2 Protocol

The schematic of 96-well method is the same as that of 6-well method (Fig. 2). The procedures of 96-well method is fundamentally the same as those of 6-well method as shown in Fig. 1. The differences from the 6-well method are as follows:

- The cell number inoculated per well are proportional relative to the well-size.
- The number of wells having transformed foci is quantified relative to the total number of observed wells, instead of the number of transformed foci in every

1 well, as is the case in the 6- well method. The wells having transformed foci in
2 the bottom are counted, but the wells having transformed foci only on the wall
3 are not counted. This is because the narrow shape of the well in a 96-well
4 micro-plate is not conducive to observing and distinguishing transformed foci
5 from non-transformed foci on the well wall. (In the 6-well micro-plate,
6 transformed foci are counted both on the bottom and on the wall since technical
7 problems of this nature are not encountered.)

- 8 ● The statistical analysis for the 96-well method is carried out using the
9 chi-square test with Bonferroni adjustment, instead of the multiple comparison
10 using the Dunnett test which is applied for the 6-well method.

11
12 The detailed test protocols used in the pre-validation phase, validation phase I and
13 validation phase II of the 96-well method validation study are presented in Annex 6,7
14 and 8, respectively. The protocol, Ver. 2 was used for the pre-validation phase, and
15 amended to Ver. 3 for the phase I studies and Ver. 4 for the phase II studies to reflect the
16 results, discussion and decisions made from the previous phases.

17 18 19 **4.2.1 Preparation**

20
21 Cells, materials and chemicals were prepared in the same manner as described in 3.2.1
22 of the 6-well method validation study. All materials, reagents, and FBS used are shown
23 in the test protocols annexed (Annex 6, 7 and 8). The vehicles used to dissolve test
24 chemicals and their final concentrations in the media are presented in the tables of
25 assay results (Tables 23-47).

26
27 Concerning the preparation and storage of test-chemical solutions, a section was added
28 in “I.1.3) Chemicals” in the protocol Ver. 3 (Page 1 in Annex 7). This addition was not an
29 amendment but a correction, because the procedures included were explained in the
30 workshops for technical transfer of the protocol and were performed in the 6-well
31 method validation study and the pre-validation phase of 96-well method validation
32 study. The insertion was as follows:

- 33
34 ● Preparation and storage of test-chemical solutions: Test-chemical solutions are
35 prepared before use as a general rule. Working solutions may be preserved in
36 aliquots at -20°C for less than 10 days and thawed before use, but must not be
37 re-frozen. When the amount of test chemical supplied is too little to be weighed
38 for every assay, a stock solution is prepared at a concentration as high as
39 possible or at some other appropriate concentration and stored in aliquots at
40 -20°C for up to a year and thawed before use. The stock solution is never
41 re-frozen. When suspensions of test chemicals are used, they must be prepared
42 before use and cannot be stored for the future use. The stock solutions of MCA
43 and TPA in DMSO can be stored at -20 °C for at least two years, if they are not
44 thawed. [Page 1 in Annex 7 and 8]

45 46 47 **4.2.2 Initiation assay**

48 49 **4.2.2.1 Cell growth assay**

50
51 A cell suspension at 4000 cells/mL in DF5F was distributed into each well of 96-well

1 micro-plates in a 0.05 mL volume (200 cells/well, day 0). Eight wells were prepared for
2 each treatment group. At 24 h after seeding, medium containing a test chemical at two
3 times the final concentration or containing vehicle alone was prepared, and 0.05 mL of
4 the preparation was added to each well without medium exchange so that the volume of
5 medium was 0.1 mL/well in total. On day 4, the medium containing the test chemical
6 was replaced with 0.1 mL/well of fresh DF5F. On day 7, the cells were fixed and stained
7 with CV, and the relative cell growth of cultures treated with a chemical was calculated
8 according to the procedures as described in 3.2.2.1.

9 10 **4.2.2.2 Dose setting for the transformation assay**

11
12 Five or more concentrations were selected based on the results of cell growth assays as
13 described in 3.2.2.2.

14 15 **4.2.2.3 Transformation assay to examine initiating activity**

16
17 One tube of the frozen Bhas 42 cells was thawed and grown in M10F up to about 70%
18 confluence and then subcultured in DF5F to reach about 70% confluence again as
19 described in 3.2.2.3. Thereafter, cells were cultured in DF5F. The cells were trypsinized
20 and suspended at a density of 4,000 cells/mL and seeded into each well of 96-well
21 micro-plates in 0.05 mL volumes (200 cells/well, day 0). One plate and 8 wells were
22 prepared per concentration, of which one plate was reserved for the transformation
23 assay and eight wells were reserved for the concurrent cell growth assay. At 24 h after
24 seeding, medium containing a test chemical at two times the final concentration or
25 containing vehicle alone was prepared, and 0.05 mL of the preparation was added to
26 each well without medium exchange so that the volume of medium was 0.1 mL/well in
27 total. The treatment in the initiation phase was continued for 72 h. Following the
28 exposure period, all treatment media were removed and the cells were refed with 0.1
29 mL/well of medium without the test chemical (day 4) and subsequently cultured in the
30 normal medium until day 21, receiving medium exchanges on day 7, day 11 and day 14.
31 The cells were then fixed with methanol and stained with Giemsa's solution. Each assay
32 also included MCA (1 µg/mL) as the positive control.

33 34 35 **4.2.3 Promotion assay**

36 37 **4.2.3.1 Cell growth assay**

38
39 A cell suspension of 4000 cells/mL in DF5F was distributed into each well in 0.1 mL of
40 medium (400 cells/well, day 0), and on day 4, the culture medium was replaced with a
41 medium containing a given test chemical. On day 7, the cells were fixed and stained
42 with CV, and the relative cell growth of cultures treated with a chemical (chemical vs.
43 vehicle) was calculated, as described in 3.2.2.1.

44 45 **4.2.3.2 Dose setting for the transformation assay**

46
47 Five or more concentrations were selected based on the results of cell growth assays as
48 described in 3.2.3.2.

49 50 **4.2.3.3 Transformation assay to examine promoting activity**

51
52 The promotion assay was carried out in the same manner as the initiation assay

1 (4.2.2.3) except for the following steps. The cells were seeded at a density of 400
2 cells/well in 0.1 mL of medium (day 0) and cultured for 4 days without a medium
3 exchange. On day 4, day 7, and day 11, the culture medium was replaced with fresh
4 medium containing a given test chemical or vehicle alone and the treatment in the
5 promotion phase was continued until day 14 (for a total of 10 days). The cells were then
6 cultured in normal medium without the test chemical for one week until day 21. Each
7 assay also included TPA (0.05 µg/mL) as the positive control.
8
9

10 **4.2.4 Focus count and statistical analysis**

11
12 The transformed foci were judged using the same criteria as those used in the 6-well
13 method (Annex 3). The number of wells having transformed foci versus the total
14 number of observed wells was recorded. The bottom area of each well was observed
15 under a stereoscopic microscope, but the wall of each well was excluded from
16 consideration for the reasons discussed under 4.2 above. The proportion of wells having
17 transformed foci was statistically analyzed by the chi-square test with Bonferroni
18 adjustment. "Toxic" was recorded for plates or wells that were not confluent at the end
19 of transformation assay because of cytotoxicity resulting from chemical treatment.
20

- 21 ● The p-value of the chi-square test was tentatively set at <0.05 or <0.025
22 (upper-sided) in protocol Ver. 2 and Ver. 3. The transformation frequency was
23 analyzed with two p-values, <0.025 and <0.05 (upper-sided), in the phase I
24 study, and the p-value was finally fixed at <0.5 (upper-sided) in protocol Ver. 4,
25 based on the results in the phase I study (Page 8 in Annex 7-8).
26
27

28 **4.2.5 Judgment**

29
30 The assay results were judged positive when there existed two or more sequential doses
31 that induced statistically significant increases in the proportion of wells having
32 transformed foci, and negative when there was no dose showing a statistically
33 significant increase in the proportion of wells having transformed foci. When the
34 statistically significant increase was at only one dose, the assay result was regarded as
35 equivocal. In such cases, the initiation or promotion assay together with the
36 concomitant cell growth assay was repeated and included the positive dose in the first
37 assay. The chemical was judged to be positive if a statistically significant increase in the
38 proportion of wells having transformed foci resulted at one or more concentrations in
39 the second assay.
40

- 41 ● In the pre-validation phase and the validation phase I study, the repeat of
42 assay to make a final judgment was compulsory for the participating
43 laboratory when the statistically significant increase was at only one dose. In
44 the validation phase II study, that requirement was amended so that the
45 decision to re-test was left up to each laboratory. However, in the absence of the
46 repeat test, the judgment remained equivocal. This amendment was
47 introduced in the interest of time and cost savings. (Page 8 in Annex 7-8)).
48
49

50 **4.2.6 Acceptance criteria**

51
52 The assay acceptance criteria were established in protocol Ver. 2 for the pre-validation

1 study, as follows (citation from Annex 6, protocol Ver. 2):

2
3 The initiation or promotion assay is repeated independently, as needed, to satisfy
4 the assay acceptance criteria.

5 1. Concurrent cell growth assay

- 6 ▪ At least one concentration should be included near the NOEL which gives
7 80-120% cell growth compared to solvent/vehicle control.
- 8 ▪ When contamination or technical problems are observed, four undamaged
9 wells are necessary at the minimum for each concentration.

10 2. Transformation assay

11 • Initiation assay and promotion assay

- 12 ▪ If the cells are killed and/or not confluent at the end of transformation
13 assay, the concentration is not valid and excluded from focus-counting,
14 statistical analysis and judgment. In such cases, “toxicity” is entered in the
15 data sheet.
- 16 ▪ When contamination or technical problems are observed, if, for a given
17 concentration, the number of damaged wells is ≥ 7 , the concentration is not
18 considered valid and is excluded from focus-counting, statistical analysis
19 and judgment. In such cases, “contamination”, “accident”, “technical
20 error”, etc. are entered in the data sheet.

21 • Initiation assay

- 22 ▪ Negative control: The number of wells having transformed foci should be
23 10 wells/plate or less.
- 24 ▪ Positive control: The number of wells having transformed foci should be 30
25 wells/plate or more.
- 26 ▪ Test chemical concentrations: Four valid concentrations are necessary, at a
27 minimum, to accept the transformation assay for evaluating a chemical.
28 Those concentrations should include at least one concentration near the
29 NOEL and three concentrations in the range between the NOEL and IC₉₀.

30 • Promotion assay

- 31 ▪ Negative control: The number of wells having transformed foci should be
32 20 wells/plate or less.
- 33 ▪ Positive control: The number of wells having transformed foci should be 30
34 wells/plate or more. Furthermore, the proportion of wells having
35 transformed foci should be significantly increased in the positive control
36 plate compared to the solvent control plate (chi-square test, $p < 0.05$).
- 37 ▪ Test chemical concentrations: Four valid concentrations are necessary, at
38 the minimum, to accept the transformation assay for evaluating a
39 chemical. Those concentrations should include at least one concentration
40 near the NOEL and two concentrations in the range of growth
41 enhancement when the chemical enhances cell growth (increases cell
42 density). The durations of chemical exposure to the cells are different
43 between the cell growth assay and the transformation assay. Consequently,
44 chemical toxicity is sometimes accumulated over the 10 day duration of the
45 promotion assay and plates may be lost because of chemical toxicity. In
46 such cases, four valid plates are also necessary in the concentration range
47 where cells are not killed and are confluent at the end of the
48 transformation assay. If excessive toxicity is encountered, it may be
49 necessary to repeat the assay in a lower concentration range.

50
51
52 The transformation frequency in the CTAs is known to be influenced by the batch of

1 FBS [Sakai *et al.*, 2002]. The parts of the assay acceptance criteria that are underlined
2 (Annex 6, section F.), which concern the transformation frequency of negative and
3 positive controls, were tentatively set based on the HRI historical data, that was
4 produced using FBS obtained from Moregate (Lot no. 7825120). The 96-well method
5 validation study, including the pre-validation phase, was carried out using FBS
6 obtained from GIBCO (Lot no. 1391481). Hence, three out of four of the underlined
7 parts of the acceptance criteria were modified in protocol Ver. 3 for phase I based on the
8 results obtained from MCA, TPA and negative controls in the pre-validation study:
9 these amendments were incorporated into protocol Ver. 4 for phase II.

10
11 The amendments of acceptance criteria concerning the transformation frequency of
12 negative and positive controls in the protocol Ver. 3 were as follows (Page 6-7 in Annexes
13 7 and 8):

14
15 2. Transformation assay

16 • Initiation assay

17 • Negative control: The number of wells having transformed foci should be
18 15 wells/plate or less.

19 • Positive control: The number of wells having transformed foci should be 40
20 wells/plate or more.

21 • Promotion assay

22 • Negative control: The number of wells having transformed foci should be
23 20 wells/plate or less.

24 • Positive control: The number of wells having transformed foci should be 40
25 wells/plate or more.

- 26
27 • An overlapping item of acceptance criteria in the protocol Ver. 2 was deleted in
28 the protocol Ver. 3. That is:

29
30 The item “• At least should exist one concentration near NOEL which gives
31 80-120% cell growth compared to solvent/vehicle control.” was deleted from
32 “II.F.1 Concurrent cell growth assay” in Ver. 2 (Page 6), and instead was
33 included in the item “• Concentrations: Four valid concentrations are
34 necessary at the minimum to accept the transformation assay for evaluating a
35 chemical: at least one concentration near the NOEL (80 – 120 %) and three
36 concentrations in the range between the NOEL and IC₉₀ in the concurrent cell
37 growth assay.” of “II F. 2.2) Initiation assay” in Ver. 3 (Page. 7 in Annex 6-8).

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42 **4.3 Results**

43
44 The validation study of the Bhas 42 CTA 96-well method was carried out on a total of 25
45 coded chemicals for the initiation and promotion assays. Each chemical was examined
46 by four laboratories in the pre-validation phase and validation phase I and by two
47 laboratories in validation phase II. The data for the negative and positive controls
48 obtained in individual laboratories are illustrated in Figures 17 and 18. The
49 transformation results of each test chemical in the 96-well method, including the
50 respective concurrent cell growth assay, are presented in Tables 23 to 47 and in Figures
51 19 to 43. The raw data submitted by individual laboratories for the pre-validation phase,

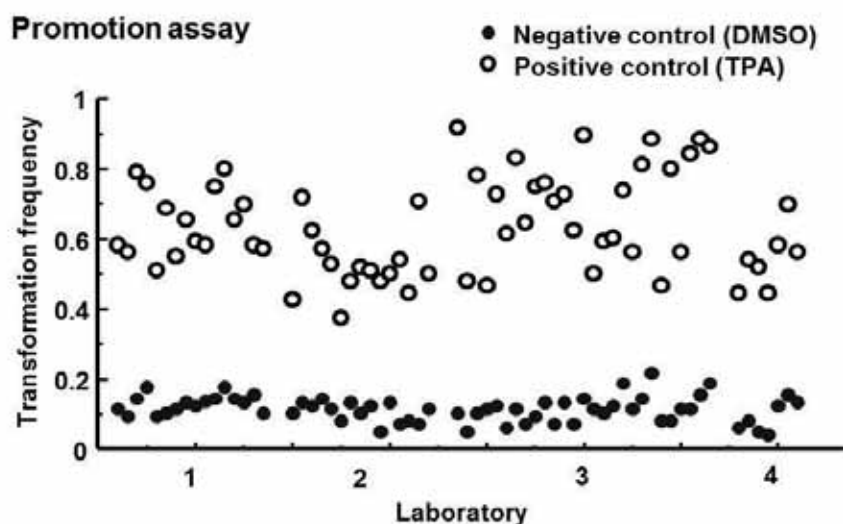
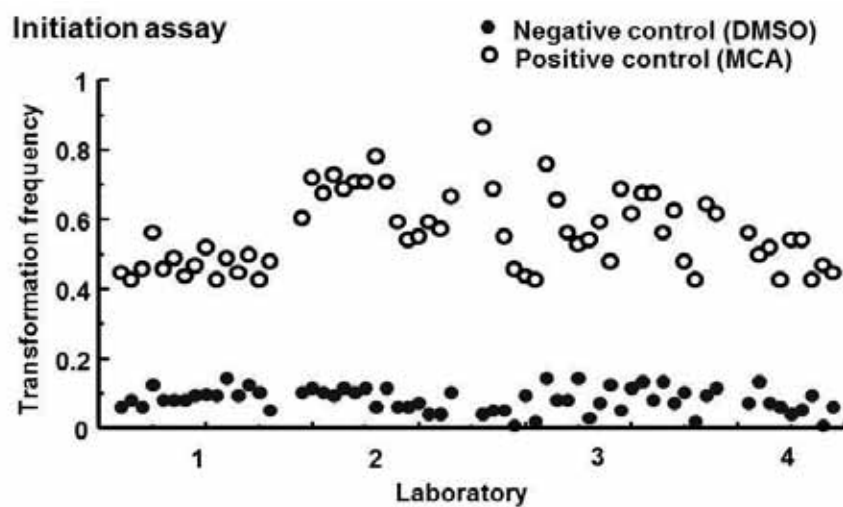
1 and for validation phase I and phase II are presented in Annex 9, 10 and 11,
2 respectively.

3 4 5 **4.3.1 Quality assurance aspects**

6
7 All of the data from all laboratories were sent at the same time to Masaya Suzuki and
8 Shoko Arai, who belong to geographically and administratively different laboratories,
9 and they assured the quality of all the data and records, by evaluating their compliance
10 with the acceptance criteria. Subsequently, M. Suzuki statistically analyzed the data. In
11 some instances, certain experiments were repeated by a particular laboratory. For such
12 cases, the data from both the first and repeated experiments are presented in the detail
13 result section for each chemical, including the reason(s) as to why the experiment was
14 repeated.

15 16 17 **4.3.2 Negative and positive controls**

18
19 The transformation frequencies for the positive and negative (DMSO) controls are
20 presented in Tables 25-47 as part of the assay results derived for each test chemicals.
21 All data of negative and positive controls satisfied assay acceptance criteria except for
22 one case: i.e., the first promotion assay of L-ascorbic acid by Lab 3 in validation phase II.
23 In Fig. 17, those control values are plotted pairwise, assay by assay, and the pairs are
24 clustered by laboratory. The transformation frequency of each positive control was
25 statistically significantly different from that of the corresponding negative control. The
26 cluster of transformation frequencies of positive controls was quite distinct from that of
27 negative controls. In Fig. 18, the transformation frequencies for the negative and
28 positive controls are averaged per individual laboratory.



1
 2 Fig. 17. Transformation frequency of the negative and positive controls in the 96-well
 3 method validation study. The negative and positive controls are paired assay by assay
 4 and the pairs are clustered laboratory by laboratory.
 5
 6

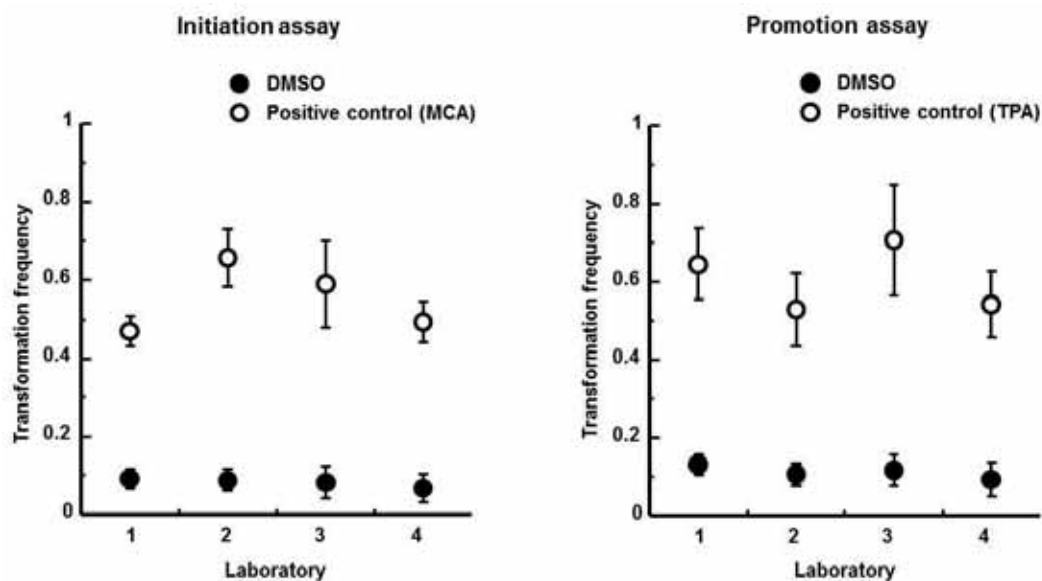


Fig. 18. Average transformation frequency of negative and positive controls per individual laboratory in the 96-well method validation study including phase I and phase II. Bars indicate standard deviation.

4.3.3 Pre-validation (transferability phase)

With the aim of assessing the transferability of the assay to the participating laboratories, MCA and TPA were examined without having been coded at the concentrations designated by the VMT.

4.3.3.1 3-Methylcholanthrene

As shown in Table 23 and Fig. 19, MCA was positive in the initiation assay in all the participating laboratories and the dose response curves were concordant between the laboratories.

Table 23.

Results of transformation assay on 3-methylcholanthrene in the pre-validation phase of the 96-well method validation study

a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	7/96	100	11/96	100	8/96	100	5/96
0.03	95	17/96	115	20/96	87	16/96	89	14/96
0.1	90	31/96 *	140	35/96 *	80	27/96 *	84	19/96 *
0.3	75	46/96 *	130	51/96 *	70	44/96 *	72	30/96 *
1	53	71/91 *	83	66/96 *	50	59/96 *	47	33/96 *

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. solvent control.

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b) Promotion assay

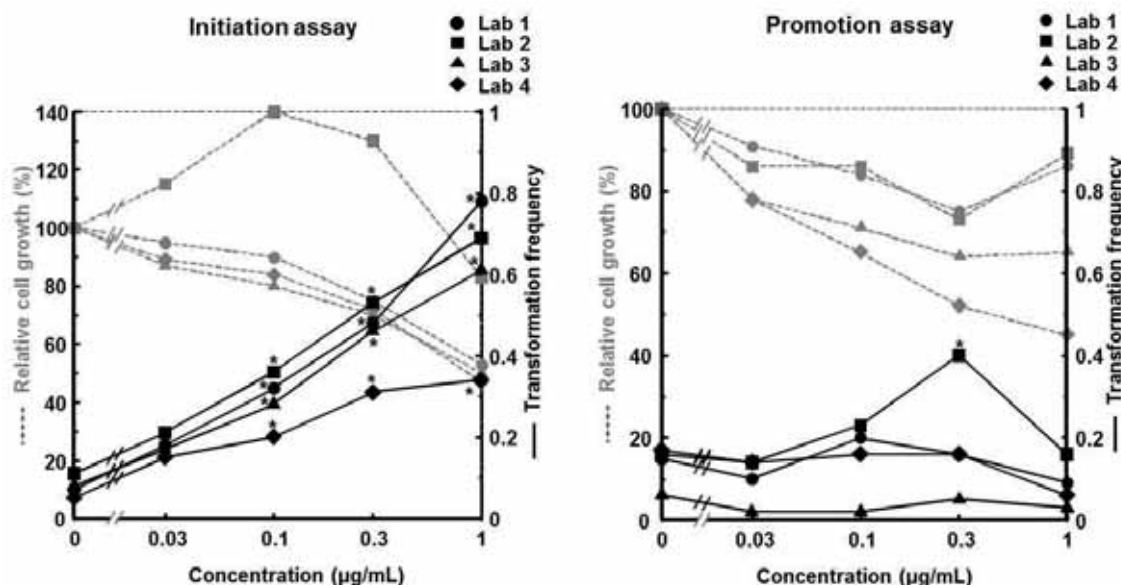
Concentration ($\mu\text{g/mL}$)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	14/94	100	15/96	100	6/96	100	16/96
0.03	91	10/96	86	13/96	78	2/96	78	13/96
0.1	84	19/96	86	22/96	71	2/96	65	15/96
0.3	75	15/96	73	38/96 *	64	5/96	52	15/96
1	86	9/96	89	15/96	65	3/96	45	6/96

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. solvent control.



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Fig. 19. Graphic view of the results of transformation assay and concurrent cell growth assay on 3-methylcholanthrene in the pre-validation phase of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

4.3.3.2 TPA

Table 24 and Fig. 20 represent the results for TPA. TPA was positive in the promotion assay in all the four laboratories and induced similar responses between laboratories.

1
2 Table 24.

3 Results of transformation assay on TPA in the pre-validation phase of the 96-well
4 method validation study

5 a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	8/96	100	10/96	100	8/96	100	7/96
0.025	95	4/96	99	4/96	91	0/96	86	2/96
0.05	97	4/96	115	4/96	94	2/96	87	0/96
0.1	100	1/96	115	5/96	97	4/96	89	3/96
0.2	111	5/96	122	6/96	107	5/96	98	2/96

13 ^a % of cell growth compared to that of solvent control.

14 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

15 ^c Solvent control.

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18 b) Promotion assay

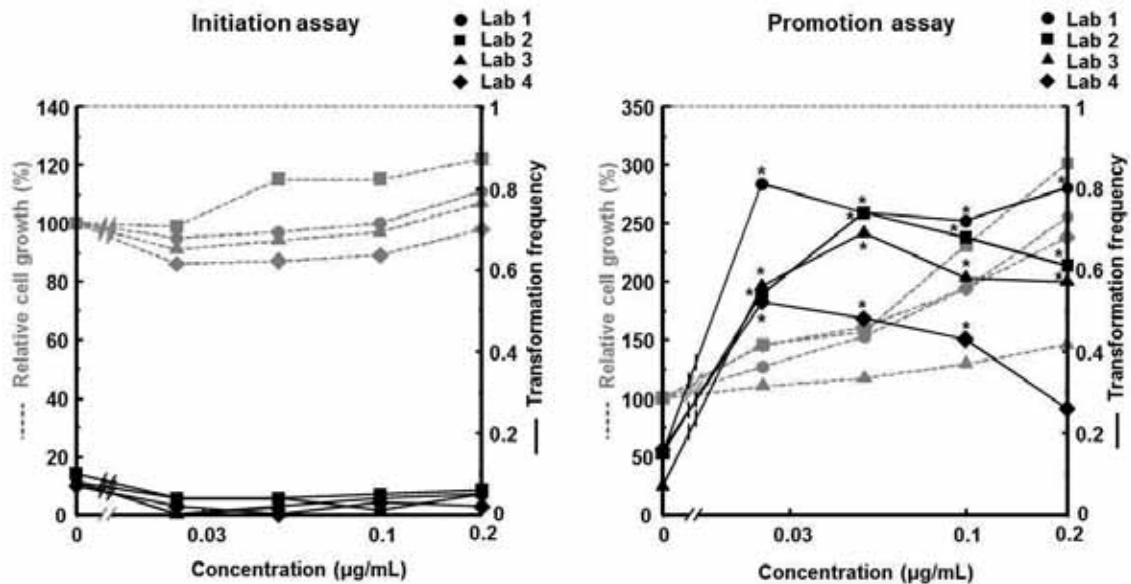
Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	14/96	100	14/96	100	7/96	100	15/96
0.025	127	78/96 *	146	52/96 *	110	54/96 *	145	50/96 *
0.05	153	71/96 *	157	71/96 *	117	66/96 *	161	46/96 *
0.1	194	69/96 *	231	65/96 *	129	56/96 *	194	41/96 *
0.2	255	77/96 *	301	59/96 *	146	55/96 *	238	25/96

26 ^a % of cell growth compared to that of solvent control.

27 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

28 ^c Solvent control.

29 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. solvent control.



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36 Fig. 20. Graphic view of the results of transformation assay and concurrent cell growth
37 assay on TPA in the pre-validation phase of the 96-well method validation study.

38 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

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Thus, all laboratories obtained comparable results from MCA and TPA and proceeded to the validation phase I study. The acceptance criteria for the transformation frequency in the negative and positive controls were amended for phase I based on the results of MCA and TPA in the pre-validation phase. The criteria applied in protocol Ver. 2 had been tentatively established based on the assays that had been carried out using a different batch of FBS prior to the 96-well validation study. Use of a new batch of FBS throughout the 96-well method validation study necessitated the revision of the acceptance criteria. These amended criteria were also employed in the validation phase II studies.

4.3.4 Validation phase I

Seven coded chemicals were tested in each laboratory.

4.3.4.1 2-Acetylaminofluorene

The results for 2-acetylaminofluorene are shown in Table 25 and Fig. 21. In the initiation assay, the chemical was negative in the Lab 1 but positive in Lab 2, Lab 3 and Lab 4. In Lab 4, there was a statistically significant increase in transformation frequency at only one concentration, 21 µg/mL, in the first run of the initiation assay. In accordance with the protocol, the assay was repeated and significant increases in transformation frequency at two serial concentrations were obtained. Hence the chemical was positive in the initiation assay in Lab 4. In the promotion assay, this chemical was positive in Lab 1, Lab 3 and Lab 4 but negative in Lab 2.

1
2 Table 25.

3 Results of transformation assay on 2-acetylaminofluorene in phase I of the 96-well method validation
4 study

5 a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4, 1st run		Lab 4, 2nd run	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	6/96	100	11/96	100	4/96	100	6/96	100	4/96
2.5	100	6/96								
5	97	10/96								
6.25			98	5/96						
8.84			102	16/96						
10	98	7/96					113	8/91	103	-/- ^d
12					104	14/96				
12.5			101	20/96						
15							90	10/93	82	6/96
16					97	17/96 *				
16.5							72	16/96	72	10/96
17.7			101	25/96 *						
18							62	15/96	45	9/96
19					75	19/96 *				
19.5							39	16/96	42	20/95 *
20	79	12/96								
21							28	25/96 *	23	18/96 *
22					53	23/96 *				
22.5							21	13/96	20	9/96
24							12	7/96	17	2/96
25			47	34/96 *	33	20/96 *				
29					20	16/96 *				
30	13	12/96					5	5/96	7	4/94
34					13	17/96 *				
35.4			14	25/96 *						
40					11	8/96				
50	6	Toxic	8	14/96	9	15/96 *				
100	4	Toxic								
MCA 1 ^e (0.1% DMSO)	26	44/96 †	48	68/96 †	49	83/96 †	56	41/96 †	47	52/96 †

37 ^a % of cell growth compared to that of solvent control.

38 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

39 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

40 ^d No data.

41 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

42 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

43 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	14/96	100	13/96	100	10/96	100	4/96
0.01							111	6/96
0.1							112	11/96
0.4							117	8/96
0.5					105	28/96 *		
0.78			108	8/96				
1					99	15/96		
1.56	90	24/96	114	16/96			119	3/96
2					103	11/96		
3					99	15/96		
3.13	84	16/96	121	18/96				
5					92	22/96		
6.25	72	43/96 *	102	19/96			104	18/96 *
9					81	29/96 *		
12.5	60	44/96 *	83	22/96			82	21/94 *
15					63	35/96 *		
25	44	16/96	57	10/96	50	22/96	53	9/96
40					47	Toxic		
50	33	0/96	37	0/96			38	0/96
100	33	Toxic					33	Toxic
TPA 0.05 ^d (0.1% DMSO)	111	76/96 †	137	46/96 †	145	88/96 †	155	43/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

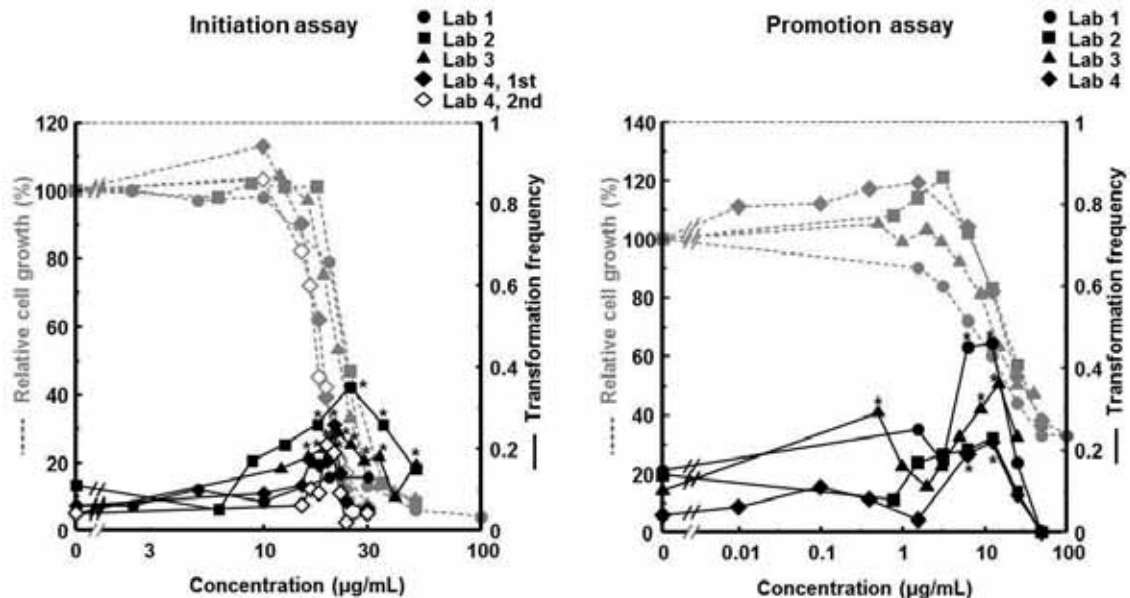


Fig. 21. Graphic view of the results of transformation assay and concurrent cell growth assay on 2-acetylaminofluorene in phase I of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

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2 **4.3.4.2 Benzo[a]pyrene**
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4 In all the laboratories, benz[a]pyrene was strongly positive in the initiation assay and
5 negative in the promotion assay as shown in Table 26 and Fig. 22.
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9 Table 26.

10 Results of transformation assay on benzo[a]pyrene in phase I of the 96-well method
11 validation study

12 a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	8/96	100	9/96	100	14/96	100	7/96
0.0016							109	11/96
0.003					93	5/96		
0.0031			92	13/96			113	8/96
0.00316	91	13/96						
0.0063			101	12/96			114	13/96
0.01	96	8/96			90	14/96		
0.0125							109	9/96
0.013			95	22/96 *				
0.025			88	29/96 *			113	18/96
0.03					80	15/96		
0.0316	67	21/96 *						
0.05			78	34/96 *	65	31/96 *	91	16/96
0.1	24	20/96 *	60	39/96 *	43	35/96 *	65	29/96 *
0.2			36	48/96 *	24	36/96 *	31	25/95 *
0.3					19	36/96 *		
0.316	9	Toxic						
0.4			24	45/96 *			12	25/96 *
0.5					12	32/96 *		
0.8			14	41/96 *				
1	7	Toxic						
3.16	3	Toxic						
MCA 1^d (0.1% DMSO)	28	41/96 †	35	70/96 †	43	73/96 †	51	54/96 †

38 ^a % of cell growth compared to that of solvent control.

39 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

40 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

41 ^d Positive control in the initiation assay: final solvent concentration in the working culture media in
42 parentheses.

43 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

44 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4		
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF	
0 ^c (0.1% DMSO)	100	9/96	100	14/96	100	11/96	100	6/96	
0.0001	95	10/96							
0.0003					93	11/96			
0.001	91	7/96			91	8/96			
0.0016			88	2/96					
0.002							107	4/96	
0.003					89	5/96			
0.0031			93	6/96					
0.0045							110	7/96	
0.0063			83	7/96					
0.009							109	4/96	
0.01	88	7/96			81	8/96			
0.013			87	2/96					
0.02					84	4/96	105	5/96	
0.025			75	5/96					
0.03					75	5/96			
0.0316	81	7/96							
0.045							91	4/96	
0.05			64	6/96	67	2/96			
0.09							79	7/96	
0.1	63	6/96	51	7/96	51	1/96			
0.2			47	11/96			59	7/96	
0.4			44	6/96					
0.45							46	Toxic	
1	45	Toxic					42	Toxic	
10	50	Toxic							
TPA	0.05 ^d (0.1% DMSO)	164	54/96 †	107	55/96 †	136	80/96 †	161	43/96 †

^a % of cell growth compared to that of solvent control.

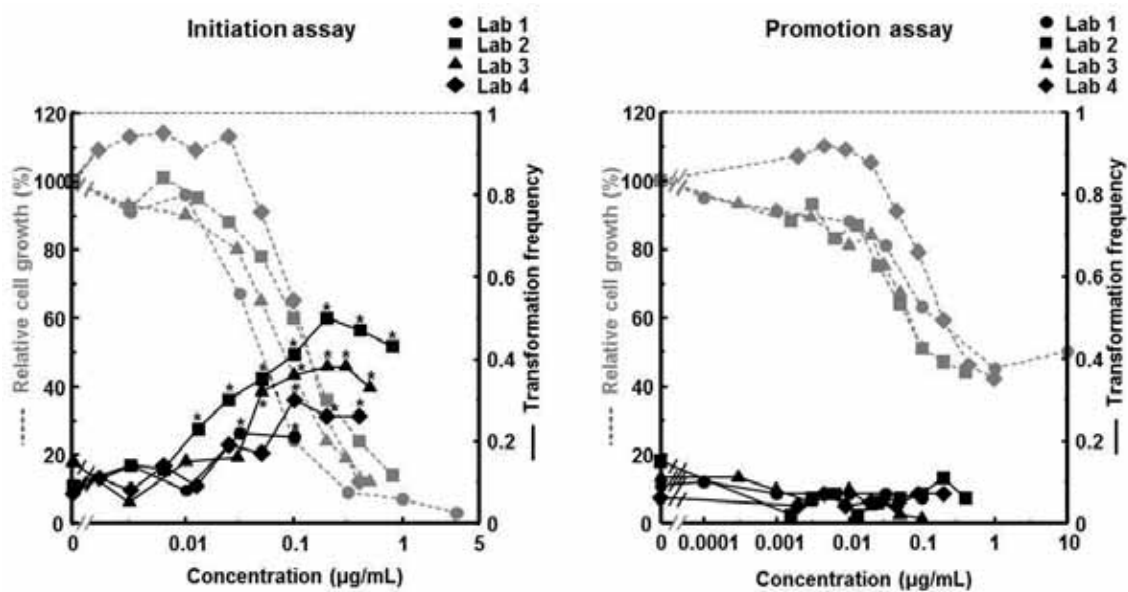
^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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Fig. 22. Graphic view of the results of transformation assay and concurrent cell growth assay on benzo[a]pyrene in phase I of the 96-well method validation study.

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* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

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4.3.4.3 3-Methylcholanthrene

The results for MCA are presented in Table 27 and Fig. 23. This chemical was clearly positive in the initiation assay in all the laboratories. In the promotion assay, it was negative in Lab 1, Lab 2 and Lab 3. In Lab 4, there was a statistically significant increase at the highest usable doses (0.2 µg/mL) before the onset of overwhelming toxicity. However, Lab 4 did not repeat the promotion assay. They considered that the initiation assay had established that the chemical was clearly positive in the Bhas 42 CTA and that it was not necessary to further differentiate the initiation vs. promotion properties of the compound.

Table 27.

Results of transformation assay on 3-methylcholanthrene in phase I of the 96-well method validation study

a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1%DMSO)			100	10/96				
0 (0.5% DMSO)	100	10/96			100	4/96	100	6/90
0.005							122	12/96
0.01	105	27/96 *						
0.025			81	20/96				
0.03					95	13/96		
0.05			80	28/96 *				
0.07							106	21/96 *
0.1	93	36/96 *	68	43/96 *	84	11/96		
0.2			61	60/96 *				
0.21							96	27/96 *
0.3					66	33/96 *		
0.316	62	44/96 *						
0.4			52	63/96 *				
0.42							76	41/96 *
0.7							65	51/96 *
0.8			46	65/96 *				
1	44	39/96 *			46	47/96 *	65	56/96 *
1.6			39	72/96 *				
2.5							53	61/96 *
3					38	31/96 *		
3.16	34	36/96 *						
3.2			41	75/96 *				
5							50	61/96 *
6.4			43	59/96 *				
10	28	Toxic			35	15/96 *	47	50/96 *
30					34	12/96		
100	25	Toxic			35	Toxic		
0 ^d (0.1%DMSO)	100	6/96			100	2/96	100	13/96
MCA 1 ^e (0.1% DMSO)	34	43/96 †	49	65/96 †	42	41/96 †	51	48/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)			100	12/96				
0 (0.5% DMSO)	100	12/96			100	8/96	100	9/95
0.00001	91	9/96						
0.0001	89	13/96						
0.001	86	4/93			96	7/96	95	13/96
0.0016			96	11/96				
0.003					93	5/96		
0.0031			90	8/96				
0.004							95	7/96
0.0063			86	12/96				
0.01	73	6/96			85	3/96		
0.013			84	10/96				
0.015							88	11/96
0.025			85	13/96				
0.03					82	1/96		
0.05			82	10/96				
0.06							86	8/96
0.1	56	10/95	70	13/96	74	2/96		
0.2			65	16/96			69	25/96 *
0.3					59	0/96		
0.4			55	12/96			66	Toxic
1	58	Toxic			61	0/96	64	Toxic
2.2							66	Toxic
3					62	0/96		
5							64	Toxic
10	58	Toxic						
100	60	Toxic						
0 ^d (0.1%DMSO)	100	11/96			100	6/96	100	8/96
TPA 0.05 ^e (0.1% DMSO)	160	56/96 †	126	60/96 †	147	59/96 †	166	52/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

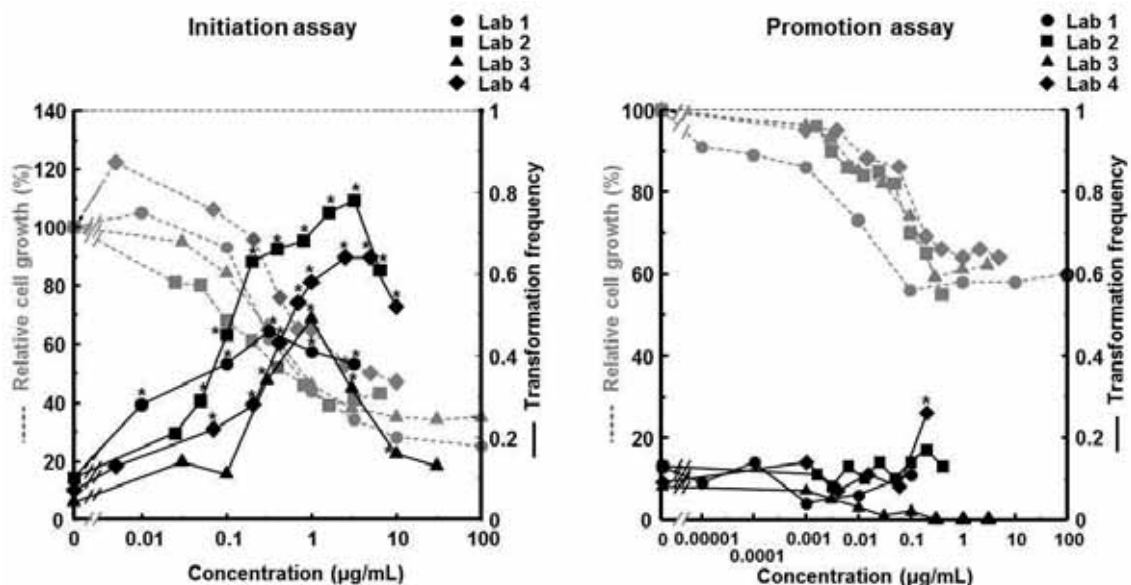


Fig. 23. Graphic view of the results of transformation assay and concurrent cell growth assay on 3-methylcholanthrene in phase I of the 96-well method validation study.
 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

4.3.4.4 σ -Toluidine

Table 28 and Fig. 24 represent the results of σ -toluidine. This hydrochloride was negative in both initiation and promotion assays.

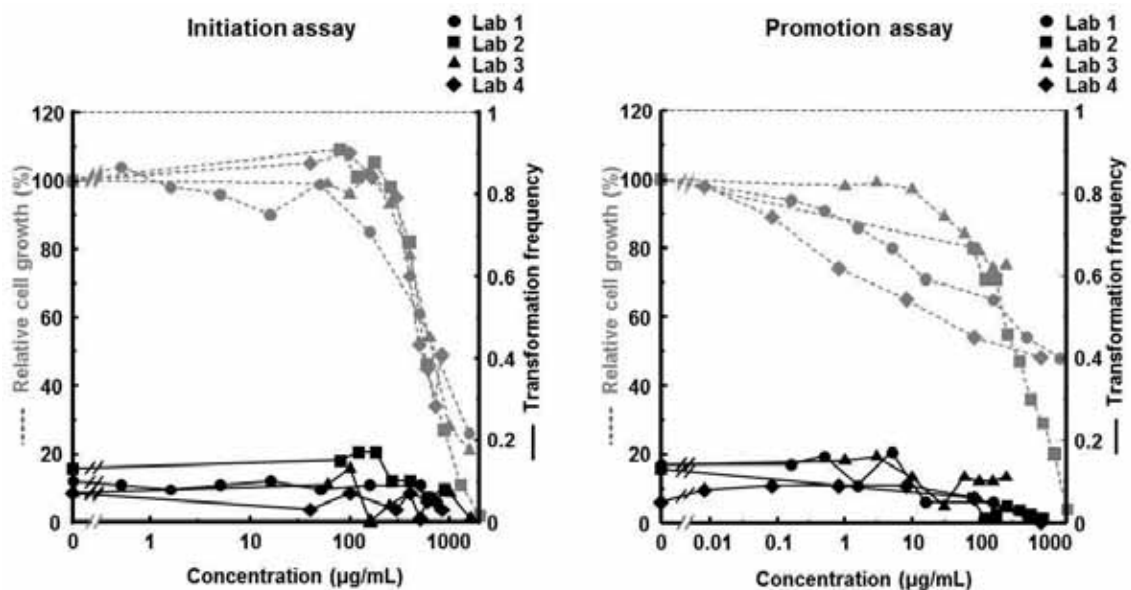


Fig. 24. Graphic view of the results of transformation assay and concurrent cell growth assay on σ -toluidine in phase I of the 96-well method validation study.

Table 28.

Results of transformation assay on *o*-toluidine in phase I of the 96-well method validation study

a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)							100	7/96
0 (0.5% DMSO)	100	10/96	100	12/96	100	7/96		
0.5	104	9/96						
1.58	98	8/96						
5	96	9/96						
15.8	90	10/96						
40							105	3/96
50	99	8/96						
60					99	9/96		
80			109	14/96				
100					96	12/96	108	7/96
120			101	16/96				
158	85	9/96						
160					102	0/96		
167							101	-/- ^d
180			105	16/96				
250					93	4/96		
260			98	10/96				
292							95	3/96
400			82	10/96	78	7/96	72	6/92
500	61	9/96					52	1/96
590			46	6/96				
600							45	5/93
630					54	5/96		
720							34	5/96
833							49	3/96
890			27	8/96				
1000					28	7/96		
1300			11	Toxic				
1580	26	Toxic						
1600					21	1/96		
2000			2	Toxic				
0 ^e (0.1% DMSO)	100	8/96	100	10/96	100	5/96		
MCA 1 ^f (0.1% DMSO)	30	44/96 †	57	68/96 †	38	66/96 †	27	50/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d No data.

^e Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^f Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs corresponding solvent control.

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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)							100	5/96
0 (0.5% DMSO)	100	13/96	100	12/96	100	13/96		
0.00000833							112	6/96
0.0000833							115	8/96
0.000833							107	7/96
0.00833							98	8/96
0.0833							89	9/96
0.158	94	13/96						
0.5	91	15/96						
0.833							74	9/96
1					98	14/96		
1.58	86	9/96						
3					99	15/96		
5	80	16/96						
8.33							65	9/96
10					97	11/96		
15.8	71	5/96						
30					89	4/96		
60					84	11/96		
80			80	6/96				
83.3							54	6/96
100					79	10/96		
120			71	1/96				
158	65	5/96						
160					74	10/96		
180			71	2/96				
250					75	11/96		
260			55	4/96				
400			47	3/96				
500	54	2/96						
590			36	2/96				
833							48	0/96
890			29	1/96				
1300			20	Toxic				
1580	48	Toxic						
2000			4	Toxic				
0 ^d (0.1% DMSO)	100	9/96	100	8/96	100	5/96		
TPA 0.05 ^e (0.1% DMSO)	110	49/96 †	137	36/96 †	171	46/96 †	89	50/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs corresponding solvent control.

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2 **4.3.4.5 TPA**
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4 The results of TPA are exhibited in Table 29 and Fig. 25. All four laboratories
5 demonstrated TPA to be clearly positive in the promotion assay. In the initiation assay,
6 TPA was negative in the concentration range pre-assigned by the VMT ($\leq 1 \mu\text{g/mL}$),
7 where cytotoxicity was not induced in the concurrent cell growth assay. To fulfill the
8 assay acceptance criteria of the initiation assay, the chemical needed to be examined up
9 to the concentrations that show cytotoxicity. However, it was considered that the
10 promotion assay had established that TPA was clearly positive in the Bhas 42 CTA and
11 that it was not necessary to further differentiate the initiation vs. promotion properties
12 of the compound for the purpose of this exercise. The results do, however, indicate that
13 for TPA, initiating activity was not detected in the concentration range at which potent
14 promoting activity was detected.
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17 Table 29.

18 Results of transformation assay on TPA in phase I of the 96-well method validation
19 study

20 a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	12/96	100	11/96	100	5/96	100	6/96
0.0001					95	3/96	98	3/96
0.0003					97	5/96		
0.000977	90	4/96						
0.001					92	1/96	89	4/96
0.003					86	2/96		
0.0032							87	2/96
0.00391	92	0/96						
0.0078			78	1/96				
0.01					89	2/96	91	1/96
0.0156	98	4/96						
0.016			85	1/96				
0.03					94	1/96		
0.031			84	1/96				
0.032							96	1/96
0.0625	103	0/96						
0.063			87	1/96				
0.1					104	1/96	104	4/96
0.13			94	4/96				
0.25	119	6/96	107	7/96				
0.3					115	4/96		
0.32							116	3/96
0.5			132	5/96				
0.64							130	-/ - ^d
1	134	12/96	147	5/96	139	9/96	131	-/ -
MCA 1 ^e (0.1% DMSO)	47	54/96 †	50	66/96 †	49	53/96 †	41	43/96 †

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49 ^a % of cell growth compared to that of solvent control.

50 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

51 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

52 ^d No data.

53 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
54 parentheses.

55 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	17/96	100	11/95	100	10/96	100	13/96
0.00001							112	14/96
0.0001					99	15/96	117	-/ - ^d
0.00014			109	10/95				
0.0003					101	10/96		
0.00041			110	12/96				
0.000977	107	30/96						
0.001					107	11/96	117	17/91
0.0012			114	12/95				
0.003					111	23/96		
0.0037			112	19/96				
0.00391	109	47/96 *						
0.01					120	43/96 *	132	47/96 *
0.011			122	31/95 *				
0.0156	118	69/96 *						
0.03					136	68/96 *		
0.032							142	59/96 *
0.033			128	34/96 *				
0.0625	134	74/96 *						
0.1			151	54/94 *	158	66/96 *	174	47/96 *
0.25	164	68/96 *					237	45/96 *
0.3			166	46/96 *	190	50/96 *		
0.5							254	-/ -
1	165	90/96 *			200	81/96 *	254	80/96 *
TPA 0.05 ^e (0.1% DMSO)	129	73/96 †	111	51/96 †	145	75/96 †	143	54/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

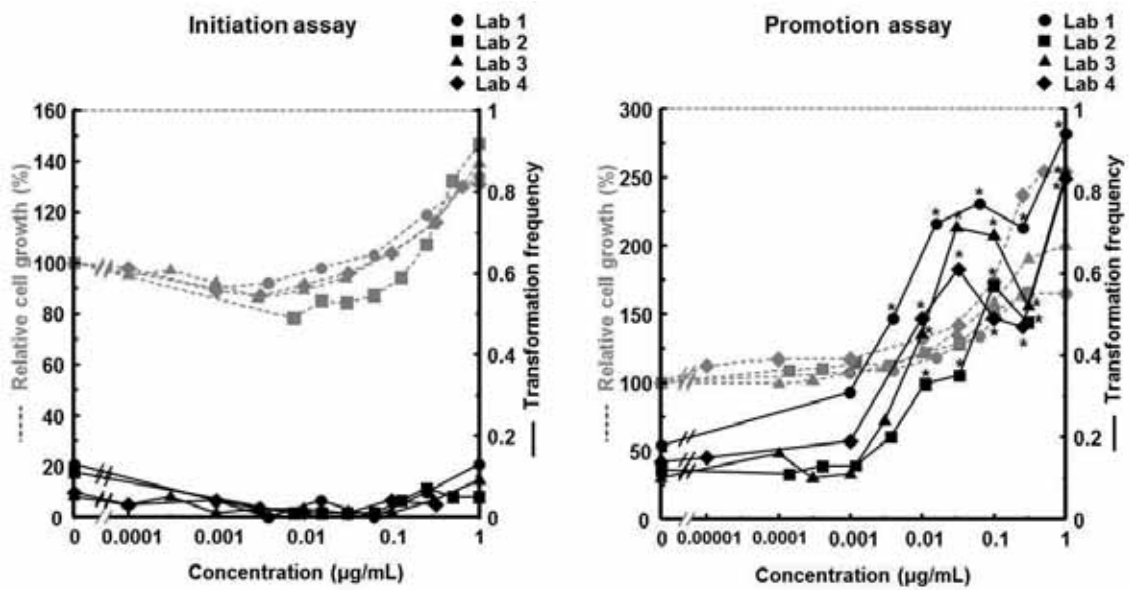
^d No data.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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Fig. 25. Graphic view of the results of transformation assay and concurrent cell growth assay on TPA in phase I of the 96-well method validation study.
* p<0.05; one-sided chi-square test with Bonferroni's adjustment.

4.3.4.6 Anthracene

The results obtained with anthracene are shown in Table 30 and Fig. 26. Anthracene was negative both in the initiation assay and in the promotion assay. There was a large difference in the maximum concentrations applied between the laboratories. Lab 1, Lab 2 and Lab 3 treated cells with the chemical up to the highest concentration soluble in DMSO, but Lab 4 applied the chemical in suspension to achieve inhibition in the cell growth assay. Despite the differences in treatment concentrations, anthracene failed to induce transformation, either as an initiator or as a promoter, in the Bhas 42 CTA.

Table 30.

Results of transformation assay on anthracene in phase I of the 96-well method validation study

a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.5% DMSO)	100	12/96	100	12/96	100	6/96	100	8/96
3					95	10/96		
4.7			101	18/96				
5					99	14/96		
7					99	1/96		
9.4			106	17/96				
10					97	6/96		
15.6	107	12/93						
19			102	18/96				
30					94	7/96		
31.3	101	16/96						
38			104	14/96				
50					104	6/96		
62.5	105	18/96						
70					97	9/96		
75			104	13/95				
100					101	9/96	118	5/96
125	132	15/96						
150			108	10/96				
250	112	14/96						
300			110	17/94				
600			111	15/95				
1000							117	12/96
1500							112	-/- ^d
2000							105	10/96
2500							84	7/96
3000							56	10/96
3500							33	1/96
4000							58	8/96
4500							17	6/96
0 ^e (0.1% DMSO)	100	8/96	100	10/96	100	1/96	100	5/93
MCA 1 ^f (0.1% DMSO)	33	47/96 †	58	58/96 †	40	44/96 †	42	52/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d No data.

^e Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^f Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF
0 ^c (0.5% DMSO)	100	12/96	100	5/96	100	12/96	100	18/96
1					103	7/96		
1.56			103	12/96				
3					103	11/96		
3.13			98	9/96				
5					103	12/96		
6.25			100	15/96				
7					102	13/96		
10					102	8/96		
12.5			100	10/96				
15.6	103	13/96						
25			101	15/96				
30					99	9/96		
31.3	106	10/96						
50			103	10/96	99	10/96		
62.5	96	14/96						
70					93	14/96		
100			93	8/96	94	7/96	116	18/96
125	103	15/96						
250	101	20/96					114	19/96
500							119	28/96
700							106	14/96
875							107	10/96
1000							102	26/96
2000							78	21/96
3500							54	Toxic
5000							47	Toxic
0 ^d (0.1% DMSO)	100	10/96	100	10/96	100	11/96	100	12/96
TPA 0.05 ^e (0.1% DMSO)	158	66/96 †	128	41/96 †	120	45/96 †	121	56/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

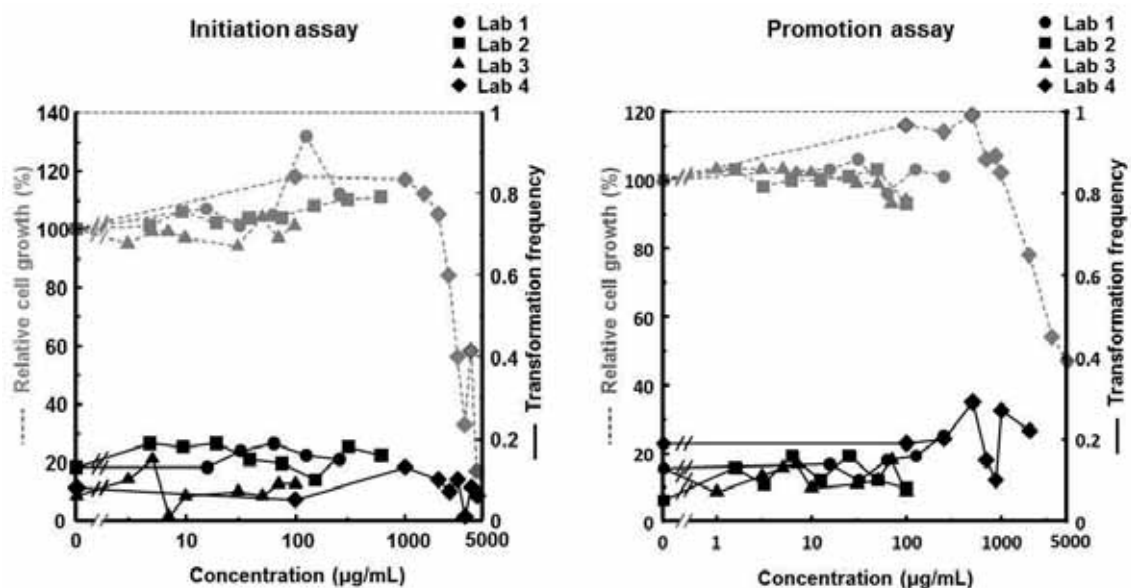
^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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5 Fig. 26. Graphic view of the results of transformation assay and concurrent cell growth
6 assay on anthracene in phase I of the 96-well method validation study.

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10 4.3.4.7 Phenanthrene

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Table 31 and Fig. 27 present the results of phenanthrene. In all four laboratories, the chemical was negative in the initiation assay. It did induce a statistically significant increase in transformation frequency at a single dose in the first run of the assay conducted by Lab 4. In accordance with the protocol, Lab 4 then repeated the initiation assay at concentrations that bracketed the positive dose, but there was no significant increase of transformation frequency at any dose in the second run. In the promotion assay, phenanthrene was negative in Lab 1, Lab 3 and Lab 4, but positive in Lab 2. Lab 1 repeated the promotion assay to verify that the chemical was negative, since it induced a statistically significant increase in the transformation frequency at one dose in the first run of promotion assay. In Lab 3 and Lab 4, the chemical did not induce a significant increase in the transformation frequency at any dose. In Lab 2, phenanthrene induced a statistically significant increase in transformation frequency at two successive concentrations.

Table 31.

Results of transformation assay on phenanthrene in phase I of the 96-well method validation study

a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 2		Lab 3		Lab 4, 1st run		Lab 4, 2nd run	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	8/96			100	9/96				
0 (0.5% DMSO)			100	12/96			100	4/96	100	7/96
0.5									106	-/ ^d
1							107	14/96	104	5/96
4									112	7/96
6	109	6/96							106	8/96
8							111	17/96 *	113	5/96
10	120	5/96							111	5/96
12									108	4/96
12.5			102	14/96						
13					103	10/96				
15									111	6/96
16							117	8/96		
17.7			102	10/96						
20	129	10/96								
24							123	7/96		
25			120	8/96	114	12/96				
30									118	5/96
35.4			77	8/96						
38					110	9/96				
40	124	9/96								
48							84	12/96		
50			34	6/96	94	3/96				
60	91	7/96								
63					76	17/96				
70.7			1	9/96						
75					59	15/96				
80							36	5/96		
88					49	5/96				
100	59	5/96	-1	7/96	45	5/96				
130					50	8/96				
160							20	10/96		
200	68	11/96								
240							17	2/96		
500							17	3/96		
0 ^e (0.1%DMSO)			100	11/96			100	9/94	100	1/96
MCA 1 ^f (0.1% DMSO)	40	42/96 †	55	69/96 †	34	42/96 †	31	41/96 †	48	45/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d No data.

^e Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^f Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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b) Promotion assay

Concentration (µg/mL)	Lab 1, 1st run		Lab 1, 2nd run		Lab 2		Lab 3		Lab 4	
	CG ^a	TF ^b	CG	TF	CG	TF	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	11/96	100	13/96			100	12/96		
0 (0.5% DMSO)					100	10/96			100	13/96
0.0001									110	12/96
0.001									118	10/96
0.01									123	6/96
0.1									108	11/96
1	102	15/96	93	10/96					108	11/96
1.56					98	10/96				
2	108	16/96								
2.5							105	10/96		
3.13					94	15/95				
5	111	16/96					102	14/96		
6.25					90	24/96 *				
10	115	18/96	87	13/96			99	3/96	101	10/96
12.5					79	30/96 *				
15							97	9/96		
20	112	23/96	77	14/96						
22							83	13/96		
25					62	13/96				
30			65	8/96			68	9/96		
40			57	6/96			57	9/96		
50	86	26/96 *	54	8/96	48	0/96				
60			55	6/96			49	0/96		
90							46	0/96		
100	86	Toxic			32	0/96			67	6/96
1000									63	Toxic
5000									62	Toxic
0 ^d (0.1% DMSO)					100	13/96			100	15/96
TPA 0.05 ^e (0.1% DMSO)	185	53/96 †	134	63/96 †	112	69/96 †	144	70/96 †	114	67/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

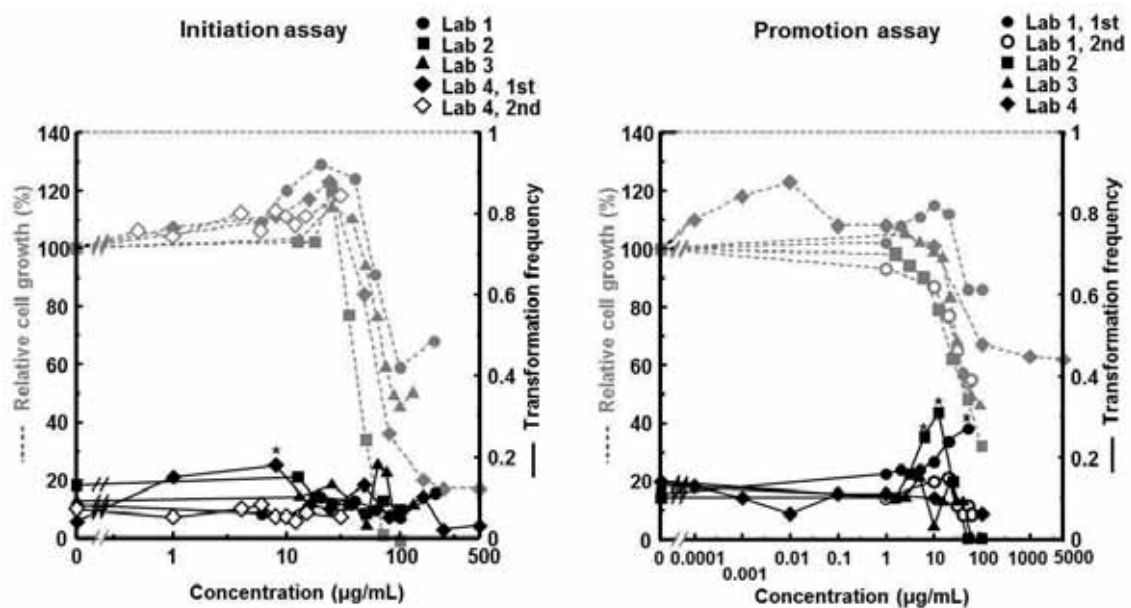
^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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Fig. 27. Graphic view of the results of transformation assay and concurrent cell growth assay on phenanthrene in phase I of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

4.3.4.8 Conclusion of phase I for the phase II study plan

As stated above, the phase I study demonstrated good reproducibility between laboratories. When we compare the results in the initiation assay or in the promotion assay between laboratories, positive and negative calls were concordant except for one call in the initiation assays and one call in the promotion assays for 2-acetylaminofluorene and one call in the promotion assays for phenanthrene (see summary data presented in Table 49). That is, the discrepancy in calls were 3 out of 55 assays (including one equivocal call in the promotion assays for MCA, which was excluded from this calculation to avoid unintended ambiguity in the absence of a confirmatory/repeat test [see 4.3.4.3]). Considering the high degree of inter-laboratory reproducibility associated with the 96 well method, it was decided that for phase II of the 96-well method the validation study for each chemical would be tested by only two laboratories.

4.3.5 Validation phase II

For the validation phase II study, three laboratories were involved; Lab 4 did not take part in this phase of study. A total of sixteen chemicals were examined; each chemical was tested by two of the three participating laboratories.

4.3.5.1 Benzo[*a*]pyrene

As shown in Table 32 and Fig. 28, benzo[*a*]pyrene was positive in the initiation assay and negative in the promotion assay in both laboratories.

Table 32

Results of transformation assay on benzo[*a*]pyrene in phase II of the 96-well method validation study

a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	9/93	100	8/96
0.001			97	8/96
0.003			99	14/96
0.01			97	14/96
0.0156	96	19/95		
0.03			89	16/96
0.0313	76	33/94 *		
0.045	61	30/95 *		
0.05			77	38/96 *
0.0625	45	28/95 *		
0.1			62	40/96 *
0.125	29	21/95		
0.15			44	26/96 *
0.25	13	21/94		
0.3			25	31/96 *
0.5	9	Toxic		
1			11	20/96
MCA 1 ^d (0.1% DMSO)	45	50/96 †	55	54/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	13/95	100	9/96
0.0001			101	12/96
0.0003			101	4/96
0.000316	91	8/94		
0.001	91	9/96	100	10/96
0.003			98	6/96
0.00316	88	9/95		
0.01	79	2/95	90	0/96
0.03			81	3/96
0.0316	69	5/93		
0.1	55	Toxic	68	2/96
0.3			55	2/96
0.316	46	Toxic		
1	45	Toxic	54	Toxic
TPA 0.05 ^d (0.1% DMSO)	137	56/96 †	132	72/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

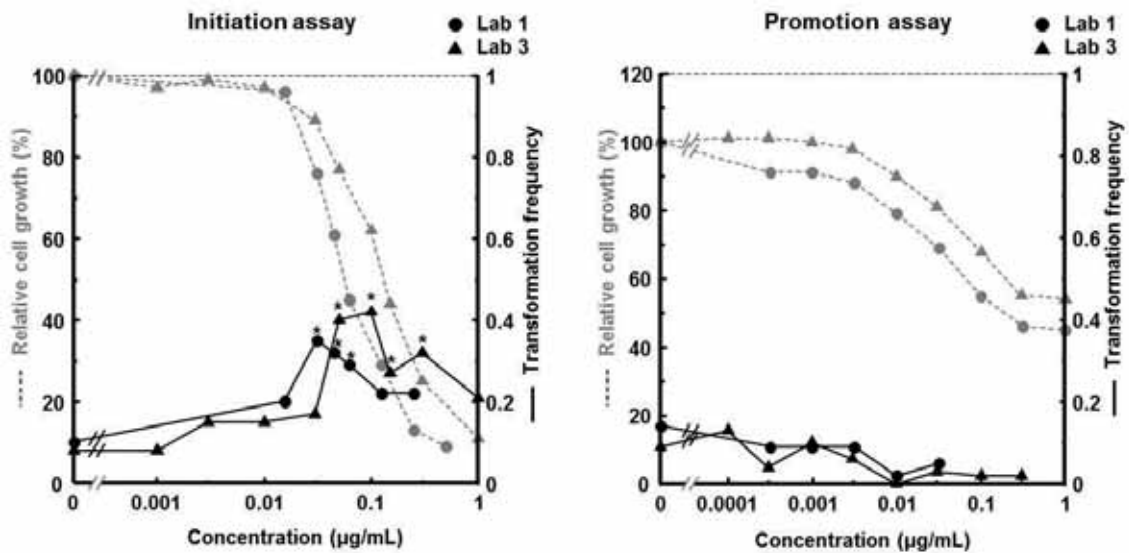


Fig. 28. Graphic view of the results of transformation assay and concurrent cell growth assay on benzo[a]pyrene in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

1 **4.3.5.2 Cadmium chloride**

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3 The results of cadmium chloride are represented in Table 33 and Fig. 29. Cadmium
4 chloride was negative in the initiation assay but positive in the promotion assay in both
5 laboratories. All but one dose of cadmium chloride used by Lab 3 were found to be toxic
6 in the first run of promotion assay. The second run of the assay performed by Lab 3 at
7 lower concentrations yielded positive results.

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10 Table 33
11 Results of transformation assay on cadmium chloride in phase II of the 96-well method
12 validation study

13 a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	9/96	100	5/96
0.335	86	8/96		
0.402	92	9/96		
0.482	90	5/96		
0.579	91	4/96		
0.6			91	7/96
0.694	85	6/96		
0.8			78	8/96
0.833	74	10/95		
1	44	5/94	43	6/96
1.2			10	3/96
1.3			4	2/94
1.4			1	Toxic
1.5			0	Toxic
1.6			0	Toxic
0 ^d (0.1% DMSO)	100	14/96	100	7/96
MCA 1 ^e (0.1% DMSO)	31	47/96 †	42	57/96 †

33 ^a % of cell growth compared to that of solvent control.

34 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

35 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

36 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
37 media in parentheses.

38 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
39 parentheses.

40 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3, 1st run		Lab 3, 2nd run	
	CG ^a	TF ^b	CG	TF	CG	TF
0 ^c (5% Water)	100	18/96	100	10/96	100	14/96
0.0625	97	17/96				
0.1					98	17/96
0.125	98	19/96				
0.15					99	13/96
0.2			86	19/96	101	24/96
0.25	96	37/96 *			106	26/96
0.3					106	43/96 *
0.35					105	53/96 *
0.4			106	Toxic	105	38/95 *
0.45					108	47/96 *
0.5	104	44/91 *				
0.6			115	Toxic		
0.75	110	Toxic				
0.8			114	Toxic		
1	103	Toxic	111	Toxic		
1.2			102	Toxic		
1.5			77	Toxic		
1.8			38	Toxic		
2	11	Toxic				
0 ^d (0.1% DMSO)	100	17/96	100	13/96	100	7/96
TPA 0.05 ^e (0.1% DMSO)	177	77/96 †	161	70/96 †	138	60/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

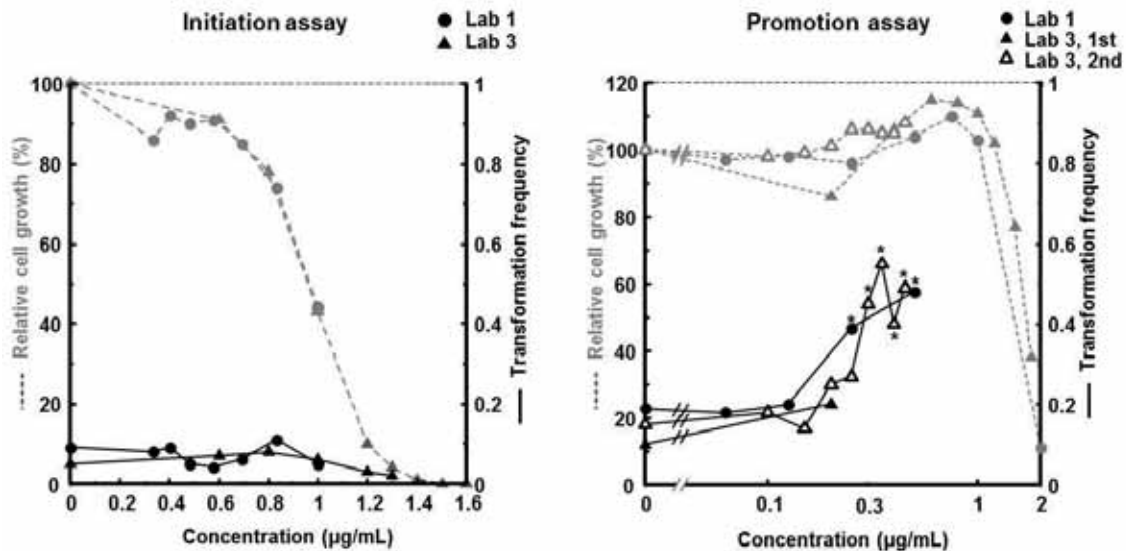


Fig. 29. Graphic view of the results of transformation assay and concurrent cell growth assay on cadmium chloride in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

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3 **4.3.5.3 Dibenz[*a,h*]anthracene**
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5 Both laboratories reported dibenz[*a,h*]anthracene to be clearly positive in the initiation
6 assay and negative in the promotion assay, as shown in Table 34 and Fig. 30.
7
8

9 Table 34.

10 Results of transformation assay on dibenz[*a,h*]anthracene in phase II of the 96-well
11 method validation study

12 **a) Initiation assay**

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	6/96	100	14/96
0.01			87	25/96
0.02	96	29/96 *		
0.03			76	34/96 *
0.04	84	42/96 *		
0.08	64	48/96 *		
0.1			59	53/96 *
0.15			52	50/96 *
0.16	50	56/96 *		
0.3			45	48/96 *
0.31	35	60/96 *		
0.5			39	51/96 *
0.63	25	63/96 *		
1			34	33/96 *
1.25	28	38/96 *		
2.5	36	34/96 *		
3			35	Toxic
5	38	29/96 *		
10			43	Toxic
MCA 1^d (0.1% DMSO)	49	75/96 †	60	51/96 †

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35 ^a % of cell growth compared to that of solvent control.

36 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

37 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

38 ^d Positive control in the initiation assay: final solvent concentration in the working culture media in
39 parentheses.

40 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

41 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	10/96	100	13/96
0.001			87	6/96
0.003			81	7/96
0.01			79	5/96
0.02	66	7/96		
0.03			73	1/87
0.04	67	5/96		
0.08	63	5/96		
0.1			67	6/96
0.16	60	3/96		
0.3			62	6/96
0.31	58	8/96		
0.63	56	5/96		
1			65	3/96
1.25	61	2/96		
2.5	60	3/96		
3			66	0/96
5	63	2/96		
10			68	0/96
TPA 0.05 ^d (0.1% DMSO)	135	50/96 †	127	73/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

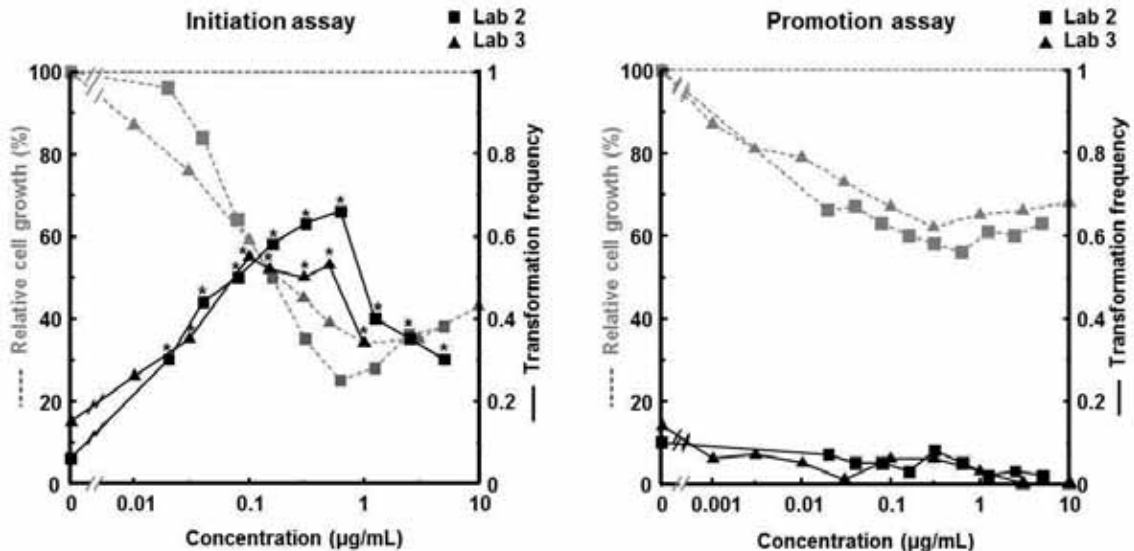


Fig. 30. Graphic view of the results of transformation assay and concurrent cell growth assay on dibenz[*a,h*]anthracene in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

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2 **4.3.5.4 Lithocholic acid**
3

4 Table 35 and Fig. 31 present the results of lithocholic acid. The chemical was negative in
5 the initiation assay but clearly positive in the promotion assay in the two laboratories.
6

7
8 Table 35
9 Results of transformation assay on lithocholic acid in phase II of the 96-well method
10 validation study.

11 a) Initiation assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	6/96	100	11/96
2.5	107	10/96		
3.5	111	6/96		
5	113	6/96		
7	113	4/96		
10	119	6/96	104	10/96
14	114	6/96		
15			110	5/96
20	121	1/96	114	1/96
22			112	3/96
24			89	4/96
26			45	4/96
28	15	2/96	19	2/95
30			2	1/91
32			1	2/89
40	2	Toxic		
MCA 1 ^d (0.1% DMSO)	63	52/96 †	55	59/96 †

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31 ^a % of cell growth compared to that of solvent control.

32 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

33 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

34 ^d Positive control in the initiation assay: final solvent concentration in the working culture media in
35 parentheses.

36 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	13/96	100	10/96
0.3125			106	21/96
0.625			102	19/96
1.25			84	15/96
2.5	111	12/96	96	13/96
3.5	107	16/96		
5	105	13/96	90	24/96 *
7	104	31/96 *		
10	104	33/96 *	74	40/96 *
14	93	42/96 *		
20	85	46/96 *	68	46/96 *
28	90	0/96		
30			70	0/96
40	14	Toxic	7	Toxic
TPA 0.05 ^d (0.1% DMSO)	155	48/96 †	125	57/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

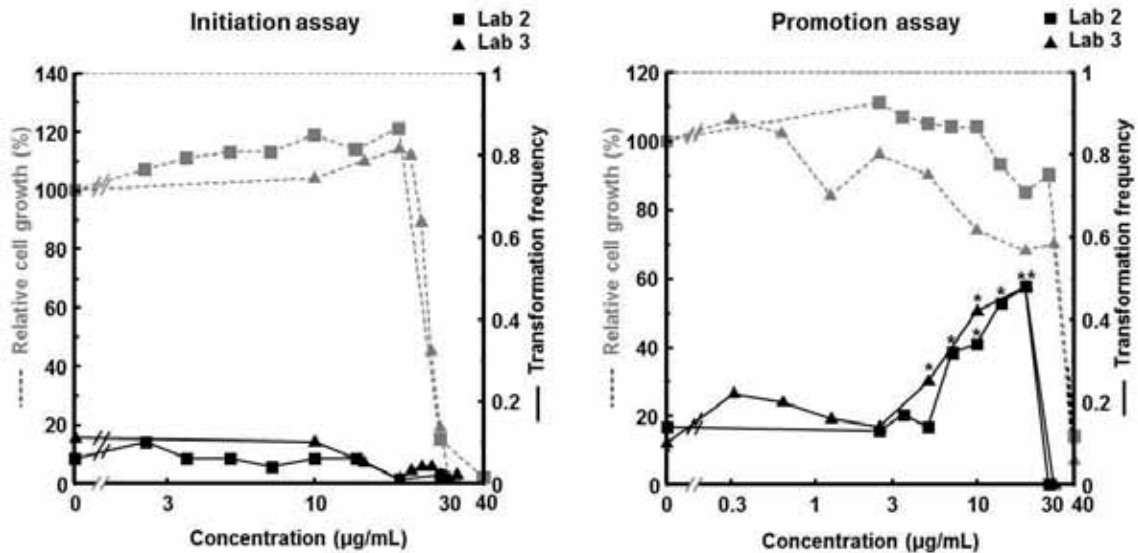


Fig. 31. Graphic view of the results of transformation assay and concurrent cell growth assay on lithocholic acid in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

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2
3 **4.3.5.5 Methapyrilene hydrochloride**
4

5 Both two laboratories demonstrated methapyrilene hydrochloride to be negative in the
6 initiation assay but clearly positive in the promotion assay, as shown in Table 36 and
7 Fig. 32.
8
9

10 Table 36
11 Results of transformation assay on methapyrilene hydrochloride in phase II of the
12 96-well method validation study.

13 a) Initiation assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	12/96	100	9/96
150	113	9/96		
180	103	5/96	110	6/96
210	80	10/96	84	6/96
240	38	7/96	38	3/96
250			44	4/96
260			37	3/96
270	38	4/96		
300	15	4/96	7	3/96
320			3	0/95
330	6	4/96		
360	5	2/96		
400	2	3/96		
500			0	Toxic
0 ^d (0.1% DMSO)	100	11/96	100	12/96
MCA 1 ^e (0.1% DMSO)	64	68/96 †	53	46/96 †

32 ^a % of cell growth compared to that of solvent control.

33 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

34 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

35 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
36 media in parentheses.

37 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
38 parentheses.

39 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	11/96	100	11/96
1			100	12/96
5			111	27/96 *
5.9	113	21/96		
8.8	108	20/96		
10			115	25/96 *
13	107	27/96 *		
20	125	30/96 *		
25			125	39/96 *
30	121	26/96 *		
44	127	44/96 *		
50			162	83/96 *
67	138	55/96 *		
100	144	46/96 *	155	78/96 *
150	126	17/96	140	24/96
280			103	Toxic
0 ^d (0.1% DMSO)	100	12/96	100	14/96
TPA 0.05 ^e (0.1% DMSO)	139	49/96 †	126	86/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

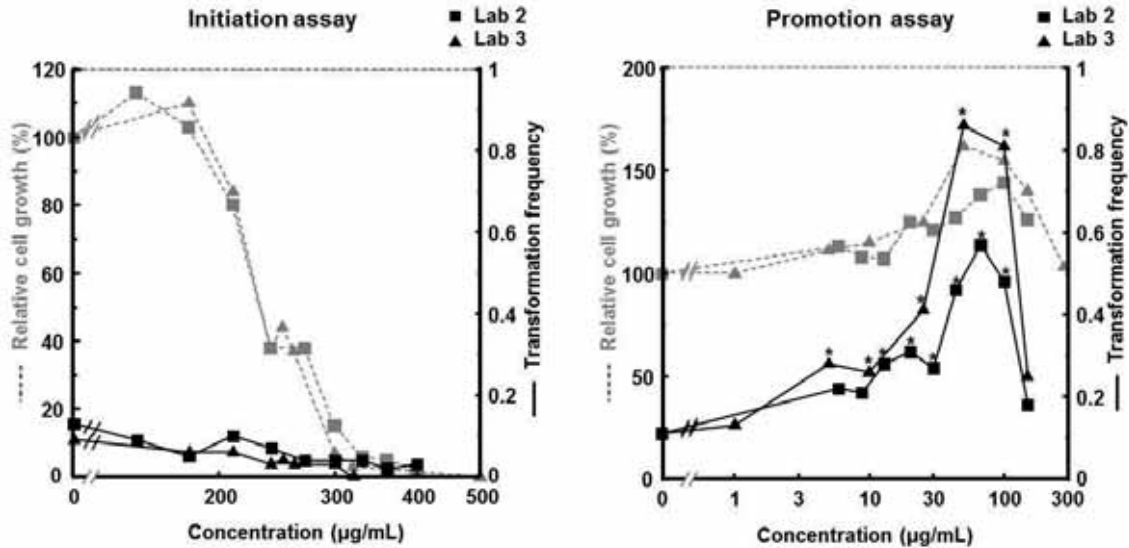


Fig. 32. Graphic view of the results of transformation assay and concurrent cell growth assay on methapyrilene hydrochloride in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

4.3.5.6 Mezerein

The results of mezerein are represented in Table 37 and Fig. 33. Mezerein proved to be positive in both the initiation assay and the promotion assay. In the promotion assay, mezerein was strongly positive in both laboratories, inducing the highest transformation response among all the chemicals tested in this validation study. The chemical caused statistically significant increase of transformation frequency at a concentration as low as 0.000156 µg/mL or 0.0003 g/mL, and produced transformed foci in almost all the wells at the highest 4 doses in either laboratory. In the initiation assay, mezerein was also positive, inducing statistically significant increases in transformation frequency in both laboratories at the highest two or four concentrations, respectively, although those concentrations were 100 times higher than the minimum dose at which promoting activity of mezerein was detected. It is noteworthy that despite the positive transformation response to mezerein in the initiation assay, the chemical was non-cytotoxic up to the maximum concentration used, which had been pre-assigned by the VMT (0.1 µg/mL).

Table 37

Results of transformation assay on mezerein in phase II of the 96-well method validation study.

a) Initiation assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	6/96	100	5/96
0.00001			104	7/96
0.00003			106	7/96
0.0001			98	6/96
0.0003			99	5/96
0.00039	96	5/96		
0.00078	97	6/96		
0.001			104	4/96
0.00156	124	4/96		
0.003			100	3/96
0.00313	110	3/96		
0.00625	144	6/96		
0.01			120	2/96
0.0125	166	18/96 *		
0.025	138	33/96 *		
0.03			94	20/96 *
0.05	137	70/96 *		
0.1	141	72/96 *	87	41/96 *
MCA 1^d (0.1% DMSO)	37	57/96 †	54	66/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

b) Promotion assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	5/96	100	11/96
0.00001			100	17/96
0.00003			101	26/96
0.000078	116	13/96		
0.0001			104	15/96
0.000156	126	18/96 *		
0.0003			111	37/96 *
0.000313	161	25/96 *		
0.000625	190	61/96 *		
0.001			139	57/96 *
0.00125	200	82/96 *		
0.0025	173	92/96 *		
0.003			180	96/96 *
0.005	158	91/96 *		
0.01	165	92/96 *	147	94/96 *
0.02	190	93/96 *		
0.03			166	94/96 *
0.1			174	96/96 *
TPA 0.05 ^d (0.1% DMSO)	175	46/96 †	135	48/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

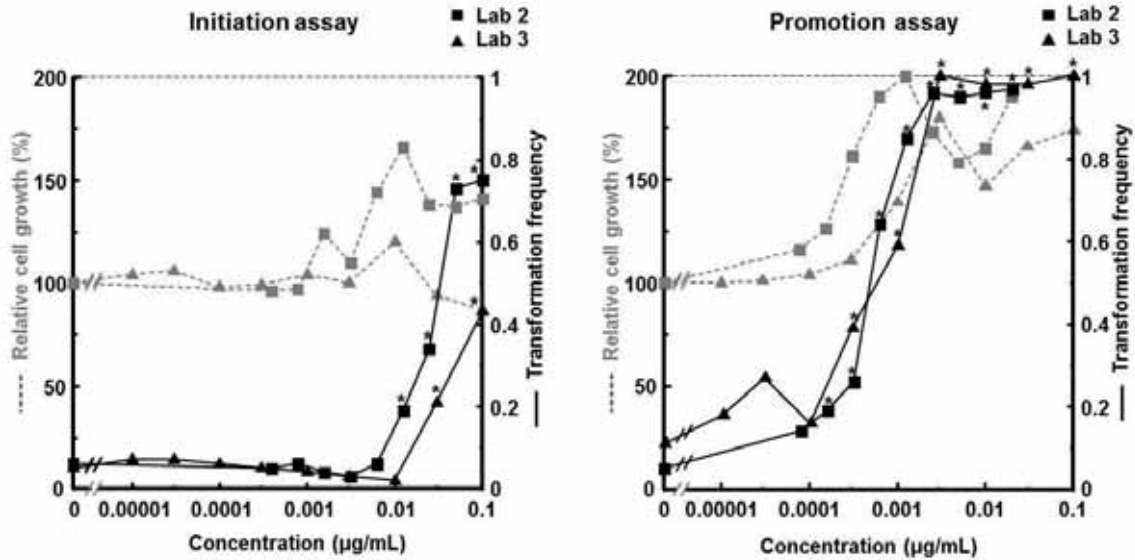


Fig. 33. Graphic view of the results of transformation assay and concurrent cell growth assay on mezerein in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

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2 **4.3.5.7 *N*-Methyl-*N*'-nitro-*N*-nitrosoguanidine**
3

4 The results of *N*-methyl-*N*'-nitro-*N*-nitrosoguanidine (MNNG) are exhibited in Table 38
5 and Fig. 34. MNNG was positive in the initiation assay and negative in the promotion
6 assay in the two laboratories.
7

8
9 Table 38.

10 Results of transformation assay on *N*-methyl-*N*'-nitro-*N*-nitrosoguanidine in phase II of
11 the 96-well method validation study.

12 a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	3/96	100	9/96
0.25			97	7/96
0.5			104	16/96
0.75			99	14/96
1			96	20/96
1.2	109	6/96		
1.25			87	23/96 *
1.5			78	31/96 *
1.55	113	10/95		
2			52	32/96 *
2.02	107	13/96 *		
2.5			22	24/96 *
2.62	101	26/95 *		
3.41	97	23/94 *		
4.43	73	18/95 *		
5.75	41	23/96 *		
7.5	13	14/94 *		
0 ^d (0.1% DMSO)	100	9/96	100	8/96
<u>MCA</u> 1 ^e (0.1% DMSO)	37	44/94 †	50	63/96 †

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34 ^a % of cell growth compared to that of solvent control.

35 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

36 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

37 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
38 media in parentheses.

39 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
40 parentheses.

41 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

42 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	10/96	100	6/96
0.1			97	10/96
0.2			102	7/96
0.5			101	9/96
0.75			99	7/96
0.78	100	12/96		
1			98	4/96
1.5			93	5/96
1.56	94	11/96		
2.5			86	3/96
3.13	93	9/96		
5			68	0/96
6.26	82	2/95		
12.5	63	0/96		
25	57	0/96		
50	13	Toxic		
0 ^d (0.1% DMSO)	100	12/95	100	7/96
TPA 0.05 ^e (0.1% DMSO)	143	57/96 †	136	62/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

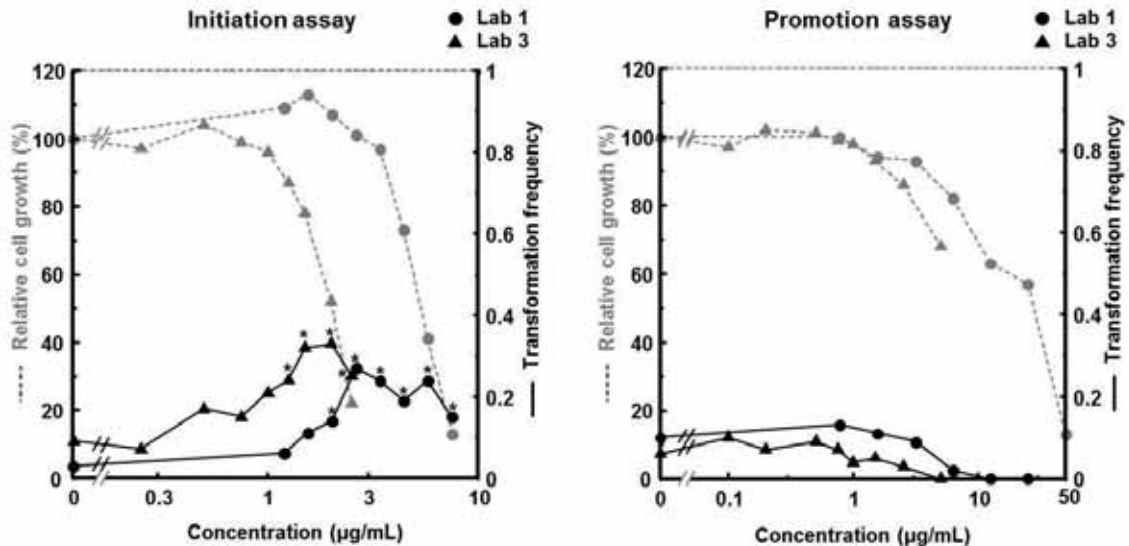


Fig. 34. Graphic view of the results of transformation assay and concurrent cell growth assay on *N*-methyl-*N*'-nitro-*N*-nitrosoguanidine in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

1
2 **4.3.5.8 Sodium arsenite**
3

4 Both of the laboratories reported negative results in the initiation and promotion assays,
5 as shown in Table 39 and Fig. 35.
6
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8 Table 39.

9 Results of transformation assay on sodium arsenite in phase II of the 96-well method
10 validation study.

11 a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	7/96	100	1/96
0.05			107	0/96
0.1			106	2/96
0.135	87	7/96		
0.175	80	1/96		
0.2			99	4/96
0.228	82	8/96		
0.296	57	3/96		
0.3			68	4/96
0.385	33	8/96		
0.4			35	5/96
0.5	12	3/96	16	3/96
0.6	9	8/96	4	4/96
0.7			3	2/96
0.72	3	6/95		
0 ^d (0.1% DMSO)	100	9/96	100	3/96
MCA 1 ^e (0.1% DMSO)	29	41/96 †	52	52/96 †

31 ^a % of cell growth compared to that of solvent control.

32 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

33 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

34 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
35 media in parentheses.

36 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
37 parentheses.

38 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	15/96	100	12/96
0.025			101	10/96
0.0469	100	21/96		
0.05			95	11/96
0.0938	93	26/96		
0.1			99	8/96
0.188	96	25/96		
0.25			100	5/96
0.375	86	7/96		
0.5			81	0/96
0.725	72	6/96		
1			63	Toxic
1.5	42	Toxic	51	Toxic
2			13	Toxic
3	6	Toxic		
0 ^d (0.1% DMSO)	100	14/96	100	7/96
TPA 0.05 ^e (0.1% DMSO)	167	72/96 †	168	68/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

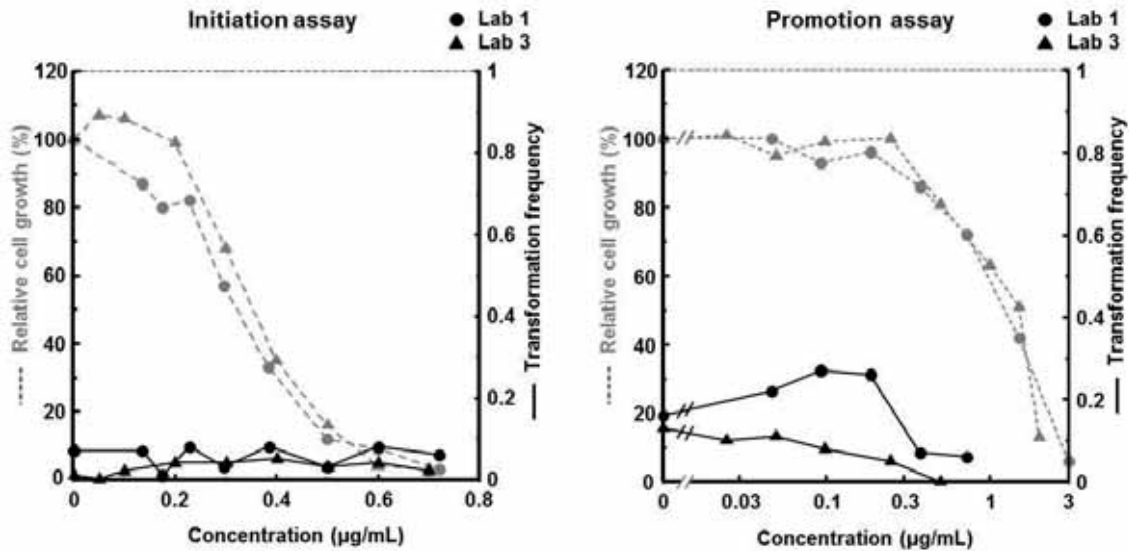


Fig. 35. Graphic view of the results of transformation assay and concurrent cell growth assay on sodium arsenite in phase II of the 96-well method validation study.

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2 **4.3.5.9 Ampicillin sodium**
3

4 The results of Ampicillin sodium were presented in Table 40 and Fig. 36. The chemical
5 was negative in the initiation and promotion assays in both laboratories.
6
7

8 Table 40.

9 Results of transformation assay on ampicillin sodium in phase II of the 96-well method
10 validation study.

11 a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	13/96	100	13/96
502	100	12/96		
603	98	10/96		
724	95	4/96		
800			98	6/96
868	95	7/96		
1000			103	6/96
1040	86	7/96		
1200			103	2/96
1250	59	7/96		
1400			86	2/96
1500	11	6/94	80	0/96
1600			55	1/96
1700			55	4/96
1800			27	5/96
0 ^d (0.1% DMSO)	100	12/96	100	13/96
<u>MCA</u> 1 ^e (0.1% DMSO)	39	48/96 †	52	54/96 †

31 ^a % of cell growth compared to that of solvent control.

32 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

33 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

34 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
35 media in parentheses.

36 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
37 parentheses.

38 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	18/96	100	9/96
250	97	9/96		
500	97	9/96	91	15/96
800			93	11/96
1000	97	12/95	95	10/96
1500			91	19/96
2000	84	8/96	85	11/96
3000	72	2/96	80	12/96
4000	66	1/96	72	0/96
5000	56	3/96	64	1/96
0 ^d (0.1% DMSO)	100	13/96	100	14/96
TPA 0.05 ^e (0.1% DMSO)	138	67/96 †	144	78/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

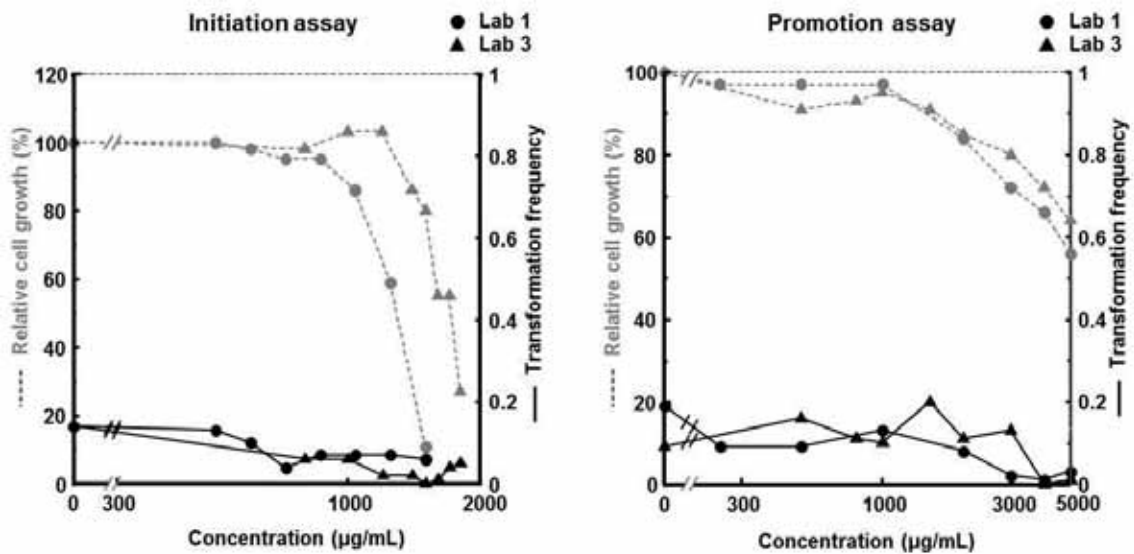


Fig. 36. Graphic view of the results of transformation assay and concurrent cell growth assay on ampicillin sodium in phase II of the 96-well method validation study.

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2 **4.3.5.10 L-Ascorbic acid**
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4 The results for L-ascorbic acid are shown in the Table 41 and Fig. 37. Ascorbic acid was
5 negative in both assays in both laboratories. In the first run of promotion assay in Lab 3,
6 only three out of eight plates treated with the chemical were observable for the
7 transformed foci, because the cells were killed by the toxic effect of chemical treatment
8 at the highest five concentrations. In addition, the incidence of transformed foci in the
9 negative control for TPA (21 wells/96 wells) exceeded the acceptance criteria (≤ 20
10 wells/96 well). Since this first run did not satisfy the assay acceptance criteria, Lab 3
11 repeated the promotion assay in a lower concentration range and confirmed L-ascorbic
12 acid to be negative.
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16 Table 41.
17 Results of transformation assay on L-ascorbic acid in phase II of the 96-well method
18 validation study.

19 a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	9/96	100	7/96
50			89	5/96
100			83	8/96
112	90	9/96		
134	89	7/96		
161	81	10/96		
193	74	4/96		
200			67	8/96
231	62	8/96		
278	43	5/96		
300			53	3/96
333	8	6/96		
350			51	1/96
380			42	6/96
400	-1	Toxic	37	2/96
450			13	1/96
0 ^d (0.1% DMSO)	100	10/96	100	7/96
MCA 1 ^e (0.1% DMSO)	40	41/96 †	56	60/96 †

40 ^a % of cell growth compared to that of solvent control.

41 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

42 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

43 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
44 media in parentheses.

45 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
46 parentheses.

47 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3, 1st run		Lab 3, 2nd run	
	CG ^a	TF ^b	CG	TF	CG	TF
0 ^c (5% Water)	100	13/96	100	7/96	100	5/96
12.5					97	4/96
25					99	0/96
50					101	2/96
96.6	90	2/96				
100					105	0/96
145	89	0/96				
200					101	0/96
217	91	1/96				
280			99	0/96		
326	86	1/96				
350			95	1/96	96	1/96
489	83	7/95				
500			92	2/96	92	Toxic
650			87	Toxic	62	Toxic
733	72	Toxic				
800			76	Toxic		
900			65	Toxic		
1000			52	Toxic		
1100	13	Toxic	40	Toxic		
0 ^d (0.1% DMSO)	100	15/96	100	21/96	100	8/96
TPA 0.05 ^e (0.1% DMSO)	137	56/96 †	128	85/96 †	120	45/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

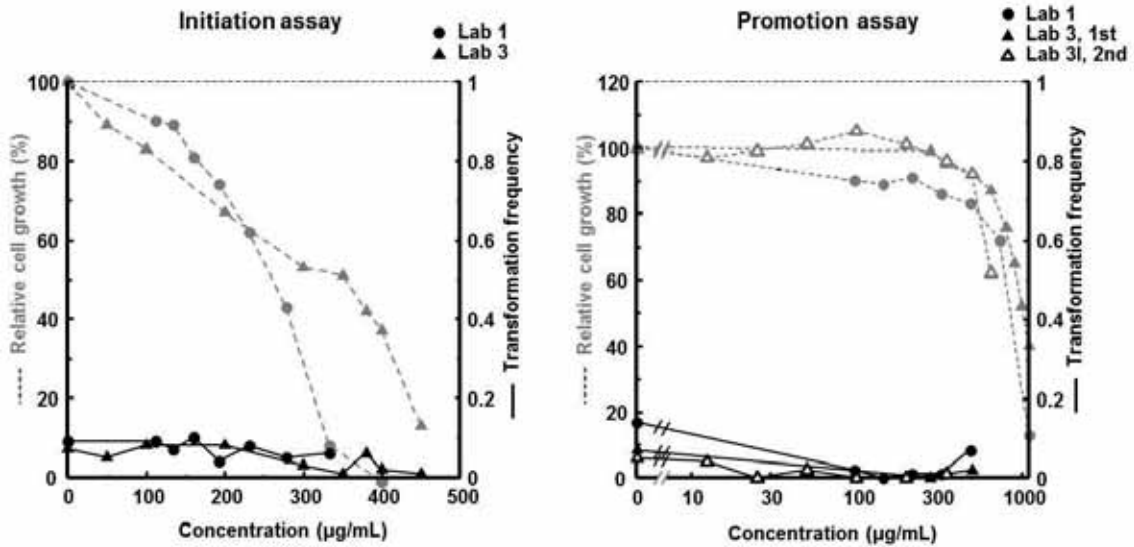


Fig. 37. Graphic view of the results of transformation assay and concurrent cell growth assay on L-ascorbic acid in phase II of the 96-well method validation study.

1
2 **4.3.5.11 Caffeine**
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4 The results of caffeine are presented in Table 42 and Fig. 38. Caffeine was negative in
5 both assays in both laboratories.
6
7

8 Table 42.

9 Results of transformation assay on caffeine in phase II of the 96-well method validation
10 study.

11 a) Initiation assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	5/96	100	3/96
200			107	7/96
226	107	8/96		
250			102	4/96
259	105	9/96		
298	105	4/96		
300			81	4/96
340			83	11/96
343	90	4/96		
370			58	12/96
395	57	4/96		
400			51	4/96
450			20	8/96
454	29	3/96		
500			7	6/96
522	13	3/96		
600	5	4/96		
0 ^d (0.1% DMSO)	100	4/96	100	2/96
MCA 1 ^e (0.1% DMSO)	56	55/96 †	44	41/96 †

33 ^a % of cell growth compared to that of solvent control.

34 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

35 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

36 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
37 media in parentheses.

38 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
39 parentheses.

40 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	6/96	100	4/96
2.5			112	6/96
5			110	7/96
10			108	7/96
18	99	5/96		
25			104	8/96
26	97	4/96		
40	94	2/96		
50			99	5/96
59	87	6/96		
89	76	3/96		
100			83	5/96
130	71	4/96		
200	57	3/96		
250			60	2/96
300	43	3/96		
500			43	0/96
0 ^d (0.1% DMSO)	100	7/96	100	11/96
TPA 0.05 ^e (0.1% DMSO)	149	68/96 †	138	54/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

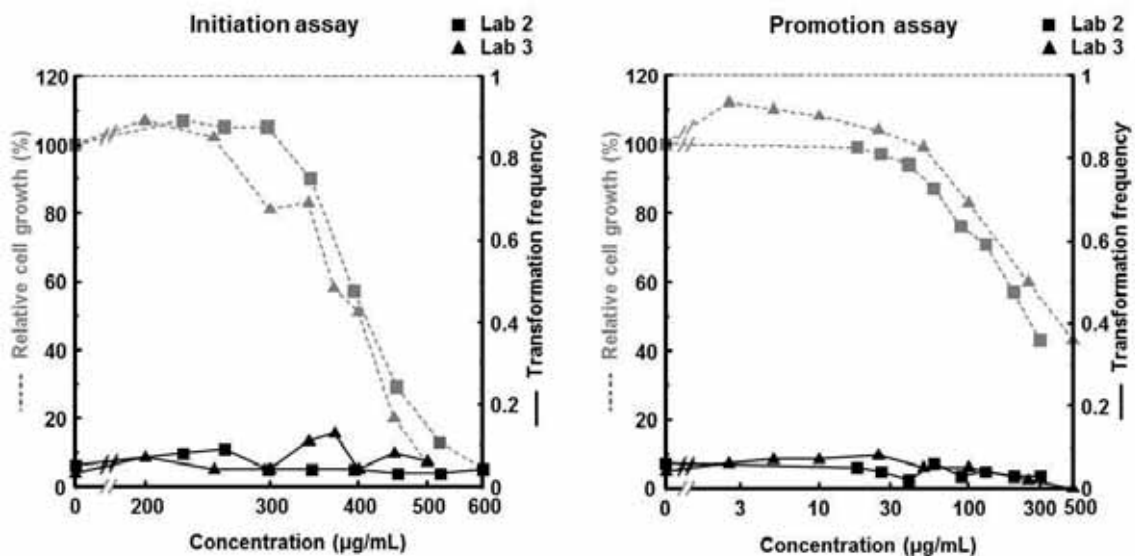


Fig. 38. Graphic view of the results of transformation assay and concurrent cell growth assay on caffeine in phase II of the 96-well validation study.

4.3.5.12 Caprolactam

Table 43 and Fig. 39 present the results of caprolactam. The chemical was determined to be negative in both the initiation and promotion assays despite the unusual outcome of the initiation assay. In the initiation assay, caprolactam induced statistically significant increases in transformation frequency at the extremely high concentrations of 2890 and 3470 µg/mL (25.54 and 30.66 mM) in Lab 1 and 3700 and 4000 µg/mL (32.70 and 35.35 mM) in Lab 3. However, in accordance with the Bhas 42 CTA protocol, those concentrations exceed the maximum allowable concentration that can be tested, i.e. 10 mM. Since the test chemical was coded for assessment in the validation study, the molecular weight was unknown to the laboratories, and so the studies were performed using these unknowingly improper chemical concentrations. The VMT concluded that the increase in transformation frequency induced by caprolactam at these excessive concentrations was, therefore, not biologically relevant and that, in fact, all test concentrations were in excess of the 10 mM maximum allowable concentration that can be tested. Thus, under the test conditions employed, caprolactam was considered to be negative in the initiation assay.

Table 43.
Results of transformation assay on caprolactam in phase II of the 96-well method validation study.

a) Initiation assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	7/96	100	9/96
1670 (14.76 mM)	113	12/96		
2010 (17.76 mM)	112	14/96		
2410 (21.30 mM)	113	18/96		
2500 (22.09 mM)			125	11/96
2890 (25.54 mM)	107	25/96 *		
3000 (26.51 mM)			117	11/96
3300 (29.16 mM)			96	16/96
3470 (30.66 mM)	87	20/96 *		
3700 (32.70 mM)			79	25/96 *
4000 (35.35 mM)			68	32/96 *
4170 (36.85 mM)	49	17/96		
4300 (38.00 mM)			56	16/96
4700 (41.53 mM)			42	11/96
5000 (44.19 mM)	15	7/96	25	19/96
0 ^d (0.1% DMSO)	100	9/96	100	8/96
MCA 1 ^e (0.1% DMSO)	36	43/96 †	40	65/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, MCA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the initiation assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	12/96	100	10/96
10			97	9/96
30			100	12/96
78.1	100	15/96		
100			96	12/96
156	103	23/96		
300			91	14/96
313	97	20/96		
625	94	17/96		
1000 (8.84 mM)			79	13/96
1250 (11.05 mM)	84	12/96		
2000 (17.67 mM)			66	13/96
2500 (22.09 mM)	58	14/96		
3000 (26.51 mM)			52	19/96
4000 (35.35 mM)			47	10/96
5000 (44.19 mM)	47	12/96		
0 ^d (0.1% DMSO)	100	14/96	100	11/96
TPA 0.05 ^e (0.1% DMSO)	136	63/96 †	137	54/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

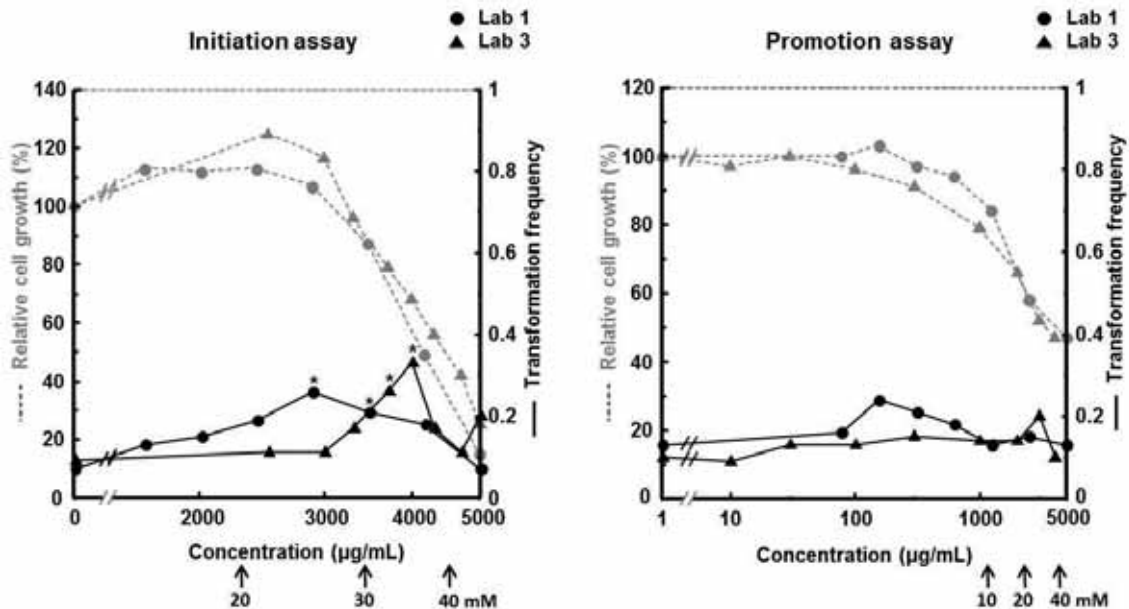


Fig. 39. Graphic view of the results of transformation assay and concurrent cell growth assay on caprolactam in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

1
2 **4.3.5.13 Eugenol**
3

4 The results of eugenol are shown in Table 44 and Fig. 40. Eugenol was negative in both
5 assays and in both laboratories. The chemical induced a statistically significant
6 increase in transformation frequency at a single concentration (2.5 µg/mL) in the first
7 run of promotion assay in Lab 3. In accordance with the protocol, Lab 3 repeated the
8 promotion assay, and in so doing, applied a narrower range of test concentrations that
9 bracketed the 2.5 µg/mL concentration. The results of the second run confirmed the
10 negative judgment.
11

12
13 Table 44.
14 Results of transformation assay on eugenol in phase II of the 96-well method validation
15 study.

16 a) Initiation assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.1% DMSO)	100	10/96	100	11/96
18	111	10/96		
25	113	12/96		
30			104	7/96
35	112	10/96		
40			116	6/96
50	106	10/96	118	3/96
60			108	9/96
70	63	7/96	92	10/96
80			72	5/96
90			41	10/96
100	28	6/96	32	4/96
120			29	6/96
140	11	4/96		
200	10	3/96		
MCA 1^d (0.1% DMSO)	21	64/96 †	18	59/96 †

35 ^a % of cell growth compared to that of solvent control.
36 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.
37 ^c Solvent control: final solvent concentration in the working culture media in parentheses.
38 ^d Positive control in the initiation assay: final solvent concentration in the working culture media in
39 parentheses.
40 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration ($\mu\text{g/mL}$)	Lab 2		Lab 3, 1st run		Lab 3, 2nd run	
	CG ^a	TF ^b	CG	TF	CG	TF
0 ^c (0.1% DMSO)	100	11/96	100	15/96	100	18/96
0.3125			105	22/96		
0.625			108	25/96		
1					105	17/96
1.25			105	20/96		
1.5					113	21/96
2					112	11/96
2.5			102	31/96 *	108	9/96
3					115	16/96
3.5					112	9/96
4					112	16/96
5			101	13/96	115	11/96
6.3	104	12/96				
7.5			99	14/96		
8.8	105	4/96				
10			92	10/96	115	4/96
13	99	7/96				
18	95	5/96				
20			89	6/96		
25	90	7/96				
35	78	4/96				
40			79	1/96		
50	65	5/96				
70	56	4/96				
100	50	6/96				
TPA 0.05 ^d (0.1% DMSO)	86	48/96 †	116	85/96 †	140	83/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

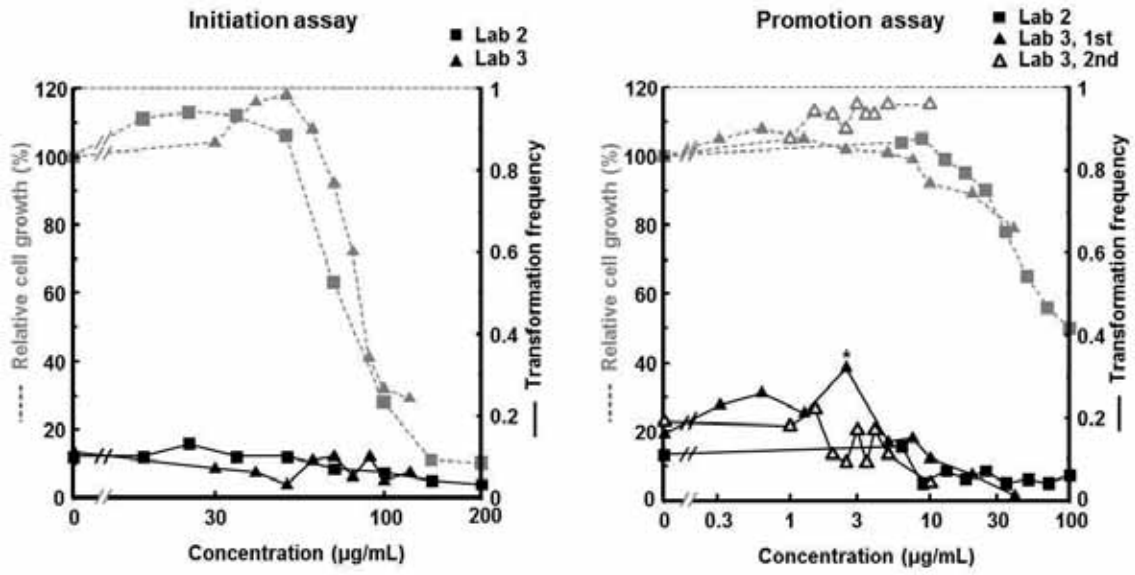
^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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Fig. 40. Graphic view of the results of transformation assay and concurrent cell growth assay on eugenol in phase II of the 96-well method validation study.
* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

1 **4.3.5.14 D-Mannitol**

2
3 Both two laboratories found D-mannitol to be negative in both assays, as shown in Table
4 45 and Fig. 41.

5
6
7 Table 45.

8 Results of transformation assay on D-mannitol in phase II of the 96-well method
9 validation study.

10 a) Initiation assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	3/96	100	6/96
290	115	3/96		
440	111	2/94		
500			96	6/96
660	122	2/96		
990	117	1/96		
1000			100	6/96
1500	118	1/96		
2000			103	1/96
2200	114	2/95		
3000			96	8/96
3300	114	3/96		
4000			98	7/96
5000	121	3/96	101	7/96
0 ^d (0.1% DMSO)	100	4/96	100	10/96
MCA 1 ^e (0.1% DMSO)	56	57/96 †	38	46/96 †

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29 ^a % of cell growth compared to that of solvent control.

30 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

31 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

32 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
33 media in parentheses.

34 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
35 parentheses.

36 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

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b) Promotion assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	5/96	100	9/96
290	105	5/96		
440	112	4/96		
500			99	7/96
660	112	11/96		
990	109	7/96		
1000			99	6/96
1500	106	9/96		
2000			102	7/96
2200	110	6/96		
3000			104	5/96
3300	101	6/96		
4000			100	3/96
5000	109	4/96	100	4/96
0 ^d (0.1% DMSO)	100	8/96	100	8/96
TPA 0.05 ^e (0.1% DMSO)	156	43/96 †	153	77/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

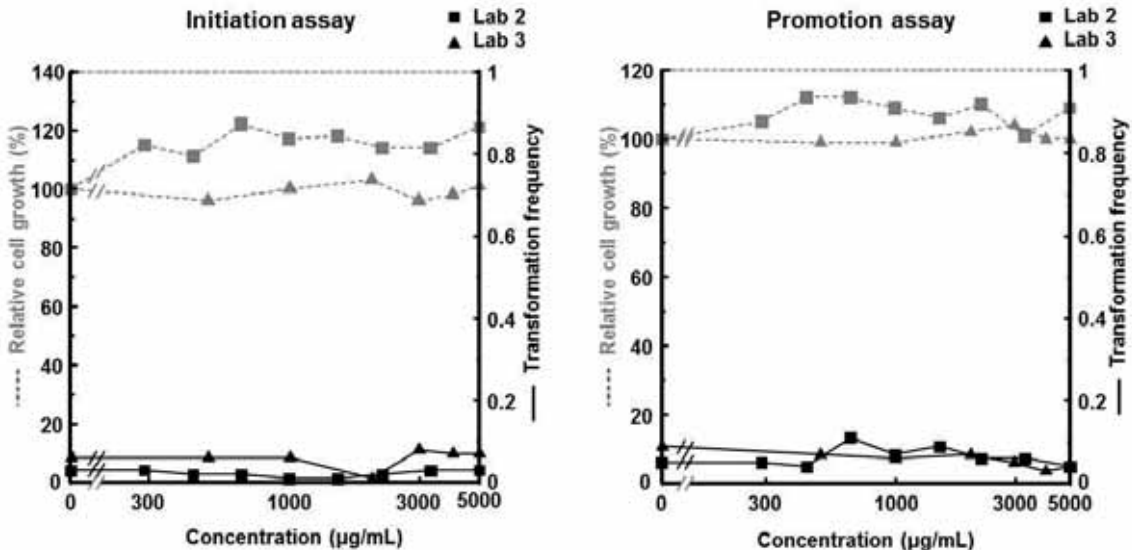


Fig. 41. Graphic view of the results of transformation assay and concurrent cell growth assay on D-mannitol in phase II of the 96-well method validation study.

1
2 **4.3.5.15 Phorbol**
3

4 The results of phorbol are represented in Table 46 and Fig. 42. The highest
5 concentration of phorbol to be tested was pre-assigned by the VMT to be $\leq 5 \mu\text{g/mL}$ in
6 the medium, because this chemical is expensive and its availability was limited. In the
7 concentration range pre-assigned by VMT, phorbol was negative in both assays in both
8 laboratories. However, cytotoxicity was also not induced at any of the concentrations in
9 the concurrent cell growth assay. Phorbol is quite soluble in polar solvents, including
10 water. In order to draw any conclusion regarding the transformation activity of phorbol,
11 this chemical should have been tested at higher concentrations that it achieved
12 cytotoxicity or induced a positive result in at least one of component assays (initiation
13 assay or promotion assay). Therefore, it was concluded that these transformation assays
14 for phorbol were incomplete because of inadequate dosing.
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16
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18 Table 46.
19 Results of transformation assay on phorbol in phase II of the 96-well method validation
20 study.

21 a) Initiation assay

Concentration ($\mu\text{g/mL}$)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	10/96	100	13/96
0.5	110	12/96	98	8/96
1	112	13/96	100	7/96
2	105	10/96	95	5/96
3	111	4/96	104	4/96
4	103	6/96	97	6/96
5	102	3/96	103	6/96
0 ^d (0.1% DMSO)	100	5/96	100	9/96
MCA 1 ^e (0.1% DMSO)	43	46/96 †	50	62/96 †

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33 ^a % of cell growth compared to that of solvent control.

34 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

35 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

36 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
37 media in parentheses.

38 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
39 parentheses.

40 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
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b) Promotion assay

Concentration (µg/mL)	Lab 1		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (5% Water)	100	9/96	100	10/96
0.5	99	12/96	100	13/96
1	98	6/96	101	12/96
2	96	8/96	100	17/96
3	96	13/96	103	13/96
4	96	7/96	103	8/96
5	97	11/96	99	12/96
0 ^d (0.1% DMSO)	100	10/96	100	11/96
TPA 0.05 ^e (0.1% DMSO)	138	55/96 †	131	81/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

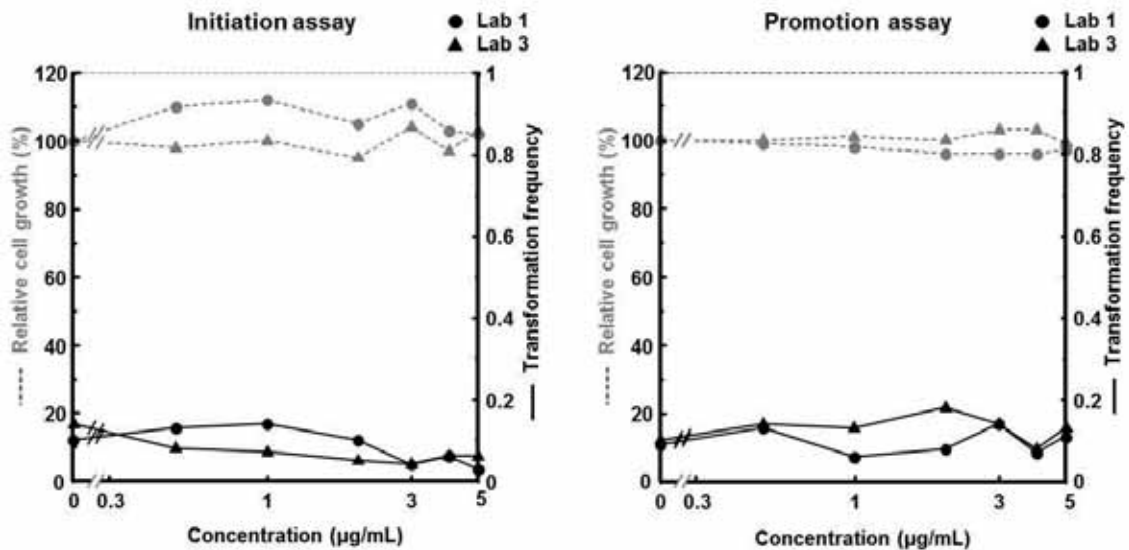


Fig. 42. Graphic view of the results of transformation assay and concurrent cell growth assay on phorbol in phase II of the 96-well method validation study.

1
2 **4.3.5.16 Pyrene**
3

4 Table 47 and Fig. 43 show the results for pyrene, which were positive in both the
5 initiation and promotion assays in both laboratories. In the initiation assay, although
6 the concentrations examined in Lab 2 were too low for the chemical to induce
7 cytotoxicity in the concurrent cell growth assay, the fact that pyrene was clearly positive
8 for transformation in that concentration range suggested that Lab 2 need not repeat the
9 assay, as determined by the VMT. In the promotion assay, pyrene was clearly positive in
10 Lab 2 as well. In contrast, Lab 3 observed a statistically significant increase in
11 transformation frequency at only a single concentration (10 µg/mL) in the first run of
12 promotion assay. Lab 3 repeated the assay at a slightly lower concentration range and
13 once again obtained the statistically significant increase in transformation frequency at
14 the same concentration. Thus, pyrene was judged to be positive in the promotion assay
15 in Lab 3.
16
17

18 Table 47
19 Results of transformation assay on pyrene in phase II of the 96-well method validation
20 study.

21 a) Initiation assay

Concentration (µg/mL)	Lab 2		Lab 3	
	CG ^a	TF ^b	CG	TF
0 ^c (0.5% DMSO)	100	5/96	100	6/96
2.8	101	8/96		
5.6	111	6/96		
11	114	8/96		
23	100	12/96		
25			106	6/96
45	103	19/96 *		
50			106	17/96
90	111	20/96 *		
100			109	21/96 *
150			96	22/96 *
180	117	25/96 *		
200			106	23/96 *
250			94	25/96 *
300			84	14/96
400			68	16/96
0 ^d (0.1% DMSO)	100	7/96	100	13/96
MCA 1 ^e (0.1% DMSO)	59	53/96 †	54	65/96 †

42 ^a % of cell growth compared to that of solvent control.

43 ^b Transformation frequency: number of wells having transformed foci/number of wells counted.

44 ^c Solvent control: final solvent concentration in the working culture media in parentheses.

45 ^d Solvent control for the positive control, MCA: final solvent concentration in the working culture
46 media in parentheses.

47 ^e Positive control in the initiation assay: final solvent concentration in the working culture media in
48 parentheses.

49 * $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

50 † $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.
51
52
53
54
55
56
57

b) Promotion assay

Concentration (µg/mL)	Lab 2		Lab 3, 1st run		Lab 3, 2nd run	
	CG ^a	TF ^b	CG	TF	CG	TF
0 ^c (0.5% DMSO)	100	13/96	100	9/96	100	14/96
2.8	101	10/96				
5			104	13/96	100	14/96
5.6	98	20/96				
10			92	25/96 *	94	31/96 *
11	92	30/96 *				
15					90	27/96
23	90	36/96 *				
25			88	17/96	89	23/96
45	97	40/96 *				
50			94	15/96	93	27/96
75					90	21/96
90	91	47/96 *				
100			96	16/96	88	26/96
150			92	7/96	85	9/96
180	95	28/96 *				
200			74	Toxic		
400			55	Toxic		
0 ^d (0.1% DMSO)	100	7/96	100	12/96	100	18/96
TPA 0.05 ^e (0.1% DMSO)	159	52/96 †	134	58/96 †	169	71/96 †

^a % of cell growth compared to that of solvent control.

^b Transformation frequency: number of wells having transformed foci/number of wells counted.

^c Solvent control: final solvent concentration in the working culture media in parentheses.

^d Solvent control for the positive control, TPA: final solvent concentration in the working culture media in parentheses.

^e Positive control in the promotion assay: final solvent concentration in the working culture media in parentheses.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment, vs. corresponding solvent control.

† $p < 0.05$; one-sided chi-square test, vs. corresponding solvent control.

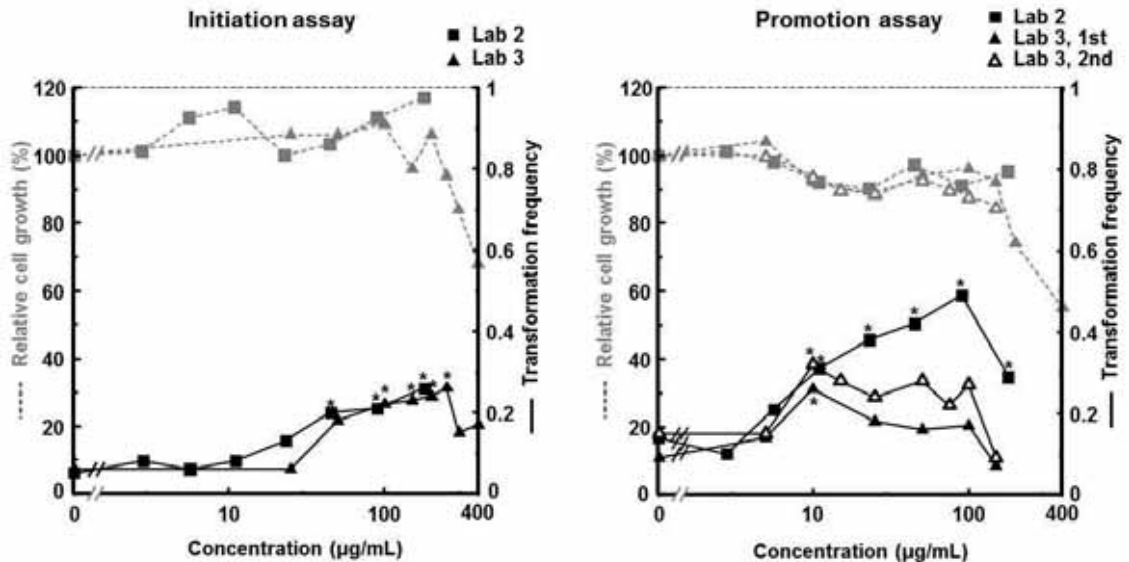


Fig. 43. Graphic view of the results of transformation assay and concurrent cell growth assay on pyrene in phase II of the 96-well method validation study.

* $p < 0.05$; one-sided chi-square test with Bonferroni's adjustment.

1
2
3 **4.3.6 Fulfillment of acceptance criteria**
4

5 The success or failure to fulfill the assay acceptance criteria for each assay conducted in each lab is
6 listed in Table 48.
7

8
9
10 Table 48.
11 Fulfillment of the assay acceptance criteria

Phase I						
Chemical	Lab	Initiation assay		Promotion assay		
		Run	All assay acceptance criteria	Run	All assay acceptance criteria	
2-Acetyl-amino-fluorene	Lab 1		Fulfilled		Fulfilled	
	Lab 2		Fulfilled		Fulfilled	
	Lab 3		Fulfilled		Fulfilled	
	Lab 4	1st		Fulfilled. However, there was a statistically significant increase in transformation frequency at one concentration. The assay was repeated.		Fulfilled
		2nd		Fulfilled		
Benzo[a]pyrene	Lab 1		Fulfilled		Fulfilled	
	Lab 2		Fulfilled		Fulfilled	
	Lab 3		Fulfilled		Fulfilled	
	Lab 4		Fulfilled		Fulfilled	
3-Methyl-cholanthrene	Lab 1		Fulfilled		Fulfilled	
	Lab 2		Fulfilled		Fulfilled	
	Lab 3		Fulfilled		Fulfilled	
	Lab 4		Fulfilled		Fulfilled	
σ-Toluidine	Lab 1		Fulfilled		Fulfilled	
	Lab 2		Fulfilled		Fulfilled	
	Lab 3		Fulfilled		Fulfilled	
	Lab 4		Fulfilled		Fulfilled	
TPA	Lab 1		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (<1 µg/mL), has not achieved cytotoxicity.		Fulfilled	
	Lab 2		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (≤1 µg/mL), had not achieved cytotoxicity		Fulfilled	
	Lab 3		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (≤1 µg/mL), had not achieved cytotoxicity.		Fulfilled	

1 Table 48.
2 (Continued)

Chemical	Lab	Initiation assay		Promotion assay		
		Run	All assay acceptance criteria	Run	All assay acceptance criteria	
	Lab 4		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (≤ 1 $\mu\text{g/mL}$), had not achieved cytotoxicity.		Fulfilled	
Anthracene	Lab 1		Fulfilled. The chemical was examined up to the maximum concentration soluble in DMSO, although it did not induced cytotoxicity.		Fulfilled	
	Lab 2		Fulfilled. The chemical was examined up to the maximum concentration soluble in DMSO, although it did not induced cytotoxicity.		Fulfilled	
	Lab 3		Fulfilled. The chemical was examined up to the maximum concentration soluble in DMSO, although it did not induced cytotoxicity.		Fulfilled	
	Lab 4		Fulfilled. The chemical was applied as suspension to the cultures and induced cytotoxicity.		Fulfilled	
Phenanthrene	Lab 1		Fulfilled	1st	Fulfilled. However, there was a statistically significant increase in transformation frequency at one concentration. The assay was repeated.	
				2nd	Fulfilled	
	Lab 2		Fulfilled		Fulfilled	
	Lab 3		Fulfilled		Fulfilled	
	Lab 4	1st	Fulfilled. However, there was a statistically significant increase in transformation frequency at one concentration. The assay was repeated.			Fulfilled
		2nd	Fulfilled except for the concentration criterion, which was considered inapplicable for the following reason: to conclude positive or negative call, this assay was carried out in a narrower range around the concentration at which there had been a statistically significant increase of transformation frequency in the 1st run.			

1 Table 48.
2 (Continued)

Phase II					
Chemical	Lab	Initiation assay		Promotion assay	
		Run	All assay acceptance criteria	Run	All assay acceptance criteria
Benzo[<i>a</i>]pyrene	Lab 1		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
Cadmium chloride	Lab 1		Fulfilled		Fulfilled
	Lab 3		Fulfilled	1st	Not fulfilled. The criterion for the number of valid concentrations was not met, i.e., there was only one valid concentration.
				2nd	Fulfilled
Dibenz[<i>a,h</i>]-anthracene	Lab 2		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
Lithocholic acid	Lab 2		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
Methapyrilene HCl	Lab 2		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
Mezerein	Lab 2		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT ($\leq 0.1 \mu\text{g/mL}$), had not achieved cytotoxicity.		Fulfilled
	Lab 3		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT ($\leq 0.1 \mu\text{g/mL}$), had not achieved cytotoxicity.		Fulfilled
MNNG	Lab 1		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
Sodium arsenite	Lab 1		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
Ampicillin sodium	Lab 1		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
L-Ascorbic acid	Lab 1		Fulfilled		Fulfilled
	Lab 3		Fulfilled	1st	Not fulfilled. The criterion for the number of valid concentrations was not met, i.e., there were only three valid concentrations. Also not fulfilled was the criterion for the negative control.
				2nd	Fulfilled
Caffeine	Lab 2		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled

1 Table 48.
2 (Continued)

Chemical	Lab	Initiation assay		Promotion assay	
		Run	All assay acceptance criteria	Run	All assay acceptance criteria
Caprolactam	Lab 1		Fulfilled		Fulfilled
	Lab 2		Fulfilled		Fulfilled
Eugenol	Lab 2		Fulfilled		Fulfilled
	Lab 3		Fulfilled	1st	Fulfilled. However, there was a statistically significant increase in transformation frequency at only one concentration. The assay was repeated.
				2nd	Fulfilled
D-Mannitol	Lab 2		Fulfilled		Fulfilled
	Lab 3		Fulfilled		Fulfilled
Phorbol	Lab 1		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (≤ 5 $\mu\text{g/mL}$), had not achieved cytotoxicity.		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (≤ 5 $\mu\text{g/mL}$), had not achieved cytotoxicity.
	Lab 2		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (≤ 5 $\mu\text{g/mL}$), had not achieved cytotoxicity.		Not fulfilled. The concentrations used, the maximum concentration of which had been pre-assigned by the VMT (≤ 5 $\mu\text{g/mL}$), had not achieved cytotoxicity.
Pyrene	Lab 2		Not fulfilled. The concentrations used had not achieved cytotoxicity. The data were considered acceptable, however, since there were three sequential concentrations that induced statistically significant increases in transformation frequency and consequently the chemical was called positive.		Fulfilled
	Lab 3		Fulfilled	1st	Fulfilled. However, there was a statistically significant increase in transformation frequency at one concentration. The assay was repeated.
				2nd	Fulfilled

3
4
5
6 A total of 62 experiments of the initiation assay were performed in the validation phase I and phase
7 II studies on the 96-well method. Out of 62 experiments, nine did not fulfill the assay acceptance
8 criteria. The failure to satisfy those acceptance criteria by eight out of nine of those experiments
9 were the result of the maximum concentrations employed, which were pre-assigned by the VMT
10 (TPA, mezerein and phorbol) being too low. Therefore, only one out of 54 experiments was judged
11 not to fulfill the assay acceptance criteria with respect to the conditions (including dose selection)
12 for which the participating laboratories were responsible. The data of this particular experiment
13 were considered acceptable, however, since there were three sequential concentrations that induced

1 statistically significant increases in transformation frequency and consequently the chemical was
2 called positive although the concentrations used had not achieved cytotoxicity (pyrene in Lab 2).

3
4 A total of 65 experiments of promotion assay were carried out. Out of 65 experiments, four did not
5 fulfill the assay acceptance criteria. The failure to satisfy those acceptance criteria by two out of
6 four of those experiments were the result of the maximum concentrations employed, which were
7 pre-assigned by the VMT (phorbol). Therefore, two out of 63 experiments were judged not to fulfill
8 the assay acceptance criteria with respect to the conditions (including dose selection) for which the
9 participating laboratories were responsible. These two particular experiments were repeated and
10 the second runs succeeded to fulfill the assay acceptance criteria (cadmium chloride in Lab 3 and
11 L-ascorbic acid in Lab 3).

12

1
2 **4.4 Conclusion**
3

4 The positive and negative calls for the test chemicals based on the results in the Bhas
5 42 CTA 96-well method validation study are summarized in Table 49. In the table, the
6 positive and negative calls by each laboratory for each chemical are made based upon
7 the integrated judgments from the initiation and promotion assay results. That is, a test
8 chemical is positive in a given laboratory if it is positive in either the initiation or
9 promotion assay. The overall judgment (AO) recorded for each chemical is determined
10 by majority rule based on the results obtained by each of the participating laboratories.
11

12
13 **4.4.1 Module 2 — Within-laboratory reproducibility**
14

15 The transformation frequencies of the negative (DMSO) and positive controls obtained
16 in all assays in phase I and phase II of the 96-well method validation study are
17 clustered by individual laboratories and plotted for the respective initiation and
18 promotion assays, as shown in Fig. 17 (section 4.3.2). The average transformation
19 frequency of the negative and positive controls per individual laboratory in phase I and
20 phase II of the 96-well method validation study is shown in Fig. 18 (section 4.3.2). The
21 results depicted in these figures demonstrated that within-laboratory reproducibility
22 was satisfactory in all the laboratories for the negative and positive controls in both the
23 initiation assay and the promotion assay.
24

25 To estimate the within-laboratory reproducibility, results of the same test chemical in
26 the same laboratory were compared between different phases.
27

28 MCA and TPA were examined in pre-validation phase and in the validation phase I by
29 each laboratory. Benzo[a]pyrene was tested in validation phases I and II. When the
30 results of these chemicals in each laboratory were compared for positive and negative
31 calls between the phases, these three chemicals are all positive in the transformation
32 assay in every laboratory in both phases (Table 49). MCA was judged to be equivocal in
33 one of the promotion assays in the pre-validation phase and in one of the promotion
34 assays in the validation phase I having induced a statistically significant increase of
35 transformation frequency at a single high concentration. Nevertheless, the other
36 judgments for all the test chemicals were concordant in both initiation and promotion
37 assays within each laboratory between phases.
38

39 A comparison of Figures 19 and 23 for MCA, Figures 20 and 25 for TPA and Figures 22
40 and 28 for benzo[a]pyrene, showed that the dose-response relationships for each test
41 chemical were similar between phases in a given laboratory. This held true even though
42 the concentration scales were different between the figures for each respective chemical
43 in the different phases.
44

45 These results demonstrated good intra-laboratory reproducibility of the 96-well method
46 of Bhas 42 CTA.
47

Table 49.

Results of validation studies on the 96-well method of Bhas 42 cell transformation assay

Compound	Pre-validation study				OA ^a	Phase I study					Phase II study				Carcinogenicity <i>in vivo</i>
	Laboratory					Laboratory					Laboratory			OA	
	I	II	III	IV		I	II	III	IV	OA	I	II	III		
2-Acetylaminofluorene															+
Initiation ^b						-	+	+	+	+					
Promotion ^c						+	-	+	+	+					
Transformation ^d						+	+	+	+	+					
Benzo[<i>a</i>]pyrene															+
Initiation						+	+	+	+	+	+		+	+	
Promotion						-	-	-	-	-	-		-	-	
Transformation						+	+	+	+	+	+		+	+	
Cadmium chloride															+
Initiation											-		-	-	
Promotion											+		+	+	
Transformation											+		+	+	
Dibenz[<i>a,h</i>]anthracene															+
Initiation												+	+	+	
Promotion												-	-	-	
Transformation												+	+	+	
Lithocholic acid															TP ^e
Initiation												-	-	-	
Promotion												+	+	+	
Transformation												+	+	+	
Methapyrilene HCl															+, TP
Initiation												-	-	-	
Promotion												+	+	+	
Transformation												+	+	+	
3-Methylcholanthrene															+
Initiation	+	+	+	+	+	+	+	+	+	+					
Promotion	-	± ^f	-	-	-	-	-	-	±	-					
Transformation	+	+	+	+	+	+	+	+	+	+					
Mezerein															TP
Initiation												+	+	+	
Promotion												+	+	+	
Transformation												+	+	+	
MNNG															+
Initiation											+		+	+	
Promotion											-		-	-	
Transformation											+		+	+	
Sodium arsenite															+
Initiation											-		-	-	
Promotion											-		-	-	
Transformation											-		-	-	
σ-Toluidine															+
Initiation						-	-	-	-	-					
Promotion						-	-	-	-	-					
Transformation						-	-	-	-	-					
TPA															TP
Initiation	-	-	-	-	-	-	-	-	-	-					
Promotion	+	+	+	+	+	+	+	+	+	+					
Transformation	+	+	+	+	+	+	+	+	+	+					
Ampicillin sodium															-
Initiation											-		-	-	
Promotion											-		-	-	
Transformation											-		-	-	
Anthracene															-
Initiation						-	-	-	-	-					
Promotion						-	-	-	-	-					
Transformation						-	-	-	-	-					

Table 49.
(Continued)

Compound	Pre-validation study				OA ^a	Phase I study				OA	Phase II study			Carcinogenicity <i>in vivo</i>
	Laboratory					Laboratory					Laboratory			
	I	II	III	IV		I	II	III	IV		I	II	III	
L-Ascorbic acid														
Initiation										-	-	-		
Promotion										-	-	-		
Transformation										-	-	-		
Caffeine														
Initiation											-	-	-	
Promotion											-	-	-	
Transformation											-	-	-	
Caprolactam														
Initiation											-g	-g	-	
Promotion											-	-	-	
Transformation											-	-	-	
Eugenol														
Initiation											-	-	-	
Promotion											-	-	-	
Transformation											-	-	-	
D-Mannitol														
Initiation											-	-	-	
Promotion											-	-	-	
Transformation											-	-	-	
Phenanthrene														
Initiation						-	-	-	-	-				
Promotion						-	+	-	-	-				
Transformation						-	+	-	-	-				
Phorbol														
Initiation											i ^h	i		
Promotion											i	i		
Transformation											i	i		
Pyrene														
Initiation												+	+	+
Promotion												+	+	+
Transformation												+	+	+

^a Overall judgment: Judgment by majority rule among laboratories.

^b Judgment in the initiation assay.

^c Judgment in the promotion assay.

^d Judgment in the Bhas 42 CTA into which judgments in initiation and promotion assays were integrated.

^e Tumor-promoter.

^f Equivocal: there was a statistically significant increase in transformation frequency at only one concentration.

^g <21.3 mM and <29.2 mM, negative in Lab I and III, respectively; 25.5 mM< and 32.7 mM<, positive in Lab I and Lab III (see section 4.3.5.12 above).

^h Incomplete: unable to judge the results due to inadequate dosing, which was the result of the VMT pre-assignment of too low a dose as the highest concentration to be tested (see section 4.3.5.15).

* Carcinogenicity in mice (leukaemogenic action) is positive or negative depending on the strains [Berenblum and Vlasta, 1970; Armuth, 1976]. Tumor-promoting activity in mouse skin is positive or negative depending on the strains [Baird and Boutwell, 1971; Slaga et al., 1976, 1980].

1 4.4.2 Module 3 — Transferability

2
3 Basically, the transferability of the 96-well method Bhas 42 CTA is the same as that of
4 the 6-well method (section 3.4.2). The Bhas 42 CTA can be performed in a laboratory
5 that has experience in routine cell culture techniques. General cell culture laboratory
6 equipment and instruments are sufficient to perform the proposed test method. All
7 supplies and reagents are readily available commercially. Like the 6-well method Bhas
8 42 CTA, however, the 96-well method requires staff training beyond that of general cell
9 culture techniques in order to distinguish transformed foci from non-transformed foci.
10 Training acquired in mastering focus discrimination using the 6-well method Bhas 42
11 CTA is directly applicable to that for the 96-well method. The identification of
12 transformed foci is carried out using the same photo catalog that is used for the 6-well
13 method Bhas 42 CTA (Annex 3), since the criteria of transformed foci are the same
14 between the 6-well method and the 96-well method.

15
16 Although all the participating laboratories had experience in the 6-well method Bhas 42
17 CTA, a one-day workshop in November, 2008 was held at HRI for the technical transfer
18 of the 96-well method. All the participating laboratories joined the workshop which was
19 designed to reinforce their previous technical training and to apply it to the 96-well
20 method Bhas 42 CTA, thereby ensuring proper identification of Bhas 42 transformed
21 foci produced in a 96-well plate.

22
23 After the workshop, to estimate transferability, MCA and TPA at the designated
24 concentrations were examined in the pre-validation phase (Tables 23 and 24, and
25 Figures 19 and 20). MCA was positive in the initiation assay and negative in the
26 promotion assay, showing concordant dose-response curves between the laboratories.
27 TPA was positive in the promotion assay and negative in the initiation assay, inducing
28 similar responses between the laboratories. These results demonstrated that with
29 adequate training, the Bhas 42 CTA 96-well method is readily transferable between
30 laboratories and because of its portability, the assay is one that can be efficiently and
31 successfully performed by laboratories outside of that of the test method developer.

32
33 Although transferability had been demonstrated in the pre-validation phase study, both
34 the VMT and the participating laboratories convened a joint meeting in January, 2009
35 in preparation for the subsequent validation phase I and phase II studies. At that
36 meeting, experiential information was exchanged on various technical issues and the
37 harmonization of focus counting (identification and discrimination) in order to ensure
38 methodological consistency and pre-empt possible unanticipated issues that might
39 arise.

40 41 42 4.4.3 Module 4 — Between-laboratory reproducibility

43
44 As shown in Figures 17 and 18, the transformation frequencies of the negative controls
45 for the initiation assay and the promotion assay were similar between laboratories and
46 those of the positive controls were also similar in range between laboratories. These
47 data indicate that the between-laboratory reproducibility was satisfactory for negative
48 and positive controls.

49
50 Table 49, which presents the summary results of 96-well method validation study,
51 shows the good reproducibility obtained between laboratories. Positive or negative calls
52 based upon the integrated judgments from the initiation and promotion assay results

per laboratory were concordant between laboratories, except for phenanthrene. Therefore, the between-laboratory reproducibility of the Bhas 42 CTA was 95% (21/22) with respect to a total of 22 chemicals which were examined in the validation phase I and phase II studies. Phorbol, of which the initiation and promotion assays were incomplete, was excluded from this calculation. Even if we compare the results of each of the component assays, i.e., initiation assay or promotion assay, the judgments were comparable between laboratories except for one of the initiation assays and one of the promotion assays for 2-acetylaminofluorene and one of the promotion assays for phenanthrene. Thus, the between-laboratory reproducibility of initiation assay was 95% (21/22) and that of promotion assay was 91% (20/22).

Between-laboratory reproducibility	
Assay	%
Initiation assay	95 (21/22)
Promotion assay	91 (20/22)
Transformation assay (Bhas 42 CTA)	95 (21/22)

Furthermore, the dose-response curves produced by positive chemicals were similar between laboratories in many instances, e.g., MCA, TPA, benzo[*a*]pyrene, cadmium chloride, dibenz[*a,h*]anthracene, lithocholic acid, methapyrilene hydrochloride, mezerein and pyrene.

These results demonstrated a high degree of inter-laboratory reproducibility of Bhas 42 CTA 96-well method.

4.4.4 Module 5 — Predictive capacity

As shown in Table 49, overall judgments (i.e., the judgment by majority rule) in the transformation assays were consistent with the reported *in vivo* carcinogenicity results except for sodium arsenite, *o*-toluidine and pyrene. The concordance for the prediction of chemical carcinogenicity was 86 % (18/21) in the 96-well method validation study: the sensitivity was 83% (10/12) and the specificity was 89% (8/9). A 2x2 contingency table of 21 tested chemicals and the performance indices of 96-well method Bhas 42 CTA for the prediction of chemical carcinogenicity are presented below (phorbol, of which the initiation and promotion assays were incomplete, was excluded from this calculation):

2x2 Contingency table of the results in the 96-well method Bhas 42 CTA validation study

		<i>In vivo</i> carcinogenicity		Total
		Carcinogen	Non-carcinogen	
Bhas 42 cell transformation assay	+	10	1	11
	–	2	8	10
Total		12	9	21

1 The performance of 96-well method Bhas 42 CTA for the
2 prediction of chemical carcinogenicity

3 <u>Performance index</u>	<u>%</u>
4 Concordance	86 (18/21)
5 Sensitivity	83 (10/12)
6 Specificity	89 (8/9)
7 Positive predictivity	91 (10/11)
8 Negative predictivity	80 (8/10)
9 False negative	17 (2/12)
10 <u>False positive</u>	<u>11 (1/9)</u>

15 4.4.5 Positive and negative controls

16
17 The transformation frequency of each positive control was statistically significantly
18 different from that of corresponding negative control in phase I and phase II of the
19 validation study, and the cluster of transformation frequencies for the positive controls
20 was quite distinct from that of the negative controls (Fig. 17). Therefore, the positive
21 control responses are considered to be robust, and the assay acceptance criteria
22 established for transformation frequency in the negative and positive controls were
23 considered to be appropriate.

24
25

5 Discussion

5.1 Module 1— Test definition

As mentioned above, the Bhas 42 CTA is a short-term system to predict chemical carcinogenicity. The assay can detect genotoxic and non-genotoxic carcinogens. To detect genotoxic activity, cells are treated with a test chemical in the beginning of growth phase and then allowed to undergo several rounds of cell division until further growth is inhibited as a result of confluence (initiation assay). To detect tumor-promoting activity, cells are repetitively treated with a test chemical beginning at sub-confluence and continuing beyond confluence achievement of cell growth (promotion assay). The assay endpoint is the formation of transformed foci. The judgments in the initiation assay and the promotion assay for a test chemical are integrated into the positive or negative call in the Bhas 42 CTA. That is, the chemical is positive in the Bhas 42 CTA if it is positive in either the initiation or promotion assay. The two Bhas 42 CTA methods, i.e., the 6-well method and the 96-well method, are fundamentally the same. The 6-well method follows the conventional procedures developed for focus transformation assays and the 96-well method has been developed to be utilized for high throughout automated applications in addition to manual procedures. The data obtained in the 6-well method are statistically analyzed by multiple comparison using the Dunnett test and those in the 96-well method are statistically analyzed by the chi-square test with Bonferroni adjustment. The respective protocols for the 6-well method and the 96-well method have undergone refinement and validation and are both available for use by other laboratories.

5.2 Module 5 — Predictive capacity

The extent of chemical carcinogenicity predictivity of the Bhas 42 CTA was demonstrated by validation studies of the 6-well and 96-well methods as described in sections 3.4.4 and 4.4.4 (Tables 16 and 49). The strong evidence for the predictive capacity of the Bhas 42 CTA was derived using 23 test chemicals in this validation effort. Those results are further supported by an in-house study conducted by HRI in which 98 chemicals were assessed with the Bhas 42 CTA. The study showed performance indices superior or similar to those of conventional genotoxicity assays [Sakai *et al.*, 2010]. As shown in Table 50, HRI applied the Bhas 42 CTA to 52 *in vivo* carcinogens and obtained positive calls from 38 carcinogens and negative calls from 14. In addition, 37 *in vivo* non-carcinogens were tested, of which 6 were positive and 31 were negative in the Bhas 42 CTA. (The other nine out of 98 chemicals were of unknown carcinogenicity). Table 50 presents a 2 x 2 contingency table of the 89 reported chemicals from which the performance indices of Bhas 42 CTA for predicting the carcinogenicity of chemicals were calculated (Table 51).

Table 50.

2x2 Contingency table of the results of Bhas 42 cell transformation assay in an in-house study by HRI [Sakai *et al.*, 2010]

		<i>In vivo</i> carcinogenicity		Total
		Carcinogen	Non-carcinogen	
Bhas 42 cell transformation assay	+	38	6	44
	-	14	31	45
Total		52	37	89

1
2
3 Table 51.

4 The performance of Bhas 42 cell transformation assay for the
5 prediction of chemical carcinogenicity [Sakai *et al.*, 2010]

6 Performance index	%
7 Concordance	78 (69/89)
8 Sensitivity	73 (38/52)
9 Specificity	84 (31/37)
10 Positive predictivity	86 (38/44)
11 Negative predictivity	69 (31/45)
12 False negative	27 (14/52)
13 <u>False positive</u>	<u>16 (6/37)</u>

14
15
16
17 **5.3 Quantification of transformation frequency**

18
19 The endpoint of the Bhas 42 CTA, i.e., focus formation, is quantified by the number of
20 transformed foci per well in the 6-well method, and by the number of wells having
21 transformed foci in the 96-well method. This latter 96-well micro-plate method of
22 quantifying transformation frequency is considerably more advantageous. In the 6-well
23 method, all foci appearing in all wells have to be individually judged as to whether they
24 are transformed or not and then the number of transformed foci/well is recorded. In
25 contrast, in the 96-well method a given well is counted if it contains at least one
26 transformed focus irrespective of the presence of other clearly transformed, marginally
27 transformed and/or non-transformed foci in the same well: the transformation
28 frequency is then expressed as the number of wells having transformed foci/number of
29 wells treated with a test chemical at a given concentration. This latter method has
30 proven to be much more expedient and efficient, thereby holding out promise for
31 adaptation to a high throughput system for carcinogenesis screening. Based on the data
32 generated in this study, it seems that the different scoring approaches do not affect the
33 final results.

34
35
36
37 **5.4 Negative and positive controls**

38
39 Comparisons of the transformation frequencies for negative and positive controls,
40 respectively, (Figures 3, 4, 17 and 18) demonstrate greater comparability of results as
41 well as similar standard deviations between laboratories in the 96-well method than in
42 the 6-well method. The improvement of between-laboratory reproducibility observed in
43 the 96-well method validation study could be methodology related and/or experience
44 related, among other possibilities. Thus, the more straightforward method of
45 quantifying transformation frequency in the 96-well method, as described above in
46 section 5.3 might be one contributing factor, while the accumulated experience of Bhas
47 42 CTA by the participating laboratories may be another. Nevertheless, it is clear that
48 the 96-well micro-plate method offers advantages that warrant additional follow up.

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50
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Table 52.
Continued.

Compound	6-well pre-validation study						6-well validation study						96-well validation study						Carcinogenicity <i>in vivo</i>						
	Laboratory			OA			Laboratory			OA			Laboratory			OA				Phase II study					
	A	B	C	D	E	F	I	II	III	IV	V	VI	1	2	3	4	OA	1		2	3	1	2	3	OA
Ampicillin sodium																									
Initiation																									
Promotion																									
Transformation																									
Anthracene																									
Initiation																									
Promotion																									
Transformation																									
L-Ascorbic acid																									
Initiation																									
Promotion																									
Transformation																									
Caffeine																									
Initiation																									
Promotion																									
Transformation																									
Caprolactam																									
Initiation																									
Promotion																									
Transformation																									
Eugenol																									
Initiation																									
Promotion																									
Transformation																									
D-Mannitol																									
Initiation																									
Promotion																									
Transformation																									
Phenanthrene																									
Initiation																									
Promotion																									
Transformation																									
Phorbol ^m																									
Initiation																									
Promotion																									
Transformation																									
Pyrene																									
Initiation																									
Promotion																									
Transformation																									

^a References: [1] *Yanaka et al. (2009)*; [2] *Sakai et al. (2001)*; [3] *Sakai et al. (in preparation)*

^b Lab A, Lab IV and Lab 2 represent the same laboratory between the validation studies. Likewise, Lab B, Lab III and Lab 1 represent the same laboratory; Lab C, Lab II and Lab 4 represent the same laboratory; Lab F, Lab I and Lab 3 represent the same laboratory.

^c Overall judgment: Judgment by majority rule among laboratories.

^d Judgment in the initiation assay.

^e Judgment in the promotion assay.

^f Judgment in the Bhas 42 CTA into which judgments in initiation and promotion assays were integrated.

^g Tumor-promoter.

^h [Lijinsky et al., 1992]

ⁱ Equivocal; there was a statistically significant increase in transformation frequency at only one concentration.

^j Incomplete; unable to judge the results due to inadequate dosing.

^k *o*-Toluidine hydrochloride was tested in the 6-well method validation study, and free *o*-toluidine was tested in the 96-well method validation study.

^l <21.3 mM and <29.2 mM, negative in Lab 1 and 3, respectively; 25.5 mM< and 32.7 mM<, positive in Lab 1 and Lab 3

^m Phorbol was unable to be judged for its transformation activity in the Bhas 42 CTA since the concentrations used were considered to be too low (see text, 4.3.5.15).

^{*} Carcinogenicity in mice (leukaemogenic action) is positive or negative depending on the strains [Berenblum and Vlasta, 1970; Armuth, 1976]. Tumor-promoting activity in mouse skin is positive or negative depending on the strains [Baird and Boutwell, 1971; Slaga et al., 1976, 1980].

5.5 Similarity of results between the 6-well method and 96-well method

The results in all three validation studies on the Bhas 42 CTA, i.e., the pre-validation study of the 6-well method, the validation study of 6-well method and the validation study of 96-well method, all of which were carried out chronologically, are summarized and compared in Table 52. The data presented illustrate that the assay results derived from the 6-well method and those derived from the 96-well method were concordant with exceptions of *o*-toluidine and sodium arsenite.

Mezerein, which is a potent tumor-promoter in mouse skin, induced marked transformation of Bhas 42 cells in the promotion assay, both in the 6-well method and the 96-well method. In the initiation assay, the chemical resulted in positive calls with the 96-well method, while it caused a statistically significant increase of transformation frequency only at the highest concentration tested in two of three laboratories with 6-well method (Tables 9 and 37; Figures 10 and 33). These results of mezerein in the initiation assay appear to be discordant between the 6-well method and the 96-well method (see Table 52). However, this discordance may have simply resulted from the different maximum concentrations of mezerein used in the 6-well method validation study (0.01 µg/mL) and the 96-well method validation study (0.1 µg/mL), which were pre-assigned by the VMT. Rather, those data show the complete agreement of results in the initiation assay between both methods, provided the results are compared at the same doses. That is, (a) in the 6-well method, at a concentration of 0.01 µg/mL, the numbers of transformed foci in the initiation assay were statistically significantly increased in the two of three laboratories, and in the other laboratory the number was increased but the increase was not statistically significant (Table 9 and Fig. 10); (b) in the 96-well method, the increase of transformation frequency in the initiation assay started at 0.0125 µg/mL in one of the laboratories and at 0.03 µg/mL in the other (Table 37 and Fig. 33). Meanwhile, it was also reasoned that these restricted positive results obtained in the initiation assay could be explained by the possible failure to thoroughly remove residual mezerein present in the treatment medium used for the initiation stage of the assay, thereby continuing to remain available to the target cells during the promotion stage. Thus, the transformation positives observed may not have been “initiation positives” per se, but rather “promotion positives” that were incorrectly assumed to have been generated during the initiation stage.

5.6 Discrepancy of results between the 6-well method and 96-well method

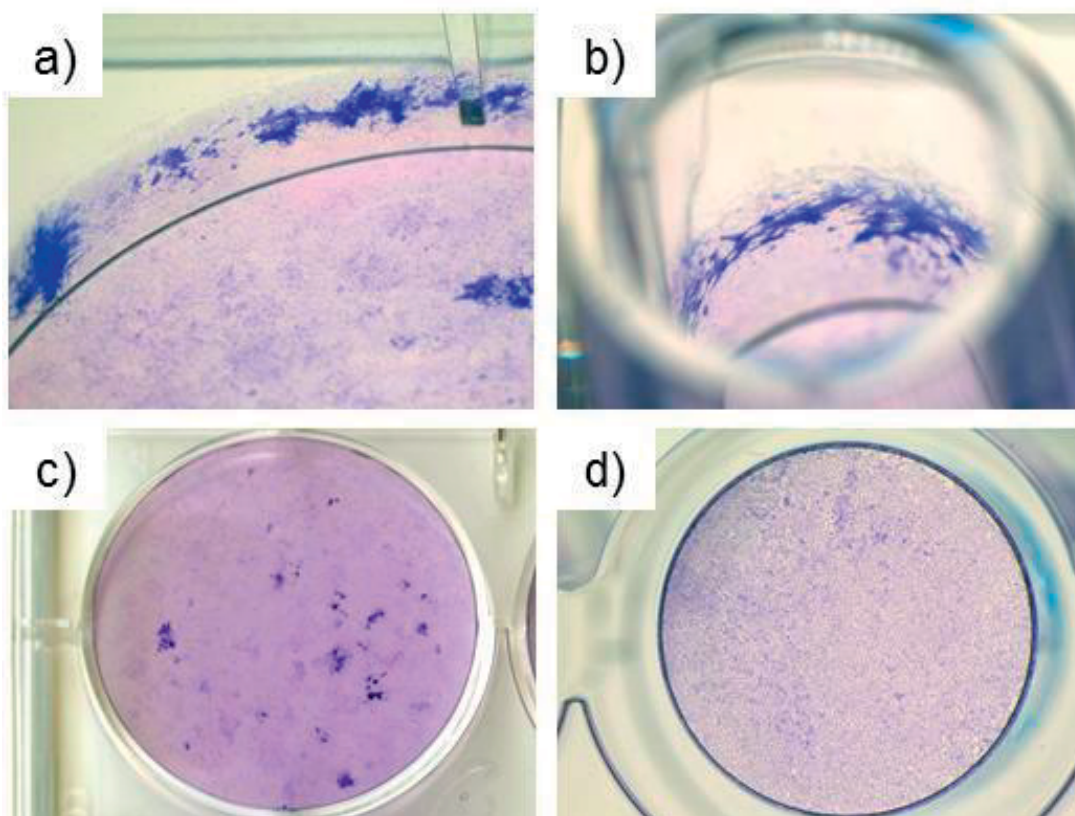
o-Toluidine hydrochloride was positive in two of three laboratories in the 6-well method validation study, but free *o*-toluidine was negative in all four laboratories in the 96-well method validation study. HRI, which was the lead laboratory, found that *o*-toluidine often yielded varied or equivocal results, i.e., was borderline positive/negative in the Bhas 42 CTA. According to HRI's repeated experiments, *o*-toluidine, including its hydrochloride, is always negative in the initiation assay, but its results in the promotion assay are inconsistent, although more often tended to be negative. This trend is similar between the 6-well method and the 96-well method and also between hydrochloride and free amine..

In the 6-well method validation study, sodium arsenite was positive in the promotion assay in all the three laboratories, although results were ambiguous in the initiation

1 assay. In the 96-well method validation study, however, the chemical was negative in
2 both the initiation and promotion assays. The consistent sodium arsenite results
3 obtained in the HRI in-house studies corroborated the positive results in the promotion
4 assay of the 6-well method [Muramatsu *et al.*, 2009; Sakai *et al.*, 2010] and the
5 predominantly negative results in the promotion assay of the 96-well method. According
6 to the information from HRI, sodium arsenite produces transformed foci mainly on the
7 side-wall rather than on the bottom of the 6-well and 96-well micro-plates, and this
8 tendency is more prevalent in the 96-well method (Fig. 44). Only the foci that formed
9 on the bottom of the well were counted in the 96-well method because it is difficult to
10 microscopically discern the morphology of foci attached to the side-wall of such a narrow
11 well, as is the case in a 96-well micro-plate. Since such difficulties were not encountered
12 in the 6-well method, the foci that formed both on the side-wall and the bottom of the
13 well were counted. Consequently, sodium arsenite was judged as negative in the 96-well
14 method. The other chemicals, including MCA and TPA, occasionally produced only a few
15 foci on the side-wall, and therefore were not problematic. Currently there is no available
16 information suggesting the mechanism that underlies the affinity for sodium
17 arsenite-induced foci to preferentially collect on the side-wall of the micro-plates.
18
19
20

6-well method

96-well method



21
22
23 Fig. 44. Foci produced by 0.25 µg/mL of sodium arsenite with the 6-well method and the
24 96-well method (photos contributed by HRI).
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5.7 Phorbol and TPA

Phorbol, the parent alcohol of TPA, was inactive at the concentrations tested in this validation study (Table 46 and Fig. 42). Because the chemical was expensive and scarce, the VMT limited the highest concentration to be $\leq 5 \mu\text{g/mL}$, which was too low a concentration to achieve cytotoxicity, thereby precluding a valid judgment of transformation activity in the Bhas 42 cells. However, it was shown that the promotion activity of phorbol was much weaker, even if active, than that of TPA in the Bhas 42 CTA. That was likewise the case in the two-stage carcinogenesis system of mouse skin [Baird and Boutwell, 1971; Slaga et al., 1976]. In the Bhas 42 CTA, TPA was active at around $0.01 \mu\text{g/mL}$, but phorbol was inactive at $5 \mu\text{g/mL}$ in the promotion assay (Tables 29 and 46, and Figures 25 and 42).

6 Recommended protocol

The protocols attached as Annex 2, and Annexes 6, 7 and 8 had been prepared for individual validation studies on the Bhas 42 CTA, the validation study of 6-well method, and the pre-validation, validation phase I and validation phase II studies of 96-well method, respectively. The integrated recommended protocol for the Bhas 42 CTA (protocol Ver. 5) has been developed on the basis of the outcome of this validation study, which was also partly complemented with historical data derived by HRI. Protocols for the 6-well method and the 96-well method have been combined into a single recommended protocol to reduce redundancy. The definitive recommended protocol is presented in Annex 12.

Assay acceptance criteria are essential components of any assay protocol. Although initially absent in the protocol for the 6-well method validation study (Annex 2), appropriate assay acceptance criteria have since been defined, emulating those for the 96-well method (see below). The acceptance criteria for the negative and positive controls have been determined based on the results of this validation study and historical data derived by HRI (see Table 57 of Annex 13). The acceptance criteria for the 6-well method provided in the recommended protocol are as follows (Annex 12, protocol Ver. 5):

The following assay acceptance criteria must be fulfilled for a given assay to be considered valid. The initiation or promotion assay is repeated independently, as needed, to satisfy the assay acceptance criteria.

1. Concurrent cell growth assay

- When contamination or technical problems are observed, two undamaged wells are necessary at the minimum for each concentration for cell growth assessment.

2. Transformation assay

- Initiation assay and promotion assay
 - If a given chemical concentration results in excessive cell death and/or cells fail to reach confluence at the end of transformation assay because of chemical toxicity, the concentration is not valid for transformation assessment and is excluded from focus-counting, statistical analysis and judgment, and “toxicity” is recorded in the data sheet.
 - When contamination or technical problems are observed, if, for a given concentration, the number of damaged wells is two or more, the concentration is not considered valid for transformation assessment and is excluded from focus-counting, statistical analysis and judgment. In such cases, “contamination”, “accident”, “technical error”, etc. are recorded in the data sheet.
 - For the positive control, there must be a statistically significant increase in the number of transformed foci per well compared to the corresponding negative control (one-sided t-test or Aspin-Welch test, $p < 0.05$).
- Initiation assay
 - In the negative control, the number of transformed foci must be ten or less per well.
 - If there is no statistically significant increase in the number of transformed foci at any dose (in case of negative results), four valid test

1 chemical concentrations are necessary, at a minimum, to accept the
2 transformation assay for evaluating a chemical. Those concentrations
3 should include at least one concentration near the NOEL and three
4 concentrations in the range between the NOEL and IC₉₀ in the concurrent
5 cell growth assay.

- 6 • Promotion assay
 - 7 ▪ In the negative control, the number of transformed foci must be twelve or
8 less per well.
 - 9 ▪ If there is no statistically significant increase in the number of
10 transformed foci at any dose (in case of negative results), four valid test
11 chemical concentrations are necessary, at a minimum, to accept the
12 transformation assay for evaluating a chemical. Those concentrations
13 should include at least one concentration near the NOEL and two
14 concentrations in the range of growth enhancement when the chemical
15 enhances cell growth (increases cell density in the cell growth assay).
16 When the chemical does not induce growth enhancement but induces
17 cytotoxicity instead, the cytotoxicity observed in the concurrent cell growth
18 assay may not be similar to that observed in the transformation assay,
19 since the durations of chemical exposure to the cells are different between
20 the cell growth assay (3 days) and the transformation assay (10 days).
21 Consequently, chemical toxicity is sometimes accumulated over the 10 day
22 duration of the transformation promotion assay and the otherwise valid
23 plates may be lost because of chemical toxicity. In such cases, four valid
24 plates are also necessary in the concentration range where cells are not
25 killed and are confluent at the end of the transformation assay. If excessive
26 toxicity is encountered, it may be necessary to repeat the assay in a lower
27 concentration range.

28
29
30 The assay acceptance criteria for the transformation frequency of negative and positive
31 controls in the 96-well method have been made more general in the recommended
32 protocol in order to avoid any presuppositions that may result from use of specific assay
33 components, materials and supplies. In this validation study, the batch of FBS used had
34 been pre-screened. For the validation phase I and II studies, the acceptance criteria for
35 the transformation frequency of negative and positive controls were based upon
36 absolute numbers, which, in turn, were based on the results obtained from DMSO, MCA
37 and TPA in the pre-validation phase, all of which presupposed the use of the specific lot
38 of FBS (GIBCO; Lot no. 1391481) (see 4.2.6).

39 The transformation frequency in any CTA is known to be affected by FBS batch [Sakai
40 et al., 2002]. Generally, FBS lots that induce lower transformation frequencies in the
41 vehicle control tend to produce the fewer numbers of transformed foci in the
42 chemically-treated cultures. As shown in Table 57 of Annex 13, Moregate FBS resulted
43 in lower transformation frequencies of Bhas 42 cells than GIBCO FBS in both negative
44 and positive controls, in both the initiation and promotion assays and in both the 6-well
45 and 96-well methods. Meanwhile, most FBS lots tested have been found to be usable
46 with the Bhas 42 CTA. Actually the absolute numbers of transformation frequency of
47 negative and positive controls are less important in the transformation assays than the
48 relative transformation frequencies of the positive and negative controls in each assay.
49 Therefore, the acceptance criteria for positive controls in the 96-well method have been
50 amended and no longer consider the absolute numbers of wells having transformed
51 foci/plate. Rather, the recommended acceptance criteria now rely upon statistically
52 significant increases in transformation frequency compared to the corresponding

1 negative control. The acceptance criteria for negative controls, which were established
2 in 96-well method protocols, Ver. 3 (for the phase I study) and Ver. 4 (for the phase II
3 study) (initiation assay, ≤ 15 wells/plate; promotion assay, ≤ 20 wells/plate), were
4 incorporated into the recommended protocol. These criteria match the historical data
5 derived at HRI, which were produced using GIBCO FBS lot no. 13914814 and Moregate
6 FBS lot no. 7825120 (see Table 57 of Annex 13).

7 The acceptance criteria provided in the recommended protocol for the 96-well method
8 are as follows (Annex 12, protocol Ver. 5):
9

10
11 The following criteria must be fulfilled for an assay to be considered valid. The
12 initiation or promotion assay is repeated independently, as needed, to satisfy the
13 assay acceptance criteria.
14

15 **1. Concurrent cell growth assay**

- 16 • When contamination or technical problems are observed, four undamaged
17 wells are necessary at the minimum for each concentration for cell growth
18 assessment.
19

20 **2. Transformation assay**

- 21 • Initiation assay and promotion assay
 - 22 • If a given chemical concentration results in excessive cell death and/or
23 cells fail to reach confluence at the end of transformation assay because of
24 chemical toxicity, the concentration is not considered valid for
25 transformation assessment and is excluded from focus-counting, statistical
26 analysis and judgment, and “toxicity” is recorded in the data sheet.
 - 27 • When contamination or technical problems are observed, such that for a
28 given concentration, the number of damaged wells is ≥ 7 , the concentration
29 is not considered valid for transformation assessment and is excluded from
30 focus-counting, statistical analysis and judgment. In such cases,
31 “contamination”, “accident”, “technical error”, etc. are recorded in the data
32 sheet.
 - 33 • For the positive control, there must be a statistically significant increase
34 in the proportion of wells having transformed foci compared to the
35 corresponding negative control (one-sided chi-square test, $p < 0.05$,
36 upper-sided).
- 37 • Initiation assay
 - 38 • In the negative control, the number of wells having transformed foci must
39 be ≤ 15 wells/plate (if there exist damaged wells, $\leq 15.625\%$ of undamaged
40 wells).
 - 41 • If there is no statistically significant increase in the proportion of wells
42 having transformed foci at any dose (in case of negative results), four valid
43 test chemical concentrations are necessary, at a minimum, to accept the
44 transformation assay for evaluating a chemical. Those concentrations
45 should include at least one concentration near the NOEL and three
46 concentrations in the range between the NOEL and IC_{90} in the concurrent
47 cell growth assay.
- 48 • Promotion assay
 - 49 • In the negative control, the number of wells having transformed foci must
50 be ≤ 20 wells/plate (if there exist damaged wells, $\leq 20.833\%$ of undamaged
51 wells).

- 1 · If there is no statistically significant increase in the proportion of wells
2 having transformed foci at any dose (in case of negative results), four valid
3 test chemical concentrations are necessary, at a minimum, to accept the
4 transformation assay for evaluating a chemical. Those concentrations
5 should include at least one concentration near the NOEL and two
6 concentrations in the range of growth enhancement when the chemical
7 enhances cell growth (increases cell density in the cell growth assay).
8 When the chemical does not induce growth enhancement but induces
9 cytotoxicity instead, the cytotoxicity observed in the concurrent cell growth
10 assay may not be similar to that observed in the transformation assay,
11 since the durations of chemical exposure to the cells are different between
12 the cell growth assay (3 days) and the transformation assay (10 days).
13 Consequently, chemical toxicity is sometimes accumulated over the 10 day
14 duration of the transformation promotion assay and the otherwise valid
15 plates may be lost because of chemical toxicity. In such cases, four valid
16 plates are also necessary in the concentration range where cells are not
17 killed and are confluent at the end of the transformation assay. If excessive
18 toxicity is encountered, it may be necessary to repeat the assay in a lower
19 concentration range.
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23 The protocols for validation studies did not include instructions for FBS selection, since
24 the batch of FBS used had been screened by the lead laboratory, HRI, prior to the
25 studies, and the batch of FBS had been specified for each validation study of the 6-well
26 method (Moregate lot no. 7825120) and the 96-well method (GIBCO lot no. 13914814). A
27 section for the selection of FBS has been inserted into the recommended protocol.
28

29 The amendments in the recommended protocol relate only to (a) refinement of the assay
30 description, (b) the amalgamation of overlapping aspects of the 6-well method and the
31 96-well method, and (c) the standardization of assay acceptance criteria among FBS lots.
32 These revisions in no way undermine the validation of the Bhas 42 CTA.
33
34

7 Overall conclusion by the validation management team

The aim of the study was to validate the Bhas 42 CTA, in a formal inter-laboratory study, following the modular approach of Hartung et al. (2004) and concentrating on modules 1-5: test definition, within-laboratory reproducibility, transferability, between-laboratory reproducibility and predictive capacity.

Two protocol variations of the Bhas 42 CTA were evaluated in this validation study, i.e., that of the 6-well method and that of the 96-well method. In its guidance and assessment of the various phases of the study, the VMT noted minor differences in study design, conduct and assessment practices had been implemented in the two protocols, much of which were resultant subsequent improvements to previous methodologies. Despite these minor variances, the two methods were found to yield similar results. Experience gained from the 6-well method protocol helped guide the evolution of those procedures ultimately implemented in the 96-well method validation study. As a result, the latter protocol came to be more closely aligned with internationally acknowledged test method validation criteria, and consequently provided highly credible assay results that satisfied the established validation criteria.

Table 53 summarizes the conclusion by the VMT on the assessment of 6-well method and 96-well method in the Bhas 42 CTA.

Table 53 Conclusions of the Validation Management Team for the different modules

6-Well method			
Module		Summary & conclusion	
Module 1	Test definition	<ul style="list-style-type: none"> • Clear definition of the scientific basis of the method • Description of the endpoint induced by genotoxic and non-genotoxic mechanisms • Protocol available with clear definition of a valid study (acceptance criteria have been defined based on the data derived from this validation study and historical data derived by HRI, following those of the 96-well method) 	yes
Module 2	Within-laboratory reproducibility	<p>The within-laboratory reproducibility was shown to be satisfactory in all laboratories for</p> <ul style="list-style-type: none"> • the vehicle control • the positive control <p>and in the laboratories which had retrospective data in addition to those data obtained in this prospective validation study for</p> <ul style="list-style-type: none"> • the test chemicals* 	yes
Module 3	Transferability	<p>The test method is transferable between laboratories:</p> <ul style="list-style-type: none"> • basic cell cultivation experience is needed • training in procedures and transformed-foci identification is essential • photo catalog of foci is available 	yes
Module 4	Between-laboratory reproducibility	<p>The between-laboratory reproducibility was shown to be satisfactory for</p> <ul style="list-style-type: none"> • the vehicle control • the positive control • the test chemicals** 	yes

1
2

Table 53 Continued

6-Well method			
Module		Summary & conclusion	
Module 5	Predictive capacity	The capacity to predict chemical carcinogenicity is satisfactory in • this validation study (12 chemicals) • the in-house HRI study in which 98 chemicals were tested	yes
96-Well method			
Module		Summary & conclusion	
Module 1	Test definition	• Clear definition of the scientific basis of the method • Description of the endpoint induced by genotoxic and non-genotoxic mechanisms • Protocol available with clear definition of a valid study (some acceptance criteria have been refined based on the data derived from this validation study and historical data derived by HRI)	yes
Module 2	Within-laboratory reproducibility	The within-laboratory reproducibility was shown to be satisfactory in all laboratories for • the vehicle control • the positive control • the test chemicals	yes
Module 3	Transferability	The test method is transferable between laboratories • basic cell cultivation experience is needed • training in procedures and transformed-foci identification is essential • Photo catalog of foci is available	yes
Module 4	Between-laboratory reproducibility	The between-laboratory reproducibility was shown to be satisfactory for • the vehicle control • the positive control • the test chemicals	yes
Module 5	Predictive capacity	The capacity to predict chemical carcinogenicity is satisfactory in • this validation study	yes

3 * The results of one chemical (*o*-toluidine) obtained by HRI were not concordant
4 between its in-house study and this validation study but the chemical was considered
5 by the VMT to be equivocal in the Bhas 42 CTA.

6 ** The results submitted for two out of twelve test chemicals were not concordant
7 between the naive and experienced laboratories.

8
9

10

11 In addition to the conclusions drawn by the VMT regarding satisfaction of the modules
12 cited, the VMT also considered the following issues noteworthy.

13

14 The Bhas 42 CTA has the following advantages:

- 15 1. The culture period (3 weeks) is shorter than that of a conventional BALB/c 3T3 CTA
16 (6 weeks).
- 17 2. Less labor and reduced amounts of materials are necessary compared with those of
18 the conventional CTAs.

- 1 3. Either the 6-well method or the 96-well method can be used with equal assurance.
- 2 4. Bhas 42 CTA can detect both tumor-initiators and tumor-promoters and discriminate
- 3 them depending on a protocol modification.

4
5 The validation studies on the Bhas 42 CTA have verified the following:

- 6 1. The 6-well method and the 96-well method produced similar results.
- 7 2. The Bhas 42 CTA is transferable between laboratories.
- 8 3. The Bhas 42 CTA is reliable, i.e., is reproducible within and between laboratories.
- 9 4. The Bhas 42 CTA is relevant, i.e., is capable of correctly predicting chemical rodent
- 10 carcinogenicity.

11
12 The in-house HRI study in which the Bhas 42 CTA was applied to 98 chemicals has

13 shown the following:

- 14 1. The Bhas 42 CTA could detect a considerable number of Ames-negative and Ames
- 15 discordant carcinogens.
- 16 2. The performance of the Bhas 42 CTA to predict the carcinogenicity of chemicals was
- 17 superior or equal to that of conventional genotoxicity assays.

18
19 In view of the results reported herein and the conclusions asserted by the VMT, we

20 consider that the overall accuracy of detecting potential chemical carcinogens would be

21 improved and the tumor-promoting activity of chemicals could be better predicted by

22 incorporating the Bhas 42 CTA into the battery of *in vitro* assays customarily employed.

23 The Bhas 42 CTA can also conceivably help reduce the number of animals routinely

24 used for *in vivo* chronic carcinogenicity testing by identifying likely positive *in vivo*

25 carcinogens, and thus allow for the possible waiving of subsequent animal testing for

26 carcinogenicity. In addition, the 96-well method has the potential to be easily adapted

27 for automation with further relatively minimal refinement. The VMT recommends that

28 an OECD Test Guideline for the Bhas 42 CTA to be drafted to encourage regulatory

29 acceptance and universal implementation of this validated, reliable, cost- and

30 time-efficient test method.

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Annex 1

Table 54.

Supplier, catalog number, and lot number of test chemicals in the 6-well method validation study.

Chemical	CAS no. ^a	Supplier	Catalog no.	Lot no.	
2-Acetylaminofluorene	53-96-3	Sigma	A7015	076K0677	
Cadmium chloride	10108-64-2	Aldrich	439800	11869TE	
Dibenz[<i>a,h</i>]anthracene	53-70-3	Wako	040-00901	CEL1090	
Lithocholic acid	434-13-9	Sigma	L6250	022K1392	
Methapyrilene HCl	135-23-9	Sigma	M9125	037F09291	
Mezerein	34807-41-5	Sigma	M5518	113K0993	
Sodium arsenite	7784-46-5	Wako	191-01241	LTM0420	
<i>o</i> -Toluidine HCl	636-21-3	Aldrich	327115	12915DD	
Anthracene	120-12-7	Aldrich	331481	12223EE	
L-Ascorbic acid	50-81-7	Sigma	A5960	106K0053	
Caffeine	58-08-2	Aldrich	C0750	126K0705	
D-Mannitol	69-65-8	Aldrich	M4125	086K0103	
3-Methylcholanthrene	56-49-5	Aldrich	213942	01115HE	Positive control
TPA ^b	16561-29-8	Sigma	P1585	037K1322	Positive control

^a Chemical Abstract Service registry number.

^b 12-*O*-tetradecanoylphorbol-13-acetate.

Annex 2

Protocol of Bhas 42 Cell Transformation Assay (2007.10.26) 6-Well Method Ver. 2

This protocol is described for the validation study of Bhas 42 cell transformation assay. Used materials and reagents with their catalogue numbers are listed in annex 2.1.

I. Preparation

1. Materials

1) Cell line

Bhas 42 cells (v-Ha-*ras*-transfected Balb/c 3T3 clone A31-1-1 cells)^{1,2}: Free from bacteria, fungi and mycoplasma, supplied from HRI (Hatano Research Institute, Food and Drug Safety Center, Japan) Cell Bank.

2) Media

MEM: Minimum essential medium with 2.2 g/L NaHCO₃.

DMEM/F12: Dulbecco's modified Eagle's medium/F12 with 1.2 g/L NaHCO₃.

FBS: Fetal bovine serum should be selected showing a low spontaneous focus formation and a high focus formation in the positive control.

PS: Penicillin G sodium (10000 units/mL) and streptomycin sulfate (10 mg/mL).

M10F: MEM + 10% FBS + 1% PS (M10F is used for the expansion of provided cells, cell storage, and the first culture after thawing.).

DF5F: DMEM/F12 + 5% FBS + 1% PS (DF5F is used for routine passages, cell growth assays and transformation experiments.).

3) Chemicals

Test chemicals and solvent/vehicle: Test chemicals are dissolved or suspended in an appropriate solvent or vehicle and diluted with the solvent/vehicle to each individual concentration before added to culture media so that all chemical treatment media contain an equal concentration of the solvent/vehicle. The solvent/vehicle should neither interact with the test chemicals nor affect survival and focus formation of the cells. The final concentration of the solvent/vehicle in the medium is $\leq 5\%$ with distilled water and $\leq 0.1\%$ with DMSO (permissible up to 0.5% when a test chemical does not dissolve).

Negative and positive controls: Negative and positive control cultures are included in each experiment. The solvent/vehicle for a test chemical is used as the negative control. A known initiator, 3-methylcholanthrene (MCA: final concentration of 1 $\mu\text{g/mL}$), and a promoter, 12-*O*-tetradecanoylphorbol-13-acetate (TPA: final concentration of 50 ng/mL), are used for the positive controls.

Blank control: Wells added with the medium alone are prepared for the blank control in cell growth assays.

4) Fixatives and staining solutions

Formalin (37% formaldehyde): Used for fixing cells in cell growth assays.

CV solution (0.1% crystal violet solution): One gram of crystal violet is dissolved in 50 mL of ethanol. The ethanol solution is diluted with distilled water to make a final concentration of 0.1%, and used for staining cells in

cell growth assays.

Extraction solution (containing 0.02 mol/L HCl and 50% ethanol): Used for extracting CV from the stained cells.

Methanol: Used for fixing cells in transformation assays.

5% Giemsa solution: Used for staining cells in transformation assays.

5) Culture vessels

φ100-mm Petri dishes: Used for routine passage.

6-well plates: Used for cell growth assays and transformation assays.

2. Cell culture and storage

1) Cell passage

Bhas 42 cells are cultured in a humidified 5% CO₂ incubator at 37°C. The passage should be done at about 70% confluence of cell growth.

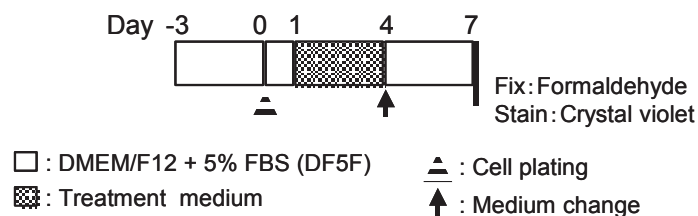
2) Cell freezing

The provided cells are expanded with M10F and cryopreserved at an early passage generation. The cells are suspended to make 5×10^5 cells/mL in cold M10F containing 5% DMSO. Aliquots of the cell suspension are transferred into freezing tubes, frozen in a -80°C deep freezer, and stored in liquid nitrogen. For each transformation experiment the cells are thawed from the frozen stock.

II. Experimental procedures

A. Initiation assay³⁾

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

The highest concentration is 5 mg/mL or 10 mM, whichever is the lowest^{4,5)}. In the case of chemicals difficult to dissolve, the highest concentration may be one or two level higher than the concentration showing the utmost solubility. Five or more concentration levels are set by an appropriate serial dilution factor such as 10, square root 10 or 2. In some chemicals the cell growth assay should be repeated in a narrower concentration range. Three wells of 6-well plates are used for each concentration.

2) Procedure of cell growth assay

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cell suspension is transferred at a volume of 10 mL to φ100-mm Petri dishes. (Cells with a high passage number can be used for the cell growth assay.)

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.2×10^4 cells/mL. The cell

suspension is transferred at a volume of 2 mL to each well of 6-well plates. Three wells are prepared for each treatment concentration.

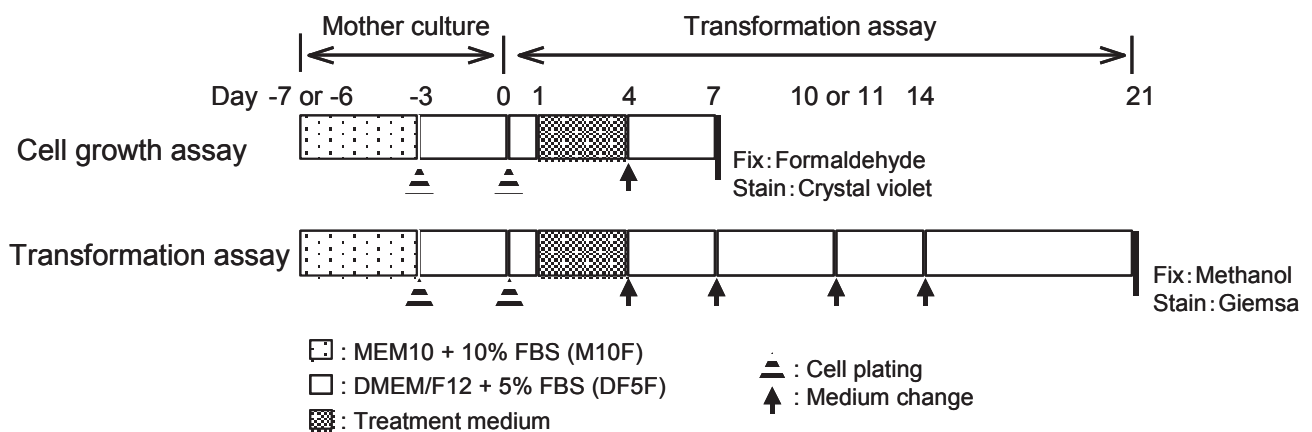
Day 1: Media containing various concentrations of test chemicals are prepared, and used for medium change. It is also practicable to prepare concentrated test chemical solutions and add them to wells without medium change.

Day 4: Medium is changed with fresh DF5F, and the cultivation is continued in the incubator.

Day 7: The cells are fixed with direct addition of 0.2 mL of formalin to the culture medium, or with 10 % formalin or methanol after medium is removed. After 30 min, the cells are washed and dried. The cells are stained with 1.5 mL of CV solution for 15 min, rinsed well with water and dried.

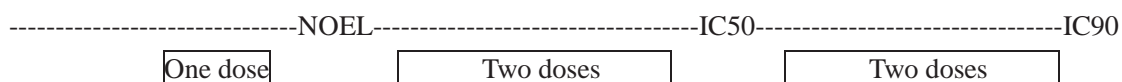
Measurement: The stained dye of each well is extracted with a constant volume of extraction solution (usually 2 mL) for 10 min, and the optical density of each extract is measured at a wavelength between 540 and 570 nm. Growth rates relative to the control culture are calculated from the absorbance.

2. Transformation assay



1) Test concentrations and used vessels

Five or more concentrations are set up based on the results of cell growth assays. These concentrations cover a range from highest toxicity (less than 20% survival compared to the control culture) to little or no toxicity. Ideally, one dose below NOEL, two doses between NOEL and IC50 and two doses between IC50 and IC90 are assessed in the initiation assay, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For low cytotoxic chemicals the highest concentration is 5 mg/mL or 10 mM, whichever is the lowest^{4,5}.

At least six wells of 6-well plates for the transformation assay, and at least three wells of 6-well plates for the concurrent cell growth assay are prepared at each concentration.

2) Procedure of transformation assay and concurrent cell growth assay

Day -7 or -6: Frozen stock cells are thawed, suspended in M10F and cultured in ϕ 100-mm Petri dishes at a volume of 10 mL.

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cells are cultured in ϕ 100-mm Petri dishes.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.2×10^4 cells/mL. The cell suspension is distributed into each well of 6-well plates at a volume of 2 mL for the transformation assay and the concurrent cell growth assay. One plate is prepared for each treatment concentration of transformation assay. Three wells are prepared for each treatment concentration of concurrent cell growth assay.

Day 1: Media containing various concentrations of test chemicals are prepared, and used for medium change. It is also practicable to prepare concentrated test chemical solutions and add them to wells without medium change.

Day 4: The medium is changed with fresh DF5F.

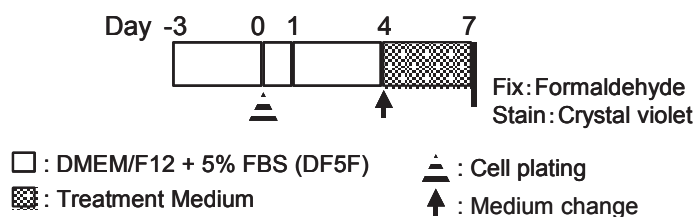
Day 7: The medium of transformation assay is changed with fresh DF5F. The cells for the concurrent growth assay are fixed and processed according to the procedure described above (II.A.1.2).

Day 10 or 11, and 14: The medium of the transformation assay is changed with fresh DF5F.

Day 21: The cells are fixed with methanol for 10 min and stained with 5% Giemsa solution for 30 min.

B. Promotion assay^{6,7)}

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

The highest concentration is 5 mg/mL or 10 mM, whichever is the lowest^{4,5)}. In the case of chemicals difficult to dissolve, the highest concentration may be one or two level higher than the concentration showing the utmost solubility. Five or more concentration levels are set by an appropriate serial dilution factor such as 10, square root 10 or 2. In some chemicals the cell growth assay should be repeated in a narrower concentration range. Three wells of 6-well plates are used for each concentration.

2) Procedure of cell growth assay

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cell suspension is transferred at a volume of 10 mL to ϕ 100-mm Petri dishes. (Cells with a high passage number can be used for the cell growth assay.)

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7×10^4 cells/mL. The cell suspension is transferred at a volume of 2 mL to each well of 6-well plates. Three wells are prepared for each treatment concentration.

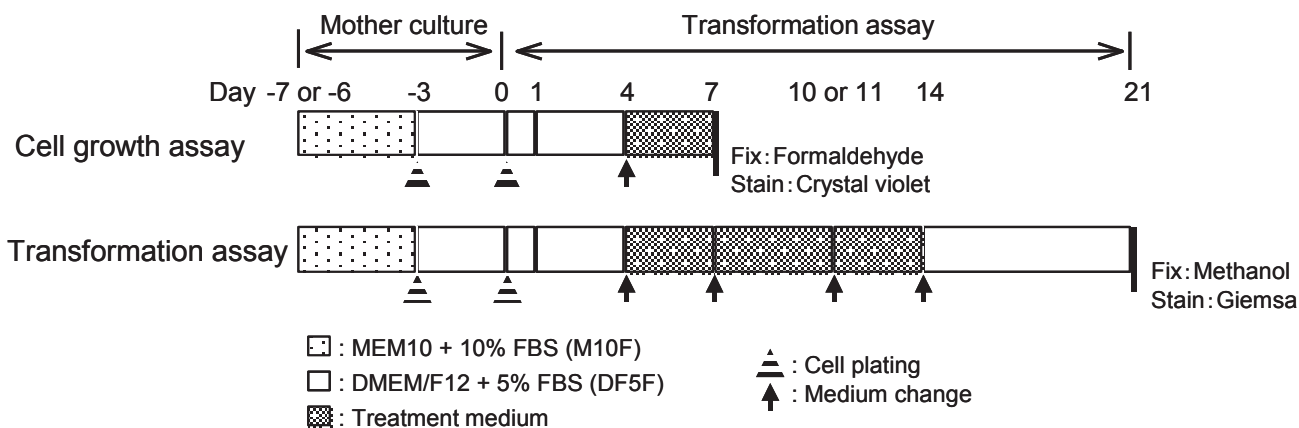
Day 4: Media containing various concentrations of test chemicals are prepared, and used for medium change.

Day 7: The cells are fixed with direct addition of 0.2 mL of formalin to the culture medium, or with 10 % formalin or methanol after medium is removed. After 30 min, the cells are washed and dried. The cells are stained with

1.5 mL of CV solution for 15 min, rinsed well with water and dried.

Measurement: The stained dye of each well is extracted with a constant volume of extraction solution (usually 2 mL) for 10 min, and the optical density of each extract is measured at a wavelength between 540 and 570 nm. Growth rates relative to the control culture are calculated from the absorbance.

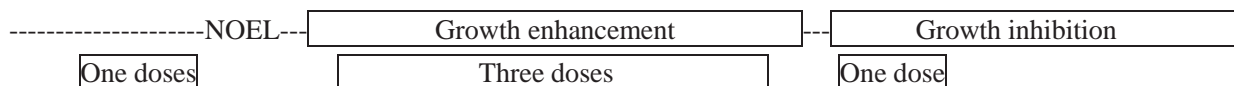
2. Transformation assay



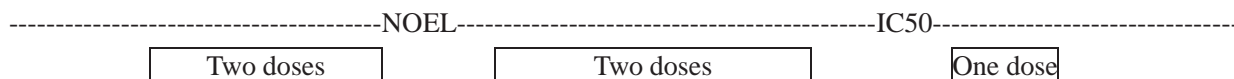
1) Test concentrations and used vessels

Five or more concentrations are set up based on the results of cell growth assays.

For the chemicals that exhibit marked growth enhancement, test concentrations are selected to cover from growth enhancement to little effect on cell growth. Ideally, one dose below NOEL, three doses in the range of growth enhancement, and one dose in the range of weak growth inhibition are assessed in the promotion assay, as follows;



For the chemicals that do not induce marked growth enhancement, test concentrations are selected ranging from a dose exhibiting below 50% growth level to that two or three levels lower than the non-effective concentration. Ideally, two doses below NOEL, two doses between NOEL and IC50 and one dose above IC50 are assessed, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For little cytotoxic compounds the highest concentration is 5 mg/mL or 10 mM, whichever is the lowest^{4,5}.

At least six wells of 6-well plates for the transformation assay, and at least three wells of 6-well plates for the concurrent cell growth assay are prepared at each concentration.

2) Procedure of transformation assay and concurrent cell growth assay

Day -7 or -6: Frozen stock cells are thawed, suspended in M10F and cultured in ϕ 100-mm Petri dishes at a volume of 10 mL.

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cells are cultured in ϕ 100-mm Petri dishes.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7×10^4 cells/mL. The cell suspension is distributed into each well of 6-well plates at a volume of 2 mL for the transformation assay and the concurrent cell growth assay. One plate is prepared for each treatment concentration of transformation assay. Three wells are prepared for each treatment concentration of concurrent cell growth assay.

Day 4: Media containing various concentrations of test chemicals are prepared, and used for medium change.

Day 7: The medium of transformation assay is changed with media containing test chemicals. The cells for the concurrent cell growth assay are fixed and processed according to the procedure described above (**II.B.1.2**).

Day 10 or 11: The medium of transformation assay is changed with media containing test chemicals.

Day 14: The medium is changed with fresh DF5F without test chemicals.

Day 21: The cells are fixed with methanol for 10 min and stained with 5% Giemsa solution for 30 min.

C. Focus count

Transformed foci are featured by the following morphological characteristics; (a) more than 100 cells, (b) spindle-shaped cells different from the contact-inhibited monolayer cells (spindle-shaped), (c) deep basophilic staining (basophilic), (d) random orientation of cells at the edge of foci (criss-cross), (e) dense multilayering of cells (piling up) and (f) invasive growth into the monolayer of surrounding contact-inhibited cells. There are transformed foci not prominent in some of these characteristics. The number of transformed foci is recorded for each well.

D. Judgment

The statistical analysis is carried out for increase in the number of transformed foci per well using the one-side Dunnett test with significant level of 5% ($p < 0.05$). The test chemical is judged positive (+), when there exist two or more doses that induce statistically significant increases in the number of transformation foci. When the statistically significant increase is observed in only one dose, the first judgment for the chemical is equivocal (+/-). Then the transformation assay together with the concomitant cell growth assay is repeated in a narrower concentration range including the positive dose of the first assay. The chemical is judged positive (+) when in the second assay the chemical again produces statistically significant increases in the number of transformed foci at one or more doses, and otherwise negative. When low transformation frequency in the positive control or high transformation frequency in the negative control is obtained, the experiment should be repeated using optimized test conditions. When biologically suspicious results are obtained, a re-experiment may be requested after discussion.

III. References

- (1) K. Sasaki, H. Mizusawa and M. Ishidate, Isolation and characterization of ras-transfected BALB/3T3 clone showing morphological transformation by 12-*O*-tetradecanoyl-phorbol-13-acetate, *Jpn. J. Cancer Res.* 79 (1988) 921-930.
- (2) K. Sasaki, H. Mizusawa, M. Ishidate and N. Tanaka, Establishment of a highly reproducible transformation assay of a ras-transfected BALB/3T3 clone by treatment with promoters, *Basic Life Sci.* 52 (1990) 411-416.
- (3) S. Asada, K. Sasaki, N. Tanaka, K. Takeda, M. Hayashi and M. Umeda, Detection of initiating activities of

chemicals using v-Ha-ras-transfected BALB/c 3T3 cells (Bhas 42 cells), *Mutat. Res.* 588 (2005) 7-21.

- (4) OECD Guidelines for the Testing of Chemicals. Test No. 473: *In vitro* Mammalian Chromosome Aberration Test.
- (5) OECD Guidelines for the Testing of Chemicals. Test No. 476: *In vitro* Mammalian Cell Gene Mutation Test.
- (6) K. Ohmori, K. Sasaki, S. Asada, N. Tanaka and M. Umeda, An assay method for the prediction of tumor promoting potential of chemicals by the use of Bhas 42 cells, *Mutat. Res.* 557 (2004) 191-202.
- (7) K. Ohmori *et al.*, Inter-laboratory collaborative study of cell transformation assay for tumor promoters using Bhas 42 cells by non-genotoxic carcinogen study group in Japan, *ATRA* 33 (2005) 1-21.

Annex 2.1 Materials and Reagents Used and their Catalogue Numbers (and Lot Numbers)

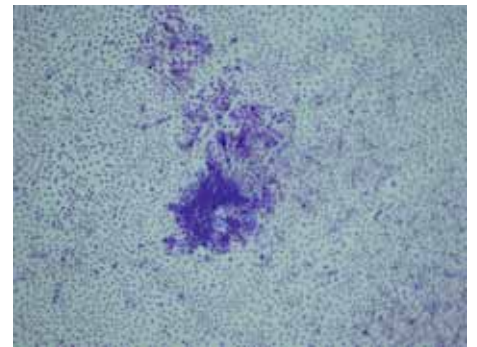
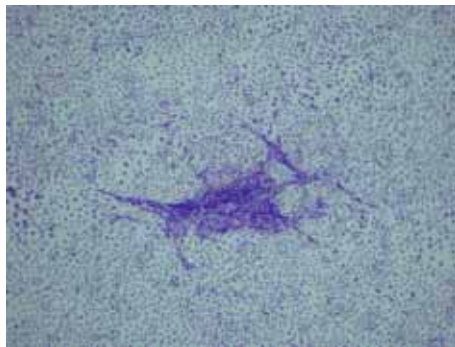
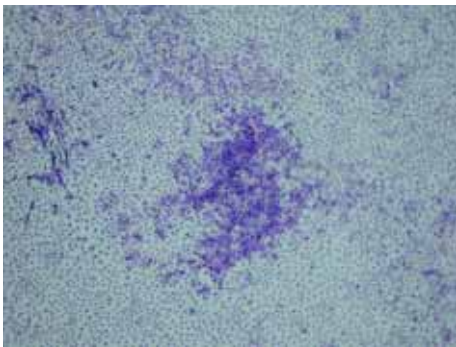
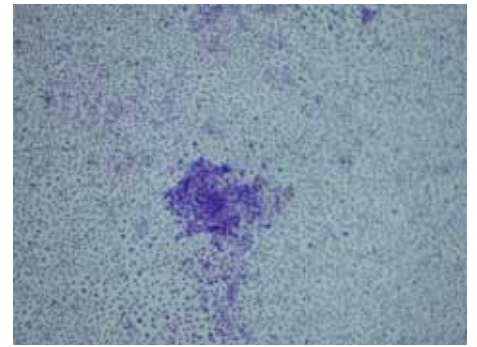
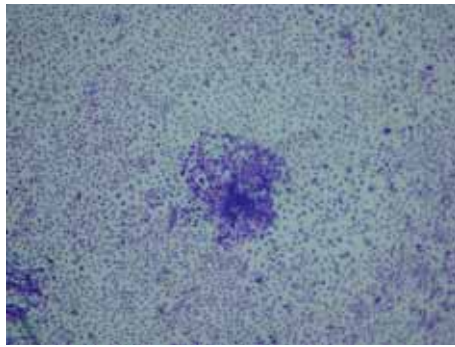
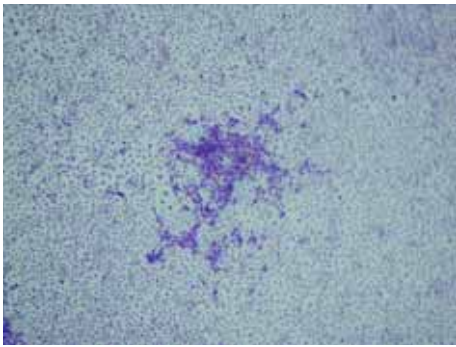
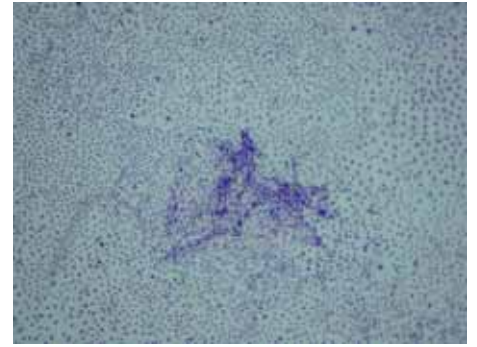
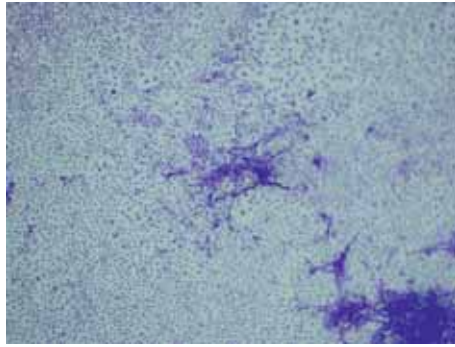
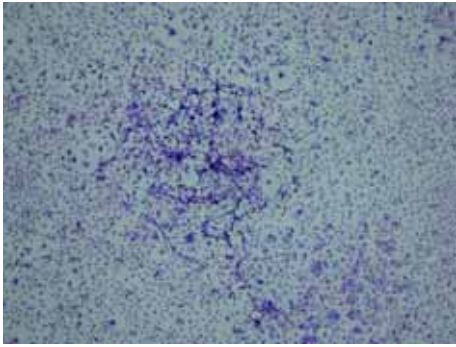
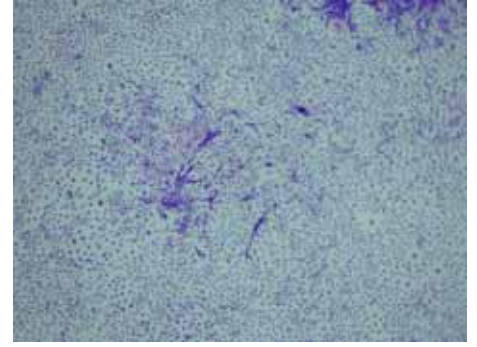
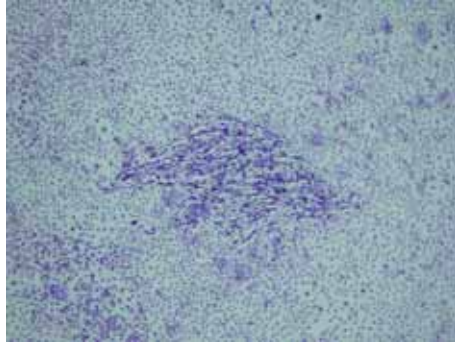
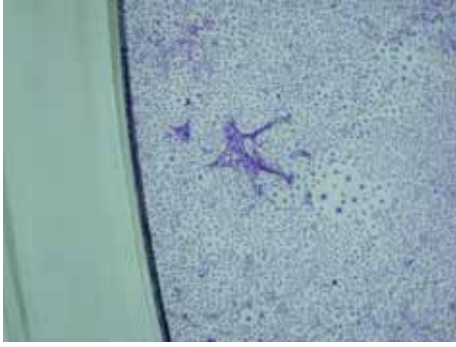
	Supplier	Cat. No.	Lot No.
Positive control			
3-Methylcholanthrene (MCA)	Aldrich	213942-100MG	01115HE
Phorbol 12-myristate 13-acetate (TPA)	SIGMA	P-1585-1MG	037K1322
Culture reagents			
Minimum Essential Medium	GIBCO	11095-080(500mL) (Japan) 11095-098(500mLx10) 31095-029 (Europe)	----
Fetal bovine serum	Moregate		7825120
Dulbecco's modified Eagle's medium/F12	GIBCO	11330-032(500mL) (Japan) 11330-057(500mLx10) (Japan) 31330-038 (Europe)	----
Penicillin(10000units/mL)-Streptomycin (10 mg/mL)	GIBCO	15140-122	----
Other reagent			
DMSO	SIGMA	D8418	----
Fixation and staining solutions			
Methanol	----	----	----
Giemsa solution *	MERK	1.09204	----
Giemsa solution (0.4%)*	Sigma	GS 500	----
Formalin (37% formaldehyde)	SIGMA	F8775	----
Crystal violet	SIGMA	C3886	----
Ethanol	----	----	----
HCl	----	----	----
Culture vessels			
6-well microplates	Falcon or Costar		----
	can be used		

*: Giemsa solution purchased from Merck is used after X20 dilution. Either Giemsa solution can be used.

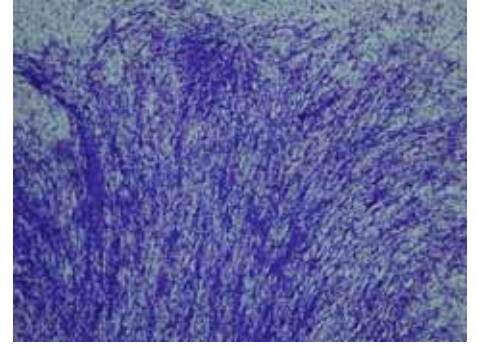
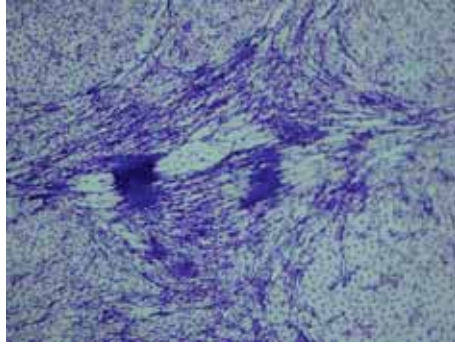
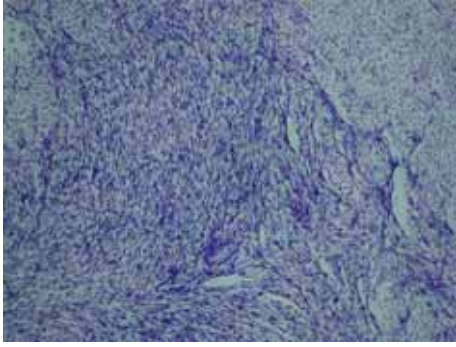
Annex 3

Photo Catalog for the Judgment of Foci in the Bhas 42 Cell Transformation Assay

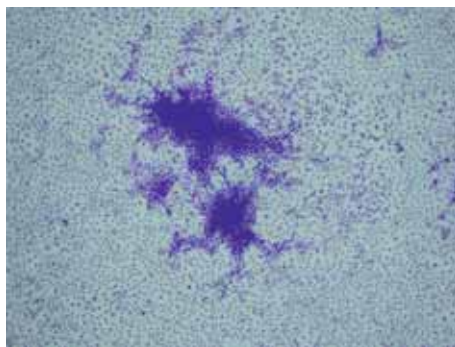
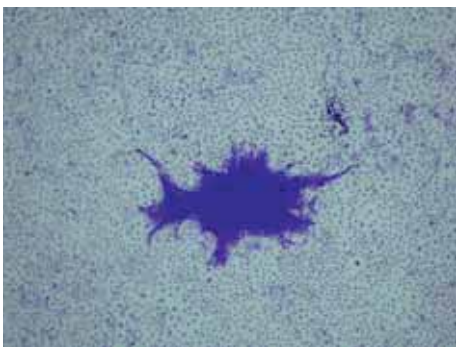
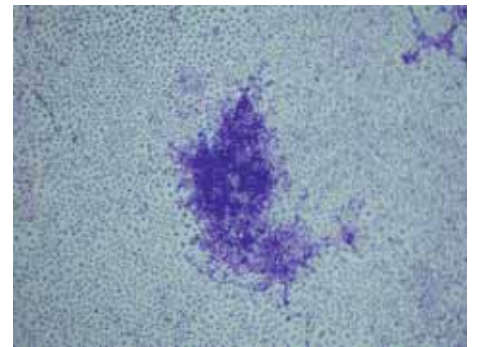
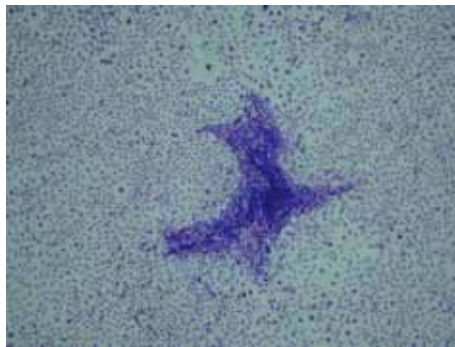
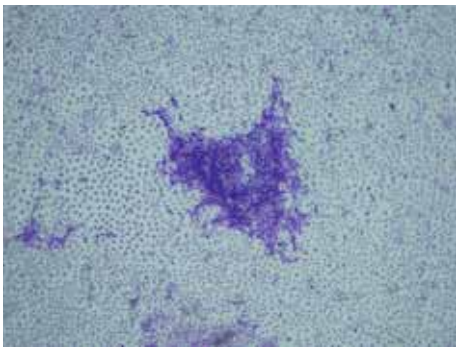
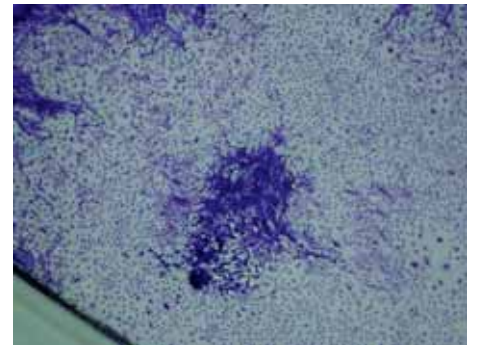
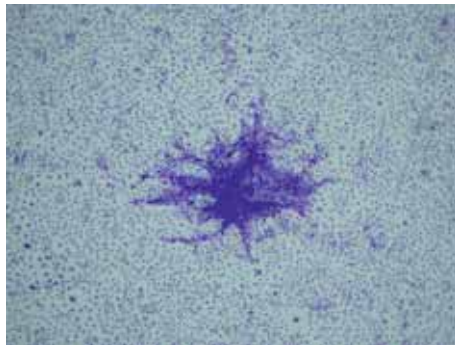
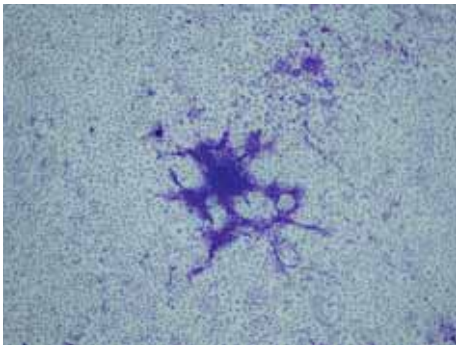
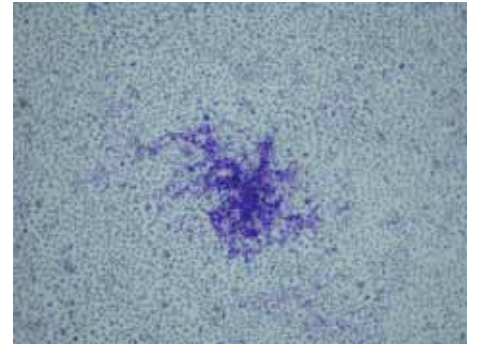
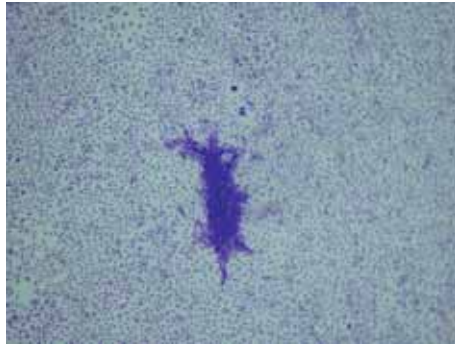
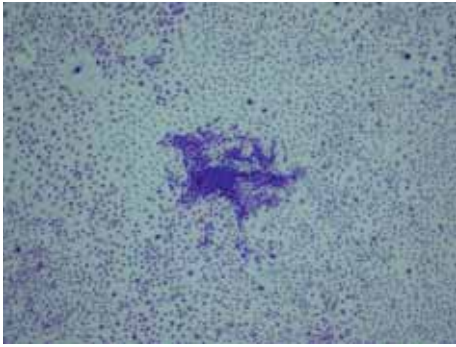
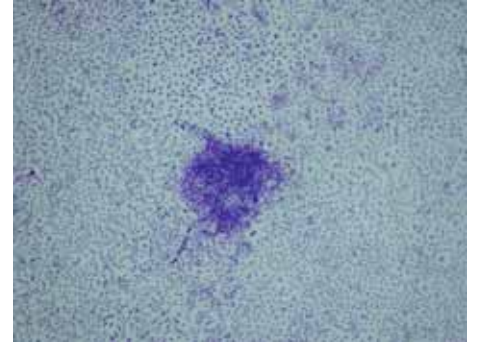
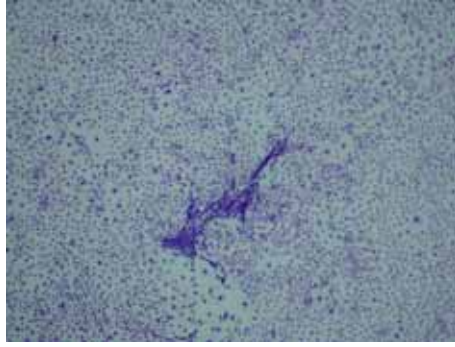
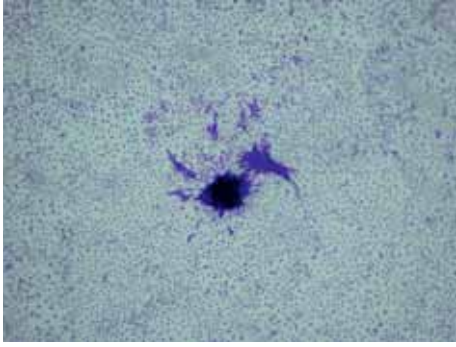
Negative foci



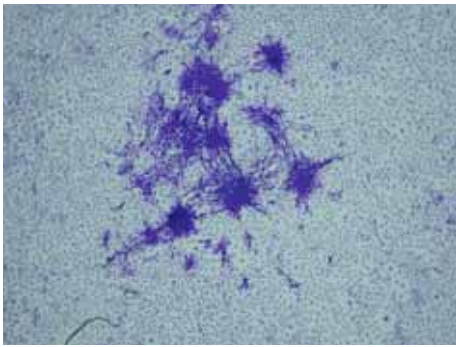
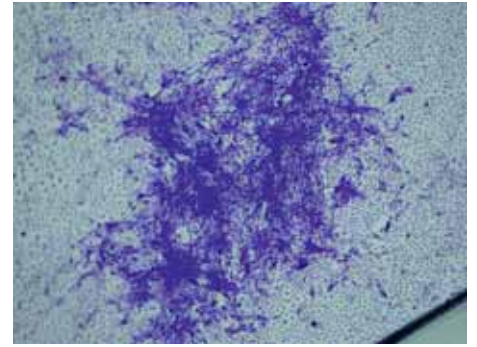
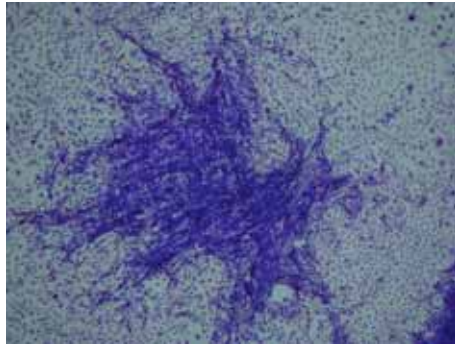
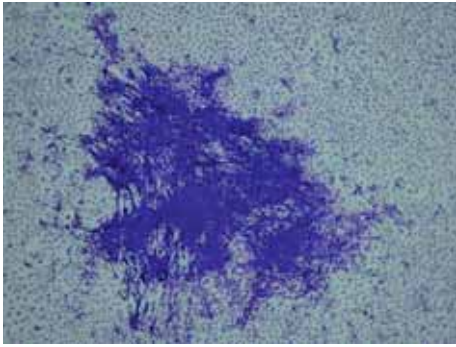
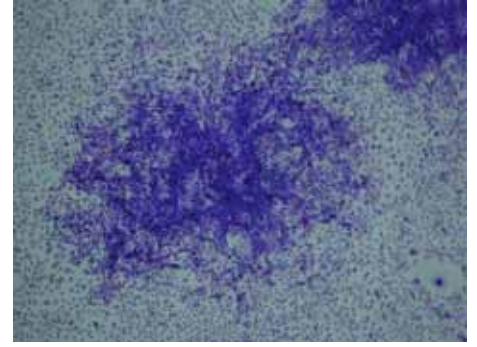
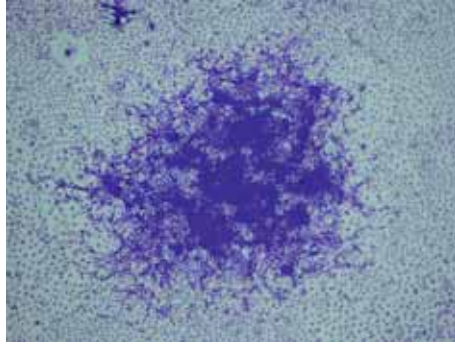
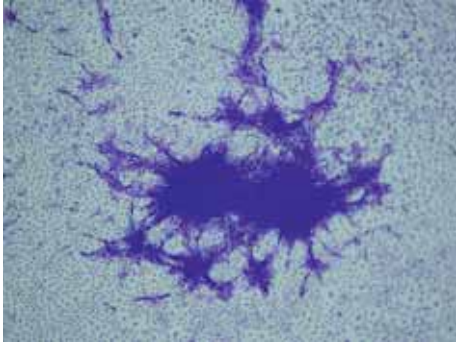
Positive foci (sparse)



Positive foci (dense and small-sized)



Positive foci (dense and middle-sized)



Annex 4

Results Submitted from Laboratories in the Validation Study of 6-Well Method

Assay Laboratory: Lab 1

Compound: 2AAF

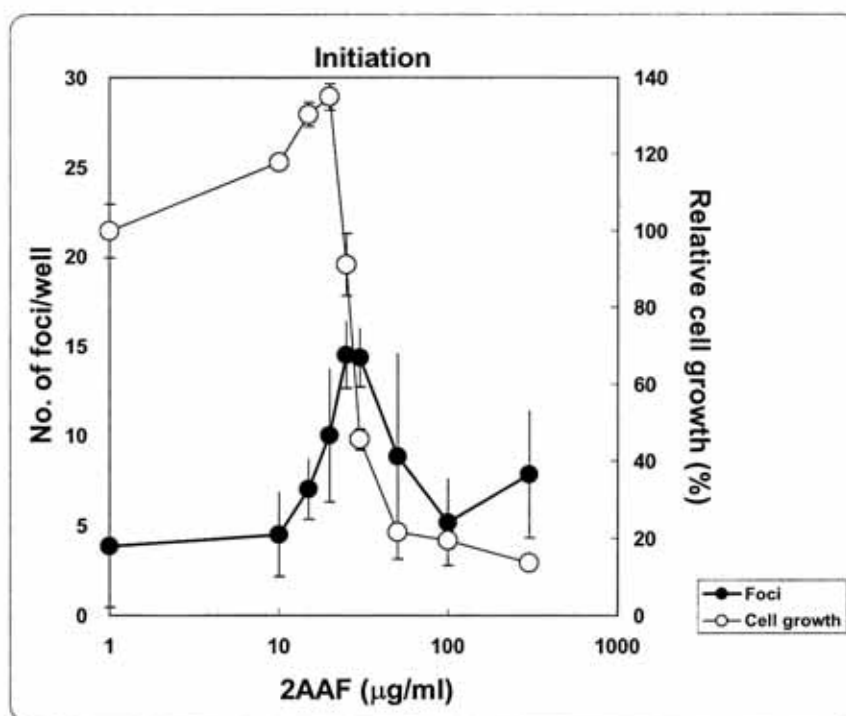
Transformation Assay

Initiation

Cell Growth Assay	Solvent: DMSO 0.1%		Compound (ug/ml)										DMSO 0.1%	MCA 1 ug/mL
	Blank	1	10	15	20	25	30	50	100	300				
OD	0.073	0.813	0.868	0.963	1.014	0.655	0.406	0.224	0.223	0.161	0.161	0.813	0.633	
	0.073	0.742	0.896	0.991	0.972	0.762	0.369	0.222	0.192	0.173	0.173	0.742	0.625	
	0.077	0.723	0.882	0.948	1.01	0.681	0.385	0.222	0.208	0.169	0.169	0.723	0.64	
Average	0.074	0.759	0.882	0.967	0.999	0.699	0.387	0.223	0.208	0.168	0.168	0.759	0.633	
SD	0.002	0.047	0.014	0.022	0.023	0.056	0.019	0.001	0.016	0.006	0.006	0.047	0.008	
Average-Blank	0.000	0.685	0.808	0.893	0.924	0.625	0.312	0.148	0.133	0.093	0.093	0.685	0.558	
Relative cell growth (%)		100.0	117.9	130.4	134.9	91.2	45.6	21.7	19.5	13.6	13.6	100.0	81.5	
SD of relative cell growth(%)		6.9	2.0	3.2	3.4	8.1	2.7	0.2	2.3	0.9	0.9	6.9	1.1	

Transformation Assay	Solvent: DMSO 0.1%		Compound (ug/ml)										DMSO 0.1%	MCA 1 ug/mL
	Well No.	1	10	15	20	25	30	50	100	300				
No of foci	1	5	4	7	9	12	14	10	8	14	5	43		
	2	6	2	6	7	13	17	1	7	8	6	43		
	3	9	8	7	14	14	14	18	7	4	9	81		
	4	1	5	7	15	17	12	7	3	9	1	49		
	5	1	2	5	9	16	15	6	3	5	1	46		
	6	1	6	10	6	15	14	11	3	7	1	38		
Average		3.8	4.5	7.0	10.0	14.5	14.3	8.8	5.2	7.8	3.8	50.0		
SD		3.4	2.3	1.7	3.7	1.9	1.6	5.7	2.4	3.5	3.4	15.6		
Dunnnett test					*	***	***	*						
t-Test (Aspin-Welch)												***		

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: 2AAF

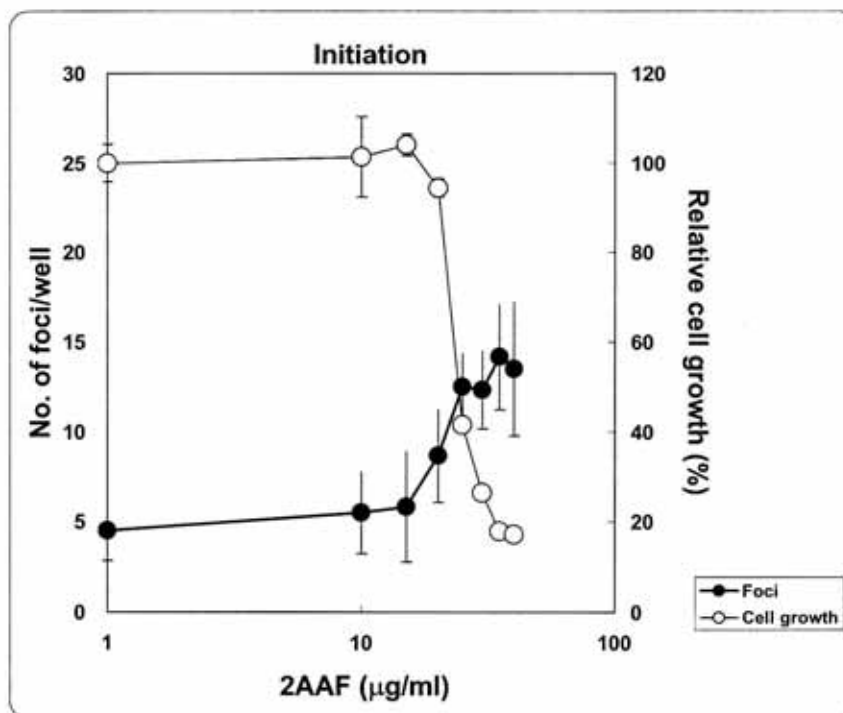
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Blank	1	10	15	20	25	30	35	40	DMSO 0.1%	MCA 1 ug/mL
OD	0.032	0.264	0.289	0.266	0.255	0.127	0.095	0.073	0.072	0.264	0.212
	0.035	0.271	0.253	0.277	0.248	0.13	0.096	0.076	0.073	0.271	0.204
	0.033	0.252	0.254	0.272	0.246	0.128	0.091	0.074	0.073	0.252	0.177
Average	0.033	0.262	0.265	0.272	0.250	0.128	0.094	0.074	0.073	0.262	0.198
SD	0.002	0.010	0.021	0.006	0.005	0.002	0.003	0.002	0.001	0.010	0.018
Average-Blank	0.000	0.229	0.232	0.238	0.216	0.095	0.061	0.041	0.039	0.229	0.164
Relative cell growth (%)		100.0	101.3	104.1	94.5	41.5	26.5	17.9	17.2	100.0	71.8
SD of relative cell growth(%)		4.2	9.0	2.4	2.1	0.7	1.2	0.7	0.3	4.2	8.0

Transformation Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Well No.	1	10	15	20	25	30	35	40	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	7	2	4	11	10	11	9	20	7	42
	2	4	7	9	9	15	15	14	9	4	38
	3	6	7	10	11	13	15	16	12	6	39
	4	3	4	3	5	12	12	13	15	3	44
	5	4	5	6	6	14	10	16	13	4	33
	6	3	8	3	10	11	11	17	12	3	35
Average		4.5	5.5	5.8	8.7	12.5	12.3	14.2	13.5	4.5	38.5
SD		1.6	2.3	3.1	2.6	1.9	2.2	2.9	3.7	1.6	4.1
Dunnett test				*	***	***	***	***			
t-Test (Aspin Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: 2AAF

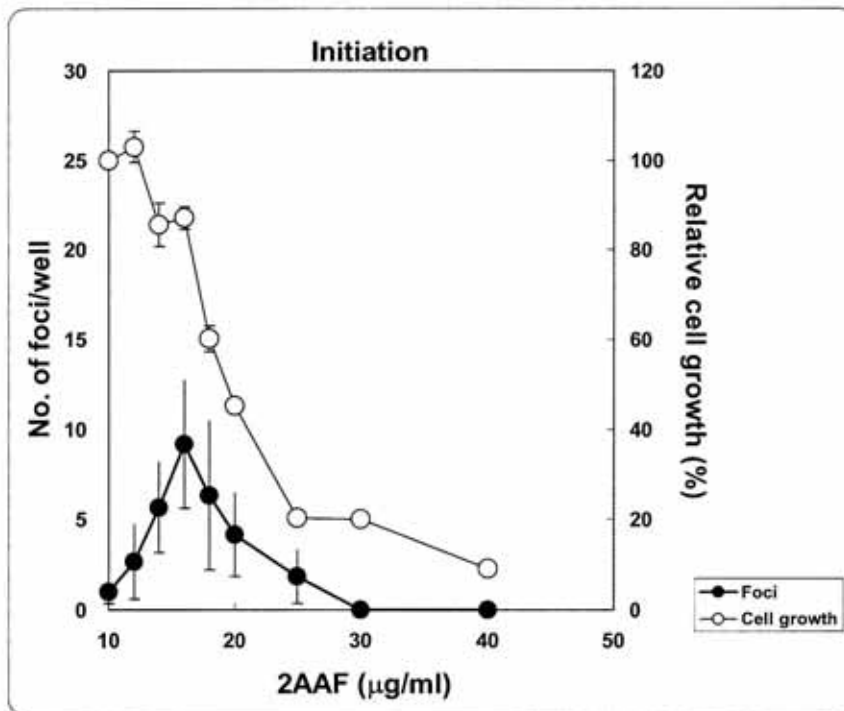
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.5%										DMSO 0.1% MCA 1 ug/mL	
Compound (ug/ml)	Blank	10	12	14	16	18	20	25	30	40			
OD	0.032	0.266	0.283	0.228	0.241	0.170	0.137	0.079	0.075	0.052	0.266	0.149	
	0.033	0.265	0.267	0.246	0.230	0.181	0.139	0.080	0.081	0.053	0.265	0.161	
	0.03	0.270	0.272	0.225	0.239	0.169	0.138	0.080	0.081	0.054	0.270	0.162	
Average	0.032	0.267	0.274	0.233	0.237	0.173	0.138	0.080	0.079	0.053	0.267	0.157	
SD	0.002	0.003	0.008	0.011	0.006	0.007	0.001	0.001	0.003	0.001	0.003	0.007	
Average-Blank	0.000	0.235	0.242	0.201	0.205	0.142	0.106	0.048	0.047	0.021	0.235	0.126	
Relative cell growth (%)		100.0	103.0	85.6	87.1	60.2	45.2	20.4	20.1	9.1	100.0	53.4	
SD of relative cell growth(%)		1.1	3.5	4.8	2.5	2.8	0.4	0.2	1.5	0.4	1.1	3.1	

Transformation Assay		Solvent: DMSO 0.5%										DMSO 0.1% MCA 1 ug/mL	
Compound (ug/ml)	Well No.	10	12	14	16	18	20	25	30	40			
No of foci	1	1	5	9	9	3	4	1 toxic	toxic		1	18	
	2	1	0	8	10	14	4	4 toxic	toxic		1	18	
	3	0	3	3	5	4	8	1 toxic	toxic		0	25	
	4	1	2	6	10	8	5	0 toxic	toxic		1	27	
	5	2	1	5	15	5	1	2 toxic	toxic		2	29	
	6	1	5	3	6	4	3	3 toxic	toxic		1	25	
Average		1.0	2.7	5.7	9.2	6.3	4.2	1.8 #DIV/0!	#DIV/0!		1.0	23.7	
SD		0.6	2.1	2.5	3.5	4.1	2.3	1.5 #DIV/0!	#DIV/0!		0.6	4.6	
Dunnnett test				**	***	**							
t-Test (Aspin-Weichi)												***	

The true value is zero. This value was temporarily put



Assay Laboratory: Lab I

Compound: 2AAF

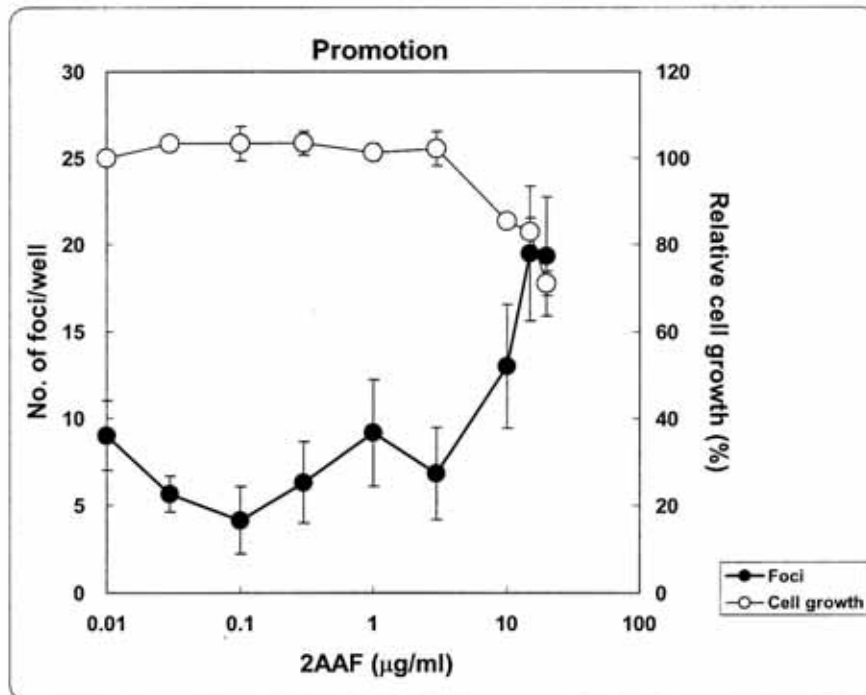
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Blank	0.01	0.03	0.1	0.3	1	3	10	15	20	DMSO 0.1%	TPA 50 ng/mL	
OD	0.073	0.931	0.965	0.989	0.937	0.942	0.981	0.797	0.801	0.658	0.931	1.074	
	0.073	0.922	0.939	0.923	0.944	0.916	0.927	0.794	0.749	0.704	0.922	1.172	
	0.077	0.918	0.952	0.946	0.98	0.948	0.919	0.81	0.788	0.673	0.918	1.16	
Average	0.074	0.924	0.952	0.953	0.954	0.935	0.942	0.800	0.779	0.678	0.924	1.135	
SD	0.002	0.007	0.013	0.034	0.023	0.017	0.034	0.009	0.027	0.023	0.007	0.053	
Average-Blank	0.000	0.849	0.878	0.878	0.879	0.861	0.868	0.726	0.705	0.604	0.849	1.061	
Relative cell growth (%)		100.0	103.3	103.4	103.5	101.4	102.2	85.5	83.0	71.1	100.0	124.9	
SD of relative cell growth(%)		0.8	1.5	3.9	2.7	2.0	4.0	1.0	3.2	2.8	0.8	6.3	

Transformation Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Well No.	0.01	0.03	0.1	0.3	1	3	10	15	20	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	11	6	2	10	7	6	19	23	18	11	39	
	2	8	7	5	7	10	7	11	24	19	8	45	
	3	12	6	7	6	14	12	13	13	14	12	41	
	4	7	5	5	5	6	5	15	19	19	7	30	
	5	8	4	2	3	11	5	11	19	22	8	39	
	6	8	6	4	7	7	6	9	19	24	8	37	
Average		9.0	5.7	4.2	6.3	9.2	6.8	13.0	19.5	19.3	9.0	38.5	
SD		2.0	1.0	1.9	2.3	3.1	2.6	3.6	3.9	3.4	2.0	5.0	
Dunnett test								*	***	***			
t-Test (Aspin-Weichi)												***	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: 2AAF

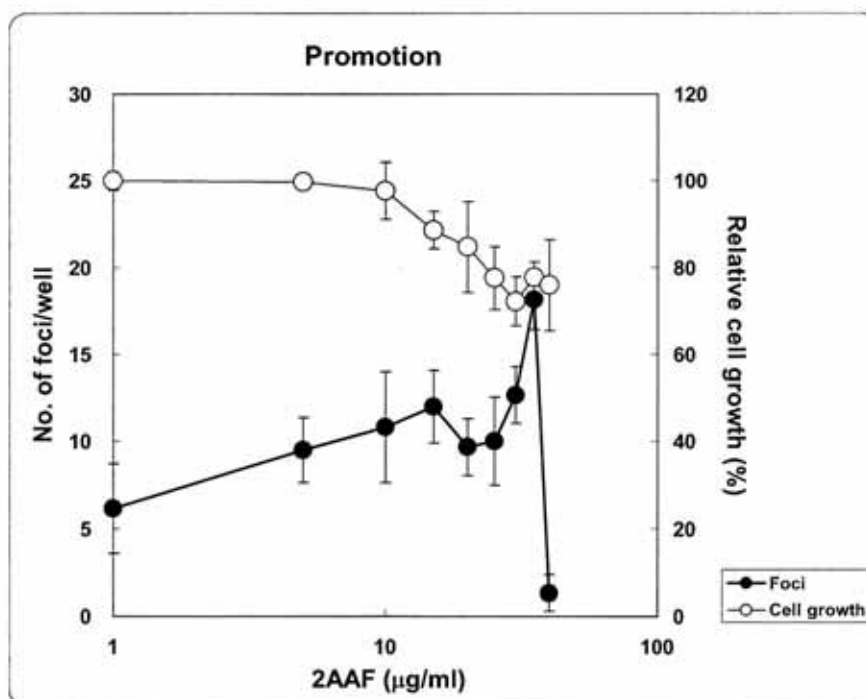
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.1%										
Compound (ug/ml)	Blank	1	5	10	15	20	25	30	35	40	DMSO 0.1%	TPA 50 ng/mL
OD	0.032	0.18	0.174	0.179	0.163	0.162	0.144	0.142	0.146	0.16	0.18	0.393
	0.035	0.174	0.178	0.18	0.166	0.166	0.156	0.142	0.15	0.136	0.174	0.337
	0.033	0.178	0.179	0.163	0.154	0.138	0.135	0.128	0.14	0.132	0.178	0.316
Average	0.033	0.177	0.177	0.174	0.161	0.155	0.145	0.137	0.145	0.143	0.177	0.349
SD	0.002	0.003	0.003	0.010	0.006	0.015	0.011	0.008	0.005	0.015	0.003	0.040
Average-Blank	0.000	0.144	0.144	0.141	0.128	0.122	0.112	0.104	0.112	0.109	0.144	0.315
Relative cell growth (%)		100.0	99.8	97.7	88.7	84.7	77.5	72.2	77.8	75.9	100.0	219.0
SD of relative cell growth(%)		2.1	1.8	6.6	4.3	10.5	7.3	5.6	3.5	10.5	2.1	27.6

Transformation Assay		Solvent:										
Compound (ug/ml)	Well No.	1	5	10	15	20	25	30	35	40	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	3	10	6	15	8	14	10	18	1	3	23
	2	8	11	13	10	8	8	15	18	0	8	22
	3	7	10	9	11	9	10	13	19	1	7	29
	4	9	6	12	12	12	8	12	17	1	9	25
	5	3	9	10	10	11	8	13	21	2	3	28
	6	7	11	15	14	10	12	13	16	3	7	21
Average		6.2	9.5	10.8	12.0	9.7	10.0	12.7	18.2	1.3	6.2	24.7
SD		2.6	1.9	3.2	2.1	1.6	2.5	1.6	1.7	1.0	2.6	3.3
Dunnett test			*	**	***	*	*	***	***			
t-Test (Assuming Equal Variances)												***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: 2AAF

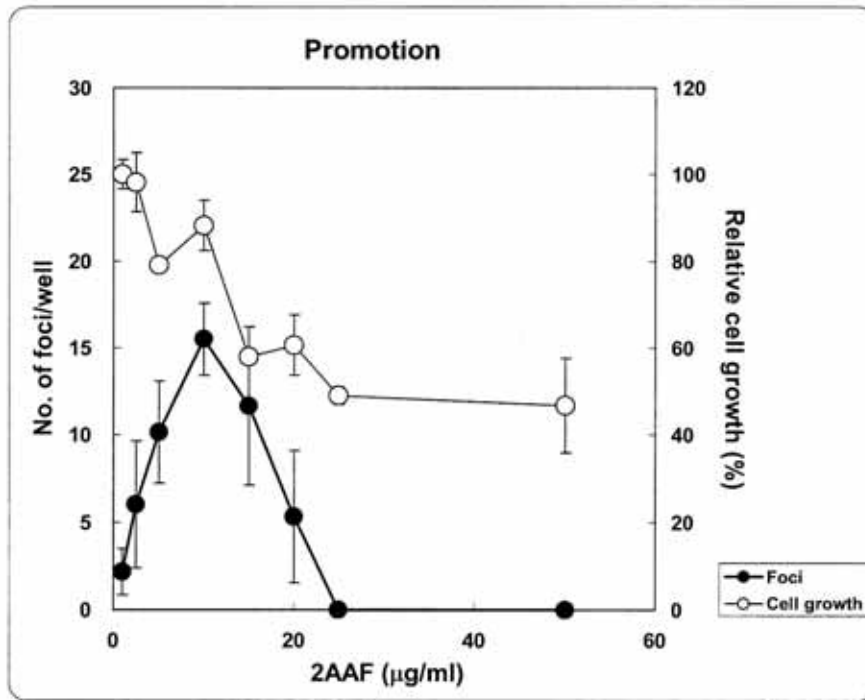
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.5%									
Compound (µg/ml)	Blank	1	2.5	5	10	15	20	25	50	DMSO 0.5%	TPA 50 ng/mL
OD	0.03	0.244	0.265	0.202	0.212	0.158	0.173	0.138	0.142	0.244	0.253
	0.032	0.259	0.241	0.206	0.228	0.156	0.174	0.144	0.153	0.259	0.277
	0.03	0.252	0.237	0.208	0.237	0.162	0.147	0.135	0.107	0.252	0.250
Average	0.031	0.252	0.248	0.205	0.226	0.159	0.165	0.139	0.134	0.252	0.260
SD	0.001	0.008	0.015	0.003	0.013	0.003	0.015	0.005	0.024	0.008	0.015
Average-Blank	0.000	0.221	0.217	0.175	0.195	0.128	0.134	0.108	0.103	0.221	0.229
Relative cell growth (%)		100.0	98.2	79.0	88.2	57.9	60.6	49.0	46.8	100.0	103.8
SD of relative cell growth(%)		3.4	6.9	1.4	5.7	1.4	6.9	2.1	10.9	3.4	6.7

Transformation Assay		Solvent: DMSO 0.5%									
Compound (µg/ml)	Well No.	1	2.5	5	10	15	20	25	50	DMSO 0.5%	TPA 50 ng/mL
No of foci	1	4	4	7	17	14	12	Toxic	Toxic	4	16
	2	2	4	9	16	19	7	Toxic	Toxic	2	17
	3	2	4	12	18	8	2	Toxic	Toxic	2	23
	4	2	9	15	15	13	2	Toxic	Toxic	2	17
	5	0	12	8	15	7	5	Toxic	Toxic	0	20
	6	3	3	10	12	9	4	Toxic	Toxic	3	12
Average		2.2	6.0	10.2	15.5	11.7	5.3	#DIV/0!	#DIV/0!	2.2	17.5
SD		1.3	3.6	2.9	2.1	4.5	3.8	#DIV/0!	#DIV/0!	1.3	3.7
Dunnett test				***	***	***					
t-Test (Aspin-Welchi)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Cadmium chloride

Transformation Assay

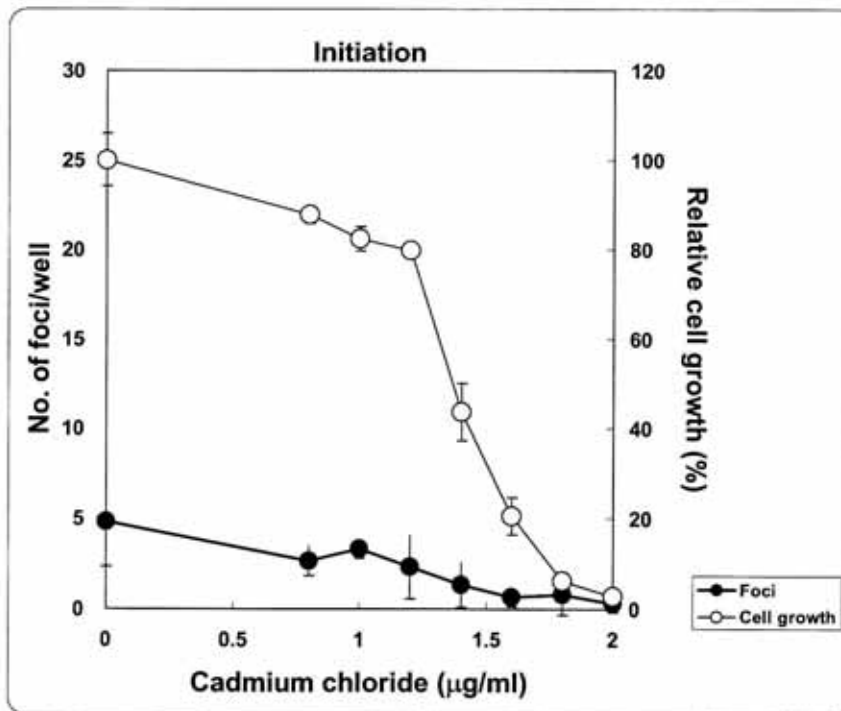
Initiation

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	0	0.8	1	1.2	1.4	1.6	1.8	2	DMSO 0.1%	MCA 1ug/mL
OD	0.073	1.019	0.897	0.857	0.809	0.526	0.255	0.14	0.109	0.875	0.603
	0.073	0.931	0.862	0.807	0.817	0.49	0.305	0.119	0.097	0.835	0.667
	0.077	1.029	0.881	0.829	0.795	0.411	0.231	0.136	0.095	0.822	0.669
Average	0.074	0.993	0.880	0.831	0.807	0.476	0.264	0.132	0.100	0.844	0.646
SD	0.002	0.054	0.018	0.025	0.011	0.059	0.038	0.011	0.008	0.028	0.038
Average-Blank	0.000	0.919	0.806	0.757	0.733	0.401	0.189	0.057	0.026	0.770	0.572
Relative cell growth (%)		100.0	87.7	82.4	79.8	43.7	20.6	6.2	2.8	100.0	74.3
SD of relative cell growth(%)		5.9	1.9	2.7	1.2	6.4	4.1	1.2	0.8	3.6	4.9

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	0	0.8	1	1.2	1.4	1.6	1.8	2	DMSO 0.1%	MCA 1ug/mL
No of foci	1	6	2	3	3	1	1	3	1	5	42
	2	8	2	4	0	3	1	1	1	6	46
	3	3	3	3	3	2	1	1	0	4	42
	4	5	4	3	2	0	1	0	0	7	50
	5	1	2	3	5	2	0	0	0	2	41
	6	6	3	4	1	0	0	0	0	2	40
Average		4.8	2.7	3.3	2.3	1.3	0.7	0.8	0.3	4.3	43.5
SD		2.5	0.8	0.5	1.8	1.2	0.5	1.2	0.5	2.1	3.8
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)											***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Cadmium chloride

Transformation Assay

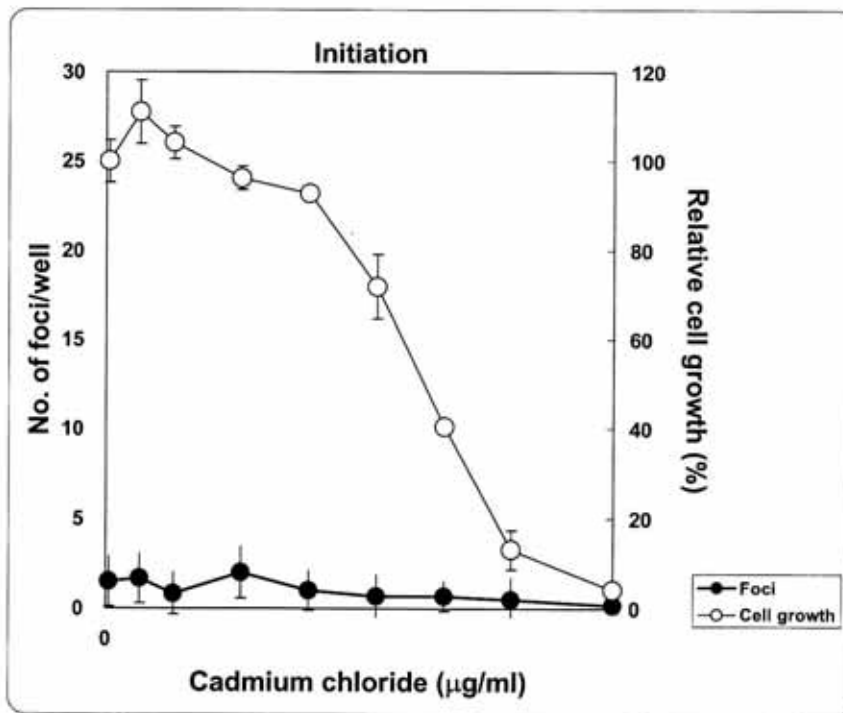
Initiation

Cell Growth Assay		Solvent:	Water 5%											
Compound (ug/ml)	Blank		0.01	0.1	0.2	0.4	0.6	0.8	1	1.2	1.5	DMSO 0.1%	MCA 1ug/mL	
OD	0.0681	0.7008	0.7818	0.7161	0.6433	0.6334	0.4608	0.3233	0.122	0.0914				
	0.0709	0.6533	0.7194	0.6726	0.6315	0.6158	0.546	0.3037	0.1733	0.102				
	0.0711	0.6501	0.7009	0.6894	0.6622	0.6257	0.4924	0.3098	0.149	0.0904				
Average	0.070	0.668	0.734	0.693	0.646	0.625	0.500	0.312	0.148	0.095	#DIV/0!	#DIV/0!		
SD	0.002	0.028	0.042	0.022	0.015	0.009	0.043	0.010	0.026	0.006	#DIV/0!	#DIV/0!		
Average-Blank	0.000	0.598	0.664	0.623	0.576	0.555	0.430	0.242	0.078	0.025	#DIV/0!	#DIV/0!		
Relative cell growth (%)		100.0	111.0	104.1	96.3	92.8	71.9	40.5	13.1	4.1	#DIV/0!	#DIV/0!		
SD of relative cell growth(%)		4.7	7.1	3.7	2.6	1.5	7.2	1.7	4.3	1.1	#DIV/0!	#DIV/0!		

Transformation Assay		Solvent:	Water 5%											
Compound (ug/ml)	Well No.		0.01	0.1	0.2	0.4	0.6	0.8	1	1.2	1.5	DMSO 0.1%	MCA 1ug/mL	
No of foci	1	1	2	1	2	0	1	0	0	0	0	2	9	
	2	3	4	3	0	0	0	1	3	0	0	2	16	
	3	0	0	0	3	2	3	0	0	0	0	0	23	
	4	2	2	0	2	2	0	2	0	0	0	1	16	
	5	0	1	0	4	2	0	1	0	0	0	1	16	
	6	3	1	1	1	0	0	0	0	0	1	0	15	
Average		1.5	1.7	0.8	2.0	1.0	0.7	0.7	0.5	0.2	1.0	15.8		
SD		1.4	1.4	1.2	1.4	1.1	1.2	0.8	1.2	0.4	0.9	4.4		
Dunnett test														
t-Test for Unequal Variances (Aspin-Weich)														

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Cadmium chloride

Transformation Assay

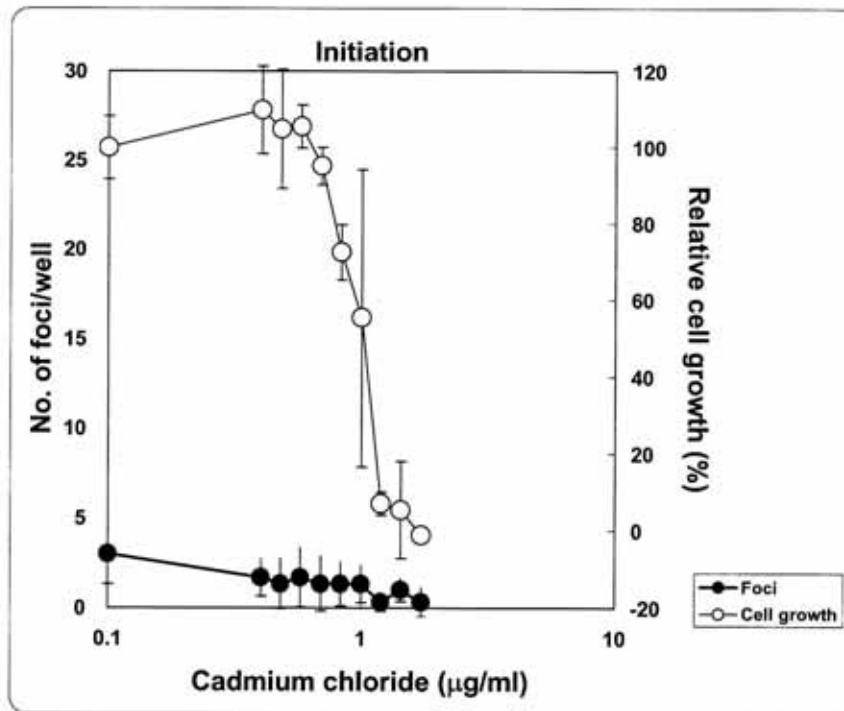
Initiation

Cell Growth Assay		Solvent: D.W. 5 %												
Compound (ug/ml)	Blank	0.1	0.402	0.482	0.579	0.694	0.833	1	1.2	1.44	1.73	DMSO 0.1%	MCA 1ug/mL	
OD	0.176	1.509	1.879	1.859	1.722	1.561	1.225	1.556	0.268	0.412	0.14	-	0.366	
	0.106	1.687	1.639	1.593	1.564	1.472	1.217	0.593	0.233	0.099	0.092	-	0.375	
	0.102	1.466	1.565	1.415	1.615	1.423	1.043	0.602	0.182	0.102	0.101	-	0.253	
Average	0.128	1.554	1.694	1.622	1.634	1.485	1.162	0.917	0.228	0.204	0.111	#DIV/0!	0.331	
SD	0.042	0.117	0.164	0.223	0.081	0.070	0.103	0.553	0.043	0.180	0.026	#DIV/0!	0.068	
Average-Blank	0.000	1.426	1.566	1.494	1.506	1.357	1.034	0.789	0.100	0.076	-0.017	#DIV/0!	0.203	
Relative cell growth (%)		100.0	109.8	104.8	105.6	95.2	72.5	55.3	7.0	5.4	-1.2	#DIV/0!	#DIV/0!	
SD of relative cell growth(%)		8.2	11.5	15.7	5.7	4.9	7.2	38.8	3.0	12.6	1.8	#DIV/0!	#DIV/0!	

Transformation Assay		Solvent: D.W. 5 %												
Compound (ug/ml)	Well No.	0.1	0.402	0.482	0.579	0.694	0.833	1	1.2	1.44	1.73	DMSO 0.1%	MCA 1ug/mL	
No of foci	1	5	2	0	4	2	2	3	0	1	0	-	11	
	2	5	2	3	1	4	1	1	1	1	0	-	8	
	3	1	1	0	2	0	3	0	1	2	0	-	7	
	4	3	2	3	0	1	0	2	0	0	2	-	11	
	5	2	0	1	3	1	0	1	0	1	0	-	11	
	6	2	3	1	0	0	2	1	0	1	0	-	7	
Average		3.0	1.7	1.3	1.7	1.3	1.3	1.3	0.3	1.0	0.3	#DIV/0!	9.2	
SD		1.7	1.0	1.4	1.6	1.5	1.2	1.0	0.5	0.6	0.8	#DIV/0!	2.0	
Dunnett test														
t-Test (Assuming Equal Variances) vs 5% Water													***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Cadmium chloride

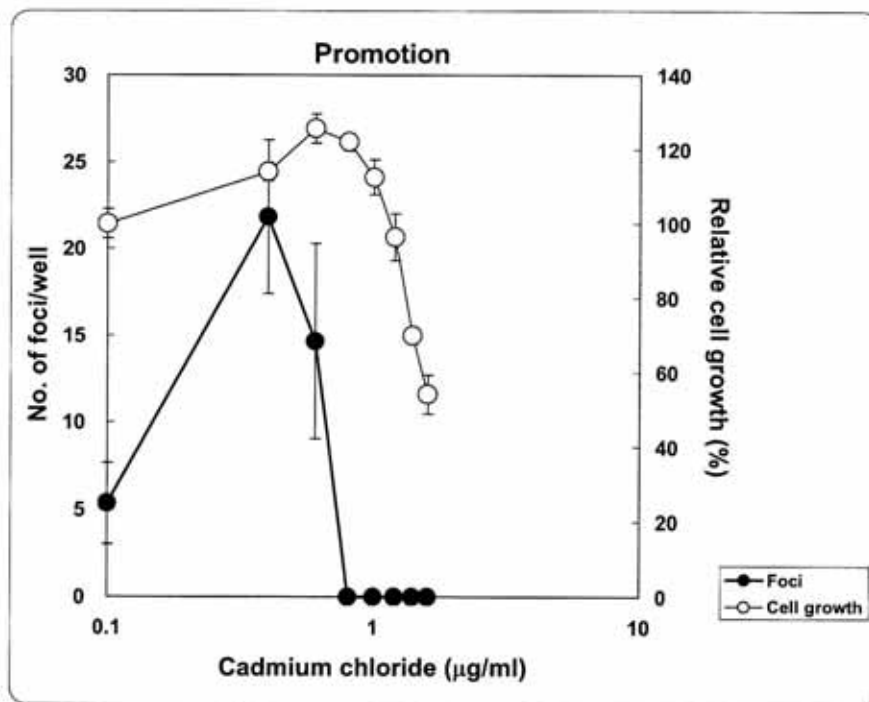
Torsformation Assay

Promotion

Cell Growth Assay		Solvent: water 5%										
Compound (ug/ml)	Blank	0.1	0.4	0.6	0.8	1	1.2	1.4	1.6	DMSO 0.1%	TPA 50 ng/mL	
OD		0.073	0.962	1.047	1.175	1.134	1.005	0.938	0.67	0.527	1.038	1.287
		0.073	0.897	1.077	1.116	1.1	1.086	0.931	0.66	0.589	1.032	1.281
		0.077	0.943	1.038	1.174	1.135	1.035	0.841	0.697	0.503	1.021	1.254
Average		0.074	0.934	1.054	1.155	1.123	1.042	0.903	0.676	0.540	1.030	1.274
SD		0.002	0.033	0.020	0.034	0.020	0.041	0.054	0.019	0.044	0.009	0.018
Average-Blank		0.000	0.860	0.980	1.081	1.049	0.968	0.829	0.601	0.465	0.956	1.200
Relative cell growth (%)			100.0	114.0	125.7	122.0	112.6	96.4	69.9	54.1	100.0	125.5
SD of relative cell growth(%)			3.9	2.4	3.9	2.3	4.8	6.3	2.2	5.2	0.9	1.8

Transformation Assay		Solvent: water 5%										
Compound (ug/ml)	Well No.	0.1	0.4	0.6	0.8	1	1.2	1.4	1.6	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	6	20	18	toxic	toxic	toxic	toxic	toxic	11	46	
	2	7	22	19	toxic	toxic	toxic	toxic	toxic	6	42	
	3	8	15	17	toxic	toxic	toxic	toxic	toxic	12	31	
	4	3	25	8	toxic	toxic	toxic	toxic	toxic	13	34	
	5	6	21	7	toxic	toxic	toxic	toxic	toxic	5	42	
	6	2	28	19	toxic	toxic	toxic	toxic	toxic	7	37	
Average		5.3	21.8	14.7	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	9.0	38.7	
SD		2.3	4.4	5.6	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	3.4	5.6	
Dunnett test			***	**								
t-Test for Unequal Variances (Aspin-Welch)											***	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Cadmium chloride

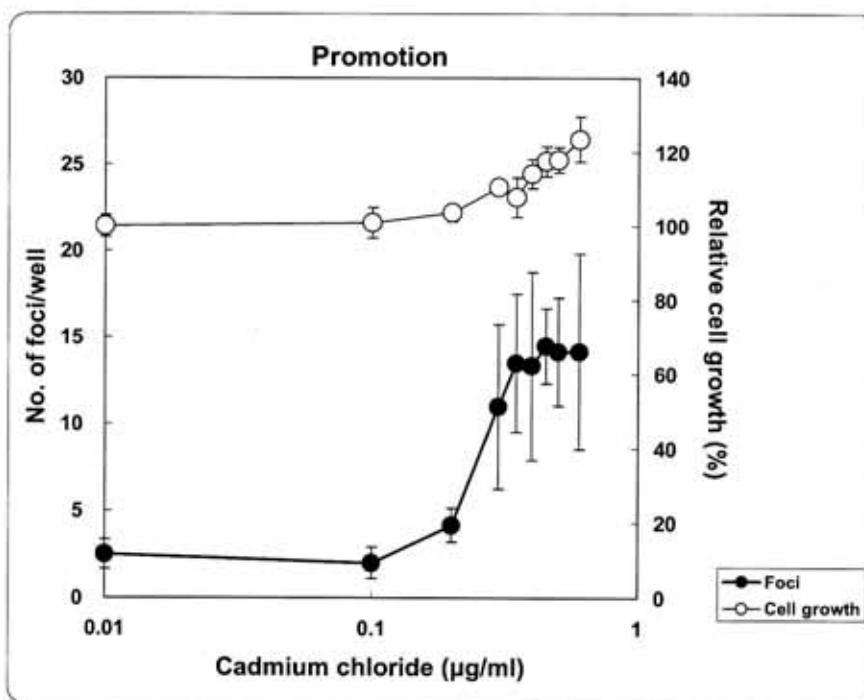
Transformation Assay

Promotion

Cell Growth Assay		Solvent: water 5%											
Compound (ug/ml)	Blank	0.01	0.1	0.2	0.3	0.35	0.4	0.45	0.5	0.6	DMSO 0.1%	TPA 50 ng/mL	
OD	0.065	0.98	0.999	1.01	1.053	1.072	1.117	1.152	1.118	1.226	0.998	1.308	
	0.065	0.956	0.964	0.975	1.049	1.022	1.078	1.092	1.142	1.14	0.956	1.265	
	0.063	0.928	0.925	0.974	1.042	0.978	1.046	1.086	1.082	1.124	0.92	1.167	
Average	0.064	0.955	0.963	0.986	1.048	1.024	1.080	1.110	1.114	1.163	0.958	1.247	
SD	0.001	0.026	0.037	0.021	0.006	0.047	0.036	0.036	0.030	0.055	0.039	0.072	
Average-Blank	0.000	0.890	0.898	0.922	0.984	0.960	1.016	1.046	1.050	1.099	0.894	1.182	
Relative cell growth (%)		100.0	100.9	103.6	110.5	107.8	114.1	117.4	117.9	123.4	100.0	132.3	
SD of relative cell growth(%)		2.9	4.2	2.3	0.6	5.3	4.0	4.1	3.4	6.2	4.4	8.1	

Transformation Assay		Solvent: water 5%											
Compound (ug/ml)	Well No.	0.01	0.1	0.2	0.3	0.35	0.4	0.45	0.5	0.6	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	2	2	5	12	14	14	13	17	17	10	21	
	2	2	1	3	10	12	14	16	17	11	4	16	
	3	2	3	5	17	12	16	14	16	24	5	28	
	4	3	1	5	14	8	5	12	13	8	7	20	
	5	2	3	4	3	15	21	14	9	12	0	11	
	6	4	2	3	10	20	10	18	13	13	4	20	
Average		2.5	2.0	4.2	11.0	13.5	13.3	14.5	14.2	14.2	5.0	19.3	
SD		0.8	0.9	1.0	4.7	4.0	5.4	2.2	3.1	5.6	3.3	5.6	
Dunnett test					**	***	***	***	***	***			
t-Test for Unequal Variances (Aspin-Welch)												***	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Cadmium chloride

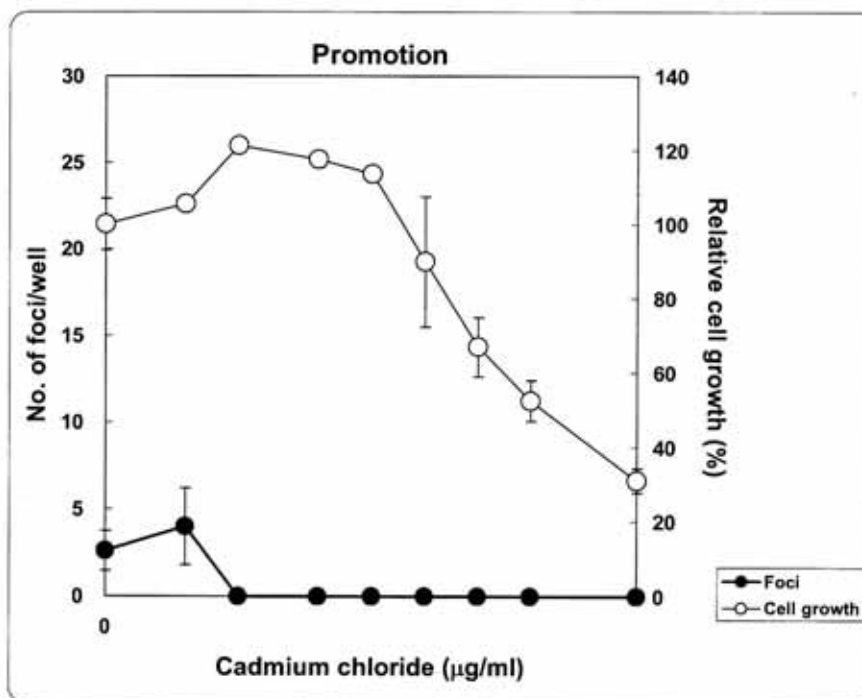
Transformation Assay

Promotion

Cell Growth Assay		Solvent: Water 5%											
Compound (ug/ml)	Blank	0	0.3	0.5	0.8	1	1.2	1.4	1.6	2	DMSO 0.1%	TPA 50 ng/mL	
OD	0.0727	0.5683	0.5679	0.6309	0.6153	0.6033	0.513	0.3993	0.3228	0.2062			
	0.0723	0.5318	0.5613	0.6265	0.6254	0.594	0.5543	0.4067	0.335	0.2082			
	0.0729	0.5042	0.5525	0.6436	0.606	0.5951	0.3975	0.34	0.2863	0.2334			
平均	0.073	0.535	0.561	0.634	0.616	0.597	0.488	0.382	0.315	0.216	#DIV/0!	#DIV/0!	
SD	0.000	0.032	0.008	0.009	0.010	0.005	0.081	0.037	0.025	0.015	#DIV/0!	#DIV/0!	
Average-Blank	0.000	0.462	0.488	0.561	0.543	0.525	0.416	0.309	0.242	0.143	#DIV/0!	#DIV/0!	
Relative cell growth (%)		100.0	105.6	121.4	117.5	113.6	89.9	66.9	52.4	31.0	#DIV/0!	#DIV/0!	
SD of relative cell growth(%)		7.0	1.7	1.9	2.1	1.1	17.6	7.9	5.5	3.3	#DIV/0!	#DIV/0!	

Transformation Assay		Solvent: Water 5%											
Compound (ug/ml)	Well No.	0	0.3	0.5	0.8	1	1.2	1.4	1.6	2	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	3	5	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	1	10	
	2	3	3	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	4	4	
	3	4	8	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	3	6	
	4	1	2	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	2	13	
	5	2	3	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	0	8	
	6	-	3	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	4	9	
Average		2.6	4.0	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	2.3	8.3	
SD		1.1	2.2	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	1.6	3.1	
t-Test for Unequal Variances (Aspin-Welch)		N.S.											
t-Test for Unequal Variances (Aspin-Welch)													**

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Cadmium chloride

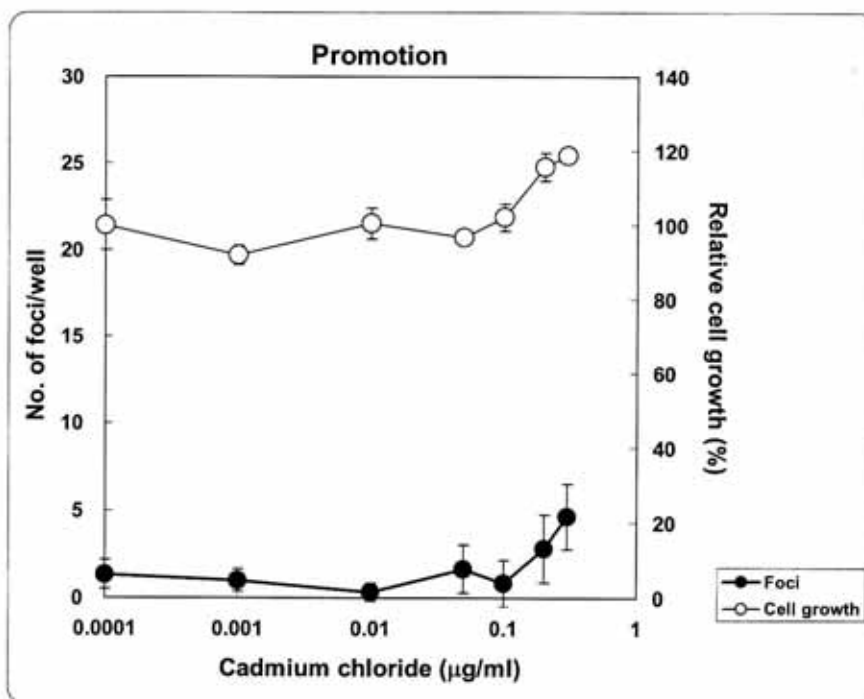
Transformation Assay

Promotion

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	0.0001	0.001	0.01	0.05	0.1	0.2	0.3	DMSO 0.1%	TPA 50 ng/mL	
OD	0.0574	0.3741	0.3829	0.3906	0.3941	0.3946	0.445	0.4693	0.4138	0.5288	
	0.0564	0.4182	0.3699	0.3975	0.3859	0.4096	0.4695	0.4646	0.4311	0.5402	
	0.0567	0.4097	0.3655	0.4184	0.3876	0.4199	0.45	0.4626	0.4127	0.5479	
Average	0.057	0.401	0.373	0.402	0.389	0.408	0.455	0.466	0.419	0.539	
SD	0.001	0.023	0.009	0.014	0.004	0.013	0.013	0.003	0.010	0.010	
Average-Blank	0.000	0.344	0.316	0.345	0.332	0.351	0.398	0.409	0.362	0.482	
Relative cell growth (%)		100.0	91.9	100.4	96.7	102.1	115.8	118.9	100.0	133.1	
SD of relative cell growth(%)		6.8	2.6	4.2	1.3	3.7	3.8	1.0	2.8	2.7	

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	0.0001	0.001	0.01	0.05	0.1	0.2	0.3	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	2	2	1	4	3	2	5	1	14	
	2	0	1	1	1	0	4	3	3	12	
	3	2	1	0	2	0	6	2	2	11	
	4	1	1	0	1	2	1	7	2	8	
	5	2	1	0	0	0	1	6	3	15	
	6	1	0	0	2	0	3	5	1	9	
Average		1.3	1.0	0.3	1.7	0.8	2.8	4.7	2.0	11.5	
SD		0.8	0.8	0.5	1.4	1.3	1.9	1.9	0.9	2.7	
Dunnett test								***			
t-Test (Aspin-Welch)										***	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Cadmium chloride

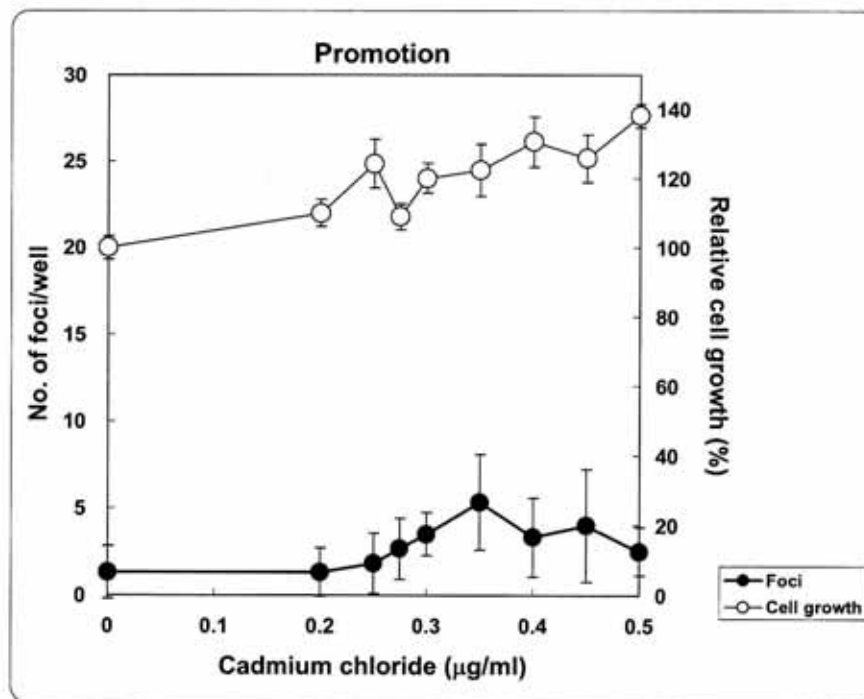
Transformation Assay

Promotion

Cell Growth Assay		Solvent:	DW 5%										DMSO 0.1%		TPA 50 ng/mL									
Compound (ug/ml)	Blank	0	0.2	0.25	0.275	0.3	0.35	0.4	0.45	0.5	0.5	0.5	0.5	0.5	0.5	0.5								
OD	0.0618	0.5231	0.5837	0.6645	0.5625	0.639	0.677	0.7126	0.6848	0.7214	0.5434	0.7011	0.059	0.5536	0.6021	0.6771	0.598	0.6478	0.6462	0.6885	0.6705	0.7317	0.5911	0.753
	0.0585	0.5306	0.5644	0.6138	0.5753	0.6082	0.6057	0.6439	0.622	0.6994	0.521	0.7044	0.0585	0.5306	0.5644	0.6138	0.5753	0.6082	0.6057	0.6439	0.622	0.6994	0.521	0.7044
Average	0.060	0.536	0.583	0.652	0.579	0.632	0.643	0.682	0.659	0.718	0.552	0.720	0.060	0.536	0.583	0.652	0.579	0.632	0.643	0.682	0.659	0.718	0.552	0.720
SD	0.002	0.016	0.019	0.034	0.018	0.021	0.036	0.035	0.033	0.016	0.036	0.029	0.002	0.016	0.019	0.034	0.018	0.021	0.036	0.035	0.033	0.016	0.036	0.029
Average-Blank	0.000	0.476	0.524	0.592	0.519	0.572	0.583	0.622	0.599	0.658	0.492	0.660	0.000	0.476	0.524	0.592	0.519	0.572	0.583	0.622	0.599	0.658	0.492	0.660
Relative cell growth (%)		100.0	110.0	124.4	109.0	120.1	122.5	130.7	125.9	138.2	100.0	134.1		100.0	110.0	124.4	109.0	120.1	122.5	130.7	125.9	138.2	100.0	134.1
SD of relative cell growth(%)		3.3	4.0	7.0	3.8	4.4	7.5	7.3	6.9	3.5	7.3	5.9		3.3	4.0	7.0	3.8	4.4	7.5	7.3	6.9	3.5	7.3	5.9

Transformation Assay		Solvent:	DW 5%										DMSO 0.1%		TPA 50 ng/mL	
Compound (ug/ml)	Well No.	0	0.2	0.25	0.275	0.3	0.35	0.4	0.45	0.5	0.5	0.5	0.5	0.5	0.5	0.5
No of foci	1	3	1	1	1	3	3	1	2	0	2	14	1	3	1	1
	2	0	4	4	0	3	7	1	9	4	2	10	2	0	4	4
	3	0	1	0	3	2	4	5	7	2	2	11	2	1	0	3
	4	2	1	4	4	3	3	5	3	3	2	7	2	2	4	4
	5	3	0	1	4	5	5	2	1	3	1	8	2	3	1	1
	6	0	1	1	4	5	10	6	2	3	2	7	2	0	1	1
Average		1.3	1.3	1.8	2.7	3.5	5.3	3.3	4.0	2.5	1.8	9.5	1.3	1.3	1.8	2.7
SD		1.5	1.4	1.7	1.8	1.2	2.7	2.3	3.2	1.4	0.4	2.7	1.5	1.4	1.7	1.8
Dunnett test							**									
t-Test (Aspin-Welch)																***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Cadmium chloride

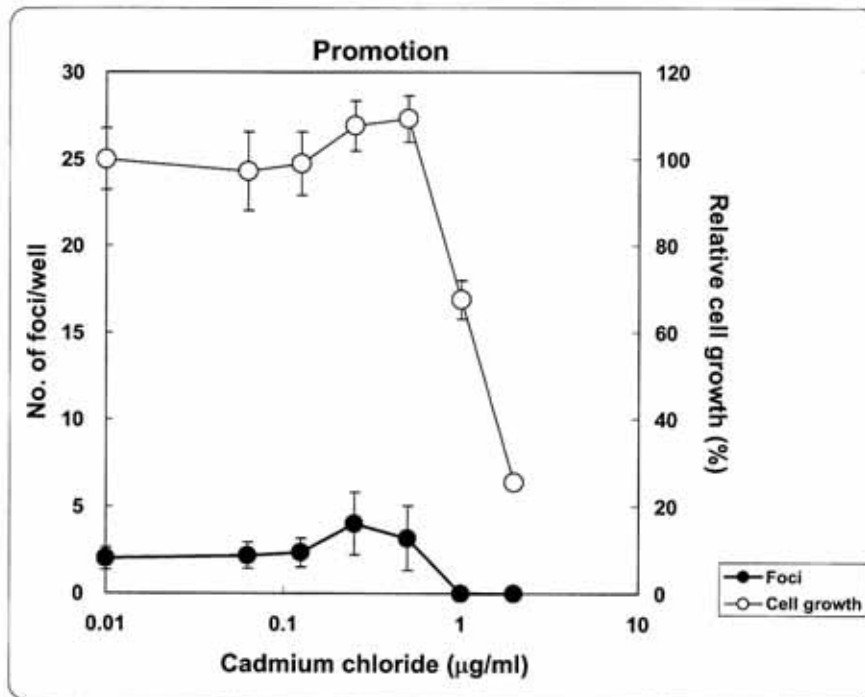
Transformation Assay

Promotion

Cell Growth Assay		Solvent: D.W. 5 %									
Compound (ug/ml)	Blank	0.01	0.0625	0.125	0.25	0.5	1	2	DMSO 0.1%	TPA 50 ng/mL	
OD	0.183	1.954	1.856	1.935	2.128	2.172	1.459	0.701	-	-	3.007
	0.188	2.193	2.149	2.125	2.31	2.323	1.585	0.706	-	-	3.287
	0.225	2.183	2.164	2.209	2.33	2.367	1.423	0.648	-	-	3.212
平均	0.199	2.110	2.056	2.090	2.256	2.287	1.489	0.685	#DIV/0!	#DIV/0!	3.169
SD	0.023	0.135	0.174	0.140	0.111	0.102	0.085	0.032	#DIV/0!	#DIV/0!	0.145
Average-Blank	0.000	1.911	1.858	1.891	2.057	2.089	1.290	0.486	#DIV/0!	#DIV/0!	2.970
Relative cell growth (%)		100.0	97.2	98.9	107.6	109.3	67.5	25.4	#DIV/0!	#DIV/0!	
SD of relative cell growth(%)		7.1	9.1	7.3	5.8	5.4	4.5	1.7	#DIV/0!	#DIV/0!	

Transformation Assay		Solvent: D.W. 5 %									
Compound (ug/ml)	Well No.	0.01	0.0625	0.125	0.25	0.5	1	2	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	2	2	3	1	1	Tox	Tox	-	-	16
	2	3	3	2	4	2	Tox	Tox	-	-	17
	3	2	3	3	3	4	Tox	Tox	-	-	23
	4	1	1	3	5	6	Tox	Tox	-	-	12
	5	2	2	1	6	4	Tox	Tox	-	-	16
	6	2	2	2	5	2	Tox	Tox	-	-	10
Average		2.0	2.2	2.3	4.0	3.2	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	15.7
SD		0.6	0.8	0.8	1.8	1.8	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	4.5
Dunnett test					*						
t-Test for Unequal Variances (Aspin-Welch) vs 5% Water											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Cadmium chloride

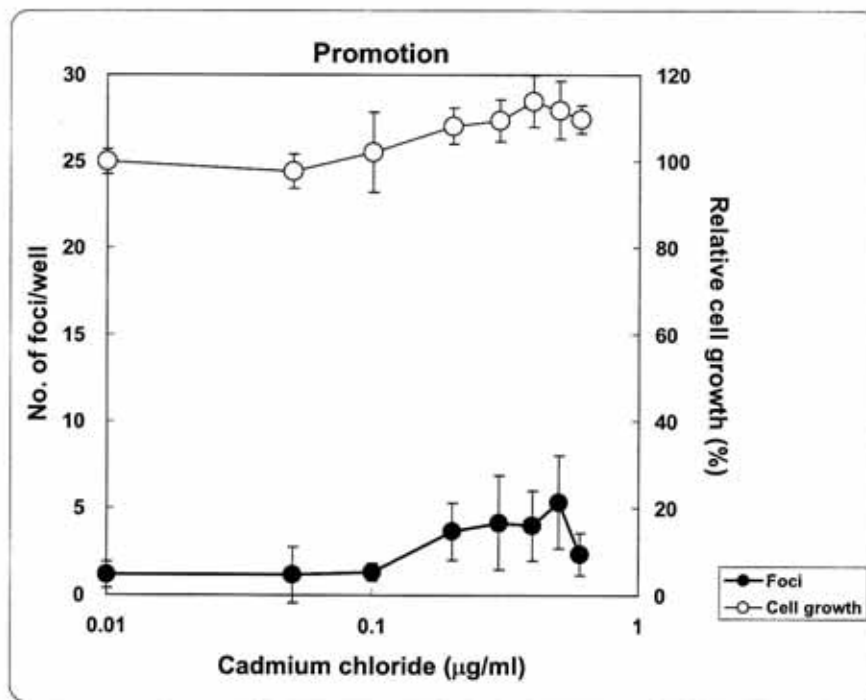
Transformation Assay

Promotion

Cell Growth Assay		Solvent: D.W. 5%									
Compound (ug/ml)	Blank	0.01	0.05	0.1	0.2	0.3	0.4	0.5	0.6	DMSO 0.1%	TPA 50 ng/mL
OD	0.267	2.091	2.006	1.992	2.351	2.192	2.473	2.193	2.267	1.984	3.004
	0.297	2.077	2.15	2.135	2.224	2.366	2.377	2.369	2.359	2.083	3.393
	0.281	2.174	2.061	2.331	2.216	2.304	2.255	2.429	2.251	2.195	3.415
Average	0.282	2.114	2.072	2.153	2.264	2.287	2.368	2.330	2.292	2.087	3.271
SD	0.015	0.052	0.073	0.170	0.076	0.088	0.109	0.123	0.058	0.106	0.231
Average-Blank	0.000	1.832	1.791	1.871	1.982	2.006	2.087	2.049	2.011	1.806	2.989
Relative cell growth (%)		100.0	97.7	102.1	108.2	109.5	113.9	111.8	109.7	100.0	165.5
SD of relative cell growth(%)		2.9	4.0	9.3	4.1	4.8	6.0	6.7	3.2	5.8	12.8

Transformation Assay		Solvent: D.W. 5%									
Compound (ug/ml)	Well No.	0.01	0.05	0.1	0.2	0.3	0.4	0.5	0.6	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	0	2	1	1	1	4	3	2	2	24
	2	1	0	1	4	3	3	7	1	6	22
	3	2	0	1	4	4	3	7	4	3	24
	4	1	4	2	6	3	3	9	3	4	18
	5	1	0	2	4	5	8	3	3	4	21
	6	2	1	1	3	9	3	3	1	4	13
Average		1.2	1.2	1.3	3.7	4.2	4.0	5.3	2.3	3.8	20.3
SD		0.8	1.6	0.5	1.6	2.7	2.0	2.7	1.2	1.3	4.2
Dunnett test						*	*	**			
t-Test for Unequal Variances (Aspin-Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: DB[a,h]A

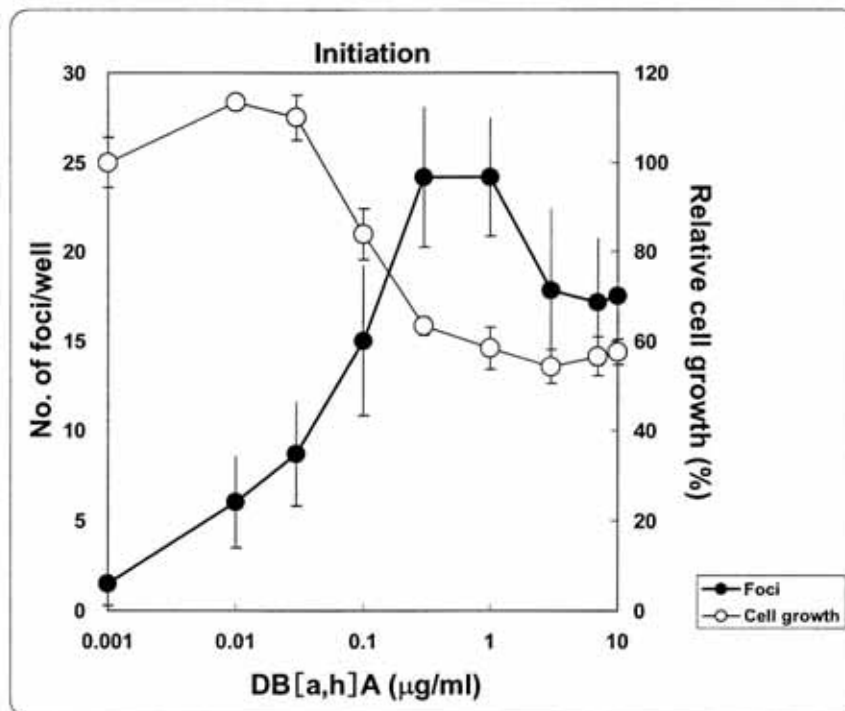
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Blank	0.001	0.01	0.03	0.1	0.3	1	3	7	10	DMSO 0.1%	MCA 1 ug/mL	
OD	0.073	0.913	0.983	0.948	0.743	0.588	0.565	0.51	0.483	0.53	0.913	0.735	
	0.073	0.847	0.972	0.899	0.777	0.577	0.546	0.47	0.53	0.505	0.847	0.695	
	0.077	0.83	0.954	0.979	0.688	0.556	0.493	0.528	0.549	0.55	0.83	0.669	
Average	0.074	0.863	0.970	0.942	0.736	0.574	0.535	0.503	0.521	0.528	0.863	0.700	
SD	0.002	0.044	0.015	0.040	0.045	0.016	0.037	0.030	0.034	0.023	0.044	0.033	
Average-Blank	0.000	0.789	0.895	0.868	0.662	0.499	0.460	0.428	0.446	0.454	0.789	0.625	
Relative cell growth (%)		100.0	113.5	110.0	83.9	63.3	58.3	54.3	56.6	57.5	100.0	79.3	
SD of relative cell growth(%)		5.6	1.9	5.1	5.7	2.1	4.7	3.8	4.3	2.9	5.6	4.2	

Transformation Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Well No.	0.001	0.01	0.03	0.1	0.3	1	3	7	10	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	2	10	7	19	27	23	25	21	14	2	16	
	2	0	6	12	11	24	18	15	15	20	0	17	
	3	2	4	12	17	30	25	15	16	22	2	24	
	4	2	4	7	15	24	26	14	18	17	2	22	
	5	3	4	5	9	20	27	22	12	13	3	25	
	6	0	8	9	19	20	26	16	21	19	0	16	
Average		1.5	6.0	8.7	15.0	24.2	24.2	17.8	17.2	17.5	1.5	20.0	
SD		1.2	2.5	2.9	4.2	3.9	3.3	4.5	3.5	3.5	1.2	4.1	
Dunnett test				**	***	***	***	***	***	***			
t-Test for Unequal Variances (Aspin-Welch)												***	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: DB[a,h]A

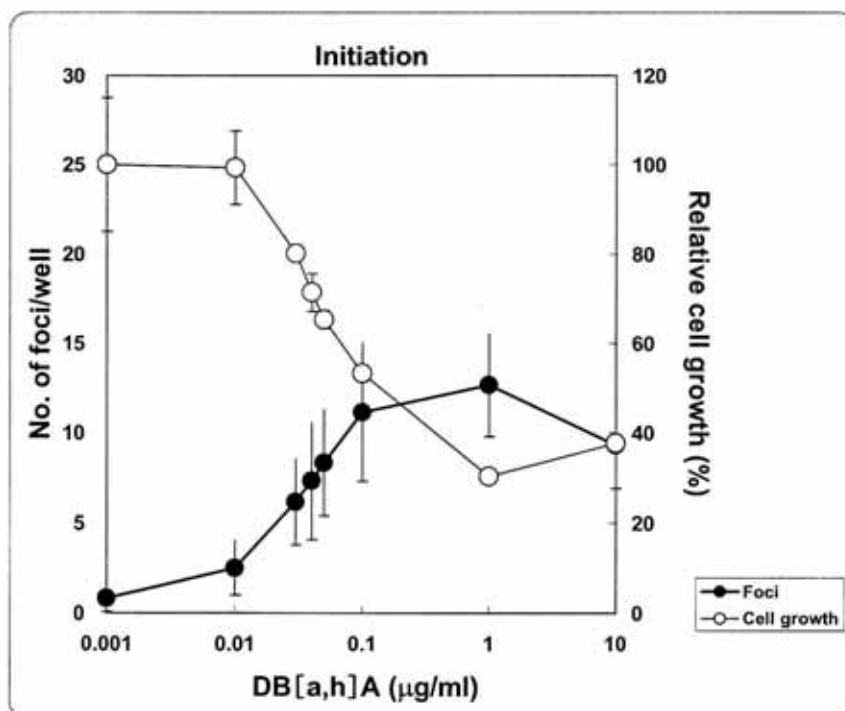
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.2%									
Compound (ug/ml)	Blank	0.001	0.01	0.03	0.04	0.05	0.1	1	10	DMSO 0.1%	MCA 1 ug/mL
OD	0.0726	0.6732	0.6899	0.5962	0.5762	0.4897	0.4299	0.268	0.3285	0.6529	0.3818
	0.0767	0.8473	0.705	0.6073	0.531	0.5126	0.4123	0.2737	0.325	0.67	0.389
	0.0755	0.6783	0.7907	0.603	0.5263	0.5114	0.4358	0.2814	0.3174	0.6404	0.3676
平均	0.075	0.733	0.729	0.602	0.545	0.505	0.426	0.274	0.324	0.654	0.379
SD	0.002	0.099	0.054	0.006	0.028	0.013	0.012	0.007	0.006	0.015	0.011
Average-Blank	0.000	0.658	0.654	0.527	0.470	0.430	0.351	0.199	0.249	0.580	0.305
Relative cell growth (%)		100.0	99.3	80.1	71.4	65.3	53.4	30.3	37.8	100.0	52.6
SD of relative cell growth(%)		15.1	8.3	0.9	4.2	2.0	1.9	1.0	0.9	2.6	1.9

Transformation Assay		Solvent: DMSO 0.2%									
Compound (ug/ml)	Well No.	0.001	0.01	0.03	0.04	0.05	0.1	1	10	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	0	2	4	6	9	15	17	13	2	19
	2	2	4	4	4	4	8	11	7	3	16
	3	1	0	10	8	13	9	12	11	3	11
	4	1	2	5	4	8	8	9	8	0	20
	5	1	3	8	12	7	17	15	10	3	19
	6	0	4	6	10	9	10	12	7	1	15
Average		0.8	2.5	6.2	7.3	8.3	11.2	12.7	9.3	2.0	16.7
SD		0.8	1.5	2.4	3.3	2.9	3.9	2.9	2.4	1.3	3.4
Dunnett test				**	***	***	***	***	***		
t-Test (Aspin-Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab V

Compound: DB[a,h]A

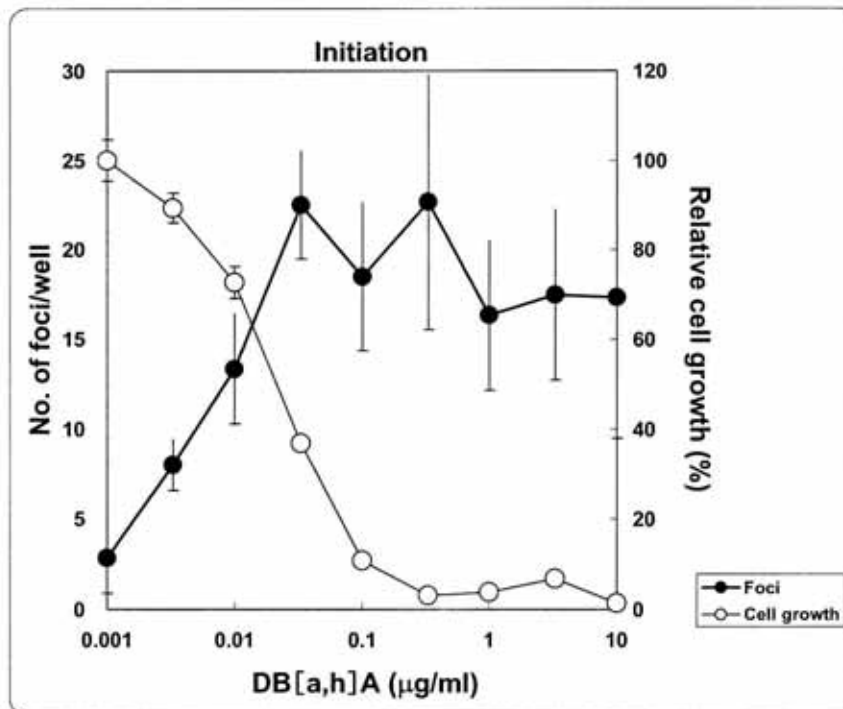
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.1%										
Compound (ug/ml)	Blank	0.001	0.0033	0.01	0.033	0.1	0.33	1	3.3	10	DMSO 0.1%	MCA 1 ug/mL
OD	0.0828 0.127 0.1006	0.4646 0.4991 0.4885	0.429 0.4532 0.4485	0.3666 0.3935 0.3796	0.2448 0.2438 0.2421	0.1455 0.1436 0.1442	0.1157 0.1159 0.1143	0.1165 0.1169 0.1206	0.1328 0.1276 0.1279	0.1091 0.1091 0.1082	0.4743 0.4714 0.4669	0.1833 0.1885 0.1897
Average	0.103	0.484	0.444	0.380	0.244	0.144	0.115	0.118	0.129	0.109	0.471	0.187
SD	0.022	0.018	0.013	0.013	0.001	0.001	0.001	0.002	0.003	0.001	0.004	0.003
Average-Blank	0.000	0.381	0.340	0.276	0.140	0.041	0.012	0.015	0.026	0.005	0.367	0.084
Relative cell growth (%)	100.0	89.4	72.6	36.8	10.8	3.1	3.8	6.8	1.4	100.0	22.8	
SD of relative cell growth(%)		4.6	3.4	3.5	0.4	0.3	0.2	0.6	0.8	0.1	1.0	0.9

Transformation Assay		Solvent: DMSO 0.1%										
Compound (ug/ml)	Well No.	0.001	0.0033	0.01	0.033	0.1	0.33	1	3.3	10	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	1	9	16	22	18	15	11	15	15	2	42
	2	3	7	10	25	17	13	19	27	14	1	43
	3	0	10	10	20	23	25	13	14	20	1	42
	4	4	8	17	22	14	29	14	16	11	1	40
	5	5	8	12	27	15	30	20	17	32	1	48
	6	4	6	15	19	24	24	21	16	12	1	33
Average		2.8	8.0	13.3	22.5	18.5	22.7	16.3	17.5	17.3	1.2	41.3
SD		1.9	1.4	3.1	3.0	4.1	7.1	4.2	4.8	7.8	0.4	4.9
Dunnett test				**	***	***	***	***	***	***		
t-Test for Unequal Variances (Aspin-Welch)												***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: DB[a,h]A

Transformation Assay

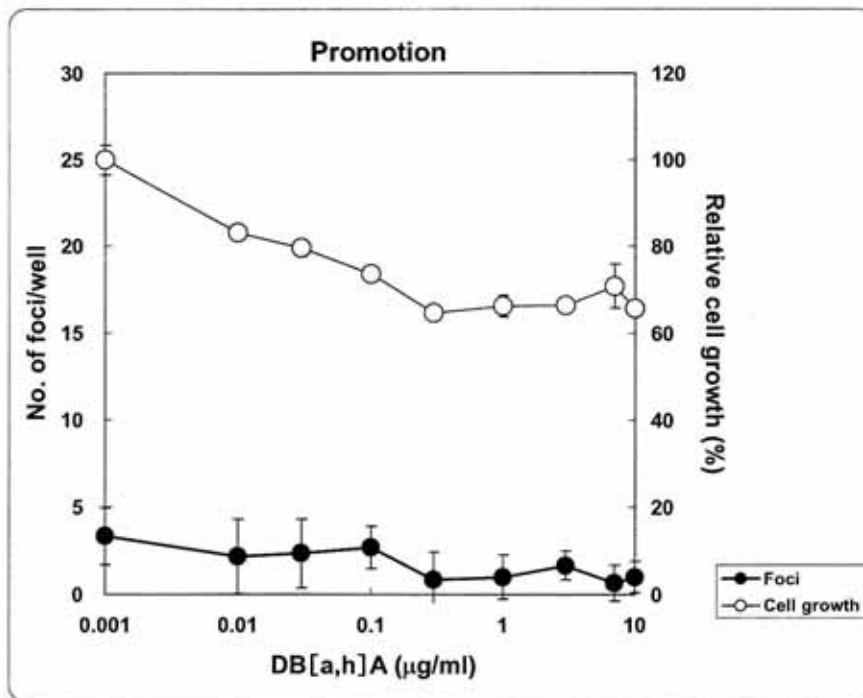
Promotion

Cell Growth Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Blank	0.001	0.01	0.03	0.1	0.3	1	3	7	10	DMSO 0.1%	TPA 50 ng/mL	
OD	0.073	0.978	0.826	0.805	0.749	0.655	0.651	0.688	0.761	0.679	0.978	1.14	
	0.073	1.012	0.835	0.804	0.736	0.655	0.675	0.678	0.717	0.662	1.012	1.148	
	0.077	0.95	0.819	0.776	0.737	0.669	0.697	0.659	0.67	0.666	0.95	1.14	
Average	0.074	0.980	0.827	0.795	0.741	0.660	0.674	0.675	0.716	0.669	0.980	1.143	
SD	0.002	0.031	0.008	0.016	0.007	0.008	0.023	0.015	0.046	0.009	0.031	0.005	
Average-Blank	0.000	0.906	0.752	0.721	0.666	0.585	0.600	0.601	0.642	0.595	0.906	1.068	
Relative cell growth (%)		100.0	83.1	79.6	73.6	64.6	66.2	66.3	70.9	65.7	100.0	118.0	
SD of relative cell growth(%)		3.4	0.9	1.8	0.8	0.9	2.5	1.6	5.0	1.0	3.4	0.5	

Transformation Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Well No.	0.001	0.01	0.03	0.1	0.3	1	3	7	10	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	3	2	4	2	0	1	2	0	1	3	21	
	2	1	1	0	2	0	2	1	0	1	1	20	
	3	3	3	4	2	4	3	1	0	2	3	18	
	4	3	1	2	3	0	0	1	2	0	3	21	
	5	4	6	4	2	0	0	2	0	2	4	16	
	6	6	0	0	5	1	0	3	2	0	6	10	
Average		3.3	2.2	2.3	2.7	0.8	1.0	1.7	0.7	1.0	3.3	17.7	
SD		1.6	2.1	2.0	1.2	1.6	1.3	0.8	1.0	0.9	1.6	4.2	
Dunnett test													
t-Test for Unequal Variances (Aspin-Welch)												***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: DB[a,h]A

Transformation Assay

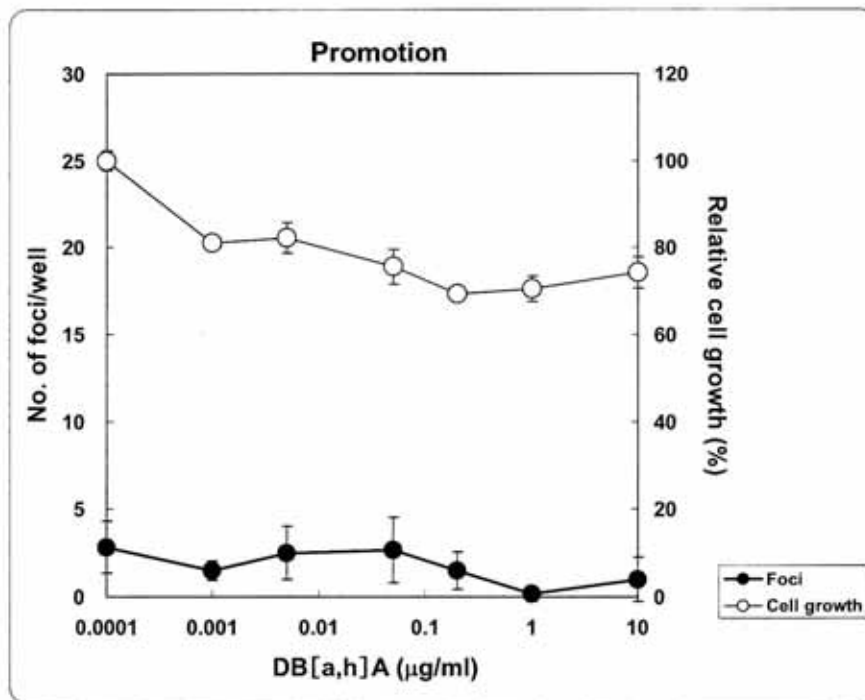
Promotion

Cell Growth Assay		Solvent: DMSO 0.2%									
Compound (ug/ml)	Blank	0.0001	0.001	0.005	0.05	0.2	1	10	DMSO 0.1%	TPA 50 ng/mL	
OD	0.0888	0.524	0.4486	0.468	0.4247	0.3949	0.3842	0.3992	0.5387	0.6938	
	0.0847	0.5422	0.447	0.4488	0.4383	0.3988	0.4055	0.4185	0.5543	0.6414	
	0.0813	0.5257	0.4436	0.4369	0.403	0.3887	0.4081	0.4304	0.4965	0.6272	
平均	0.085	0.531	0.446	0.451	0.422	0.394	0.399	0.416	0.530	0.654	
SD	0.004	0.010	0.003	0.016	0.018	0.005	0.013	0.016	0.030	0.035	
Average-Blank	0.000	0.446	0.361	0.366	0.337	0.309	0.314	0.331	0.445	0.569	
Relative cell growth (%)		100.0	81.1	82.2	75.6	69.4	70.5	74.3	100.0	127.9	
SD of relative cell growth(%)		2.3	0.6	3.5	4.0	1.1	2.9	3.5	6.7	7.9	

Transformation Assay		Solvent: DMSO 0.2%									
Compound (ug/ml)	Well No.	0.0001	0.001	0.005	0.05	0.2	1	10	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	5	1	3	5	2	0	0	3	8	
	2	1	2	2	2	1	0	2	0	13	
	3	2	2	0	2	0	0	3	0	10	
	4	4	1	4	1	3	0	0	4	7	
	5	3	1	4	5	2	0	1	5	11	
	6	2	2	2	1	1	1	0	3	4	
Average		2.8	1.5	2.5	2.7	1.5	0.2	1.0	2.5	8.8	
SD		1.5	0.5	1.5	1.9	1.0	0.4	1.3	2.1	3.2	
Dunnnett test											
t-Test (Assuming Equal variance)										**	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab V

Compound: DB[a,h]A

Transformation Assay

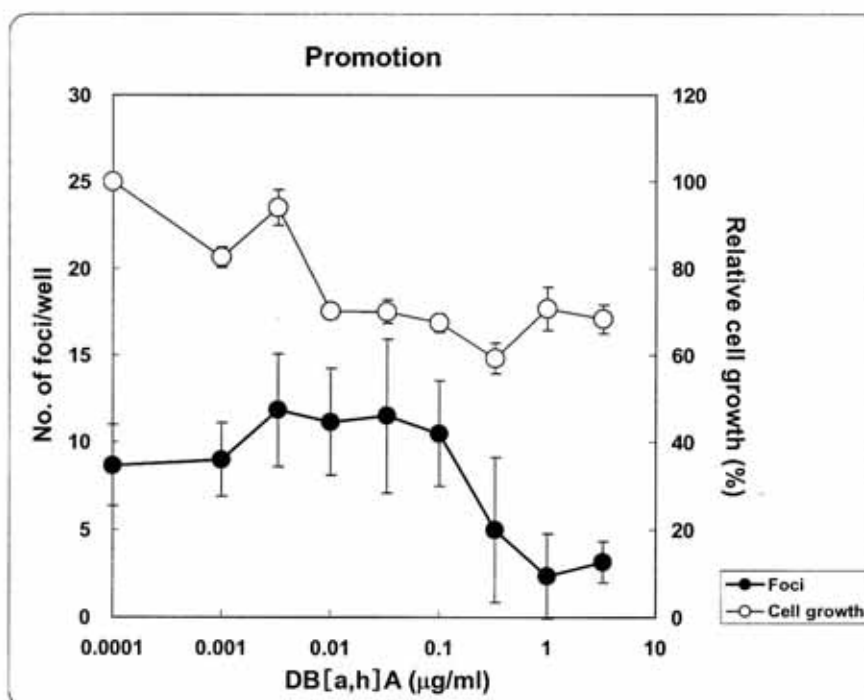
Promotion

Cell Growth Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Blank	0.0001	0.001	0.0033	0.01	0.033	0.1	0.33	1	3.3	DMSO 0.1%	TPA 1 ug/mL	
OD	0.1064	0.5565	0.4771	0.5489	0.4208	0.4333	0.4194	0.3902	0.4454	0.4257	0.5395	0.6993	
	0.1084	0.5556	0.4872	0.5155	0.4236	0.4199	0.4008	0.3596	0.401	0.3971	0.5138	0.6537	
	0.1096	0.5521	0.4662	0.519	0.42	0.4091	0.4076	0.3689	0.425	0.4167	0.5176	0.6675	
Average	0.108	0.555	0.477	0.528	0.421	0.421	0.409	0.373	0.424	0.413	0.524	0.674	
SD	0.002	0.002	0.011	0.018	0.002	0.012	0.009	0.016	0.022	0.015	0.014	0.023	
Average-Blank	0.000	0.447	0.369	0.420	0.313	0.313	0.301	0.265	0.316	0.305	0.416	0.565	
Relative cell growth (%)		100.0	82.6	94.0	70.2	70.0	67.4	59.3	70.7	68.3	100.0	136.1	
SD of relative cell growth(%)		0.5	2.4	4.1	0.4	2.7	2.1	3.5	5.0	3.3	3.3	5.6	

Transformation Assay		Solvent: DMSO 0.1%											
Compound (ug/ml)	Well No.	0.0001	0.001	0.0033	0.01	0.033	0.1	0.33	1	3.3	DMSO 0.1%	TPA1 ug/mL	
No of foci	1	12	12	11	14	15	10	3	3	3	12	44	
	2	8	11	16	9	11	13	5	6	4	8	42	
	3	5	7	7	15	6	13	10	4	2	5	40	
	4	8	7	11	10	8	5	1	0	5	8	39	
	5	10	8	11	12	18	10	1	0	3	10	46	
	6	9	9	15	7	11	12	10	1	2	9	50	
Average		8.7	9.0	11.8	11.2	11.5	10.5	5.0	2.3	3.2	8.7	43.5	
SD		2.3	2.1	3.3	3.1	4.4	3.0	4.1	2.4	1.2	2.3	4.1	
Dunnett test													
t-Test for Unequal Variances (Aspin-Welch)													

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Methapyrilene HCl

Transformation Assay

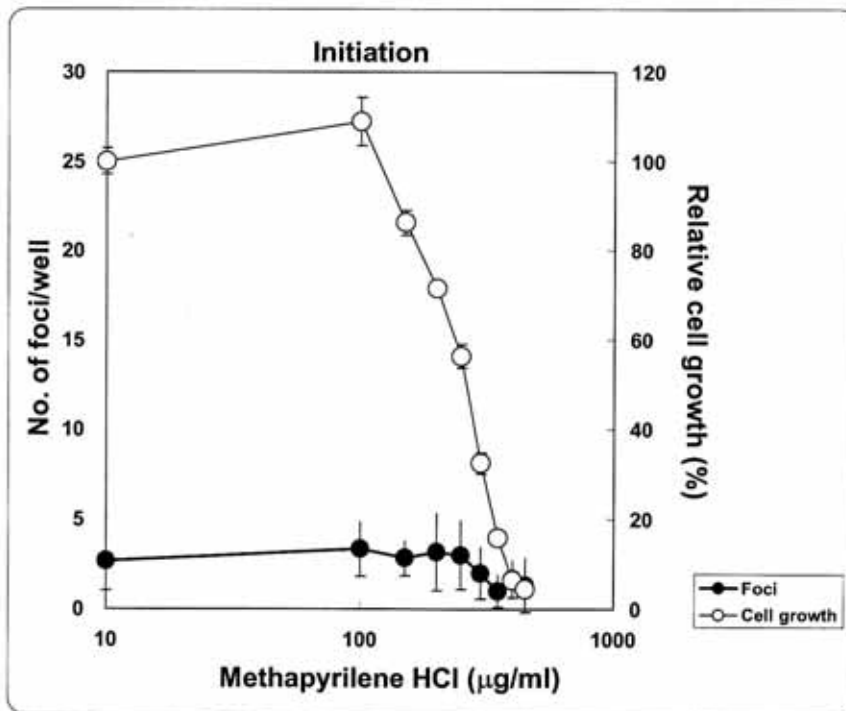
Initiation

Cell Growth Assay		Solvent: water 5%											
Compound (ug/ml)	Blank	10	100	150	200	250	300	350	400	450	DMSO 0.1%	MCA 1 ug/mL	
OD	0.08	1.284	1.359	1.123	0.994	0.822	0.516	0.299	0.174	0.151	1.26	0.637	
	0.088	1.342	1.475	1.15	0.959	0.761	0.489	0.266	0.155	0.138	1.248	0.599	
	0.09	1.352	1.475	1.192	0.964	0.769	0.458	0.282	0.169	0.134	1.286	0.607	
Average	0.086	1.326	1.436	1.155	0.972	0.784	0.488	0.282	0.166	0.141	1.265	0.614	
SD	0.005	0.037	0.067	0.035	0.019	0.033	0.029	0.017	0.010	0.009	0.019	0.020	
Average-Blank	0.000	1.240	1.350	1.069	0.886	0.698	0.402	0.196	0.080	0.055	1.179	0.528	
Relative cell growth (%)		100.0	108.9	86.2	71.5	56.3	32.4	15.8	6.5	4.4	100.0	44.8	
SD of relative cell growth(%)		3.0	5.4	2.8	1.5	2.7	2.3	1.3	0.8	0.7	1.6	1.7	

Transformation Assay		Solvent: water 5%											
Compound (ug/ml)	Well No.	10	100	150	200	250	300	350	400	450	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	1	3	4	2	5	0	1	2	4	1	26	
	2	3	2	2	3	3	2	0	2	2	2	30	
	3	2	6	2	1	0	4	1	2	1	3	35	
	4	4	2	2	2	5	3	2	1	1	4	27	
	5	5	4	4	4	7	2	1	0	0	3	26	
	6	1	3	3	4	3	2	2	2	3	0	5	26
Average		2.7	3.3	2.8	3.2	3.0	2.0	1.0	1.7	1.3	3.0	28.3	
SD		1.6	1.5	1.0	2.1	1.9	1.4	0.9	1.0	1.5	1.4	3.6	
Dunnett test													
t-Test for Unequal Variances (Aspin-Welch)												***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Methapyrilene HCl

Transformation Assay

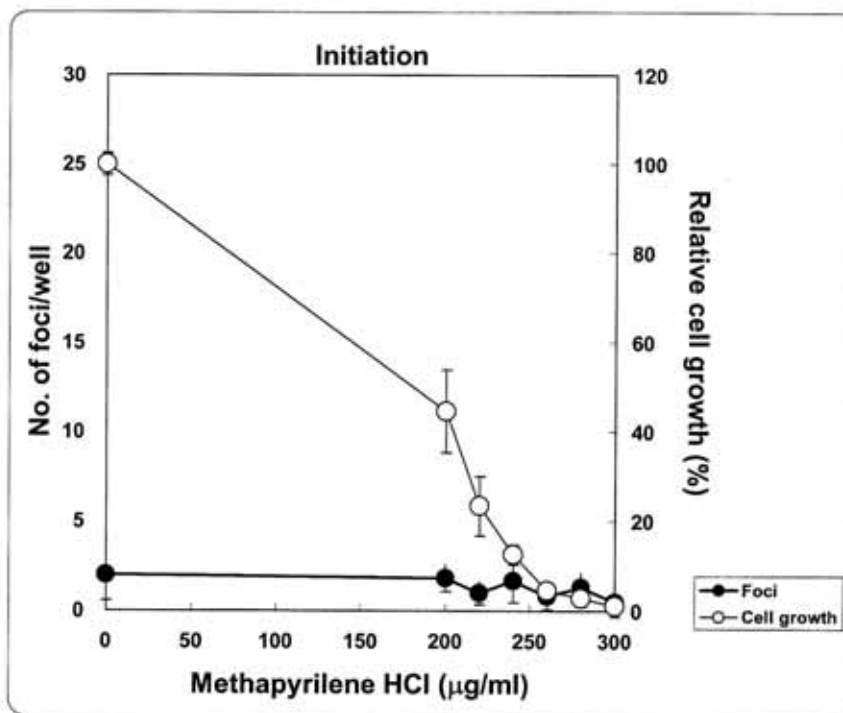
Initiation

Cell Growth Assay		Solvent: Water 5%									
Compound (ug/ml)	Blank	0	200	220	240	260	280	300	DMSO 0.1%	MCA 1 ug/mL	
OD	0.0715	0.802	0.3184	0.1948	0.1465	0.1029	0.081	0.0818	0.6566	0.3978	
	0.0764	0.7666	0.446	0.2396	0.1758	0.1023	0.1021	0.0815	0.7332	0.421	
	0.0793	0.7823	0.4083	0.2884	0.1704	0.1188	0.106	0.0877	0.6764	0.4037	
Average	0.076	0.784	0.391	0.241	0.164	0.108	0.096	0.084	0.689	0.408	
SD	0.004	0.018	0.066	0.047	0.016	0.009	0.013	0.003	0.040	0.012	
Average-Blank	0.000	0.708	0.315	0.165	0.089	0.032	0.021	0.008	0.613	0.332	
Relative cell growth (%)		100.0	44.5	23.3	12.5	4.6	2.9	1.1	100.0	54.1	
SD of relative cell growth(%)		2.5	9.3	6.6	2.2	1.3	1.9	0.5	6.5	2.0	

Transformation Assay		Solvent: Water 5%									
Compound (ug/ml)	Well No.	0	200	220	240	260	280	300	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	3	2	1	2	1	2	0	2	19	
	2	2	1	0	3	2	2	0	1	18	
	3	0	1	1	3	0	1	0	0	19	
	4	2	3	2	1	1	2	2	2	12	
	5	1	2	1	1	1	1	0	3	16	
	6	4	2	1	0	0	0	1	3	21	
Average		2.0	1.8	1.0	1.7	0.8	1.3	0.5	1.8	17.5	
SD		1.4	0.8	0.6	1.2	0.8	0.8	0.8	1.2	3.1	
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)										***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Methapyrilene HCl

Transformation Assay

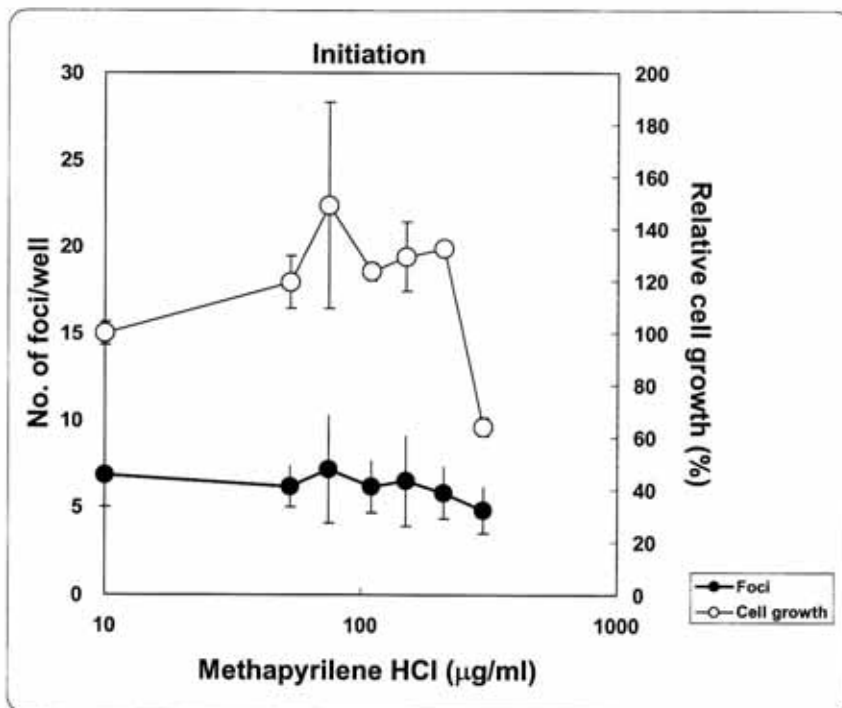
Initiation

Cell Growth Assay		Solvent: Water 5%									
Compound (ug/ml)	Blank	10	53	75	110	150	210	300	DMSO 0.1%	MCA 1 ug/mL	
OD	0.032	0.223	0.247	0.29	0.265	0.275	0.277	0.145	0.198	0.148	
	0.035	0.207	0.271	0.383	0.253	0.289	0.273	0.157	0.215	0.153	
	0.033	0.216	0.235	0.241	0.257	0.242	0.273	0.147	0.192	0.15	
Average	0.033	0.215	0.251	0.305	0.258	0.269	0.274	0.150	0.202	0.150	
SD	0.002	0.008	0.018	0.072	0.006	0.024	0.002	0.006	0.012	0.003	
Average-Blank	0.000	0.182	0.218	0.271	0.225	0.235	0.241	0.116	0.168	0.117	
Relative cell growth (%)		100.0	119.6	149.1	123.6	129.3	132.4	63.9	100.0	69.5	
SD of relative cell growth(%)		4.4	10.1	39.6	3.4	13.3	1.3	3.5	7.1	1.5	

Transformation Assay		Solvent: Water 5%									
Compound (ug/ml)	Well No.	10	53	75	110	150	210	300	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	6	8	8	6	8	6	4	4	45	
	2	8	5	6	7	5	5	7	6	40	
	3	9	6	3	8	9	5	5	8	47	
	4	8	6	11	7	7	7	5	4	41	
	5	4	5	5	4	2	8	5	6	38	
	6	6	7	10	5	8	4	3	5	40	
Average		6.8	6.2	7.2	6.2	6.5	5.8	4.8	5.5	41.8	
SD		1.8	1.2	3.1	1.5	2.6	1.5	1.3	1.5	3.4	
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)										***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Methapyrilene HCl

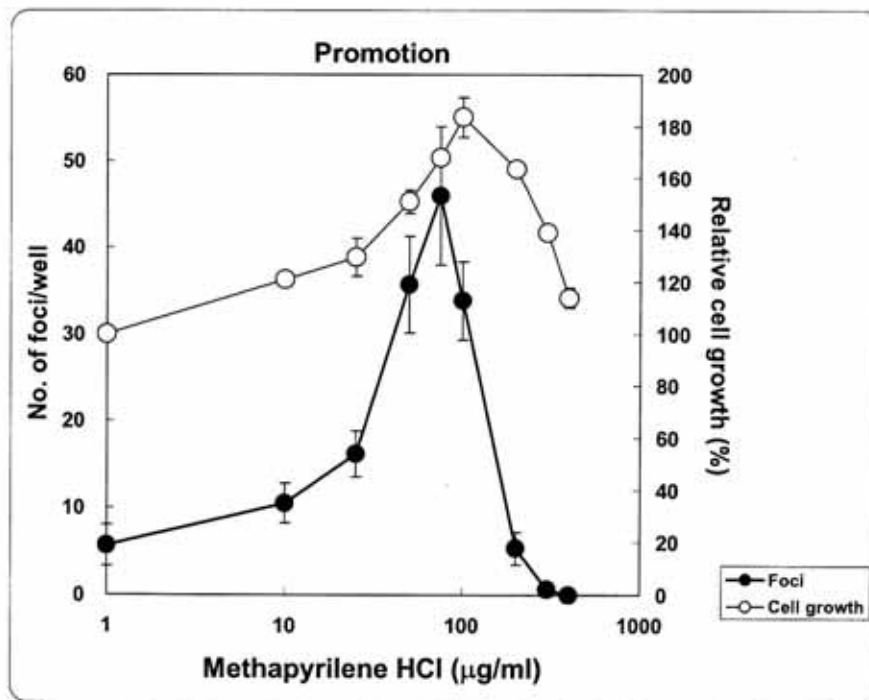
Transformation Assay

Promotion

Cell Growth Assay		Solvent: Water 5%											
Compound (ug/ml)	Blank	Water 5%	1	10	25	50	75	100	200	300	400	DMSO 0.1%	TPA 50 ng/mL
OD	0.073	1.047	1.277	1.415	1.6	1.72	1.954	1.678	1.458	1.229		1.006	1.416
	0.073	1.062	1.238	1.271	1.54	1.712	1.81	1.67	1.41	1.156		1.08	1.287
	0.077	1.048	1.259	1.337	1.513	1.72	1.844	1.673	1.438	1.179		1.094	1.258
Average	0.074	1.052	1.258	1.341	1.551	1.717	1.869	1.674	1.435	1.188		1.060	1.320
SD	0.002	0.008	0.020	0.072	0.045	0.005	0.075	0.004	0.024	0.037		0.047	0.084
Average-Blank	0.000	0.978	1.184	1.267	1.477	1.643	1.795	1.599	1.361	1.114		0.986	1.246
Relative cell growth (%)		100.0	121.0	129.5	151.0	168.0	183.5	163.5	139.2	113.9		100.0	126.4
SD of relative cell growth(%)		0.9	2.0	7.4	4.6	0.5	7.7	0.4	2.5	3.8		4.8	8.5

Transformation Assay		Solvent: Water 5%											
Compound (ug/ml)	Well No.	Water 5%	1	10	25	50	75	100	200	300	400	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	7	12	15	40	56	26	7	1	Tox		6	35
	2	6	8	14	42	39	31	5	2	Tox		6	39
	3	3	11	20	30	41	35	8	1	Tox		3	47
	4	9	9	18	38	53	36	4	0	Tox		6	40
	5	3	14	13	36	50	38	5	0	Tox		6	41
	6	6	9	17	28	37	37	3	0	Tox		3	36
Average		5.7	10.5	16.2	35.7	46.0	33.8	5.3	0.7	#DIV/0!		5.0	39.7
SD		2.3	2.3	2.6	5.6	8.0	4.5	1.9	0.8	#DIV/0!		1.5	4.3
Dunnett test					***	***	***	***					
t-Test for Unequal Variances (Aspin-Welch)													***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: **Lah II**

Compound: **Methapyrilene HCl**

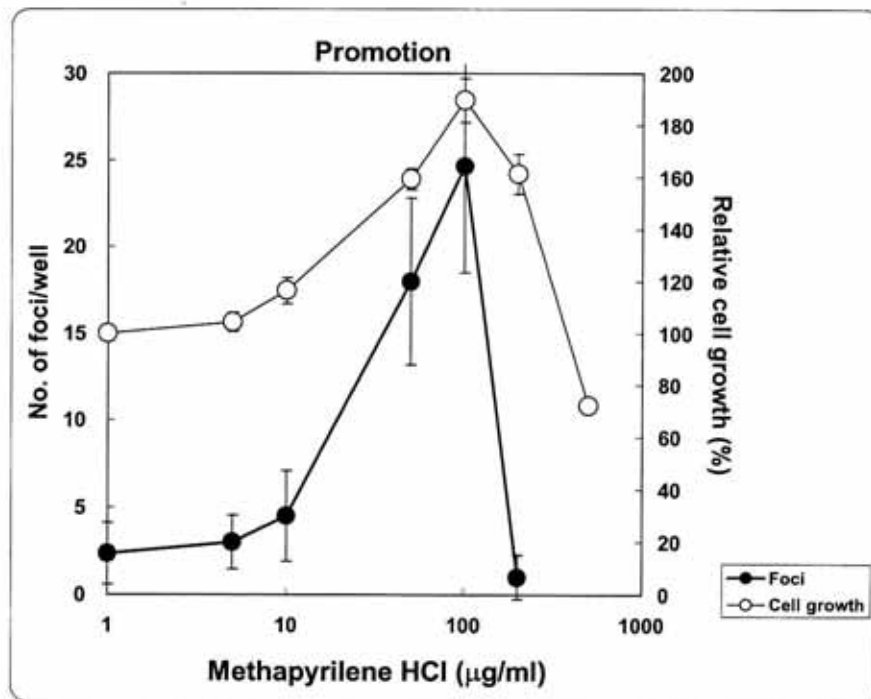
Transformation Assay

Promotion

Cell Growth Assay		Solvent: Water 5%								
Compound (ug/ml)	Blank	1	5	10	50	100	200	500	DMSO 0.1%	TPA 50 ng/mL
OD	0.0736	0.5376	0.5797	0.611	0.8302	1.0048	0.8057	0.4192		
	0.0761	0.544	0.5608	0.6455	0.8329	0.9277	0.8709	0.4056		
	0.0721	0.5455	0.5462	0.6007	0.7989	0.9542	0.8123	0.4136		
平均	0.074	0.542	0.562	0.619	0.821	0.962	0.830	0.413	#DIV/0!	#DIV/0!
SD	0.002	0.004	0.017	0.023	0.019	0.039	0.036	0.007	#DIV/0!	#DIV/0!
Average-Blank	0.000	0.468	0.488	0.545	0.747	0.888	0.756	0.339	#DIV/0!	#DIV/0!
Relative cell growth (%)		100.0	104.2	116.4	159.4	189.6	161.3	72.3	#DIV/0!	#DIV/0!
SD of relative cell growth(%)		0.9	3.6	5.0	4.0	8.4	7.7	1.5	#DIV/0!	#DIV/0!

Transformation Assay		Solvent: Water 5%								
Compound (ug/ml)	Well No.	1	5	10	50	100	200	500	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	5	2	5	22	26	3	Toxic	1	8
	2	0	2	8	15	15	0	Toxic	5	10
	3	1	3	7	15	20	1	Toxic	2	12
	4	3	2	3	15	26	0	Toxic	4	5
	5	3	3	2	15	32	2	Toxic	3	11
	6	2	6	2	26	29	0	Toxic	1	6
Average		2.3	3.0	4.5	18.0	24.7	1.0		2.7	8.7
SD		1.8	1.5	2.6	4.8	6.2	1.3		1.6	2.8
Dunnett test					***	***				
t-Test for Unequal Variances (Aspin-Welch)										**

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Methapyrilene HCl

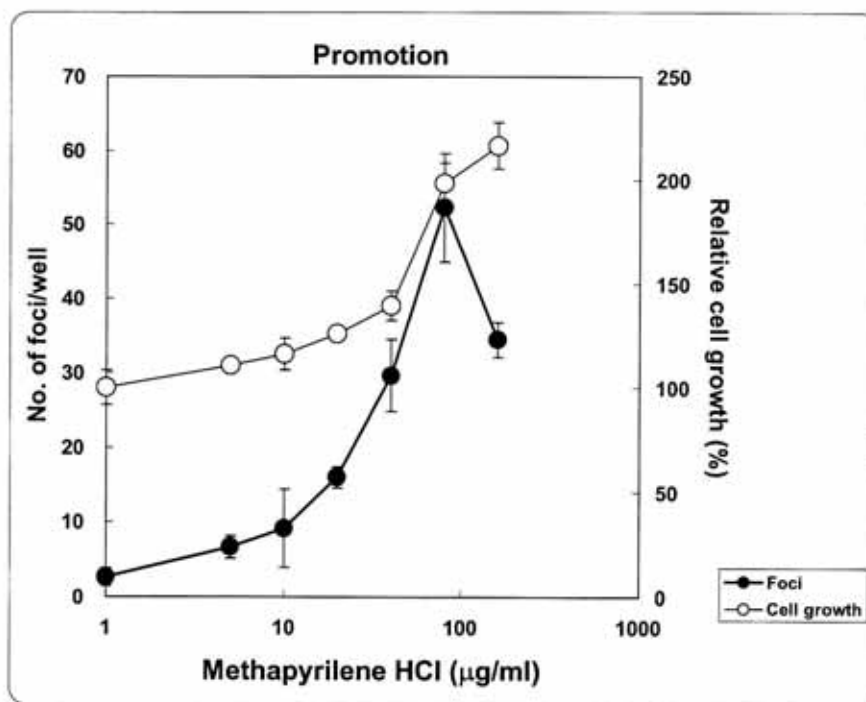
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DW 5%								
Compound (ug/ml)	Blank	1	5	10	20	40	80	160	DMSO 0.1%	TPA 50 ng/mL
OD	0.062	0.362	0.389	0.405	0.417	0.479	0.629	0.676		0.433
	0.061	0.32	0.379	0.369	0.416	0.439	0.603	0.654		0.391
	0.058	0.36	0.368	0.409	0.433	0.465	0.659	0.717		0.41
Average	0.060	0.347	0.379	0.394	0.422	0.461	0.630	0.682	#DIV/0!	0.411
SD	0.002	0.024	0.011	0.022	0.010	0.020	0.028	0.032	#DIV/0!	0.021
Average-Blank	0.000	0.287	0.318	0.334	0.362	0.401	0.570	0.622	#DIV/0!	0.351
Relative cell growth (%)		100.0	110.9	116.4	126.0	139.6	198.6	216.7	#DIV/0!	#DIV/0!
SD of relative cell growth(%)		8.3	3.7	7.7	3.3	7.1	9.8	11.1	#DIV/0!	#DIV/0!

Transformation Assay		Solvent: DW 5%								
Compound (ug/ml)	Well No.	1	5	10	20	40	80	160	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	2	7	8	18	33	59	32	3	22
	2	3	4	10	16	32	52	35	1	22
	3	1	6	19	17	30	48	32	3	13
	4	4	8	8	15	32	61	34	3	17
	5	2	7	6	16	20	53	36	2	15
	6	4	8	4	14	31	41	38	3	23
Average		2.7	6.7	9.2	16.0	29.7	52.3	34.5	2.5	18.7
SD		1.2	1.5	5.2	1.4	4.8	7.3	2.3	0.8	4.2
Dunnett test				*	***	***	***	***		
t-Test for Unequal Variances (Aspin-Welch)										***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Sodium arsenite

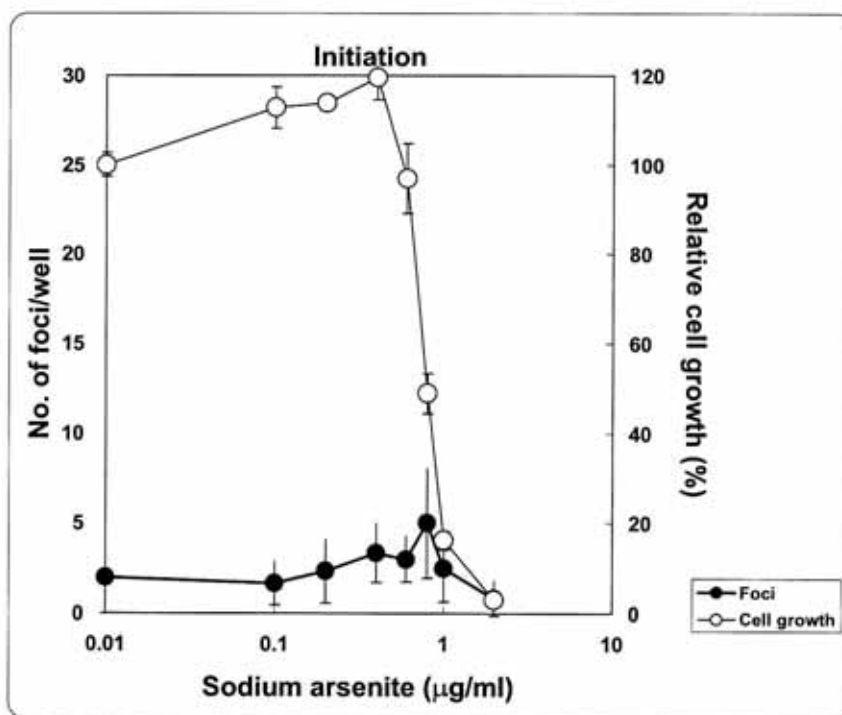
Transformation Assay

Initiation-No.: 1

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	0.01	0.1	0.2	0.4	0.6	0.8	1	2	DMSO 0.1%	MCA 1 ug/mL
OD	0.073	0.905	1.011	0.994	1.005	0.817	0.479	0.201	0.101	0.871	0.596
	0.073	0.863	1.011	0.988	1.041	0.832	0.43	0.191	0.096	0.934	0.584
	0.077	0.887	0.946	1.012	1.084	0.933	0.502	0.224	0.098	0.975	0.579
Average	0.074	0.885	0.989	0.998	1.043	0.861	0.470	0.205	0.098	0.927	0.586
SD	0.002	0.021	0.038	0.012	0.040	0.063	0.037	0.017	0.003	0.052	0.009
Average-Blank	0.000	0.811	0.915	0.924	0.969	0.786	0.396	0.131	0.024	0.852	0.512
Relative cell growth (%)		100.0	112.9	113.9	119.5	97.0	48.8	16.2	3.0	100.0	60.1
SD of relative cell growth(%)		2.6	4.6	1.5	4.9	7.8	4.5	2.1	0.3	6.1	1.0

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	0.01	0.1	0.2	0.4	0.6	0.8	1	2	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	1	2	4	3	2	5	1	2	3	39
	2	5	1	2	3	2	8	6	0	2	44
	3	1	3	1	6	4	3	2	0	5	43
	4	4	1	5	3	3	1	1	1	5	42
	5	0	3	1	1	2	4	3	2	2	42
	6	1	0	1	4	5	9	2	0	1	38
Average		2.0	1.7	2.3	3.3	3.0	5.0	2.5	0.8	3.0	41.3
SD		2.0	1.2	1.8	1.6	1.3	3.0	1.9	1.0	1.7	2.3
Dunnett test							*				
t-Test (Assuming Equal Variances)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Sodium arsenite

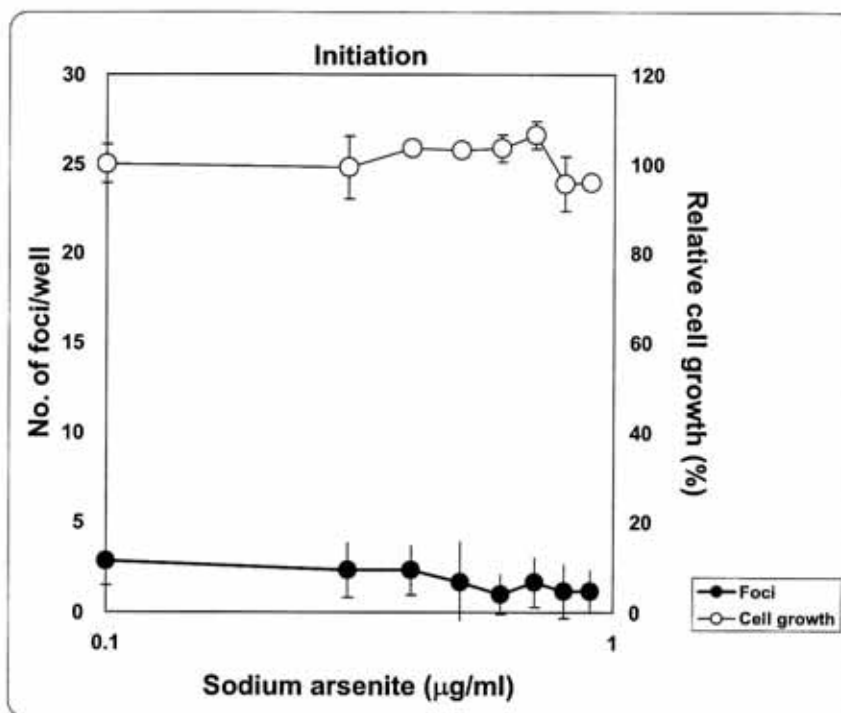
Transformation Assay

Initiation

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	0.1	0.3	0.4	0.5	0.6	0.7	0.8	0.9	DMSO 0.1%	MCA 1 ug/mL
OD	0.08 0.088 0.09	1.44 1.327 1.388	1.48 1.329 1.315	1.441 1.406 1.445	1.407 1.441 1.426	1.465 1.386 1.438	1.506 1.428 1.472	1.312 1.256 1.412	1.324 1.346 1.322	1.24 1.327 1.526	0.639 0.648 0.773
Average	0.086	1.385	1.375	1.431	1.425	1.430	1.469	1.327	1.331	1.364	0.687
SD	0.005	0.057	0.091	0.021	0.017	0.040	0.039	0.079	0.013	0.147	0.075
Average-Blank	0.000	1.299	1.289	1.345	1.339	1.344	1.383	1.241	1.245	1.278	0.601
Relative cell growth (%)		100.0	99.2	103.5	103.1	103.4	106.4	95.5	95.8	100.0	47.0
SD of relative cell growth(%)		4.4	7.0	1.7	1.3	3.1	3.0	6.1	1.0	11.5	5.9

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	0.1	0.3	0.4	0.5	0.6	0.7	0.8	0.9	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	2	3	2	6	0	1	1	1	2	33
	2	2	2	2	2	3	2	0	0	2	33
	3	2	1	3	0	0	1	1	0	2	42
	4	5	1	0	1	1	0	0	1	5	31
	5	4	2	3	1	1	4	1	3	4	21
	6	2	5	4	0	1	2	4	2	2	32
Average		2.8	2.3	2.3	1.7	1.0	1.7	1.2	1.2	2.8	32.0
SD		1.3	1.5	1.4	2.3	1.1	1.4	1.5	1.2	1.3	6.7
Dunnett test											N.S.
t-Test for Unequal Variances (Aspin-Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Sodium arsenite

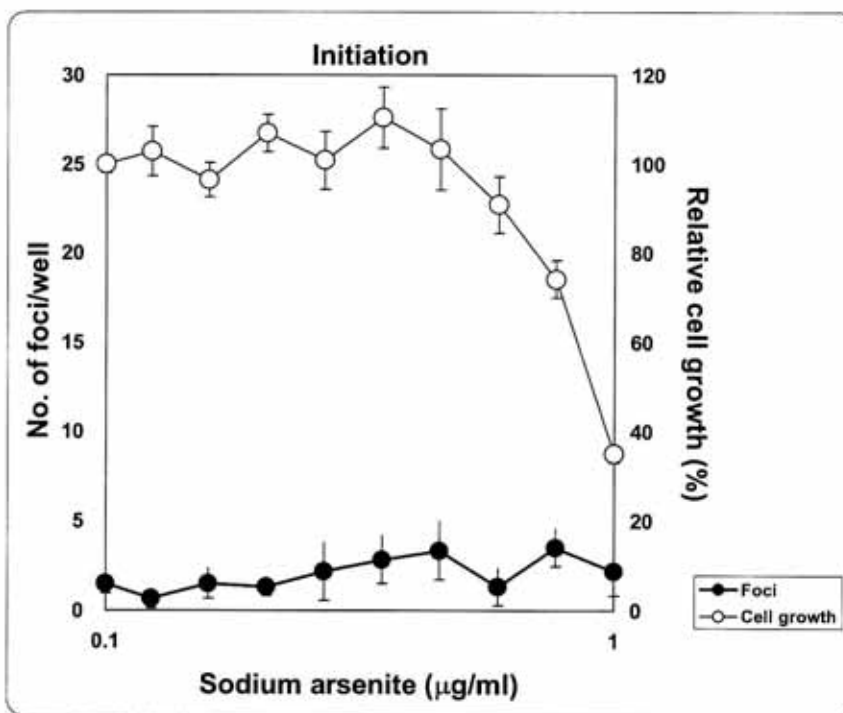
Transformation Assay

Initiation

Cell Growth Assay		Solvent: D.W. 5 %*										
Compound (ug/ml)	Blank	0.1	0.123	0.159	0.207	0.269	0.35	0.455	0.592	0.769	1	DMSO 0.1% MCA 1 ug/mL
OD	0.226	2.617	2.501	2.562	2.633	2.43	2.643	2.418	2.2	2.049	1.013	0.677
	0.26	2.533	2.743	2.544	2.798	2.7	2.908	2.809	2.471	2.022	1.093	0.761
	0.257	2.603	2.708	2.399	2.811	2.68	2.934	2.76	2.44	1.868	1.084	0.855
Average	0.248	2.584	2.651	2.502	2.747	2.603	2.828	2.662	2.370	1.980	1.063	#DIV/0!
SD	0.019	0.045	0.131	0.089	0.099	0.150	0.161	0.213	0.148	0.098	0.044	#DIV/0!
Average-Blank	0.000	2.337	2.403	2.254	2.500	2.356	2.581	2.415	2.123	1.732	0.816	#DIV/0!
Relative cell growth (%)		100.0	102.8	96.5	107.0	100.8	110.4	103.3	90.8	74.1	34.9	#DIV/0!
SD of relative cell growth(%)		1.9	5.6	3.8	4.2	6.4	6.9	9.1	6.3	4.2	1.9	#DIV/0!

Transformation Assay		Solvent: D.W. 5 %*										
Compound (ug/ml)	Well No.	0.1	0.123	0.159	0.207	0.269	0.35	0.455	0.592	0.769	1	DMSO 0.1% MCA 1 ug/mL
No of foci	1	1	1	3	1	2	2	2	0	3	2	-
	2	2	0	1	1	0	1	6	1	5	0	-
	3	1	1	1	1	4	4	2	2	4	3	-
	4	1	1	1	1	4	4	4	3	3	4	-
	5	2	0	2	2	2	4	2	1	2	2	-
	6	2	1	1	2	1	2	4	1	4	2	-
Average		1.5	0.7	1.5	1.3	2.2	2.8	3.3	1.3	3.5	2.2	#DIV/0!
SD		0.5	0.5	0.8	0.5	1.6	1.3	1.6	1.0	1.0	1.3	#DIV/0!
Parametric Dannett								*		*		
t-test for Unequal Variances (Aspin-Welch) vs 5% Water												***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Sodium arsenite

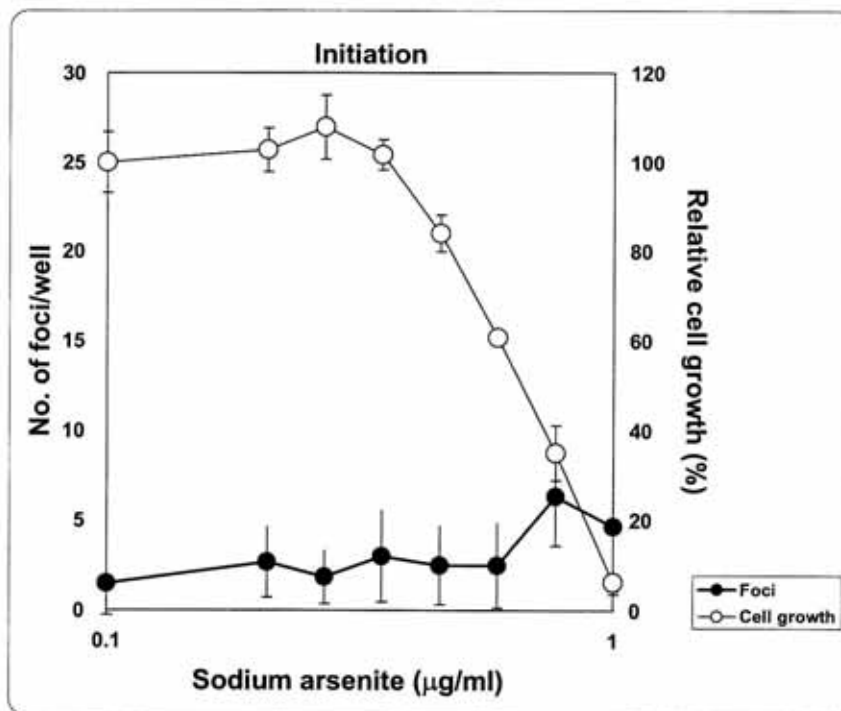
Transformation Assay

Initiation

Cell Growth Assay		Solvent: D.W. 5%											
Compound (ug/ml)	Blank	0.1	0.207	0.269	0.35	0.455	0.592	0.769	1	DMSO 0.1%	MCA 1 ug/mL		
OD	0.242	2.977	2.983	3.196	2.919	2.274	1.78	1.25	0.355	2.78	0.78		
	0.239	2.75	2.854	2.854	2.744	2.482	1.807	0.96	0.447	2.985	0.825		
	0.271	2.636	2.735	2.913	2.827	2.39	1.788	1.204	0.423	3.442	0.887		
Average	0.251	2.788	2.857	2.988	2.830	2.382	1.792	1.138	0.408	3.069	0.831		
SD	0.018	0.174	0.124	0.183	0.088	0.104	0.014	0.156	0.048	0.339	0.054		
Average-Blank	0.000	2.537	2.607	2.737	2.579	2.131	1.541	0.887	0.158	2.818	0.580		
Relative cell growth (%)		100.0	102.7	107.9	101.7	84.0	60.7	35.0	6.2	100.0	20.6		
SD of relative cell growth(%)		6.8	4.9	7.2	3.5	4.1	0.5	6.1	1.9	12.0	1.9		

Transformation Assay		Solvent: D.W. 5%											
Compound (ug/ml)	Well No.	0.1	0.207	0.269	0.35	0.455	0.592	0.769	1	DMSO 0.1%	MCA 1 ug/mL		
No of foci	1	4	2	3	0	5	2	6	2	4	5		
	2	0	5	2	1	1	1	4	8	4	8		
	3	2	4	1	3	3	7	4	10	4	9		
	4	3	0	0	6	5	3	11	3	3	10		
	5	0	1	4	6	0	1	5	0	4	16		
	6	0	4	1	2	1	1	8	5	1	10		
Average		1.5	2.7	1.8	3.0	2.5	2.5	6.3	4.7	3.3	9.7		
SD		1.8	2.0	1.5	2.5	2.2	2.3	2.7	3.8	1.2	3.6		
Dunnett test								**					
t-Test for Unequal Variances (Aspin-Welch)												**	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Sodium arsenite

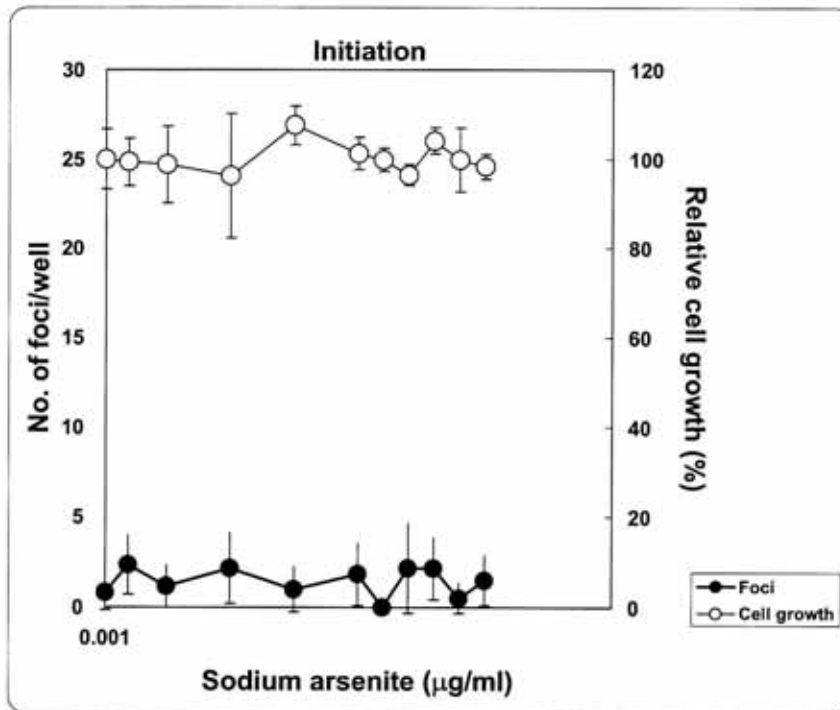
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.5%												
Compound (ug/ml)	Blank	0.001	0.01	0.025	0.05	0.075	0.1	0.11	0.12	0.13	0.14	0.15	DMSO 0.5% MCA 1 ug/ml	
OD	0.032	0.572	0.601	0.554	0.601	0.608	0.596	0.593	0.556	0.618	0.599	0.551	0.572	0.455
	0.033	0.617	0.578	0.624	0.601	0.646	0.596	0.567	0.572	0.588	0.600	0.577	0.617	0.475
	0.03	0.544	0.543	0.535	0.469	0.603	0.562	0.571	0.547	0.594	0.532	0.578	0.544	0.460
Average	0.032	0.578	0.574	0.571	0.557	0.619	0.585	0.577	0.558	0.600	0.577	0.569	0.578	0.463
SD	0.002	0.037	0.029	0.047	0.076	0.024	0.020	0.014	0.013	0.016	0.039	0.015	0.037	0.010
Average-Blank	0.000	0.546	0.542	0.539	0.525	0.587	0.553	0.545	0.527	0.568	0.545	0.537	0.546	0.432
Relative cell growth (%)		100.0	99.3	98.8	96.2	107.6	101.3	99.9	96.5	104.1	99.9	98.4	100.0	79.1
SD of relative cell growth(%)		6.7	5.3	8.6	14.0	4.3	3.6	2.6	2.3	2.9	7.1	2.8	6.7	1.9

Transformation Assay		Solvent: DMSO 0.5%												
Compound (ug/ml)	Well No.	0.001	0.01	0.025	0.05	0.075	0.1	0.11	0.12	0.13	0.14	0.15	DMSO 0.5% MCA 1 ug/ml	
No of foci	1	2	1	1	4	3	4	0	1	4	1	0	2	18
	2	1	2	0	0	0	4	0	7	3	2	3	1	14
	3	0	4	1	1	0	1	0	2	0	0	1	0	9
	4	0	3	2	2	0	1	0	1	3	0	3	0	12
	5	0	0	3	1	2	0	0	2	3	0	0	0	10
	6	2	4	0	5	1	1	0	0	0	0	0	2	9
Average		0.8	2.3	1.2	2.2	1.0	1.8	0.0	2.2	2.2	0.5	1.5	0.8	12.0
SD		1.0	1.6	1.2	1.9	1.3	1.7	0.0	2.5	1.7	0.8	1.4	1.0	3.5
Parametric Dunnett t-Test for Unequal Variances (Aspin-Welch)														N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab 1

Compound: Sodium arsenite

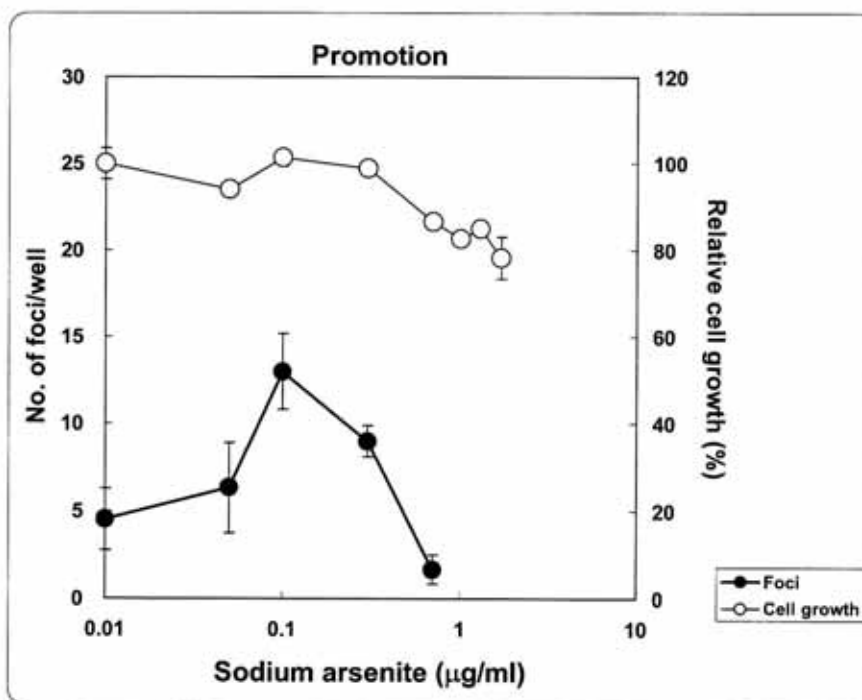
Transformation Assay

Promotion

Cell Growth Assay		Solvent: Water 5%									
Compound (ug/ml)	Blank	0.01	0.05	0.1	0.3	0.7	1	1.3	1.7	DMSO 0.1%	TPA 50 ng/mL
OD	0.073	0.954	0.871	0.948	0.922	0.82	0.771	0.787	0.773	1.003	1.21
	0.073	0.896	0.869	0.917	0.904	0.796	0.774	0.789	0.692	1.044	1.331
	0.077	0.911	0.87	0.929	0.907	0.802	0.773	0.802	0.743	1.057	1.369
Average	0.074	0.920	0.870	0.931	0.911	0.806	0.773	0.793	0.736	1.035	1.303
SD	0.002	0.030	0.001	0.016	0.010	0.012	0.002	0.008	0.041	0.028	0.083
Average-Blank	0.000	0.846	0.796	0.857	0.837	0.732	0.698	0.718	0.662	0.960	1.229
Relative cell growth (%)		100.0	94.1	101.3	98.9	86.5	82.5	84.9	78.2	100.0	128.0
SD of relative cell growth(%)		3.6	0.1	1.8	1.1	1.5	0.2	1.0	4.8	2.9	8.6

Transformation Assay		Solvent: Water 5%									
Compound (ug/ml)	Well No.	0.01	0.05	0.1	0.3	0.7	1	1.3	1.7	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	6	3	13	8	3 Tox	Tox	Tox		3	32
	2	4	6	12	9	2 Tox	Tox	Tox		3	38
	3	6	10	12	8	1 Tox	Tox	Tox		4	42
	4	2	8	16	10	1 Tox	Tox	Tox		8	36
	5	3	7	10	9	2 Tox	Tox	Tox		4	35
	6	6	4	15	10	1 Tox	Tox	Tox		8	37
Average		4.5	6.3	13.0	9.0	1.7				5.0	36.7
SD		1.8	2.6	2.2	0.9	0.8				2.4	3.3
Dunnett				***	***						***
t-Test (Assuming Equal Variances)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Sodium arsenite

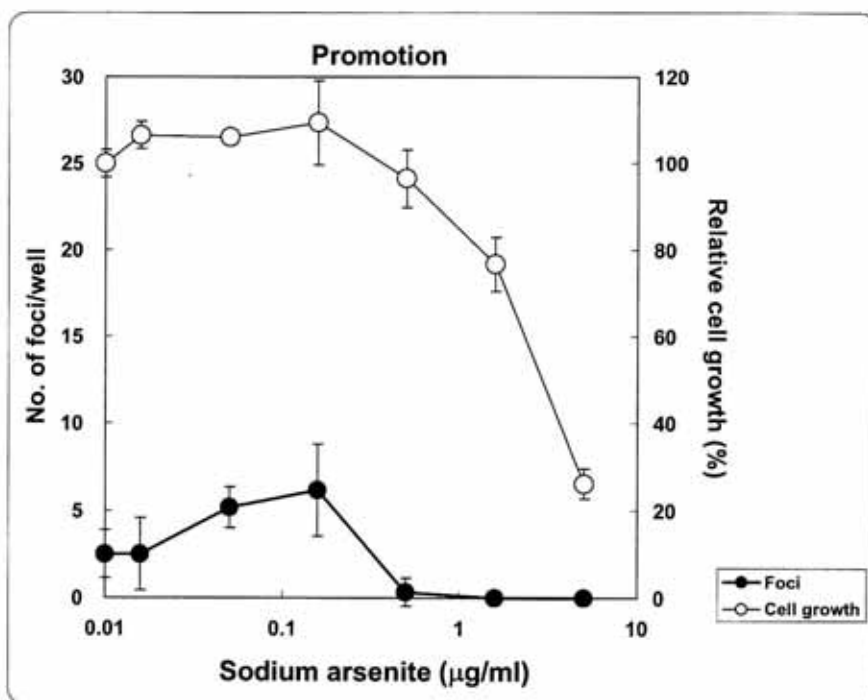
Transformation Assay

Promotion

Cell Growth Assay		Solvent: D.W. 5 %*										
Compound (ug/ml)	Blank	0.01	0.0158	0.05	0.158	0.5	1.58	5	DMSO 0.1%	TPA 50 ng/mL		
OD	0.19	1.941	2.048	2.075	2.363	1.803	1.448	0.602	-	-	2.721	
	0.206	1.986	2.153	2.094	2.084	2.04	1.619	0.724	-	-	3.017	
	0.2	2.053	2.133	2.138	2.039	1.949	1.656	0.679	-	-	2.002	
平均	0.199	1.993	2.111	2.102	2.162	1.931	1.574	0.668	#DIV/0!	#DIV/0!	2.580	
SD	0.008	0.056	0.056	0.032	0.176	0.120	0.111	0.062	#DIV/0!	#DIV/0!	0.522	
Average-Blank	0.000	1.795	1.913	1.904	1.963	1.732	1.376	0.470	#DIV/0!	#DIV/0!	2.381	
Relative cell growth (%)		100.0	106.6	106.1	109.4	96.5	76.7	26.2	#DIV/0!	#DIV/0!		
SD of relative cell growth(%)		3.1	3.1	1.8	9.8	6.7	6.2	3.4	#DIV/0!	#DIV/0!		

Transformation Assay		Solvent: D.W. 5 %*										
Compound (ug/ml)	Well No.	0.01	0.0158	0.05	0.158	0.5	1.58	5	DMSO 0.1%	TPA 50 ng/mL		
No of foci	1	4	6	4	8	0	Tox	Tox	-	-	14	
	2	4	2	7	10	2	Tox	Tox	-	-	26	
	3	1	4	5	4	0	Tox	Tox	-	-	14	
	4	3	1	4	5	0	Tox	Tox	-	-	13	
	5	2	1	6	3	0	Tox	Tox	-	-	7	
	6	1	1	5	7	0	Tox	Tox	-	-	8	
Average		2.5	2.5	5.2	6.2	0.3	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	13.7	
SD		1.4	2.1	1.2	2.6	0.8	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	6.8	
Parametric Dannett					*	**						
t-Test for Unequal Variances (Aspin-Welch) vs 5% Water											**	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Sodium arsenite

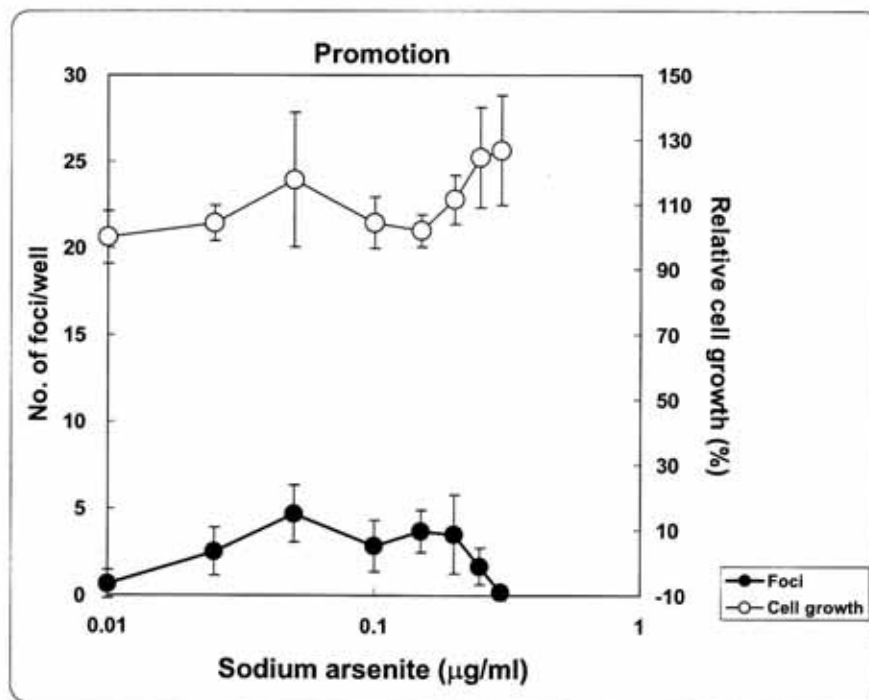
Transformation Assay

Promotion

Cell Growth Assay		Solvent: D.W. 5%									
Compound (ug/ml)	Blank	0.01	0.025	0.05	0.1	0.15	0.2	0.25	0.3	DMSO 0.1%	TPA 50 ng/mL
OD	0.103	0.952	1.015	1.393	0.987	0.993	1.053	1.396	1.436	0.979	2.185
	0.219	1.095	1.112	1.095	1.095	1.07	1.175	1.199	1.176	1.25	2.067
	0.115	1.034	1.068	1.06	1.117	1.069	1.159	1.135	1.177	1.14	1.983
Average	0.146	1.027	1.065	1.183	1.066	1.044	1.129	1.243	1.263	1.123	2.078
SD	0.064	0.072	0.049	0.183	0.070	0.044	0.066	0.136	0.150	0.136	0.101
Average-Blank	0.000	0.881	0.919	1.037	0.921	0.898	0.983	1.098	1.117	0.977	1.933
Relative cell growth (%)		100.0	104.3	117.7	104.5	101.9	111.6	124.5	126.8	100.0	197.7
SD of relative cell growth(%)		8.1	5.5	20.8	7.9	5.0	7.5	15.4	17.0	13.9	10.4

Transformation Assay		Solvent: D.W. 5%									
Compound (ug/ml)	Well No.	0.01	0.025	0.05	0.1	0.15	0.2	0.25	0.3	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	2	4	5	2	2	4	1	1	3	21
	2	0	3	5	1	5	2	2	0	1	14
	3	1	3	5	2	3	1	2	0	2	17
	4	1	0	4	3	4	5	0	0	3	19
	5	0	2	7	5	3	7	2	0	2	17
	6	0	3	2	4	5	2	3	0	4	21
Average		0.7	2.5	4.7	2.8	3.7	3.5	1.7	0.2	2.5	18.2
SD		0.8	1.4	1.6	1.5	1.2	2.3	1.0	0.4	1.0	2.7
Dunnett test				***	*	**	**				
t-Test for Unequal Variances (Aspin-Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Sodium arsenite

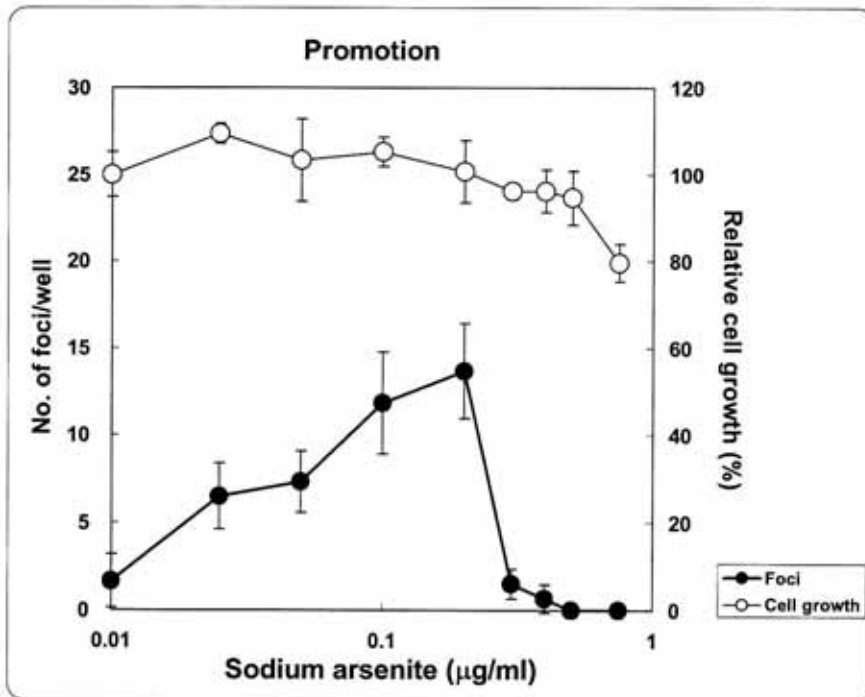
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.5%										DMSO 0.5% TPA 50 ng/mL	
Compound (ug/ml)	Blank	0.01	0.025	0.05	0.1	0.2	0.3	0.4	0.5	0.75			
OD	0.06	0.473	0.496	0.438	0.480	0.424	0.441	0.419	0.454	0.355	0.473	0.436	
	0.062	0.456	0.496	0.453	0.483	0.469	0.430	0.440	0.438	0.378	0.456	0.498	
	0.063	0.432	0.480	0.509	0.459	0.476	0.444	0.457	0.406	0.388	0.432	0.530	
Average	0.062	0.454	0.491	0.467	0.474	0.456	0.438	0.439	0.433	0.374	0.454	0.488	
SD	0.002	0.021	0.009	0.037	0.013	0.028	0.007	0.019	0.024	0.017	0.021	0.048	
Average-Blank	0.000	0.392	0.429	0.405	0.412	0.395	0.377	0.377	0.371	0.312	0.392	0.426	
Relative cell growth (%)		100.0	109.4	103.3	105.2	100.7	96.1	96.2	94.6	79.6	100.0	108.8	
SD of relative cell growth(%)		5.3	2.4	9.5	3.3	7.2	1.9	4.9	6.2	4.3	5.3	12.2	

Transformation Assay		Solvent: DMSO 0.5%										DMSO 0.5% TPA 50 ng/mL	
Compound (ug/ml)	Well No.	0.01	0.025	0.05	0.1	0.2	0.3	0.4	0.5	0.75			
No of foci	1	0	5	8	13	15	1	0	0	0	0	13	
	2	1	9	10	8	10	1	1	0	0	1	7	
	3	3	7	7	16	13	1	1	0	0	3	16	
	4	4	8	5	9	14	1	0	0	0	4	23	
	5	1	6	8	12	12	2	0	0	0	1	16	
	6	1	4	6	13	18	3	2	0	0	1	11	
Average		1.7	6.5	7.3	11.8	13.7	1.5	0.7	0.0	0.0	1.7	14.3	
SD		1.5	1.9	1.8	2.9	2.7	0.8	0.8	0.0	0.0	1.5	5.4	
Dunnnett test			***	***	***	***							
t-Test for Unequal Variances (Aspin-Welch)												**	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab 1

Compound: o-Toluidine HCL

Transformation Assay

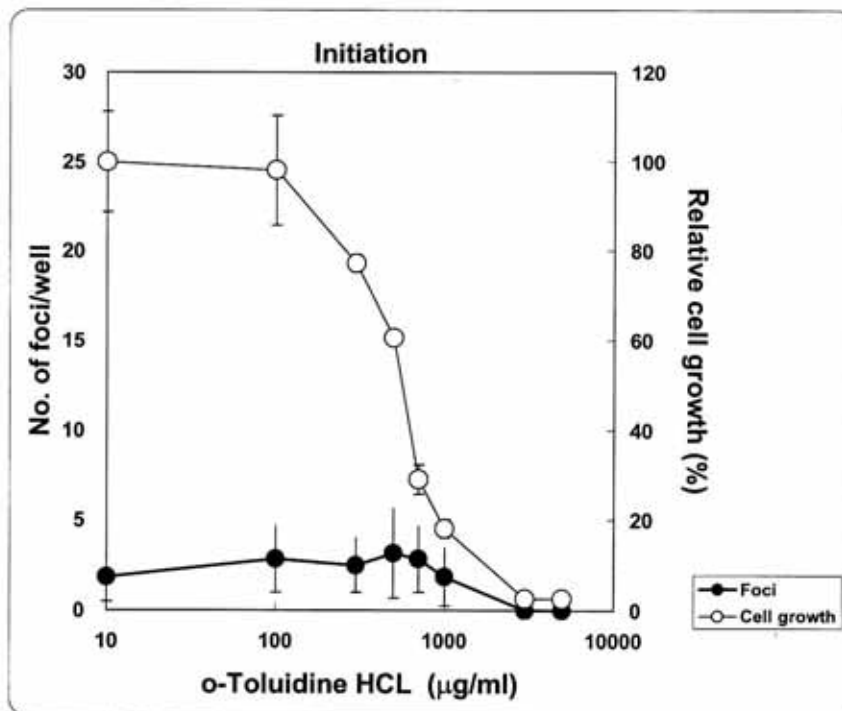
Initiation

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	10	100	300	500	700	1000	3000	5000	DMSO 0.1%	MCA 1 ug/mL
OD	0.08	1.1	1.059	0.983	0.77	0.46	0.318	0.113	0.118	1.236	0.717
	0.088	1.228	1.222	0.979	0.778	0.399	0.281	0.112	0.115	1.332	0.699
	0.09	1.357	1.338	0.944	0.787	0.393	0.277	0.119	0.115	1.216	0.709
Average	0.086	1.228	1.206	0.969	0.778	0.417	0.292	0.115	0.116	1.261	0.708
SD	0.005	0.129	0.140	0.021	0.009	0.037	0.023	0.004	0.002	0.062	0.009
Average-Blank	0.000	1.142	1.120	0.883	0.692	0.331	0.206	0.029	0.030	1.175	0.622
Relative cell growth (%)		100.0	98.1	77.3	60.6	29.0	18.0	2.5	2.6	100.0	52.9
SD of relative cell growth(%)		11.2	12.3	1.9	0.7	3.2	2.0	0.3	0.2	5.3	0.8

Transformation Assay		Solvent:									
Compound (ug/ml)	Well No.	10	100	300	500	700	1000	3000	5000	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	0	2	4	1	4	2	toxic	toxic	2	20
	2	2	2	0	6	6	0	toxic	toxic	3	18
	3	4	2	2	0	1	4	toxic	toxic	1	22
	4	2	6	2	6	2	3	toxic	toxic	2	25
	5	1	1	3	3	2	2	toxic	toxic	1	17
	6	2	4	4	3	2	0	toxic	toxic	1	14
Average		1.8	2.8	2.5	3.2	2.8	1.8	#DIV/0!	#DIV/0!	1.7	19.3
SD		1.3	1.8	1.5	2.5	1.8	1.6	#DIV/0!	#DIV/0!	0.8	3.9
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)											***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab V

Compound: o-Toluidine HCL

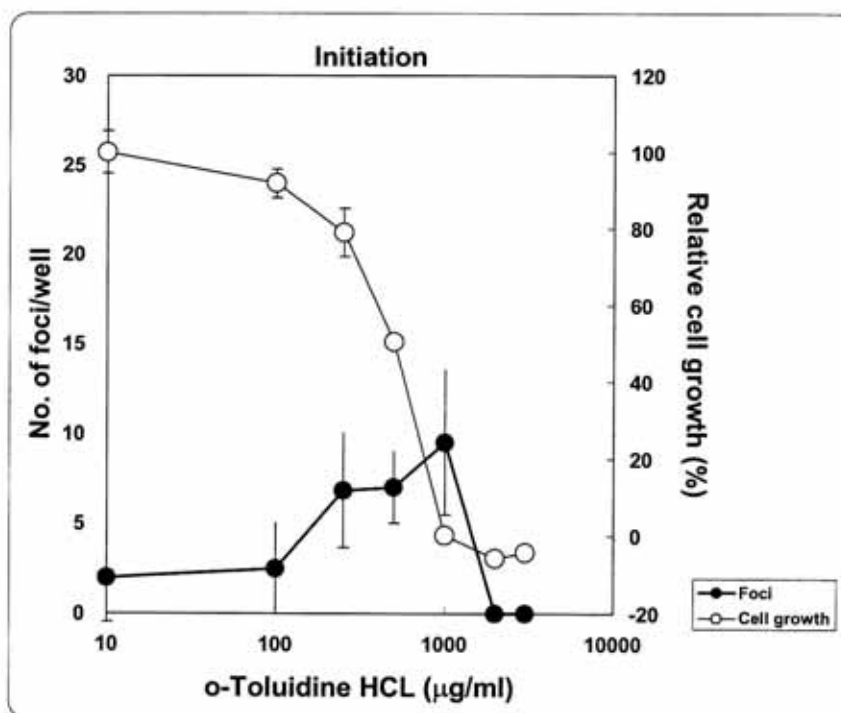
Transformation Assay

Initiation

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	10	100	250	500	1000	2000	3000	DMSO 0.1%	MCA 1 ug/mL	
OD	0.0741	0.4239	0.4031	0.3485	0.2743	0.0967	0.0752	0.081	0.4903	0.2136	
	0.1194	0.4581	0.4234	0.3919	0.2721	0.0962	0.0757	0.0815	0.4166	0.2132	
	0.0943	0.4573	0.4279	0.3787	0.2718	0.0977	0.0758	0.0802	0.4501	0.2146	
Average	0.096	0.446	0.418	0.373	0.273	0.097	0.076	0.081	0.452	0.214	
SD	0.023	0.020	0.013	0.022	0.001	0.001	0.000	0.001	0.037	0.001	
Average-Blank	0.000	0.351	0.322	0.277	0.177	0.001	-0.020	-0.015	0.356	0.118	
Relative cell growth (%)		100.0	91.9	79.1	50.4	0.3	-5.8	-4.3	100.0	33.1	
SD of relative cell growth(%)		5.6	3.8	6.3	0.4	0.2	0.1	0.2	10.4	0.2	

Transformation Assay		Solvent: Water5%									
Compound (ug/ml)	Well No.	10	100	250	500	1000	2000	3000	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	0	4	5	10	4	Toxic	Toxic	1	40	
	2	0	0	11	6	10	Toxic	Toxic	3	53	
	3	0	6	9	7	8	Toxic	Toxic	2	41	
	4	3	0	8	7	13	Toxic	Toxic	3	46	
	5	6	1	2	8	7	Toxic	Toxic	2	46	
	6	3	4	6	4	15	Toxic	Toxic	3	36	
Average		2.0	2.5	6.8	7.0	9.5	#DIV/0!	#DIV/0!	2.3	43.7	
SD		2.4	2.5	3.2	2.0	4.0	#DIV/0!	#DIV/0!	0.8	6.0	
Dunnett test				**	**	***					
t-Test for Unequal Variances (Aspin-Welch)										***	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: o-Toluidine HCL

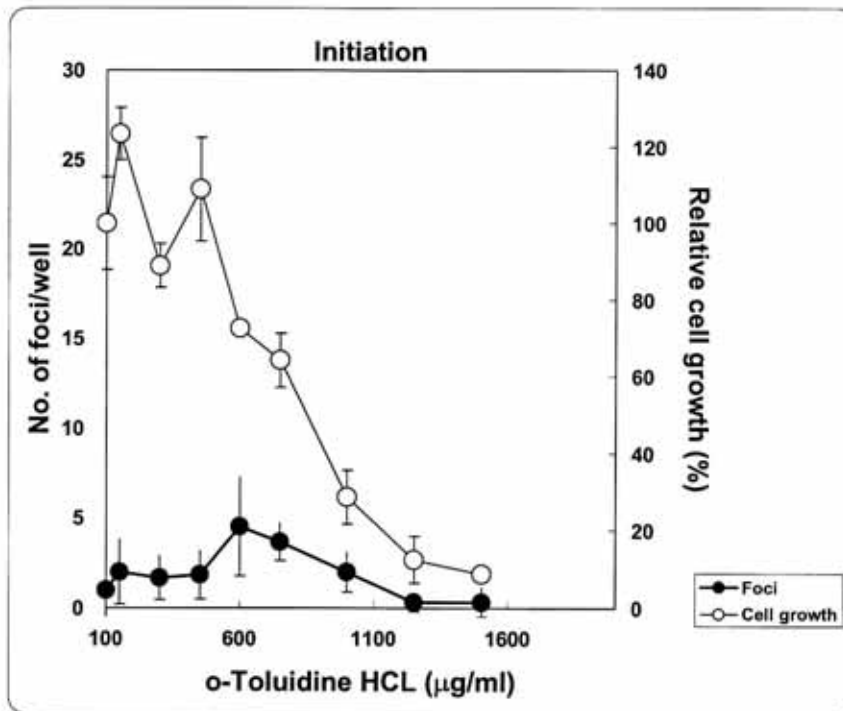
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.5%										
Compound (ug/ml)	Blank	100	150	300	450	600	750	1000	1250	1500	DMSO 0.5%	MCA 1 ug/mL
OD	0.032	0.228	0.291	0.221	0.237	0.197	0.184	0.097	0.076	0.059	0.228	0.125
	0.031	0.277	0.314	0.241	0.278	0.190	0.186	0.094	0.079	0.062	0.277	0.128
	0.071	0.245	0.289	0.220	0.290	0.195	0.160	0.120	0.056	0.067	0.245	0.130
Average	0.045	0.250	0.298	0.227	0.268	0.194	0.177	0.104	0.070	0.063	0.250	0.128
SD	0.023	0.025	0.014	0.012	0.028	0.004	0.014	0.014	0.013	0.004	0.025	0.003
Average-Blank	0.000	0.205	0.253	0.183	0.224	0.149	0.132	0.059	0.026	0.018	0.205	0.083
Relative cell growth (%)		100.0	123.4	89.0	108.9	72.7	64.3	28.7	12.5	8.8	100.0	40.4
SD of relative cell growth(%)		12.1	6.8	5.8	13.5	1.8	7.0	6.9	6.1	2.0	12.1	1.2

Transformation Assay		Solvent: DMSO 0.5%										
Compound (ug/ml)	Well No.	100	150	300	450	600	750	1000	1250	1500	DMSO 0.5%	MCA 1 ug/mL
No of foci	1	0	3	3	2	3	4	4	0	0	0	25
	2	0	5	1	1	3	3	1	0	0	0	20
	3	0	0	1	2	3	5	2	0	0	0	14
	4	1	1	2	0	4	4	2	1	0	1	22
	5	4	1	3	4	10	2	2	1	0	4	32
	6	1	2	0	2	4	4	1	0	2	1	17
Average		1.0	2.0	1.7	1.8	4.5	3.7	2.0	0.3	0.3	1.0	21.7
SD		1.5	1.8	1.2	1.3	2.7	1.0	1.1	0.5	0.8	1.5	6.3
Dunnett test						**	*					
t-Test for Unequal Variances (Aspin-Welch)												***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: o-Toluidine HCL

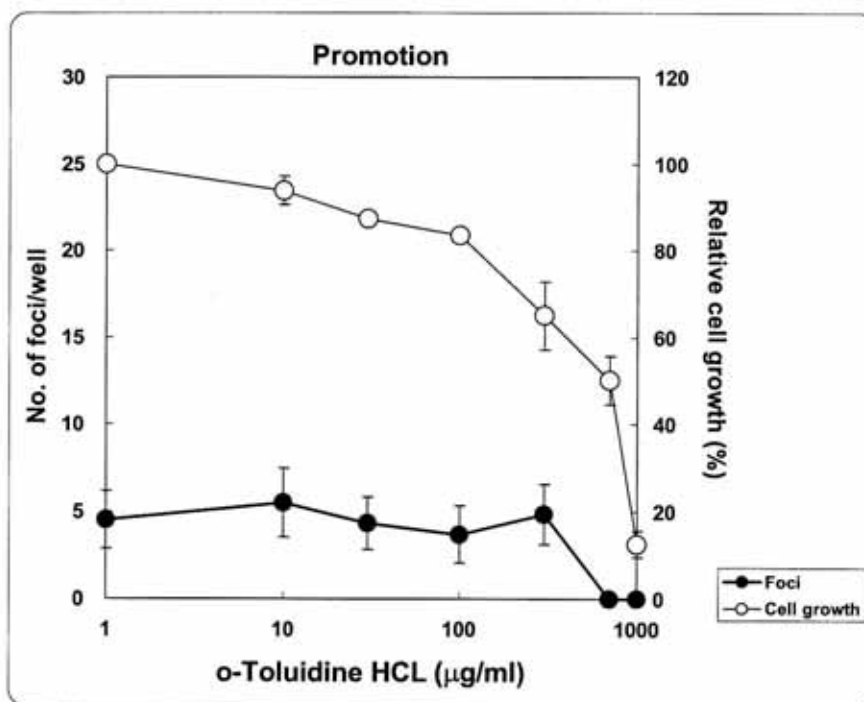
Transformation Assay

Promotion

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	1	10	30	100	300	700	1000	DMSO 0.1%	TPA 50 ng/mL	
OD	0.073	0.964	0.92	0.861	0.833	0.58	0.484	0.186	1.028	1.225	
	0.073	0.958	0.875	0.848	0.806	0.657	0.5	0.159	1.028	1.255	
	0.077	0.969	0.931	0.843	0.811	0.72	0.576	0.212	1.002	1.331	
Average	0.074	0.964	0.909	0.851	0.817	0.652	0.520	0.186	1.019	1.270	
SD	0.002	0.006	0.030	0.009	0.014	0.070	0.049	0.027	0.015	0.055	
Average-Blank	0.000	0.889	0.834	0.776	0.742	0.578	0.446	0.111	0.945	1.196	
Relative cell growth (%)		100.0	93.8	87.3	83.5	65.0	50.1	12.5	100.0	126.6	
SD of relative cell growth(%)		0.6	3.3	1.0	1.6	7.9	5.5	3.0	1.6	5.8	

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	1	10	30	100	300	700	1000	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	5	4	6	4	2	Toxic	Toxic	6	37	
	2	3	4	5	2	6	Toxic	Toxic	11	46	
	3	5	8	5	3	4	Toxic	Toxic	7	44	
	4	6	8	5	2	7	Toxic	Toxic	9	39	
	5	2	5	2	5	5	Toxic	Toxic	5	40	
	6	6	4	3	6	5	Toxic	Toxic	9	40	
Average		4.5	5.5	4.3	3.7	4.8	#DIV/0!	#DIV/0!	7.8	41.0	
SD		1.6	2.0	1.5	1.6	1.7	#DIV/0!	#DIV/0!	2.2	3.3	
Dunnet test											N.S.
t-Test (Assuming Equal Variances)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab V

Compound: o-Toluidine HCL

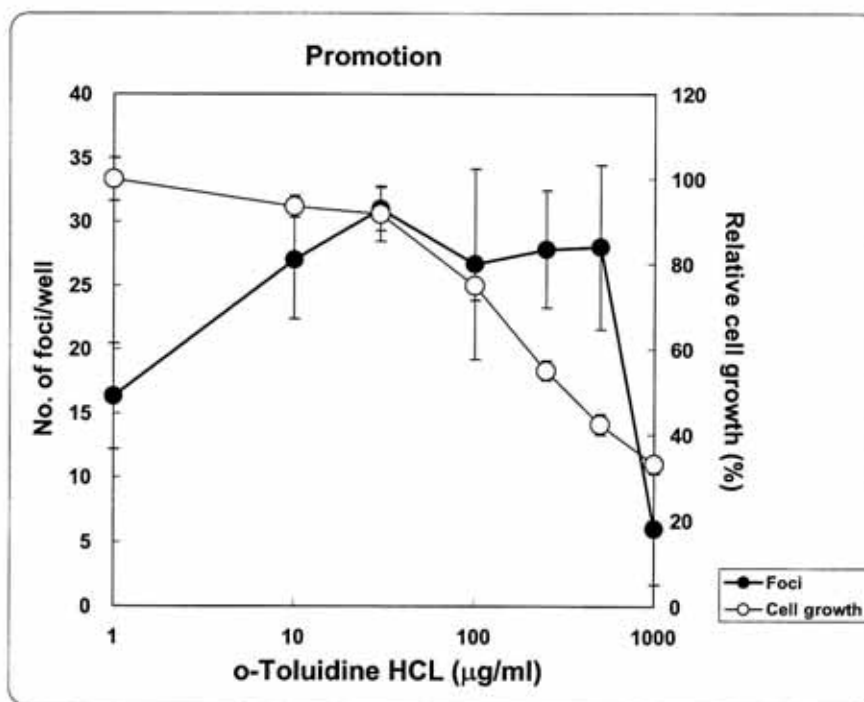
Transformation Assay

Promotion

Cell Growth Assay		Solvent: Water										
Compound (ug/ml)	Blank	Water	1	10	30	100	250	500	1000	DMSO 0.1%	TPA 50 ng/mL	
OD	0.0859	0.5773	0.5363	0.5481	0.4552	0.3553	0.2922	0.2486	0.5831	0.7353		
	0.0875	0.5508	0.5183	0.5067	0.4257	0.3406	0.2719	0.2309	0.5275	0.698		
	0.0886	0.5303	0.5144	0.4892	0.4278	0.334	0.2897	0.2442	0.5404	0.6873		
Average	0.087	0.553	0.523	0.515	0.436	0.343	0.285	0.241	0.550	0.707		
SD	0.001	0.024	0.012	0.030	0.016	0.011	0.011	0.009	0.029	0.025		
Average-Blank	0.000	0.465	0.436	0.427	0.349	0.256	0.197	0.154	0.463	0.620		
Relative cell growth (%)		100.0	93.6	91.8	75.0	55.0	42.4	33.1	100.0	133.8		
SD of relative cell growth(%)		5.1	2.5	6.5	3.5	2.3	2.4	2.0	6.3	5.4		

Transformation Assay		Solvent: Water										
Compound (ug/ml)	Well No.	Water	1	10	30	100	250	500	1000	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	12	36	31	26	26	31	2	11	40		
	2	14	25	29	19	22	20	1	21	40		
	3	12	27	34	36	31	21	11	21	45		
	4	21	25	30	35	28	27	5	17	38		
	5	19	23	31	19	25	34	6	9	60		
	6	20	26	31	25	35	35	11	10	39		
Average		16.3	27.0	31.0	26.7	27.8	28.0	6.0	14.8	43.7		
SD		4.1	4.6	1.7	7.4	4.6	6.4	4.3	5.5	8.4		
Dunnett test			**	***	**	**	**					
t-Test(Assuming Equal Variances)										***		

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: o-Toluidine HCL

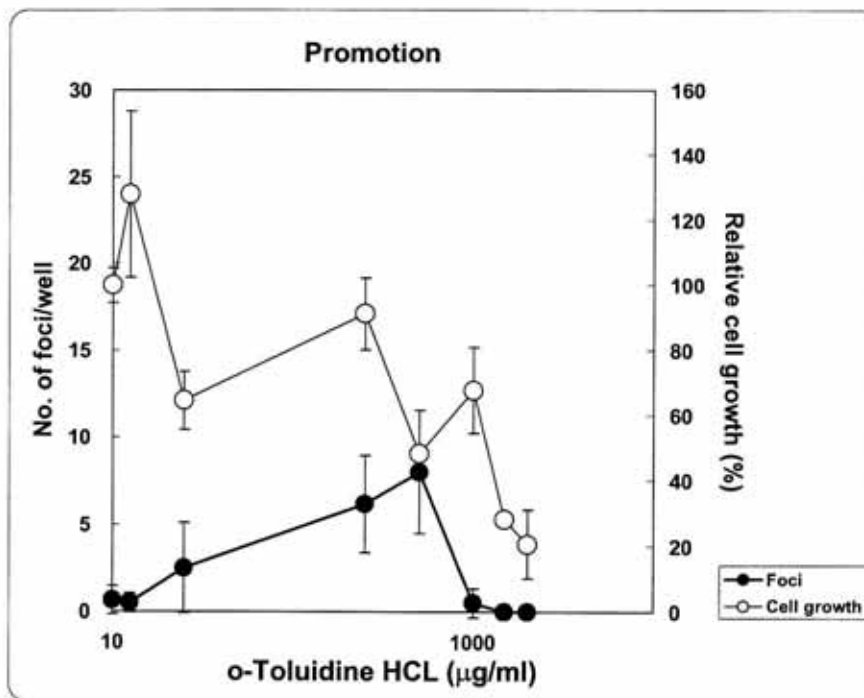
Transformation Assay

Promotion

Cell Growth Assay		Solvent:	DMSO 0.5%									
Compound (ug/ml)	Blank		10	12.5	25	250	500	1000	1500	2000	DMSO 0.5%	TPA 50 ng/mL
OD	0.032	0.226	0.296	0.145	0.197	0.128	0.141	0.090	0.085	0.085	0.226	0.234
	0.031	0.247	0.350	0.181	0.221	0.134	0.194	0.089	0.088	0.247	0.268	
	0.031	0.243	0.244	0.170	0.243	0.132	0.180	0.090	0.049	0.243	0.236	
Average	0.031	0.239	0.297	0.165	0.220	0.131	0.172	0.090	0.074	0.239	0.246	
SD	0.001	0.011	0.053	0.018	0.023	0.003	0.027	0.001	0.022	0.011	0.019	
Average-Blank	0.000	0.207	0.265	0.134	0.189	0.100	0.140	0.058	0.043	0.207	0.215	
Relative cell growth (%)		100.0	128.0	64.6	91.2	48.2	67.7	28.1	20.6	100.0	103.5	
SD of relative cell growth(%)		5.4	25.6	8.9	11.1	1.5	13.2	0.3	10.5	5.4	9.2	

Transformation Assay		Solvent:	DMSO 0.5%									
Compound (ug/ml)	Well No.		10	12.5	25	250	500	1000	1500	2000	DMSO 0.5%	TPA 50 ng/mL
No of foci	1	1	0	0	2	6	0	Toxic	Toxic	1	17	
	2	0	0	2	5	6	0	Toxic	Toxic	0	16	
	3	2	1	7	10	11	0	Toxic	Toxic	2	16	
	4	1	1	3	5	3	1	Toxic	Toxic	1	14	
	5	0	0	3	8	10	0	Toxic	Toxic	0	12	
	6	0	1	0	7	12	2	Toxic	Toxic	0	18	
Average		0.7	0.5	2.5	6.2	8.0	0.5	#DIV/0!	#DIV/0!	0.7	15.5	
SD		0.8	0.5	2.6	2.8	3.5	0.8	#DIV/0!	#DIV/0!	0.8	2.2	
Dunnett test					***	***						
t-Test for Unequal Variances (Aspin-Welch)											***	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Lithocholic acid

Transformation Assay

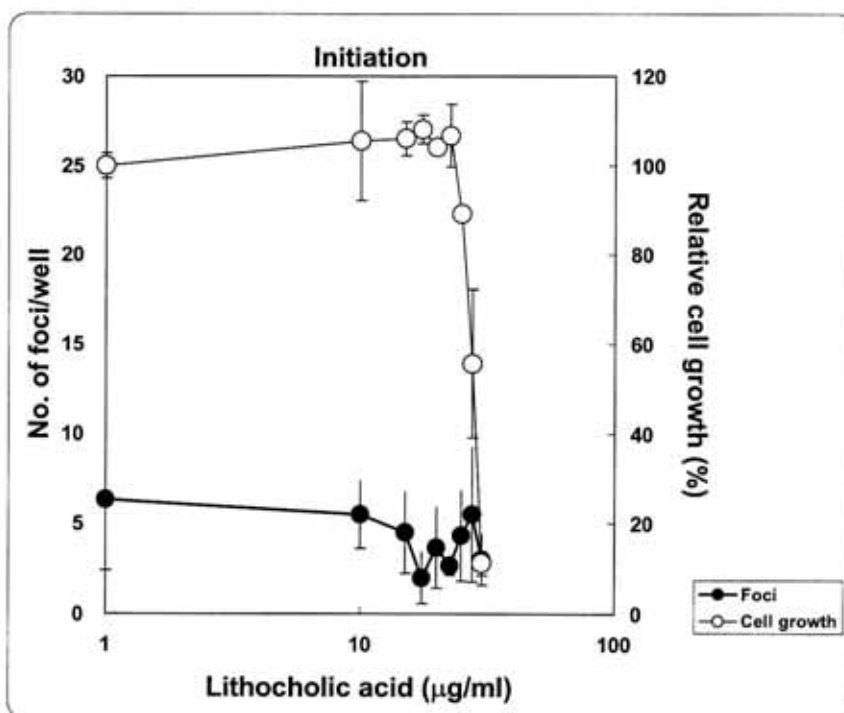
Initiation

Cell Growth Assay		Solvent: DMSO 0.1%										
Compound E (ug/ml)	Blank	1	10	15	17.5	20	22.5	25	27.5	30	DMSO 0.1% MCA 1 ug/mL	
OD	0.073	0.882	1.055	0.901	0.979	0.919	0.874	0.807	0.434	0.151	0.882	0.558
	0.073	0.865	0.879	0.959	0.928	0.918	0.973	0.801	0.463	0.19	0.865	0.53
	0.077	0.911	0.857	0.944	0.948	0.921	0.973	0.786	0.68	0.155	0.911	0.914
Average	0.074	0.886	0.930	0.935	0.952	0.919	0.940	0.798	0.526	0.165	0.886	0.667
SD	0.002	0.023	0.109	0.030	0.026	0.002	0.057	0.011	0.134	0.021	0.023	0.214
Average-Blank	0.000	0.812	0.856	0.860	0.877	0.845	0.866	0.724	0.451	0.091	0.812	0.593
Relative cell growth (%)		100.0	105.5	106.0	108.1	104.1	106.7	89.2	55.6	11.2	100.0	73.1
SD of relative cell growth(%)			2.9	13.4	3.7	3.2	0.2	7.0	1.3	16.6	2.6	26.4

Transformation Assay		Solvent: DMSO 0.1%										
Compound E (ug/ml)	Well No.	1	10	15	17.5	20	22.5	25	27.5	30	DMSO 0.1% MCA 1 ug/mL	
No of foci	1	6	5	7	1	6	3	4	2	4	6	40
	2	2	6	5	2	1	3	9	12	3	2	44
	3	10	7	2	4	4	3	5	7	2	10	48
	4	12	2	4	0	6	2	2	5	3	12	49
	5	5	6	2	3	1	3	3	5	1	5	43
	6	3	7	7	2	4	2	3	2	5	3	41
Average		6.3	5.5	4.5	2.0	3.7	2.7	4.3	5.5	3.0	6.3	44.2
SD		3.9	1.9	2.3	1.4	2.3	0.5	2.5	3.7	1.4	3.9	3.7
Dunnett test												
t-Test (Assuming Equal Variances)												***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Lithocholic acid

Transformation Assay

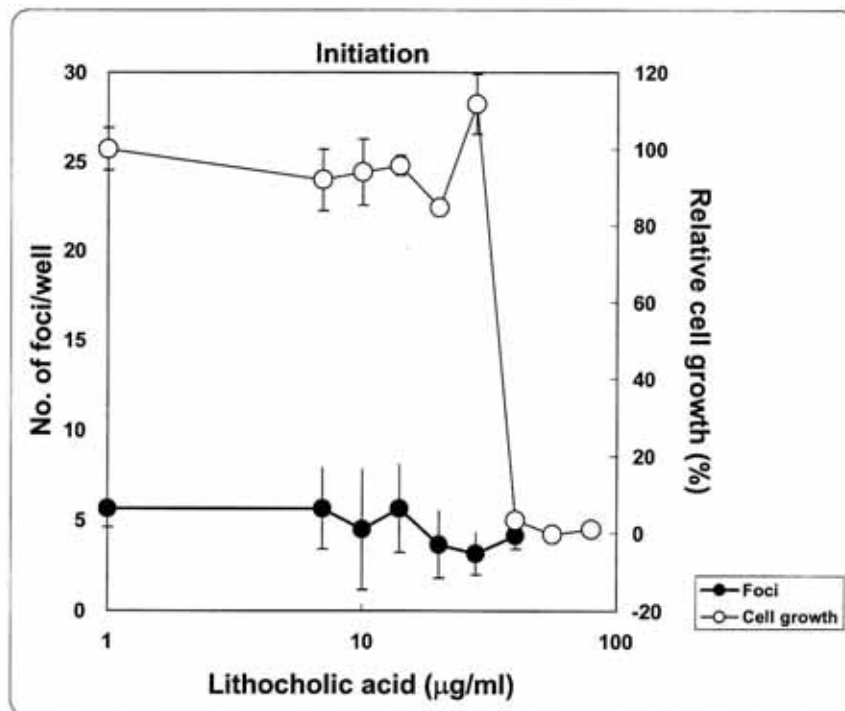
Initiation

Cell Growth Assay		Solvent: DMSO 0.1%										DMSO 0.1% MCA 1 ug/mL	
Compound (ug/ml)	Blank	1	7	10	14	20	28	40	56	80			
OD	0.059	0.37	0.372	0.388	0.381	0.339	0.441	0.075	0.061	0.069	0.37	0.225	
	0.067	0.406	0.381	0.381	0.365	0.333	0.44	0.076	0.062	0.067	0.406	0.271	
	0.063	0.388	0.332	0.336	0.376	0.343	0.397	0.071	0.063	0.063	0.388	0.253	
Average	0.063	0.388	0.362	0.368	0.374	0.338	0.426	0.074	0.062	0.066	0.388	0.250	
SD	0.004	0.018	0.026	0.028	0.008	0.005	0.025	0.003	0.001	0.003	0.018	0.023	
Average-Blank	0.000	0.325	0.299	0.305	0.311	0.275	0.363	0.011	-0.001	0.003	0.325	0.187	
Relative cell growth (%)		100.0	91.9	93.9	95.7	84.7	111.7	3.4	-0.3	1.0	100.0	57.4	
SD of relative cell growth(%)		5.5	8.0	8.7	2.5	1.5	7.7	0.8	0.3	0.9	5.5	7.1	

Transformation Assay		Solvent: DMSO 0.1%										DMSO 0.1% MCA 1 ug/mL	
Compound (ug/ml)	Well No.	1	7	10	14	20	28	40	56	80			
No of foci	1	6	2	1	2	4	2	4	Toxic	Toxic	6	37	
	2	6	5	1	5	2	3	5	Toxic	Toxic	6	39	
	3	4	6	7	8	4	2	5	Toxic	Toxic	4	41	
	4	7	5	9	8	7	5	4	Toxic	Toxic	7	43	
	5	5	8	3	4	2	3	3	Toxic	Toxic	5	36	
	6	6	8	6	7	3	4	4	Toxic	Toxic	6	38	
Average		5.7	5.7	4.5	5.7	3.7	3.2	4.2			5.7	39.0	
SD		1.0	2.3	3.3	2.4	1.9	1.2	0.8			1.0	2.6	
Dunnett test													
t-Test (Aspin-Welch)												***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Lithocholic acid

Transformation Assay

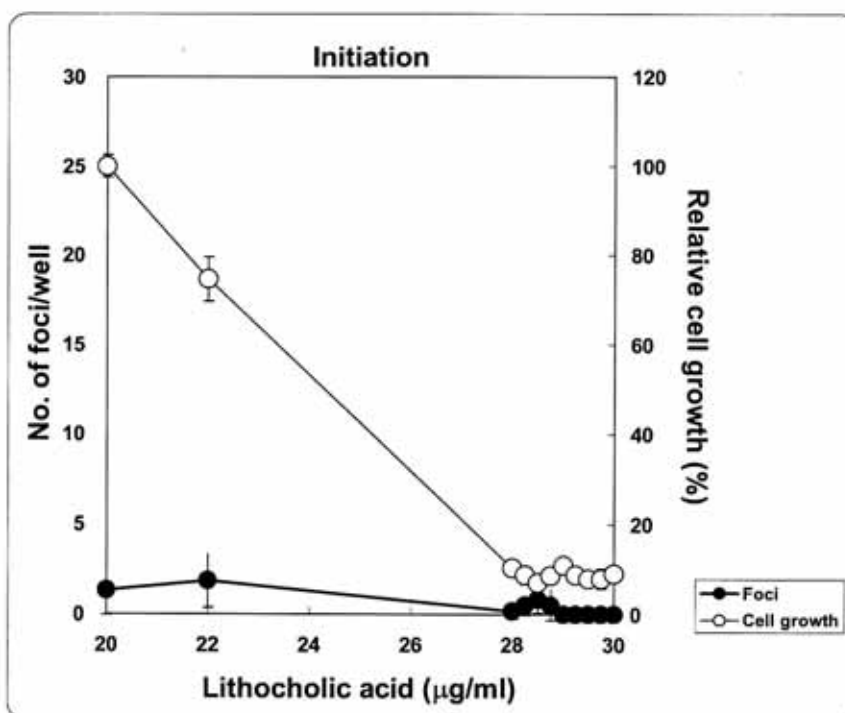
Initiation

Cell Growth Assay		Solvent: DMSO 0.5%												
Compound (ug/ml)	Blank	20	22	28	28.25	28.5	28.75	29	29.25	29.5	29.75	30	DMSO 0.5%	MCA 1 ug/mL
OD	0.033	0.303	0.226	0.061	0.054	0.051	0.053	0.059	0.060	0.051	0.051	0.056	0.303	0.176
	0.034	0.316	0.236	0.061	0.054	0.050	0.058	0.062	0.056	0.058	0.052	0.057	0.316	0.193
	0.033	0.305	0.253	0.062	0.063	0.056	0.059	0.068	0.056	0.056	0.062	0.061	0.305	0.196
Average	0.033	0.308	0.238	0.061	0.057	0.052	0.057	0.063	0.057	0.055	0.055	0.058	0.308	0.188
SD	0.001	0.007	0.014	0.001	0.005	0.003	0.003	0.005	0.002	0.004	0.006	0.003	0.007	0.011
Average-Blank	0.000	0.275	0.205	0.028	0.024	0.019	0.023	0.030	0.024	0.022	0.022	0.025	0.275	0.155
Relative cell growth (%)		100.0	74.6	10.2	8.6	6.9	8.5	10.8	8.7	7.9	7.9	9.0	100.0	56.4
SD of relative cell growth(%)		2.5	5.0	0.2	1.9	1.2	1.2	1.7	0.8	1.3	2.2	1.0	2.5	3.9

Transformation Assay		Solvent: DMSO 0.5%												
Compound (ug/ml)	Well No.	20	22	28	28.25	28.5	28.75	29	29.25	29.5	29.75	30	DMSO 0.5%	MCA 1 ug/mL
No of foci	1	3	4	1	1	1	0	0	Toxic	Toxic	Toxic	Toxic	3	8
	2	0	2	0	0	0	0	0	Toxic	Toxic	Toxic	Toxic	0	15
	3	1	1	0	0	1	0	0	Toxic	Toxic	Toxic	Toxic	1	16
	4	0	0	0	1	1	0	0	Toxic	Toxic	Toxic	Toxic	0	5
	5	1	1	0	0	0	2	0	Toxic	Toxic	Toxic	Toxic	1	9
	6	3	3	0	1	2	1	0	Toxic	Toxic	Toxic	Toxic	3	10
Average		1.3	1.8	0.2	0.5	0.8	0.5	0.0	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	1.3	10.5
SD		1.4	1.5	0.4	0.5	0.8	0.8	0.0	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	1.4	4.2
Dunnett test														
t-Test (Aspin-Weich)														**

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Lithocholic acid

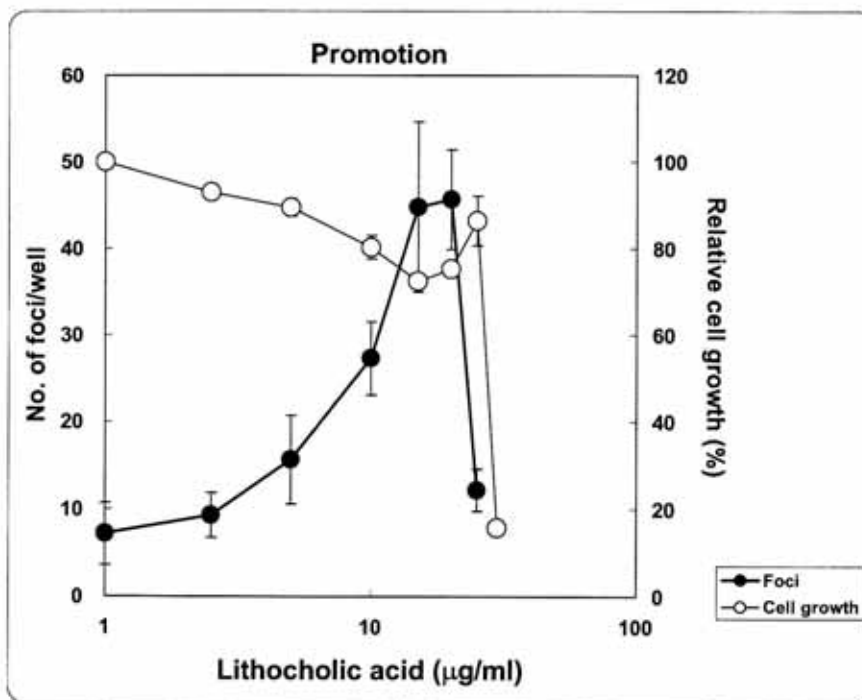
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.1%									
Compound E (ug/ml)	Blank	1	2.5	5	10	15	20	25	30	DMSO 0.1%	TPA 50 ng/mL
OD	0.073	1.001	0.923	0.891	0.78	0.732	0.765	0.839	0.219	1.001	1.123
	0.073	0.974	0.917	0.872	0.812	0.73	0.741	0.827	0.228	0.974	1.224
	0.077	0.985	0.927	0.909	0.829	0.743	0.775	0.922	0.207	0.985	1.283
Average	0.074	0.987	0.922	0.891	0.807	0.735	0.760	0.863	0.218	0.987	1.210
SD	0.002	0.014	0.005	0.019	0.025	0.007	0.017	0.052	0.011	0.014	0.081
Average-Blank	0.000	0.912	0.848	0.816	0.733	0.661	0.686	0.788	0.144	0.912	1.136
Relative cell growth (%)		100.0	92.9	89.5	80.3	72.4	75.2	86.4	15.7	100.0	124.5
SD of relative cell growth(%)		1.5	0.6	2.0	2.7	0.8	1.9	5.7	1.2	1.5	8.9

Transformation Assay		Solvent: DMSO 0.1%									
Compound E (ug/ml)	Well No.	1	2.5	5	10	15	20	25	30	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	5	13	16	33	38	49	16	tox	5	43
	2	2	9	21	29	48	35	11	tox	2	43
	3	7	8	17	23	53	51	13	tox	7	41
	4	7	12	13	27	50	45	9	tox	7	45
	5	10	7	7	22	52	49	13	tox	10	42
	6	12	7	20	30	28	45	11	tox	12	40
Average		7.2	9.3	15.7	27.3	44.8	45.7	12.2		7.2	42.3
SD		3.5	2.6	5.1	4.2	9.8	5.8	2.4		3.5	1.8
Dunnett test					*	***	***	***			***
t-Test for Unequal Variances (Aspin-Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Lithocholic acid

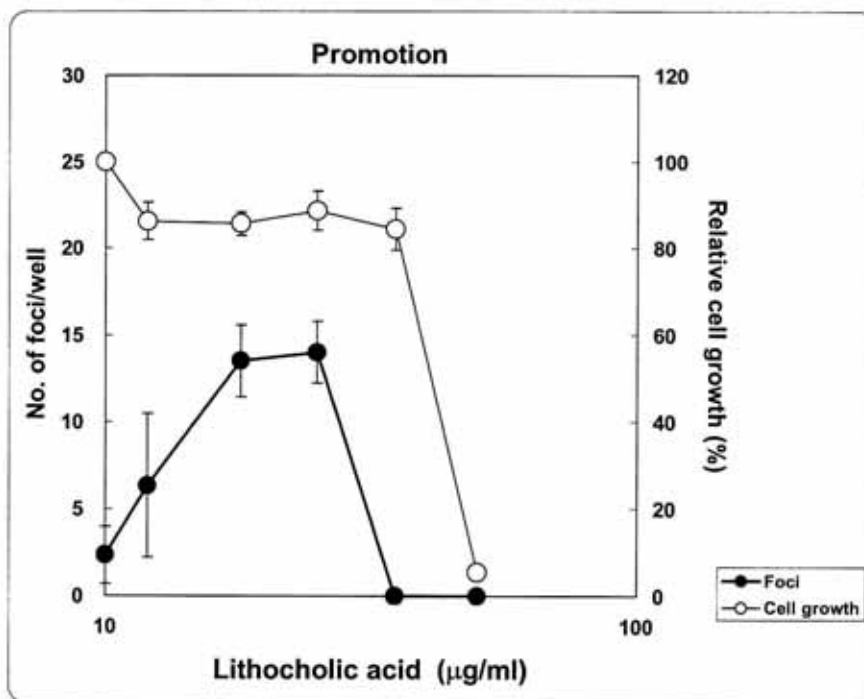
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.1%								
Compound (ug/ml)	Blank	10	12	18	25	35	50	DMSO 0.1%	TPA 50 ng/mL	
OD	0.062	0.359	0.334	0.326	0.319	0.313	0.074	0.359	0.474	
	0.061	0.362	0.312	0.31	0.342	0.299	0.08	0.362	0.447	
	0.06	0.359	0.311	0.316	0.318	0.328	0.078	0.359	0.479	
Average	0.061	0.360	0.319	0.317	0.326	0.313	0.077	0.360	0.467	
SD	0.001	0.002	0.013	0.008	0.014	0.015	0.003	0.002	0.017	
Average-Blank	0.000	0.299	0.258	0.256	0.265	0.252	0.016	0.299	0.406	
Relative cell growth (%)		100.0	86.3	85.7	88.7	84.4	5.5	100.0	135.7	
SD of relative cell growth(%)		0.6	4.3	2.7	4.5	4.9	1.0	0.6	5.8	

Transformation Assay		Solvent: DMSO 0.1%								
Compound (ug/ml)	Well No.	10	12	18	25	35	50	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	4	1	12	17	Toxic	Toxic	4	9	
	2	0	7	12	12	Toxic	Toxic	0	12	
	3	4	2	12	13	Toxic	Toxic	4	8	
	4	2	12	17	15	Toxic	Toxic	2	4	
	5	1	8	15	13	Toxic	Toxic	1	14	
	6	3	8	13	14	Toxic	Toxic	3	10	
Average		2.3	6.3	13.5	14.0	#DIV/0!	#DIV/0!	2.3	9.5	
SD		1.6	4.1	2.1	1.8	#DIV/0!	#DIV/0!	1.6	3.4	
Dunnett test			*	***	***					
t-Test for Unequal Variances (Aspin-Welch)										**

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Lithocholic acid

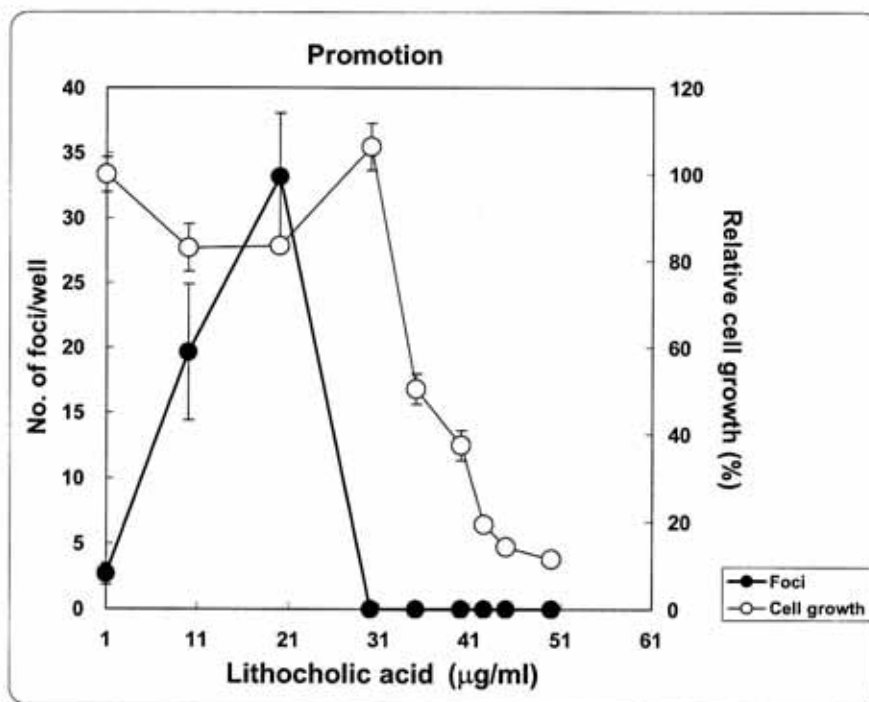
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.5%										
Compound (ug/ml)	Blank	1	10	20	30	35	40	42.5	45	50	DMSO 0.5%	TPA 50 ng/mL
OD	0.033	0.223	0.174	0.181	0.215	0.125	0.093	0.070	0.057	0.054	0.223	0.286
	0.031	0.212	0.183	0.186	0.233	0.130	0.104	0.066	0.059	0.052	0.212	0.302
	0.031	0.209	0.194	0.186	0.231	0.117	0.104	0.065	0.057	0.052	0.209	0.304
Average	0.032	0.215	0.184	0.184	0.226	0.124	0.100	0.067	0.058	0.053	0.215	0.297
SD	0.001	0.007	0.010	0.003	0.010	0.007	0.006	0.003	0.001	0.001	0.007	0.010
Average-Blank	0.000	0.183	0.152	0.153	0.195	0.092	0.069	0.035	0.026	0.021	0.183	0.266
Relative cell growth (%)		100.0	83.1	83.4	106.4	50.5	37.5	19.3	14.2	11.5	100.0	145.2
SD of relative cell growth(%)		4.0	5.5	1.6	5.4	3.6	3.5	1.4	0.6	0.6	4.0	5.4

Transformation Assay		Solvent: DMSO 0.5%										
Compound (ug/ml)	Well No.	1	10	20	30	35	40	42.5	45	50	DMSO 0.5%	TPA 50 ng/mL
No of foci	1	2	17	31	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	2	14
	2	2	14	33	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	2	17
	3	4	18	26	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	4	18
	4	2	29	37	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	2	12
	5	3	22	32	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	3	24
	6	3	18	40	Toxic	Toxic	Toxic	Toxic	Toxic	Toxic	3	16
Average		2.7	19.7	33.2	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	2.7	16.8
SD		0.8	5.2	4.9	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	0.8	4.1
Dunnett test			***	***								
t-Test for Unequal Variances (Aspin-Welch)												***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Mezelein

Transformation Assay

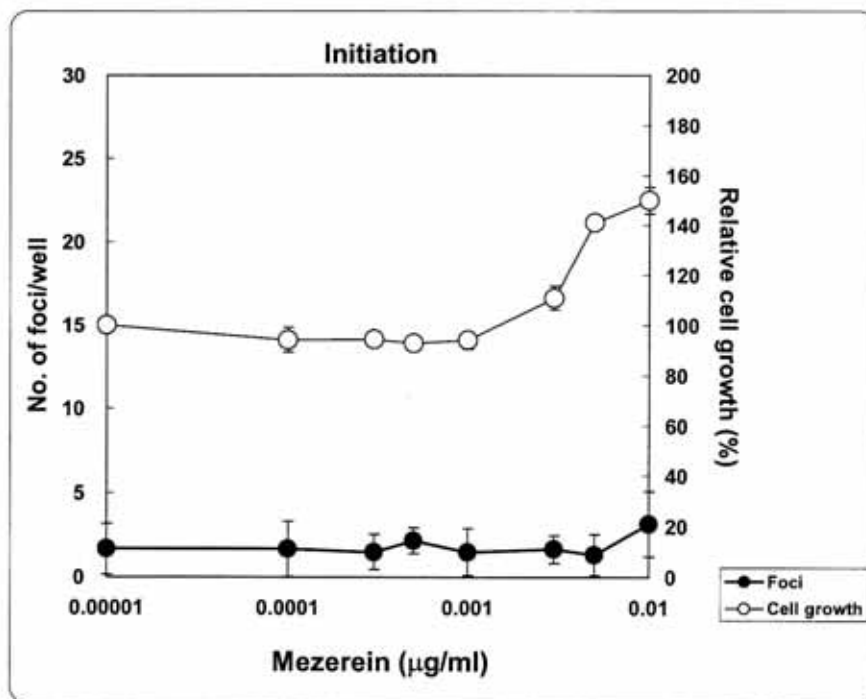
Initiation

Cell Growth Assay		Solvent:	DMSO 0.1%									
Compound (ug/ml)	Blank	0.00001	0.0001	0.0003	0.0005	0.001	0.003	0.005	0.01	DMSO 0.1%	MCA 1ug/mL	
OD	0.073	1.1	0.962	1.007	0.996	1.051	1.132	1.518	1.523	1.1	0.641	
	0.073	1.059	1.041	1.004	1.019	0.981	1.211	1.466	1.579	1.059	0.631	
	0.077	1.07	1.053	1.047	0.998	1.019	1.217	1.483	1.631	1.07	0.62	
Average	0.074	1.076	1.019	1.019	1.004	1.017	1.187	1.489	1.578	1.076	0.631	
SD	0.002	0.021	0.049	0.024	0.013	0.035	0.047	0.027	0.054	0.021	0.011	
Average-Blank	0.000	1.002	0.944	0.945	0.930	0.943	1.112	1.415	1.503	1.002	0.556	
Relative cell growth (%)		100.0	94.2	94.3	92.8	94.1	111.0	141.2	150.0	100.0	55.5	
SD of relative cell growth(%)		2.1	4.9	2.4	1.3	3.5	4.7	2.6	5.4	2.1	1.0	

Transformation Assay		Solvent:	DMSO 0.1%									
Compound (ug/ml)	Well No.	0.00001	0.0001	0.0003	0.0005	0.001	0.003	0.005	0.01	DMSO 0.1%	MCA 1ug/mL	
No of foci	1	3	3	3	3	3	1	1	6	3	17	
	2	0	2	1	3	2	1	0	3	0	25	
	3	0	0	2	2	0	2	2	1	0	19	
	4	1	1	1	2	3	3	0	4	1	22	
	5	3	0	0	2	1	1	3	4	3	20	
	6	3	4	2	1	0	2	2	1	3	30	
Average		1.7	1.7	1.5	2.2	1.5	1.7	1.3	3.2	1.7	22.2	
SD		1.5	1.6	1.0	0.8	1.4	0.8	1.2	1.9	1.5	4.7	
Dunnett test												
t-Test for Unequal Variances (Aspin-Welch)											***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.

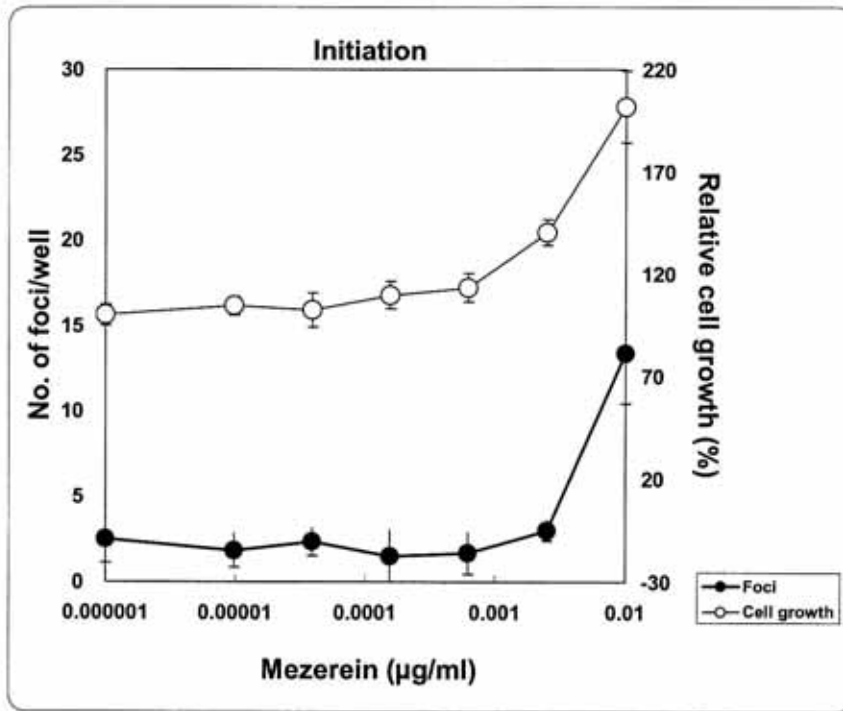


Assay Laboratory: Lab III
 Compound: Mezerein
 Transformation Assay
 Initiation

Cell Growth Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Blank	0.000001	0.00000977	0.0000391	0.000156	0.000625	0.0025	0.01	DMSO 0.1%	MCA 1 ug/mL	
OD	0.112	1.091	1.144	1.09	1.168	1.209	1.495	2.19	1.091	0.386	
	0.134	1.169	1.23	1.261	1.288	1.295	1.625	2.366	1.169	0.399	
	0.145	1.19	1.21	1.169	1.287	1.352	1.561	2.009	1.19	0.449	
Average	0.130	1.150	1.195	1.173	1.248	1.285	1.560	2.188	1.150	0.411	
SD	0.017	0.052	0.045	0.086	0.069	0.072	0.065	0.179	0.052	0.033	
Average-Blank	0.000	1.020	1.064	1.043	1.117	1.155	1.430	2.058	1.020	0.281	
Relative cell growth (%)		100.0	104.4	102.3	109.6	113.3	140.2	201.8	100.0	27.6	
SD of relative cell growth(%)		5.1	4.4	8.4	6.8	7.1	6.4	17.5	5.1	3.3	

Transformation Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Well No.	0.000001	0.00000977	0.0000391	0.000156	0.000625	0.0025	0.01	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	2	2	3	2	3	3	14	2	14	
	2	3	0	1	2	2	4	12	3	12	
	3	1	3	2	1	3	3	13	1	7	
	4	1	2	2	0	0	2	18	1	9	
	5	4	2	3	4	1	3	9	4	4	
	6	4	2	3	0	1	3	14	4	9	
Average		2.5	1.8	2.3	1.5	1.7	3.0	13.3	2.5	9.2	
SD		1.4	1.0	0.8	1.5	1.2	0.6	2.9	1.4	3.5	
Dunnett test								***			
t-Test (Aspin-Welch)										**	

The true value is zero. This value was temporarily put to make logarithm graph.

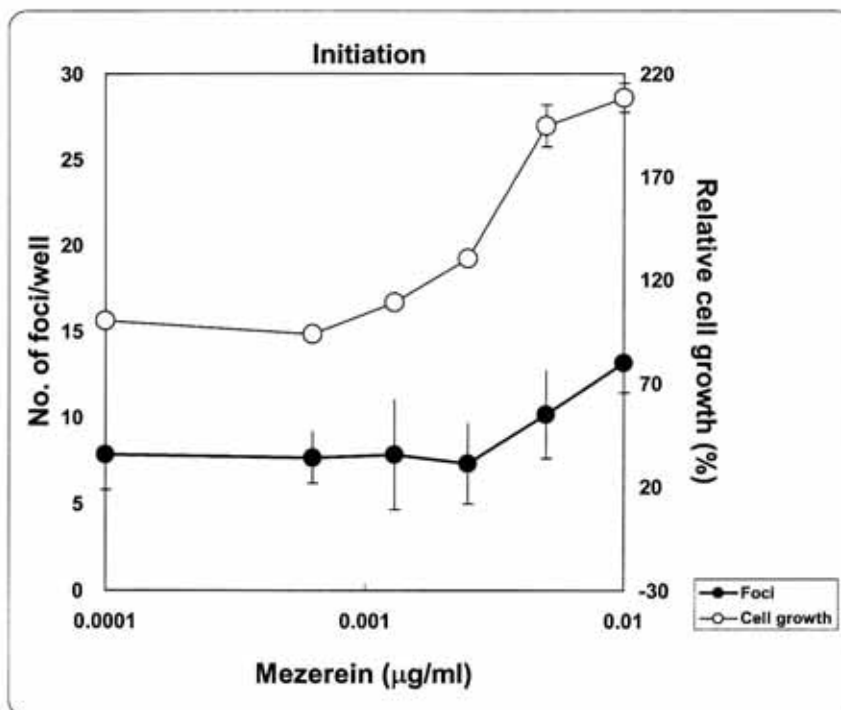


Assay Laboratory: Lab IV
 Compound: Mezelein
 Transformation Assay
 Initiation

Cell Growth Assay	Solvent:	DMSO 0.1%							
Compound (ug/ml)	Blank	0.0001	0.00063	0.0013	0.0025	0.005	0.01	DMSO 0.1%	MCA 1 ug/mL
OD	0.069	0.356	0.337	0.383	0.433	0.622	0.67	0.356	0.25
	0.068	0.364	0.332	0.378	0.438	0.597	0.641	0.364	0.269
	0.069	0.343	0.339	0.379	0.451	0.655	0.68	0.343	0.292
Average	0.069	0.354	0.336	0.380	0.441	0.625	0.664	0.354	0.270
SD	0.001	0.011	0.004	0.003	0.009	0.029	0.020	0.011	0.021
Average-Blank	0.000	0.286	0.267	0.311	0.372	0.556	0.595	0.286	0.202
Relative cell growth (%)		100.0	93.6	109.0	130.2	194.6	208.3	100.0	70.6
SD of relative cell growth(%)		3.7	1.3	0.9	3.3	10.2	7.1	3.7	7.4

Transformation Assay	Solvent:	DMSO 0.1%							
Compound (ug/ml)	Well No.	0.0001	0.00063	0.0013	0.0025	0.005	0.01	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	5	9	11	9	9	14	5	42
	2	10	5	2	6	11	11	10	43
	3	10	9	10	9	9	16	10	47
	4	8	7	9	4	8	13	8	46
	5	6	8	7	6	15	12	6	44
	6	8	8	8	10	9	13	8	52
Average		7.8	7.7	7.8	7.3	10.2	13.2	7.8	45.7
SD		2.0	1.5	3.2	2.3	2.6	1.7	2.0	3.6
Dunnett test							**		
t-Test for Unequal Variances (Aspin-Welch)									***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Mezelein

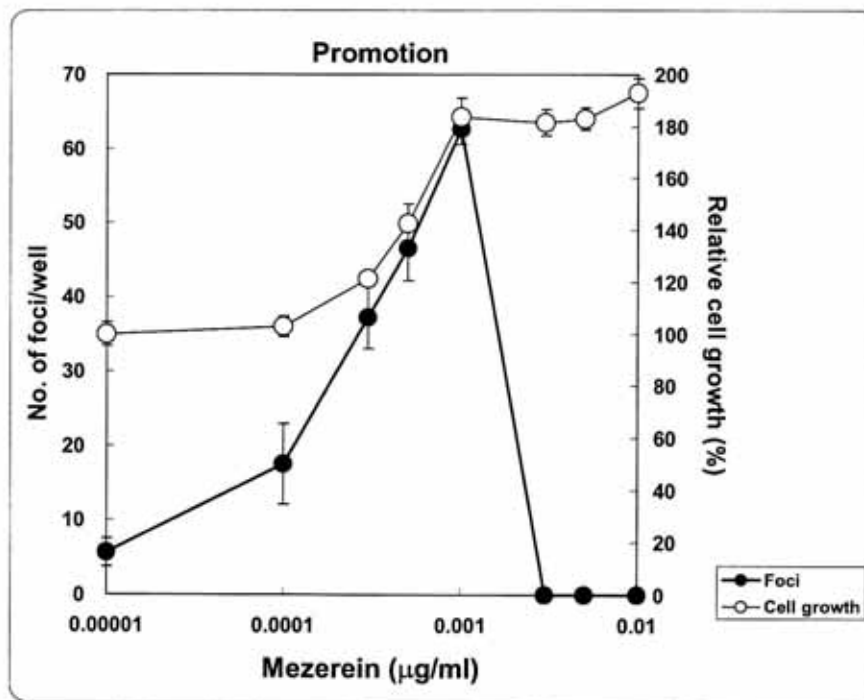
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Blank	0.00001	0.0001	0.0003	0.0005	0.001	0.003	0.005	0.01	DMSO 0.1%	TPA 50 ng/mL
OD	0.073	1.085	1.066	1.276	1.54	1.937	1.789	1.88	2.007	1.085	1.258
	0.073	1.05	1.111	1.227	1.41	1.814	1.827	1.801	1.921	1.05	1.235
	0.077	0.997	1.037	1.248	1.418	1.816	1.886	1.863	1.903	0.997	1.343
Average	0.074	1.044	1.071	1.250	1.456	1.856	1.834	1.848	1.944	1.044	1.279
SD	0.002	0.044	0.037	0.025	0.073	0.070	0.049	0.042	0.056	0.044	0.057
Average-Blank	0.000	0.970	0.997	1.176	1.382	1.781	1.760	1.774	1.869	0.970	1.204
Relative cell growth (%)		100.0	102.8	121.3	142.5	183.7	181.5	182.9	192.8	100.0	124.2
SD of relative cell growth(%)		4.6	3.8	2.5	7.5	7.3	5.0	4.3	5.7	4.6	5.9

Transformation Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Well No.	0.00001	0.0001	0.0003	0.0005	0.001	0.003	0.005	0.01	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	4	20	39	49	60	Toxic	Toxic	Toxic	4	31
	2	6	7	36	48	62	Toxic	Toxic	Toxic	6	32
	3	6	22	36	50	66	Toxic	Toxic	Toxic	6	31
	4	7	20	42	50	62	Toxic	Toxic	Toxic	7	42
	5	3	17	40	40	64	Toxic	Toxic	Toxic	3	33
	6	8	19	30	42	62	Toxic	Toxic	Toxic	8	43
Average		5.7	17.5	37.2	46.5	62.7	#DIV/0!	#DIV/0!	#DIV/0!	5.7	35.3
SD		1.9	5.4	4.2	4.4	2.1	#DIV/0!	#DIV/0!	#DIV/0!	1.9	5.6
Dunnett test			***	***	***	***					
t-Test for Unequal Variances (Aspin-Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Mezeirin

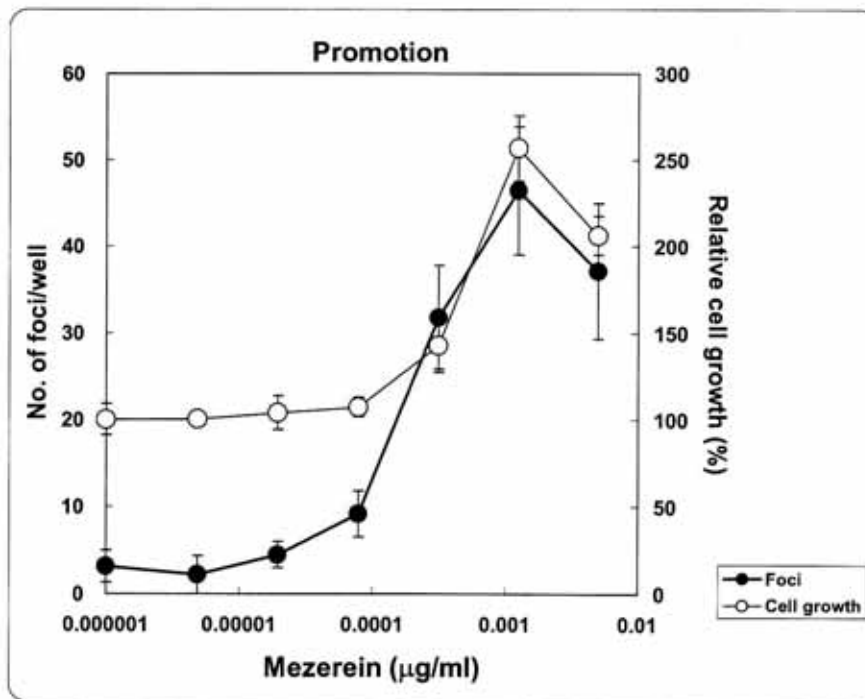
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Blank	0.000001	0.00000488	0.0000195	0.0000781	0.000313	0.00125	0.005	DMSO 0.1%	TPA 50 ng/mL	
OD	0.111	0.936	0.988	0.967	1.041	1.322	2.269	1.885	0.936	1.301	
	0.115	1.08	1.042	1.094	1.106	1.358	2.528	2.053	1.08	1.506	
	0.121	1.073	1.066	1.138	1.145	1.587	2.594	2.071	1.073	1.456	
平均	0.116	1.030	1.031	1.066	1.097	1.422	2.464	2.003	1.030	1.421	
SD	0.005	0.081	0.041	0.089	0.053	0.144	0.172	0.103	0.081	0.107	
Average-Blank	0.000	0.914	0.916	0.951	0.982	1.307	2.348	1.887	0.914	1.305	
Relative cell growth (%)		100.0	100.2	104.0	107.4	143.0	256.9	206.5	100.0	142.8	
SD of relative cell growth(%)		8.9	4.5	9.7	5.7	15.7	18.8	11.2	8.9	11.7	

Transformation Assay		Solvent: DMSO 0.1%									
Compound (ug/ml)	Well No.	0.000001	0.00000488	0.0000195	0.0000781	0.000313	0.00125	0.005	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	6	6	7	8	34	40	33	6	11	
	2	5	1	5	14	37	50	35	5	24	
	3	2	0	3	7	36	53	47	2	8	
	4	2	3	3	7	34	41	43	2	12	
	5	2	1	4	10	29	39	25	2	11	
	6	2	2	5	9	21	56	40	2	7	
Average		3.2	2.2	4.5	9.2	31.8	46.5	37.2	3.2	12.2	
SD		1.8	2.1	1.5	2.6	6.0	7.4	7.9	1.8	6.1	
Dunnett test						***	***	***			
t-Test for Unequal Variances (Aspin-Welch)										*	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Mezelein

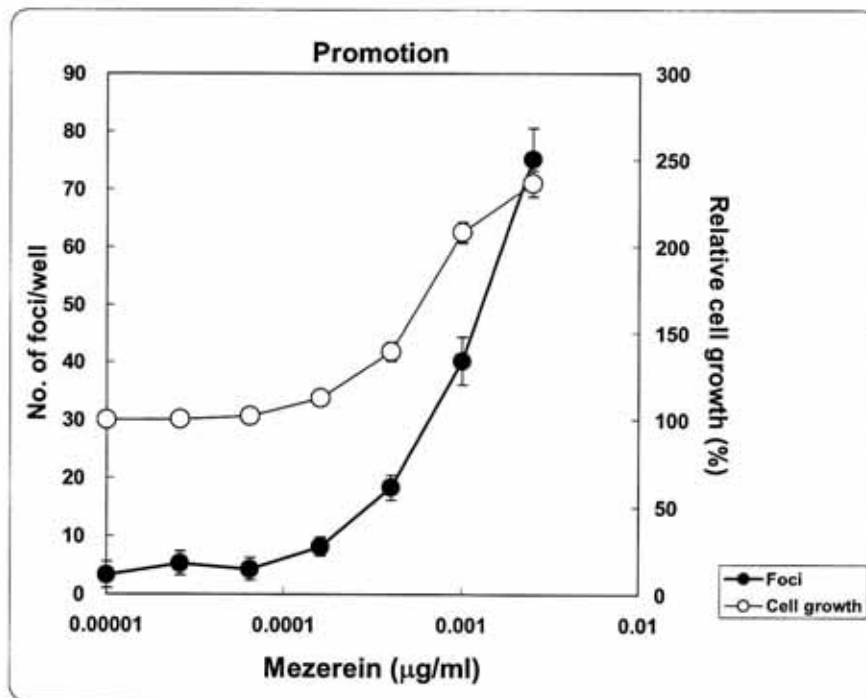
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.1%								DMSO 0.1% TPA 50 ng/mL	
Compound (ug/ml)	Blank	0.00001	0.000026	0.000064	0.00016	0.0004	0.001	0.0025			
OD	0.062	0.359	0.36	0.366	0.412	0.479	0.68	0.808	0.359	0.483	
	0.061	0.377	0.364	0.373	0.393	0.474	0.692	0.763	0.377	0.49	
	0.06	0.362	0.378	0.38	0.408	0.505	0.716	0.775	0.362	0.507	
Average	0.061	0.366	0.367	0.373	0.404	0.486	0.696	0.782	0.366	0.493	
SD	0.001	0.010	0.009	0.007	0.010	0.017	0.018	0.023	0.010	0.012	
Average-Blank	0.000	0.305	0.306	0.312	0.343	0.425	0.635	0.721	0.305	0.432	
Relative cell growth (%)		100.0	100.4	102.3	112.6	139.3	208.2	236.4	100.0	141.7	
SD of relative cell growth(%)		3.2	3.1	2.3	3.3	5.5	6.0	7.6	3.2	4.0	

Transformation Assay		Solvent: DMSO 0.1%								DMSO 0.1% TPA 50 ng/mL	
Compound (ug/ml)	Well No.	0.00001	0.000026	0.000064	0.00016	0.0004	0.001	0.0025			
No of foci	1	7	6	6	8	18	36	79	7	8	
	2	2	3	7	8	19	39	72	2	6	
	3	5	4	5	8	20	46	73	5	8	
	4	3	5	3	8	17	44	84	3	13	
	5	2	5	3	11	21	40	69	2	14	
	6	1	9	2	6	15	36	74	1	10	
Average		3.3	5.3	4.3	8.2	18.3	40.2	75.2	3.3	9.8	
SD		2.3	2.1	2.0	1.6	2.2	4.1	5.4	2.3	3.1	
Dunnett test					*	***	***	***			
t-Test (Assuming Equal Variances)										**	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Anthracene

Transformation Assay

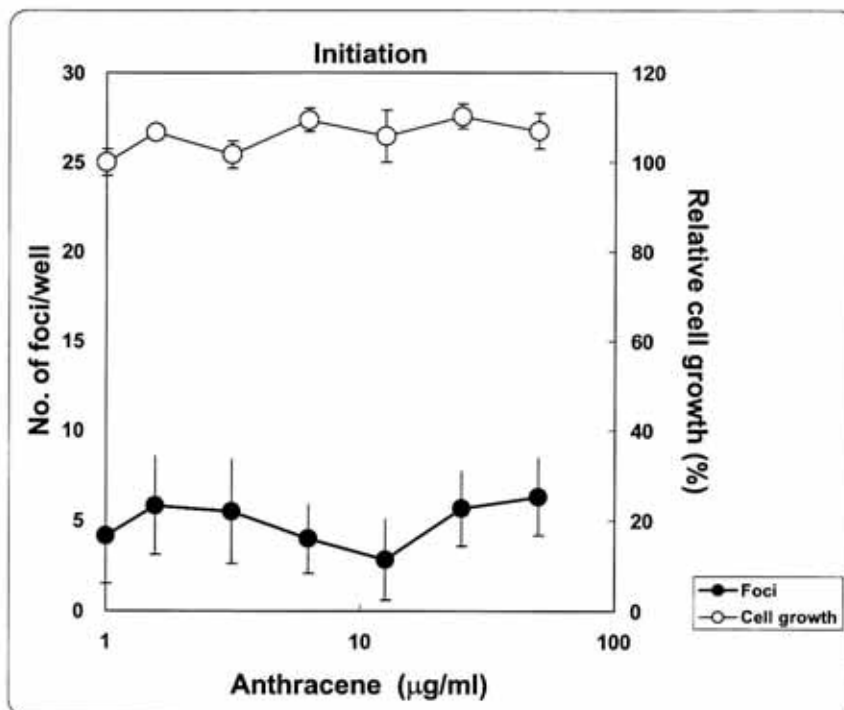
Initiation

Cell Growth Assay		Solvent: DMSO 0.5%									
Compound (ug/ml)	Blank	1	1.5625	3.125	6.25	12.5	25	50	DMSO 0.1%	MCA 1 ug/mL	
OD	0.073	1.24	1.288	1.182	1.34	1.193	1.3	1.234	1.185	0.581	
	0.073	1.193	1.286	1.248	1.307	1.295	1.355	1.323	1.137	0.601	
	0.076	1.176	1.26	1.234	1.28	1.316	1.301	1.286	1.182	0.609	
Average	0.074	1.203	1.278	1.221	1.309	1.268	1.319	1.281	1.168	0.597	
SD	0.002	0.033	0.016	0.035	0.030	0.066	0.031	0.045	0.027	0.014	
Average-Blank	0.000	1.129	1.204	1.147	1.235	1.194	1.245	1.207	1.094	0.523	
Relative cell growth (%)		100.0	106.6	101.6	109.4	105.8	110.2	106.9	100.0	47.8	
SD of relative cell growth(%)		2.9	1.4	3.1	2.7	5.8	2.8	4.0	2.5	1.3	

Transformation Assay		Solvent: DMSO 0.5%									
Compound (ug/ml)	Well No.	1	1.5625	3.125	6.25	12.5	25	50	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	8	7	8	3	1	4	5	2	24	
	2	5	6	5	2	3	3	5	3	14	
	3	3	5	3	5	5	8	10	2	24	
	4	2	1	4	2	6	6	7	4	28	
	5	6	7	3	6	1	8	4	3	30	
	6	1	9	10	6	1	5	7	2	26	
Average		4.2	5.8	5.5	4.0	2.8	5.7	6.3	2.7	24.3	
SD		2.6	2.7	2.9	1.9	2.2	2.1	2.2	0.8	5.6	
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)										***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Anthracene

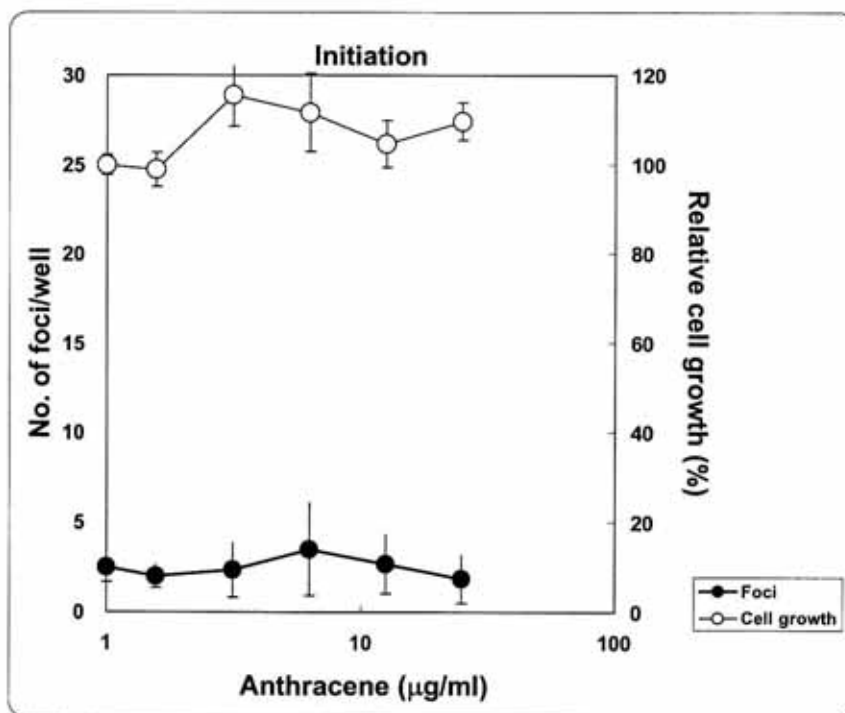
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.5%								
Compound (ug/ml)	Blank	1	1.56	3.13	6.25	12.5	25	DMSO 0.1%	MCA 1 ug/mL	
OD	0.0669	0.7751	0.7566	0.9402	0.888	0.8361	0.8315	0.7033	0.4616	
	0.0733	0.8065	0.7844	0.9293	0.9342	0.8582	0.8628	0.7356	0.4519	
	0.0665	0.7957	0.8123	0.8469	0.8062	0.7839	0.8925	0.6301	0.4411	
Average	0.069	0.792	0.784	0.905	0.877	0.826	0.862	0.690	0.452	
SD	0.004	0.016	0.028	0.051	0.064	0.038	0.031	0.054	0.010	
Average-Blank	0.000	0.724	0.716	0.837	0.808	0.757	0.793	0.621	0.383	
Relative cell growth (%)		100.0	98.9	115.6	111.7	104.6	109.7	100.0	61.6	
SD of relative cell growth(%)		2.2	3.8	7.1	8.8	5.3	4.2	8.7	1.7	

Transformation Assay		Solvent: DMSO 0.5%								
Compound (ug/ml)	Well No.	1	1.56	3.13	6.25	12.5	25	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	2	3	3	3	3	1	5	15	
	2	2	2	3	7	1	3	1	18	
	3	2	2	1	0	5	3	4	18	
	4	4	1	4	6	4	3	2	15	
	5	2	2	3	3	2	1	2	18	
	6	3	2	0	2	1	0	4	17	
Average		2.5	2.0	2.3	3.5	2.7	1.8	3.0	16.8	
SD		0.8	0.6	1.5	2.6	1.6	1.3	1.5	1.5	
Parametric Dannett t-Test (Assuming Equal Variances)									N.S.	

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Anthracene

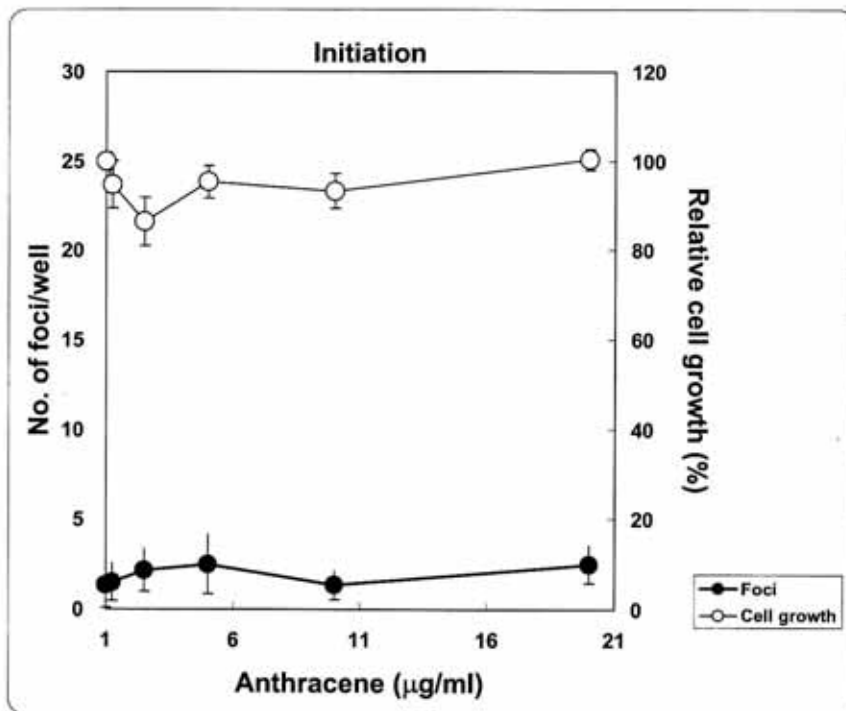
Transformation Assay

Initiation

Cell Growth Assay		Solvent: DMSO 0.5%								
Compound (ug/ml)	Blank	1	1.25	2.5	5	10	20	DMSO 0.5%	MCA 1 ug/mL	
OD	0.033	0.305	0.295	0.257	0.306	0.281	0.311	0.305	0.185	
	0.031	0.316	0.311	0.274	0.300	0.291	0.318	0.316	0.198	
	0.03	0.310	0.281	0.287	0.286	0.303	0.305	0.310	0.209	
Average	0.031	0.310	0.296	0.273	0.297	0.292	0.311	0.310	0.197	
SD	0.002	0.006	0.015	0.015	0.010	0.011	0.007	0.006	0.012	
Average-Blank	0.000	0.279	0.264	0.241	0.266	0.260	0.280	0.279	0.166	
Relative cell growth (%)		100.0	94.7	86.5	95.3	93.3	100.4	100.0	59.5	
SD of relative cell growth(%)		2.0	5.4	5.4	3.7	3.9	2.3	2.0	4.3	

Transformation Assay		Solvent: DMSO 0.5%								
Compound (ug/ml)	Well No.	1	1.25	2.5	5	10	20	DMSO 0.5%	MCA 1 ug/mL	
No of foci	1	3	1	3	4	1	2	3	13	
	2	2	3	0	1	2	2	2	14	
	3	0	1	3	3	0	3	0	17	
	4	2	0	2	0	2	1	2	16	
	5	0	2	3	3	2	3	0	10	
	6	1	2	2	4	1	4	1	16	
Average		1.3	1.5	2.2	2.5	1.3	2.5	1.3	14.3	
SD		1.2	1.0	1.2	1.6	0.8	1.0	1.2	2.6	
Dunnett test										N.S.
t-Test for Unequal Variances (Aspin-Welch)										***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Anthracene

Transformation Assay

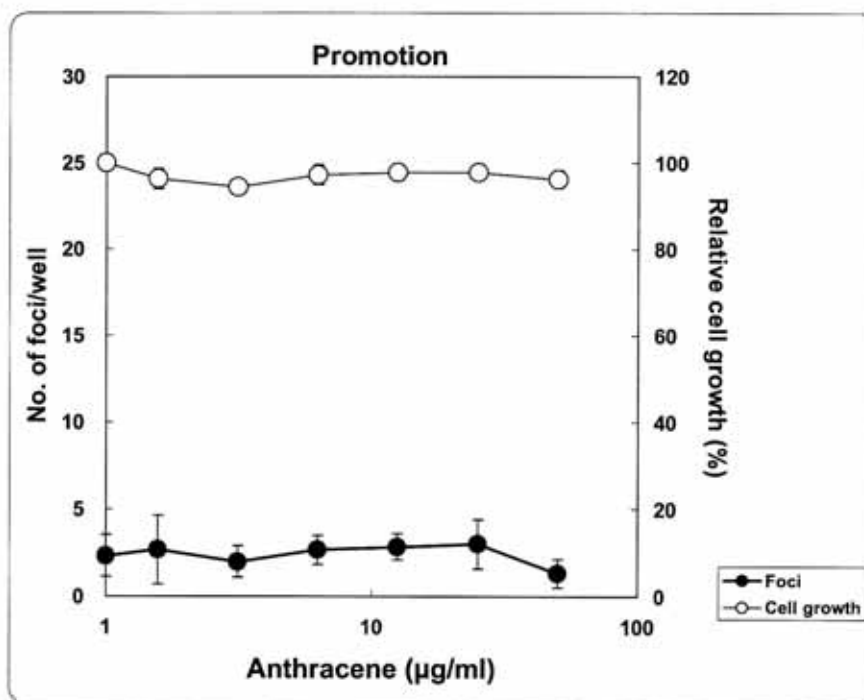
Promotion

Cell Growth Assay		Solvent: DMSO 0.5%									
Compound (ug/ml)	Blank	1	1.5625	3.125	6.25	12.5	25	50	DMSO 0.1%	TPA 50 ng/mL	
OD	0.07	0.907	0.88	0.861	0.894	0.886	0.862	0.879	0.957	1.269	
	0.067	0.886	0.871	0.845	0.863	0.885	0.891	0.867	0.969	1.239	
	0.068	0.893	0.843	0.841	0.86	0.86	0.88	0.846	0.962	1.223	
Average	0.068	0.895	0.865	0.849	0.872	0.877	0.878	0.864	0.963	1.244	
SD	0.002	0.011	0.019	0.011	0.019	0.015	0.015	0.017	0.006	0.023	
Average-Blank	0.000	0.827	0.796	0.781	0.804	0.809	0.809	0.796	0.894	1.175	
Relative cell growth (%)		100.0	96.3	94.4	97.2	97.8	97.9	96.2	100.0	131.4	
SD of relative cell growth(%)		1.3	2.3	1.3	2.3	1.8	1.8	2.0	0.7	2.6	

Transformation Assay		Solvent: DMSO 0.5%									
Compound (ug/ml)	Well No.	1	1.5625	3.125	6.25	12.5	25	50	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	2	1	3	3	3	3	2	2	9	
	2	3	1	2	3	2	4	0	4	23	
	3	4	2	2	3	2	2	2	6	15	
	4	1	2	1	3	3	1	1	2	17	
	5	1	4	3	1	4	3	1	2	27	
	6	3	6	1	3	3	5	2	6	14	
Average		2.3	2.7	2.0	2.7	2.8	3.0	1.3	3.7	17.5	
SD		1.2	2.0	0.9	0.8	0.8	1.4	0.8	2.0	6.5	
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)										**	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Anthracene

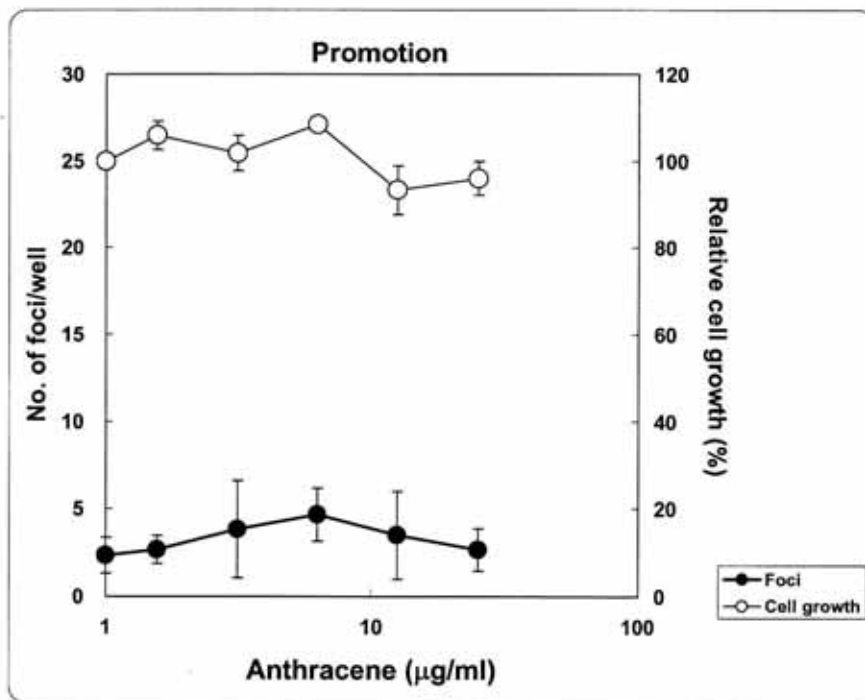
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.5%								
Compound (ug/ml)	Blank	1	1.56	3.13	6.25	12.5	25	DMSO 0.1%	TPA 50 ng/mL	
OD	0.0726	0.5185	0.558	0.5402	0.5602	0.506	0.5221	0.5127	0.6579	
	0.0775	0.5245	0.552	0.5379	0.5522	0.5048	0.4981	0.5317	0.6761	
	0.0777	0.5187	0.5303	0.5078	0.5617	0.4621	0.4891	0.4817	0.6676	
平均	0.076	0.521	0.547	0.529	0.558	0.491	0.503	0.509	0.667	
SD	0.003	0.003	0.015	0.018	0.005	0.025	0.017	0.025	0.009	
Average-Blank	0.000	0.445	0.471	0.453	0.482	0.415	0.427	0.433	0.591	
Relative cell growth (%)		100.0	105.9	101.8	108.4	93.3	96.1	100.0	136.6	
SD of relative cell growth(%)		0.8	3.3	4.1	1.1	5.6	3.8	5.8	2.1	

Transformation Assay		Solvent: DMSO 0.5								
Compound (ug/ml)	Well No.	1	1.56	3.13	6.25	12.5	25	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	3	3	1	5	1	2	2	12	
	2	2	4	4	7	7	1	3	12	
	3	2	2	3	3	6	4	3	10	
	4	4	2	2	5	1	4	8	8	
	5	2	3	9	3	3	3	3	11	
	6	1	2	4	5	3	2	3	12	
Average		2.3	2.7	3.8	4.7	3.5	2.7	3.7	10.8	
SD		1.0	0.8	2.8	1.5	2.5	1.2	2.2	1.6	
Parametric Dannett t-Test (Assuming Equal Variances)								***		N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Anthracene

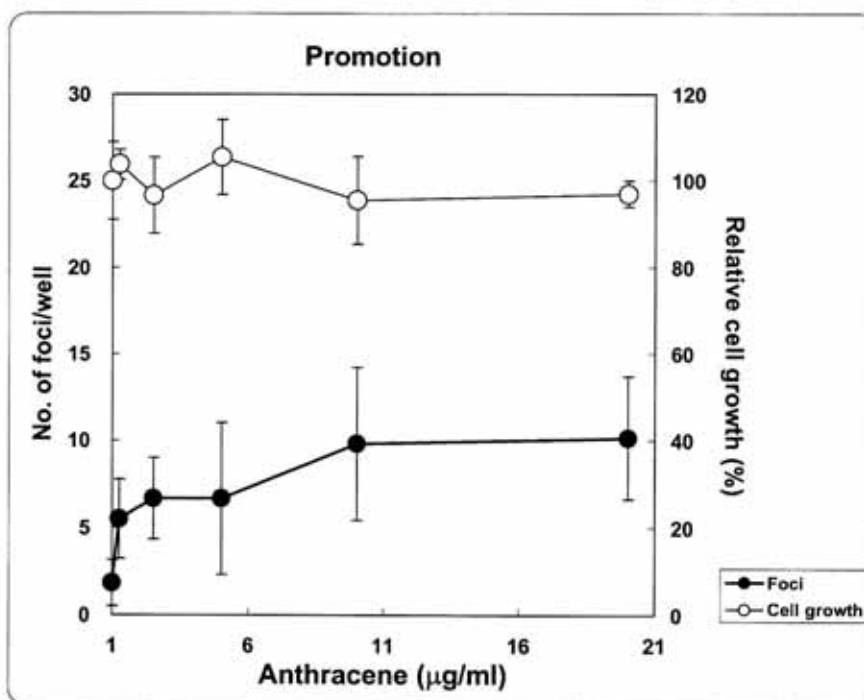
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.5%									
Compound (ug/ml)	Blank	1	1.25	2.5	5	10	20	DMSO 0.5%	TPA 50 ng/mL		
OD	0.033	0.216	0.242	0.208	0.245	0.207	0.229	0.216	0.272		
	0.031	0.252	0.248	0.23	0.262	0.22	0.233	0.252	0.314		
	0.032	0.233	0.234	0.243	0.227	0.247	0.221	0.233	0.334		
Average	0.032	0.234	0.241	0.227	0.245	0.225	0.228	0.234	0.307		
SD	0.001	0.018	0.007	0.018	0.018	0.020	0.006	0.018	0.032		
Average-Blank	0.000	0.202	0.209	0.195	0.213	0.193	0.196	0.202	0.275		
Relative cell growth (%)		100.0	103.8	96.7	105.5	95.5	97.0	100.0	136.2		
SD of relative cell growth(%)		8.9	3.5	8.8	8.7	10.1	3.0	8.9	15.7		

Transformation Assay		Solvent: DMSO 0.5%									
Compound (ug/ml)	Well No.	1	1.25	2.5	5	10	20	DMSO 0.5%	TPA 50 ng/mL		
No of foci	1	3	5	10	13	13	14	3	15		
	2	3	5	4	4	4	13	3	14		
	3	0	8	9	10	6	13	0	19		
	4	1	5	6	8	14	8	1	16		
	5	1	2	5	2	14	7	1	10		
	6	3	8	6	3	8	6	3	14		
Average		1.8	5.5	6.7	6.7	9.8	10.2	1.8	14.7		
SD		1.3	2.3	2.3	4.4	4.4	3.5	1.3	2.9		
Dunnett test				*	*	***	***				
t-Test for Unequal Variances (Aspin-Welch)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Mannitol

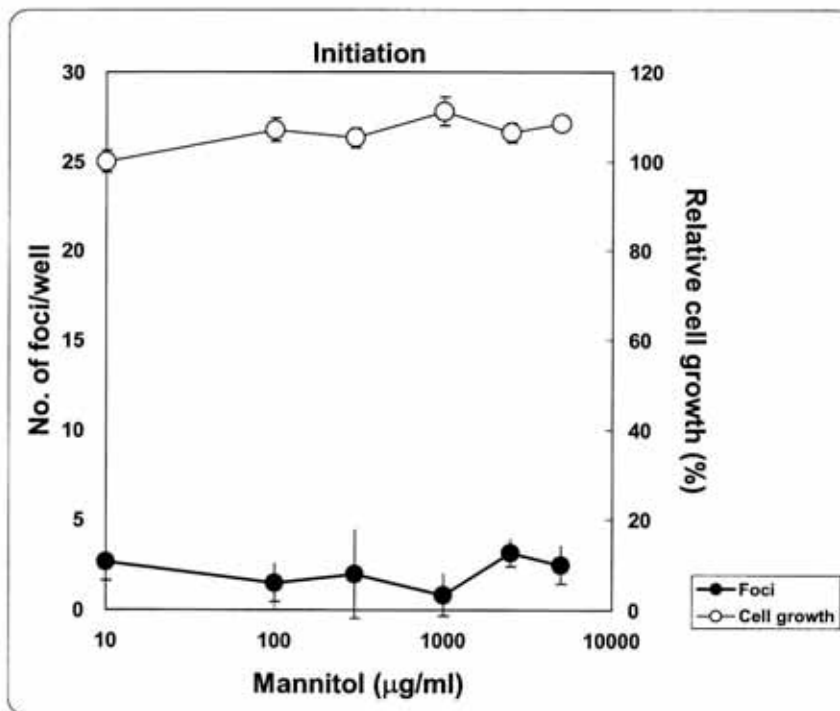
Transformation Assay

Initiation

Cell Growth Assay		Solvent: water 5%								
Compound (ug/ml)	Blank	10	100	300	1000	2500	5000	DMSO 0.1%	MCA 1 ug/mL	
OD		0.073	0.937	1.027	1.003	1.061	0.996	1.035	1.12	0.702
		0.073	0.973	0.984	1.009	1.064	1.027	1.024	1.023	0.698
		0.077	0.935	1.02	0.973	1.015	0.991	1.009	1.022	0.72
Average		0.074	0.948	1.010	0.995	1.047	1.005	1.023	1.055	0.707
SD		0.002	0.021	0.023	0.019	0.027	0.020	0.013	0.056	0.012
Average-Blank		0.000	0.874	0.936	0.921	0.972	0.930	0.948	0.981	0.632
Relative cell growth (%)			100.0	107.1	105.3	111.3	106.4	108.5	100.0	64.5
SD of relative cell growth(%)			2.4	2.6	2.2	3.1	2.2	1.5	5.7	1.2

Transformation Assay		Solvent: water 5%								
Compound (ug/ml)	Well No.	10	100	300	1000	2500	5000	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	3	0	1	3	3	3	3	3	33
	2	2	2	0	1	3	2	5	3	39
	3	1	3	6	1	2	4	3	3	35
	4	3	2	1	0	4	3	0	0	39
	5	3	1	0	0	4	1	2	2	28
	6	4	1	4	0	3	2	3	3	28
Average		2.7	1.5	2.0	0.8	3.2	2.5	2.7	33.7	
SD		1.0	1.0	2.4	1.2	0.8	1.0	1.6	5.0	
Dunnett test										N.S.
t-Test for Unequal Variances (Aspin-Welch)										***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Mannitol

Transformation Assay

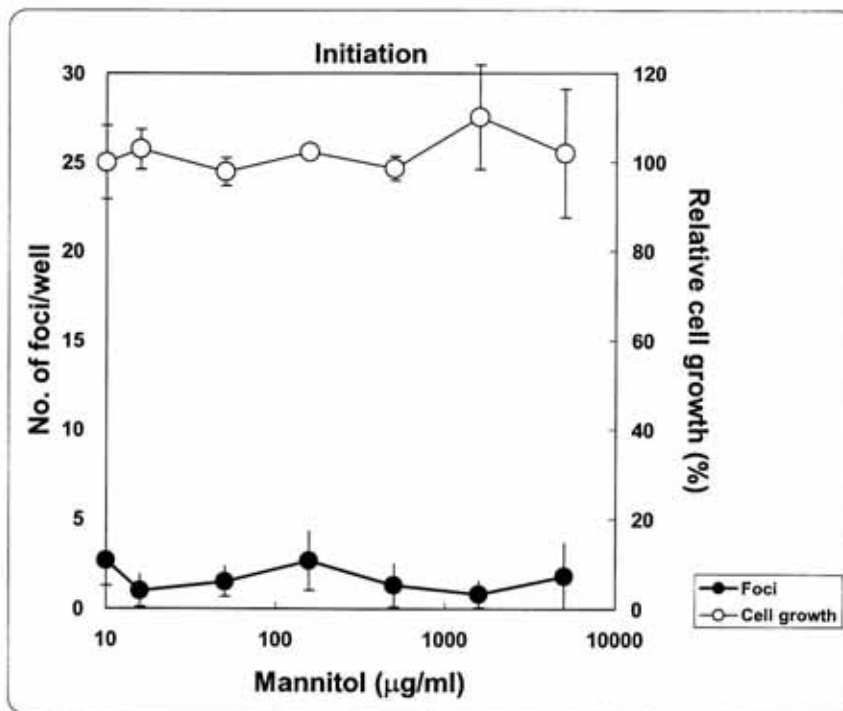
Initiation

Cell Growth Assay		Solvent: D.W. 5 %								
Compound (ug/ml)	Blank	10	15.8	50	158	500	1580	5000	DMSO 0.1%	MCA 1ug/mL
OD	0.198	2.694	2.45	2.382	2.567	2.411	3.029	2.925	-	0.937
	0.221	2.488	2.647	2.523	2.565	2.534	2.656	2.331	-	0.966
	0.227	2.318	2.606	2.449	2.525	2.459	2.51	2.385	-	1
Average	0.215	2.500	2.568	2.451	2.552	2.468	2.732	2.547	#DIV/0!	0.968
SD	0.015	0.188	0.104	0.071	0.024	0.062	0.268	0.328	#DIV/0!	0.032
Average-Blank	0.000	2.285	2.352	2.236	2.337	2.253	2.516	2.332	#DIV/0!	0.752
Relative cell growth (%)		100.0	103.0	97.9	102.3	98.6	110.1	102.1	#DIV/0!	#DIV/0!
SD of relative cell growth(%)		8.2	4.5	3.1	1.0	2.7	11.7	14.4	#DIV/0!	#DIV/0!

Transformation Assay		Solvent: D.W. 5 %								
Compound (ug/ml)	Well No.	10	15.8	50	158	500	1580	5000	DMSO 0.1%	MCA 1ug/mL
No of foci	1	2	2	2	5	0	1	0	-	11
	2	3	1	2	3	2	1	2	-	20
	3	1	0	2	2	3	0	5	-	19
	4	2	0	1	0	1	0	2	-	13
	5	5	1	0	3	2	2	2	-	10
	6	3	2	2	3	0	1	0	-	11
Average		2.7	1.0	1.5	2.7	1.3	0.8	1.8	#DIV/0!	14.0
SD		1.4	0.9	0.8	1.6	1.2	0.8	1.8	#DIV/0!	4.4
Dunnett test										
t-Test for Unequal Variances (Aspin-Welch) vs 5% Water										***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Mannitol

Transformation Assay

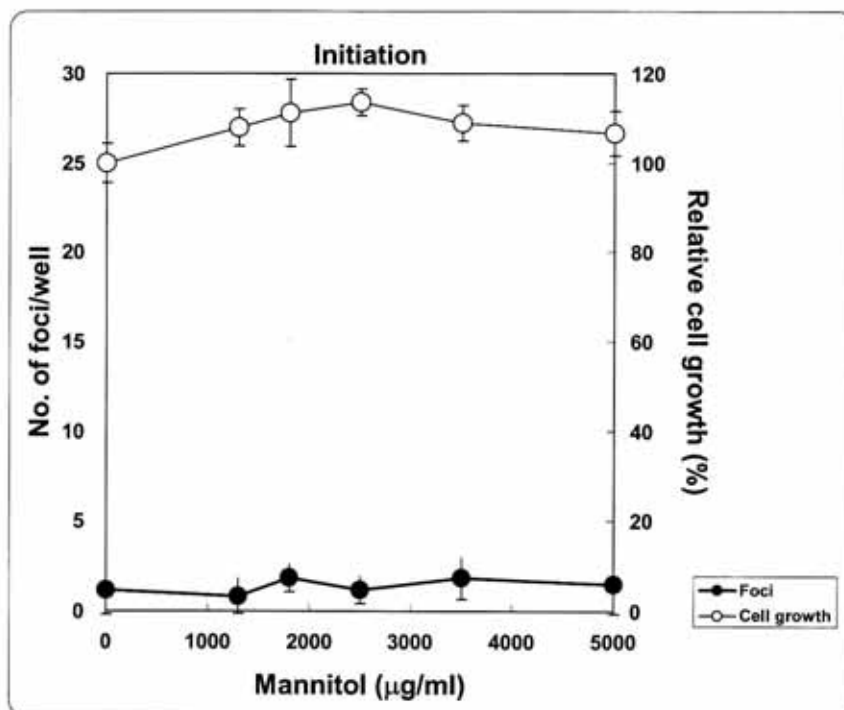
Initiation

Cell Growth Assay		Solvent:	DW 5%							
Compound (ug/ml)	Blank		0	1300	1800	2500	3500	5000	DMSO 0.1%	MCA 1 ug/mL
OD	0.063	0.285	0.324	0.307	0.336	0.322	0.321			0.224
	0.06	0.3	0.305	0.342	0.322	0.324	0.317			0.208
	0.059	0.305	0.317	0.32	0.328	0.307	0.299			0.226
Average	0.061	0.297	0.315	0.323	0.329	0.318	0.312	#DIV/0!		0.219
SD	0.002	0.010	0.010	0.018	0.007	0.009	0.012	#DIV/0!		0.010
Average-Blank	0.000	0.236	0.255	0.262	0.268	0.257	0.252	#DIV/0!		0.159
Relative cell growth (%)		100.0	107.9	111.2	113.6	108.9	106.6	#DIV/0!		#DIV/0!
SD of relative cell growth(%)		4.4	4.1	7.5	3.0	3.9	5.0	#DIV/0!		#DIV/0!

Transformation Assay		Solvent:	DW 5%							
Compound (ug/ml)	Well No.		0	1300	1800	2500	3500	5000	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	3	2	1	1	3	4		2	33
	2	0	0	2	2	0	3		2	39
	3	2	1	1	2	3	1		0	38
	4	0	2	3	0	1	0		1	35
	5	0	0	2	1	2	0		1	43
	6	2	0	2	1	2	1		0	55
Average		1.2	0.8	1.8	1.2	1.8	1.5	1.0		40.5
SD		1.3	1.0	0.8	0.8	1.2	1.6	0.9		7.9
Dunnett test										
t-Test for Unequal Variances (Aspin-Welch)										***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Mannitol

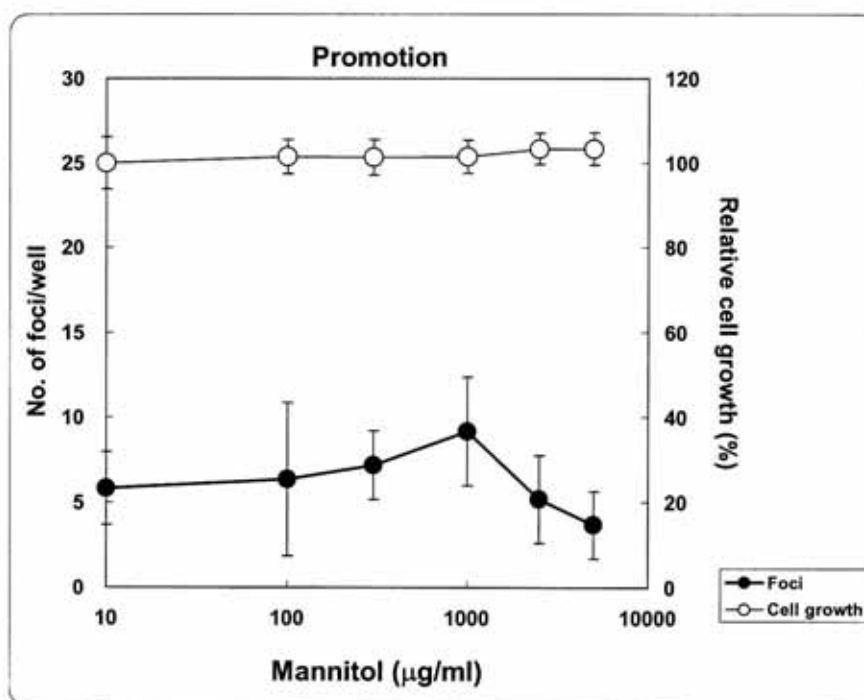
Transformation Assay

Promotion

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	10	100	300	1000	2500	5000	DMSO 0.1%	TPA 50 ng/mL		
OD	0.073	1.098	1.092	1.09	1.091	1.102	1.107	1.12	1.415		
	0.073	0.983	1.021	1.04	1.032	1.032	1.039	1.048	1.388		
	0.077	1.018	1.029	1.01	1.02	1.062	1.049	1.06	1.446		
Average	0.074	1.033	1.047	1.047	1.048	1.065	1.065	1.076	1.416		
SD	0.002	0.059	0.039	0.040	0.038	0.035	0.037	0.039	0.029		
Average-Blank	0.000	0.959	0.973	0.972	0.973	0.991	0.991	1.002	1.342		
Relative cell growth (%)		100.0	101.5	101.4	101.5	103.4	103.3	100.0	134.0		
SD of relative cell growth(%)		6.1	4.1	4.2	4.0	3.7	3.8	3.9	2.9		

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	10	100	300	1000	2500	5000	DMSO 0.1%	TPA 50 ng/mL		
No of foci	1	9	8	7	13	3	5	12	51		
	2	6	2	8	13	7	1	5	48		
	3	3	14	10	8	3	2	10	42		
	4	4	2	4	5	9	6	4	52		
	5	7	5	6	8	3	5	12	53		
	6	6	7	8	8	8	3	6	55		
Average		5.8	6.3	7.2	9.2	5.2	3.7	8.2	50.2		
SD		2.1	4.5	2.0	3.2	2.6	2.0	3.6	4.6		
Dunnett test											N.S.
t-Test (Assuming Equal Variances)											***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab III

Compound: Mannitol

Transformation Assay

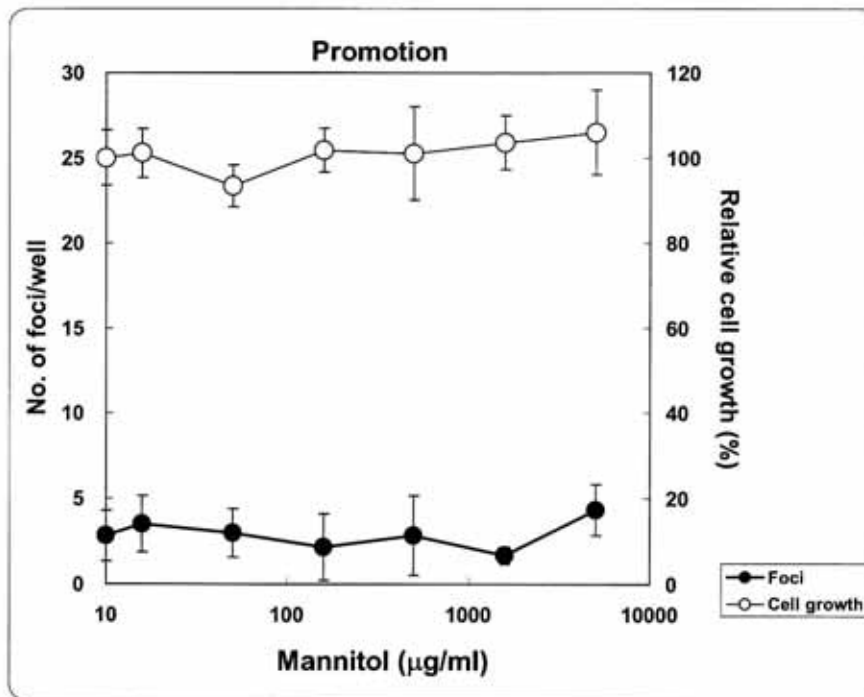
Promotion

Cell Growth Assay	Solvent:	D.W. 5 %								DMSO 0.1%	TPA 50 ng/mL
Compound (ug/ml)	Blank	10	15.8	50	158	500	1580	5000			
OD	0.171	2.388	2.393	1.984	2.151	2.513	2.172	2.59	-	3.491	
	0.196	2.172	2.209	2.164	2.336	2.099	2.431	2.259	-	3.369	
	0.218	2.146	2.172	2.154	2.33	2.158	2.325	2.222	-	3.339	
平均	0.195	2.235	2.258	2.101	2.272	2.257	2.309	2.357	#DIV/0!	3.400	
SD	0.024	0.133	0.118	0.101	0.105	0.224	0.130	0.203	#DIV/0!	0.081	
Average-Blank	0.000	2.040	2.063	1.906	2.077	2.062	2.114	2.162	#DIV/0!	3.205	
Relative cell growth (%)		100.0	101.1	93.4	101.8	101.0	103.6	106.0	#DIV/0!	#DIV/0!	
SD of relative cell growth(%)		6.5	5.8	5.0	5.2	11.0	6.4	9.9	#DIV/0!	#DIV/0!	

Transformation Assay	Solvent:	D.W. 5 %								DMSO 0.1%	TPA 50 ng/mL
Compound (ug/ml)	Well No.	10	15.8	50	158	500	1580	5000			
No of foci	1	3	3	5	0	3	1	4	-	12	
	2	2	1	3	1	3	1	3	-	12	
	3	2	3	4	2	0	2	5	-	10	
	4	1	6	2	1	2	2	4	-	6	
	5	4	4	3	4	2	2	7	-	9	
	6	5	4	1	5	7	2	3	-	3	
Average		2.8	3.5	3.0	2.2	2.8	1.7	4.3	#DIV/0!	8.7	
SD		1.5	1.6	1.4	1.9	2.3	0.5	1.5	#DIV/0!	3.6	
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch) vs 5% Water										**	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Mannitol

Transformation Assay

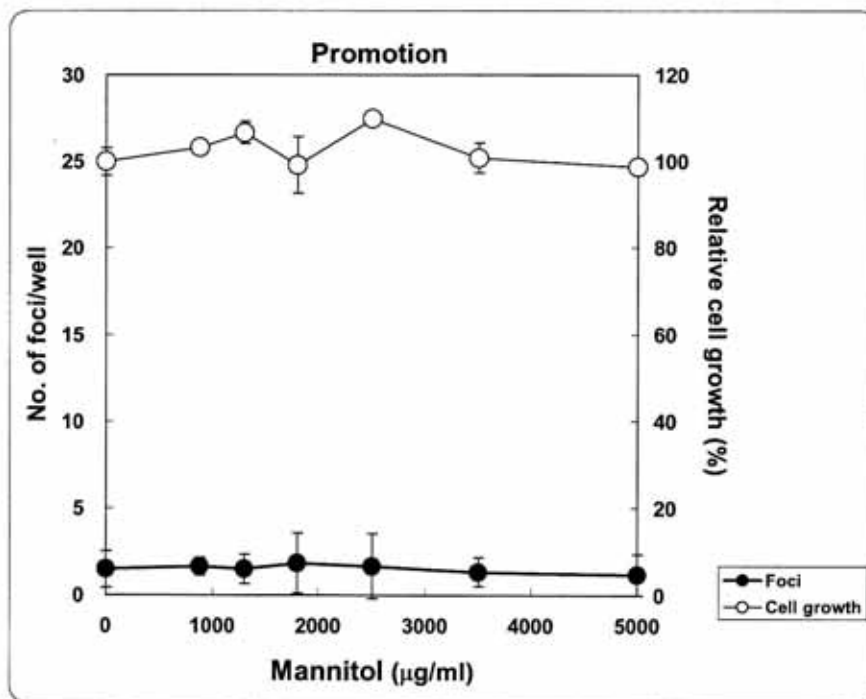
Promotion

Cell Growth Assay		Solvent: DW 5%										
Compound (ug/ml)	Blank	0	880	1300	1800	2500	3500	5000	DMSO 0.1%	TPA 50 ng/mL		
OD	0.063	0.272	0.273	0.282	0.275	0.286	0.275	0.267		0.305		
	0.06	0.259	0.274	0.282	0.267	0.282	0.262	0.261		0.305		
	0.059	0.265	0.269	0.273	0.249	0.289	0.264	0.26		0.297		
Average	0.061	0.265	0.272	0.279	0.264	0.286	0.267	0.263	#DIV/0!	0.302		
SD	0.002	0.007	0.003	0.005	0.013	0.004	0.007	0.004	#DIV/0!	0.005		
Average-Blank	0.000	0.205	0.211	0.218	0.203	0.225	0.206	0.202	#DIV/0!	0.242		
Relative cell growth (%)		100.0	103.3	106.7	99.2	109.9	100.8	98.7	#DIV/0!	#DIV/0!		
SD of relative cell growth(%)		3.2	1.3	2.5	6.5	1.7	3.4	1.8	#DIV/0!	#DIV/0!		

Transformation Assay		Solvent: DW 5%										
Compound (ug/ml)	Well No.	0	880	1300	1800	2500	3500	5000	DMSO 0.1%	TPA 50 ng/mL		
No of foci	1	1	2	2	2	0	2	1	4	9		
	2	0	2	1	1	2	1	3	1	11		
	3	3	1	2	0	1	0	0	2	12		
	4	2	1	0	1	0	2	1	3	8		
	5	1	2	2	2	5	2	0	2	13		
	6	2	2	2	5	2	1	2	2	8		
Average		1.5	1.7	1.5	1.8	1.7	1.3	1.2	2.3	10.2		
SD		1.0	0.5	0.8	1.7	1.9	0.8	1.2	1.0	2.1		
Dunnett test												
t-Test for Unequal Variances (Aspin-Welch)										***		

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Caffeine

Transformation Assay

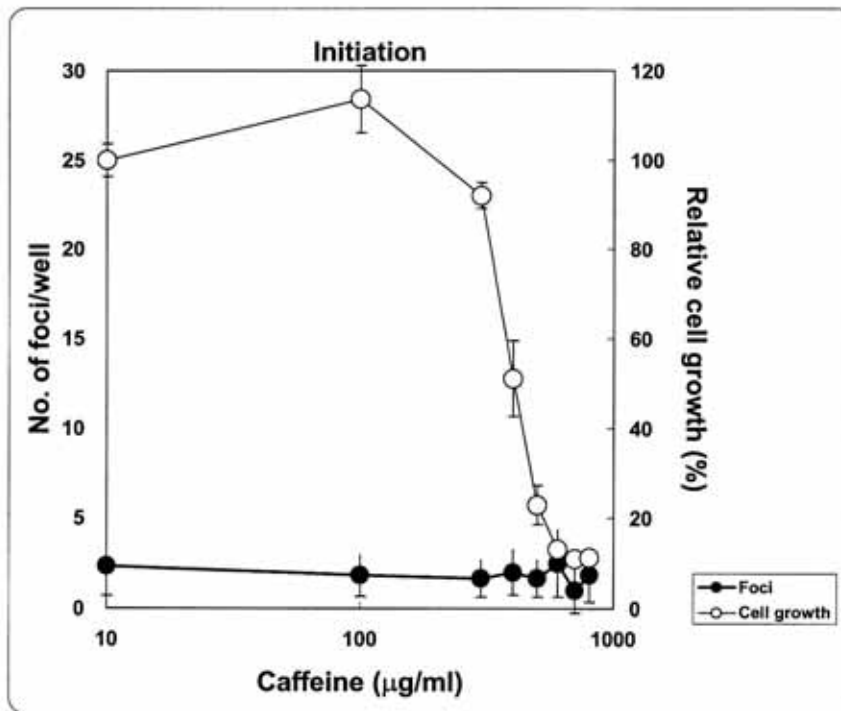
Initiation

Cell Growth Assay		Solvent: water 5%										
Compound (ug/ml)	Blank	10	100	300	400	500	600	700	800	DMSO 0.1%	MCA 1 ug/mL	
OD	0.08	1.009	1.258	1	0.667	0.341	0.213	0.202	0.204	1.159	0.639	
	0.088	1.052	1.119	0.966	0.554	0.318	0.211	0.183	0.189	1.173	0.654	
	0.09	1.079	1.155	0.944	0.51	0.26	0.213	0.189	0.188	1.243	0.529	
Average	0.086	1.047	1.177	0.970	0.577	0.306	0.212	0.191	0.194	1.192	0.607	
SD	0.005	0.035	0.072	0.028	0.081	0.042	0.001	0.010	0.009	0.045	0.068	
Average-Blank	0.000	0.961	1.091	0.884	0.491	0.220	0.126	0.105	0.108	1.106	0.521	
Relative cell growth (%)		100.0	113.6	92.0	51.1	22.9	13.2	11.0	11.2	100.0	47.2	
SD of relative cell growth(%)		3.7	7.5	2.9	8.4	4.3	0.1	1.0	0.9	4.1	6.2	

Transformation Assay		Solvent: water 5%										
Compound (ug/ml)	Well No.	10	100	300	400	500	600	700	800	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	1	1	2	4	2	1	0	4	0	19	
	2	3	3	1	2	0	2	2	1	0	15	
	3	5	2	3	0	1	2	0	3	1	17	
	4	3	2	2	2	2	3	0	0	3	18	
	5	1	3	0	2	2	1	1	1	2	25	
	6	1	0	2	2	2	3	6	3	2	24	
Average		2.3	1.8	1.7	2.0	1.7	2.5	1.0	1.8	1.3	19.7	
SD		1.6	1.2	1.0	1.3	1.0	1.9	1.3	1.5	1.2	4.0	
Dunnett test												
t-Test for Unequal Variances (Aspin-Welch)											***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab V

Compound: Caffeine

Transformation Assay

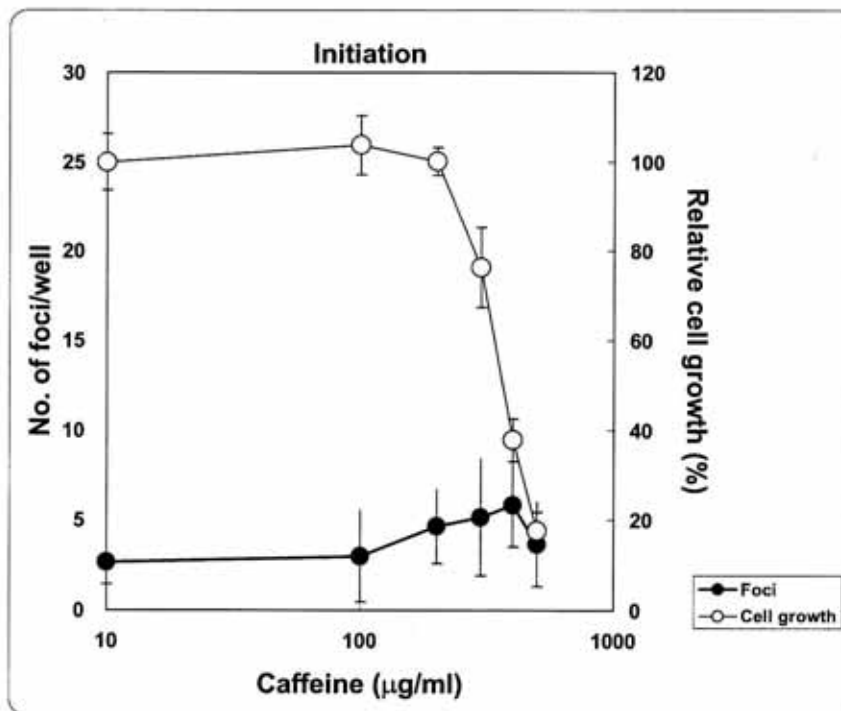
Initiation

Cell Growth Assay		Solvent: Water 5%								
Compound (ug/ml)	Blank	10	100	200	300	400	500	DMSO 0.1%	MCA 1 ug/mL	
OD	0.0628	1.0052	1.0452	1.0831	0.8255	0.4196	0.2911	0.8554	0.5062	
	0.0707	1.1261	1.1784	1.1084	0.9364	0.4261	0.2435	0.9557	0.5617	
	0.0747	1.1027	1.1232	1.0451	0.7562	0.5056	0.2084	0.9029	0.4859	
Average	0.069	1.078	1.116	1.079	0.839	0.450	0.248	0.905	0.518	
SD	0.006	0.064	0.067	0.032	0.091	0.048	0.042	0.050	0.039	
Average-Blank	0.000	1.009	1.046	1.009	0.770	0.381	0.178	0.835	0.449	
Relative cell growth (%)		100.0	103.7	100.1	76.3	37.8	17.7	100.0	53.7	
SD of relative cell growth(%)		6.4	6.6	3.2	9.0	4.7	4.1	6.0	4.7	

Transformation Assay		Solvent: Water 5%								
Compound (ug/ml)	Well No.	10	100	200	300	400	500	DMSO 0.1%	MCA 1 ug/mL	
No of foci	1	4	6	7	7	8	5	4	31	
	2	1	2	5	0	9	4	2	45	
	3	2	1	3	7	3	0	1	28	
	4	4	3	7	5	6	3	4	49	
	5	3	0	2	3	4	7	6	45	
	6	2	6	4	9	5	3	3	50	
Average		2.7	3.0	4.7	5.2	5.8	3.7	3.3	41.3	
SD		1.2	2.5	2.1	3.3	2.3	2.3	1.8	9.4	
Dunnett test										
t-Test for Unequal Variances (Aspin-Welch)									***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Caffeine

Transformation Assay

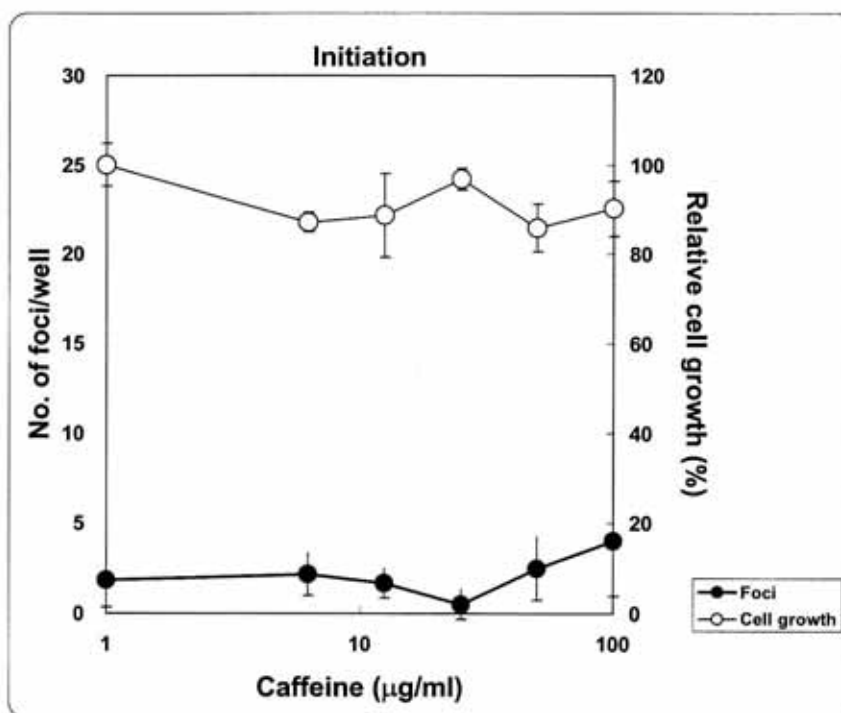
Initiation

Cell Growth Assay		Solvent: DMSO 0.5%							MCA 1 ug/mL	
Compound (ug/ml)	Blank	1	6.25	12.5	25	50	100	DMSO 0.5%	MCA 1 ug/mL	
OD	0.033	0.342	0.293	0.267	0.316	0.28	0.303	0.342	0.181	
	0.032	0.316	0.291	0.29	0.31	0.272	0.312	0.316	0.203	
	0.031	0.32	0.281	0.322	0.324	0.302	0.277	0.32	0.202	
Average	0.032	0.326	0.288	0.293	0.317	0.285	0.297	#DIV/0!	0.195	
SD	0.001	0.014	0.006	0.028	0.007	0.016	0.018	#DIV/0!	0.012	
Average-Blank	0.000	0.294	0.256	0.261	0.285	0.253	0.265	#DIV/0!	0.163	
Relative cell growth (%)		100.0	87.2	88.8	96.8	85.9	90.2	#DIV/0!	55.6	
SD of relative cell growth(%)		4.8	2.2	9.4	2.4	5.3	6.2	#DIV/0!	4.2	

Transformation Assay		Solvent: DMSO 0.5%							MCA 1 ug/mL	
Compound (ug/ml)	Well No.	1	6.25	12.5	25	50	100	DMSO 0.5%	MCA 1 ug/mL	
No of foci	1	3	3	2	0	5	3	3	19	
	2	1	1	2	0	0	2	1	16	
	3	0	2	1	0	1	10	0	17	
	4	1	4	3	0	3	2	1	23	
	5	4	2	1	1	3	3	4	15	
	6	2	1	1	2	3	4	2	13	
Average		1.8	2.2	1.7	0.5	2.5	4.0	#DIV/0!	17.2	
SD		1.5	1.2	0.8	0.8	1.8	3.0	#DIV/0!	3.5	
Dunnett test										
t-Test for Unequal Variances (Aspin-Welch)									***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Caffeine

Transformation Assay

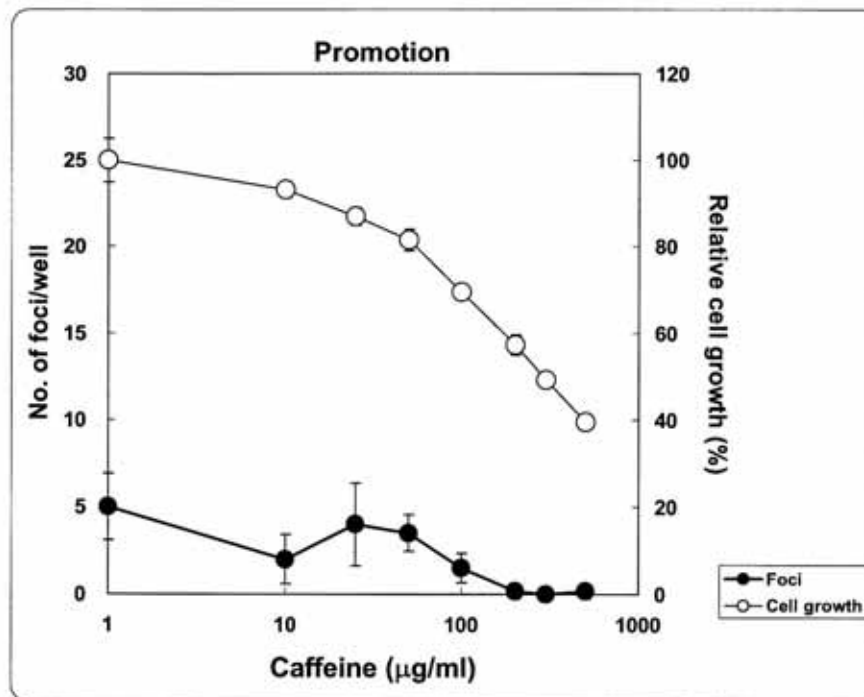
Promotion

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	1	10	25	50	100	200	300	500	DMSO 0.1%	TPA 50 ng/mL
OD	0.073	1.096	0.999	0.943	0.894	0.759	0.625	0.56	0.473	1.069	1.257
	0.073	1.061	0.964	0.927	0.874	0.757	0.661	0.55	0.452	1.048	1.238
	0.077	0.998	0.991	0.902	0.846	0.745	0.617	0.555	0.456	1.029	1.271
Average	0.074	1.052	0.985	0.924	0.871	0.754	0.634	0.555	0.460	1.049	1.255
SD	0.002	0.050	0.018	0.021	0.024	0.008	0.023	0.005	0.011	0.020	0.017
Average-Blank	0.000	0.977	0.910	0.850	0.797	0.679	0.560	0.481	0.386	0.974	1.181
Relative cell growth (%)		100.0	93.1	86.9	81.5	69.5	57.3	49.2	39.5	100.0	121.2
SD of relative cell growth(%)			5.1	1.9	2.1	2.5	0.8	2.4	0.5	1.1	2.1

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	1	10	25	50	100	200	300	500	DMSO 0.1%	TPA 50 ng/mL
No of foci	1	3	1	3	3	2	0	0	0	4	29
	2	3	0	1	4	0	0	0	1	4	31
	3	6	3	8	3	2	1	0	0	9	46
	4	5	2	3	4	1	0	0	0	6	33
	5	5	4	4	2	2	0	0	0	8	37
	6	8	2	5	5	2	0	0	0	9	37
Average		5.0	2.0	4.0	3.5	1.5	0.2	0.0	0.2	6.7	35.5
SD		1.9	1.4	2.4	1.0	0.8	0.4	0.0	0.4	2.3	6.1
Dunnet test											
t-Test for Unequal Variances (Aspin-Welch)											***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab V

Compound: Caffeine

Transformation Assay

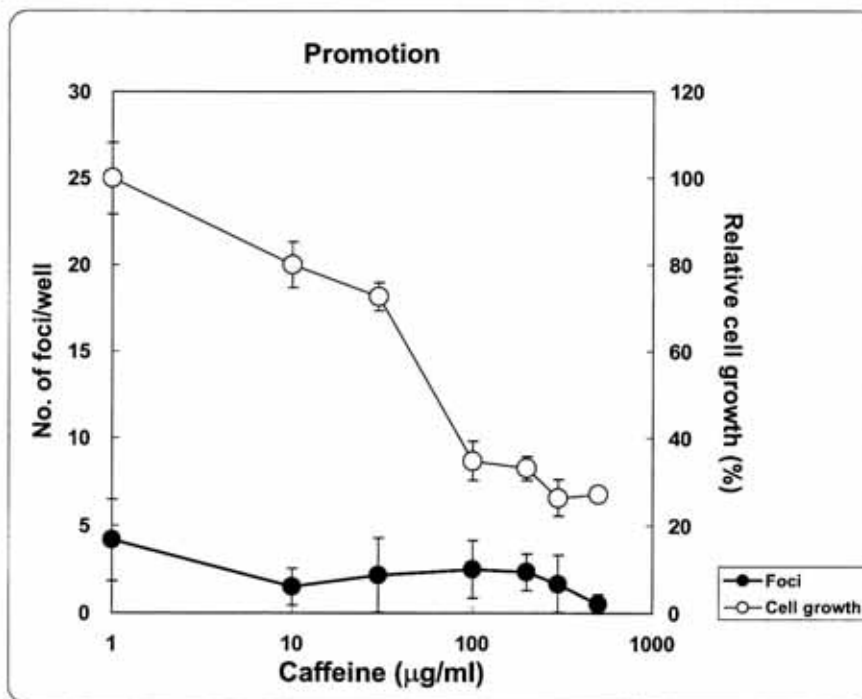
Promotion

Cell Growth Assay	Solvent: Water 5%									
	Blank	1	10	30	100	200	300	500	DMSO 0.1%	TPA 50 ng/mL
Compound (ug/ml)										
OD	0.0659	0.9931	0.794	0.7031	0.3734	0.3739	0.3161	0.2948	0.7554	0.7851
	0.0703	0.9022	0.7417	0.6978	0.323	0.349	0.2525	0.3135	0.7473	0.8139
	0.0744	0.8544	0.7052	0.6533	0.3971	0.3285	0.3106	0.2929	0.7076	0.7309
Average	0.070	0.917	0.747	0.685	0.365	0.350	0.293	0.300	0.737	0.777
SD	0.004	0.070	0.045	0.027	0.038	0.023	0.035	0.011	0.026	0.042
Average-Blank	0.000	0.846	0.677	0.615	0.294	0.280	0.223	0.230	0.667	0.706
Relative cell growth (%)		100.0	80.0	72.6	34.8	33.1	26.3	27.2	100.0	106.0
SD of relative cell growth(%)		8.3	5.3	3.2	4.5	2.7	4.2	1.3	3.8	6.3

Transformation Assay	Solvent: Water 5%									
	Well No.	1	10	30	100	200	300	500	DMSO 0.1%	TPA 50 ng/mL
Compound (ug/ml)										
No of foci	1	4	1	5	3	1	2	1	3	17
	2	1	2	4	2	3	0	0	2	25
	3	6	3	0	0	3	1	1	2	22
	4	2	0	3	5	3	0	0	1	26
	5	7	2	1	3	1	4	0	2	20
	6	5	1	0	2	3	3	1	6	19
Average		4.2	1.5	2.2	2.5	2.3	1.7	0.5	2.7	21.5
SD		2.3	1.0	2.1	1.6	1.0	1.6	0.5	1.8	3.5
Dunnett test										
t-Test for Unequal Variances (Aspin-Welch)										***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab VI

Compound: Caffeine

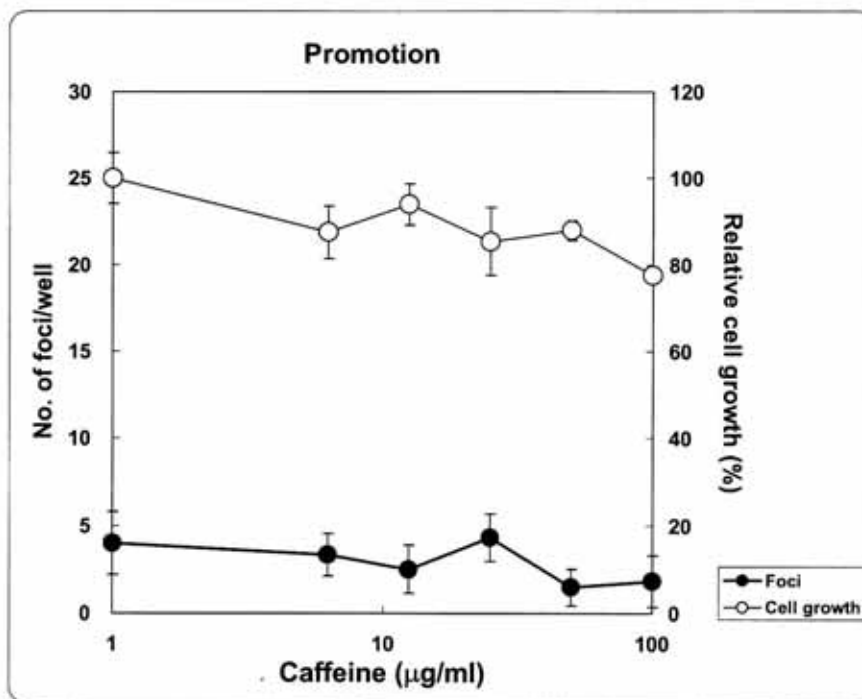
Transformation Assay

Promotion

Cell Growth Assay		Solvent: DMSO 0.5%								
Compound (ug/ml)	Blank	1	6.25	12.5	25	50	100	DMSO 0.5%	TPA 50 ng/mL	
OD	0.033	0.268	0.234	0.229	0.231	0.227	0.205	0.268	0.315	
	0.032	0.253	0.235	0.249	0.233	0.233	0.206	0.253	0.377	
	0.032	0.242	0.211	0.245	0.202	0.223	0.203	0.242	0.342	
Average	0.032	0.254	0.227	0.241	0.222	0.228	0.205	0.254	0.345	
SD	0.001	0.013	0.014	0.011	0.017	0.005	0.002	0.013	0.031	
Average-Blank	0.000	0.222	0.194	0.209	0.190	0.195	0.172	0.222	0.312	
Relative cell growth (%)		100.0	87.5	94.0	85.4	88.0	77.6	100.0	140.7	
SD of relative cell growth(%)		5.9	6.1	4.8	7.8	2.3	0.7	5.9	14.0	

Transformation Assay		Solvent: DMSO 0.5%								
Compound (ug/ml)	Well No.	1	6.25	12.5	25	50	100	DMSO 0.5%	TPA 50 ng/mL	
No of foci	1	5	2	2	2	1	2	5	18	
	2	4	3	2	6	2	0	4	15	
	3	2	4	1	4	2	4	2	11	
	4	3	5	2	5	0	1	3	14	
	5	3	4	3	5	1	1	3	25	
	6	7	2	5	4	3	3	7	16	
Average		4.0	3.3	2.5	4.3	1.5	1.8	4.0	16.5	
SD		1.8	1.2	1.4	1.4	1.0	1.5	1.8	4.8	
Dunnnett test										N.S.
t-Test for Unequal Variances (Aspin-Welch)										***

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Ascorbic acid

Transformation Assay

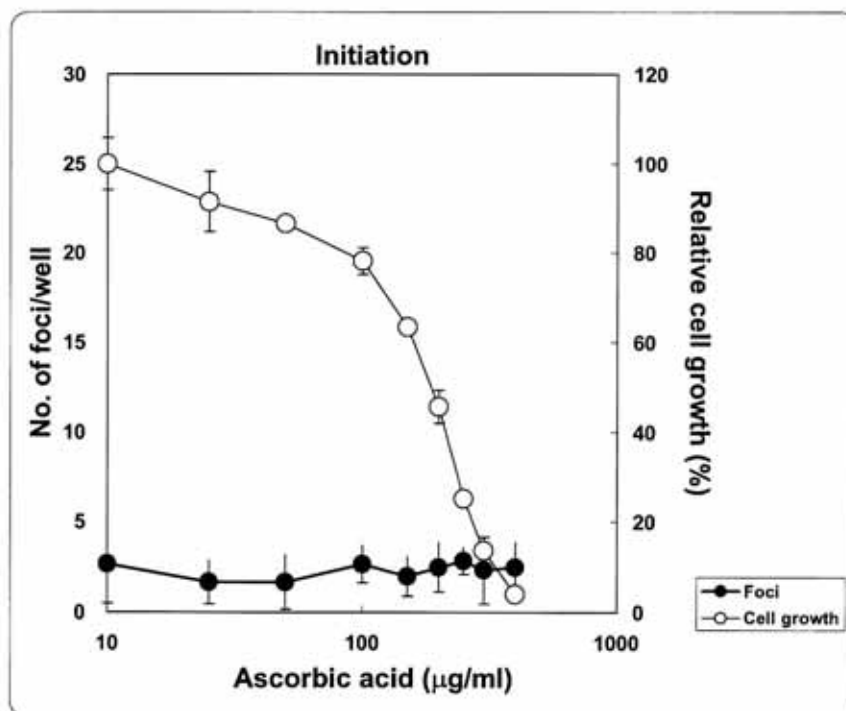
Initiation

Cell Growth Assay		Solvent: Water 5%										
Compound (ug/ml)	Blank	10	25	50	100	150	200	250	300	400	DMSO 0.1%	MCA 1 ug/mL
OD	0.08	1.339	1.26	1.1	1.045	0.847	0.677	0.398	0.209	0.14	1.265	0.613
	0.088	1.278	1.105	1.126	1.023	0.849	0.595	0.369	0.256	0.132	1.245	0.636
	0.09	1.201	1.15	1.115	0.975	0.822	0.609	0.387	0.279	0.13	1.163	0.65
Average	0.086	1.273	1.172	1.114	1.014	0.839	0.627	0.385	0.248	0.134	1.224	0.633
SD	0.005	0.069	0.080	0.013	0.036	0.015	0.044	0.015	0.036	0.005	0.054	0.019
Average-Blank	0.000	1.187	1.086	1.028	0.928	0.753	0.541	0.299	0.162	0.048	1.138	0.547
Relative cell growth (%)	100.0	91.5	86.6	78.2	63.5	45.6	25.2	13.7	4.0	100.0	48.1	1.6
SD of relative cell growth(%)		5.8	6.7	1.1	3.0	1.3	3.7	1.2	3.0	0.4	4.7	1.6

Transformation Assay		Solvent: Water										
Compound (ug/ml)	Well No.	10	25	50	100	150	200	250	300	400	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	4	1	4	3	0	3	3	1	4	5	35
	2	0	1	1	3	2	1	3	6	1	2	25
	3	4	2	0	2	2	4	3	2	4	1	27
	4	3	0	1	4	3	4	2	1	2	3	23
	5	5	3	1	1	3	1	4	2	1	1	20
	6	0	3	3	3	2	2	2	2	3	2	30
Average		2.7	1.7	1.7	2.7	2.0	2.5	2.8	2.3	2.5	2.3	26.7
SD		2.2	1.2	1.5	1.0	1.1	1.4	0.8	1.9	1.4	1.5	5.3
Dunnett test												
t-Test for Unequal Variances (Aspin-Welch)												***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Ascorbic acid

Transformation Assay

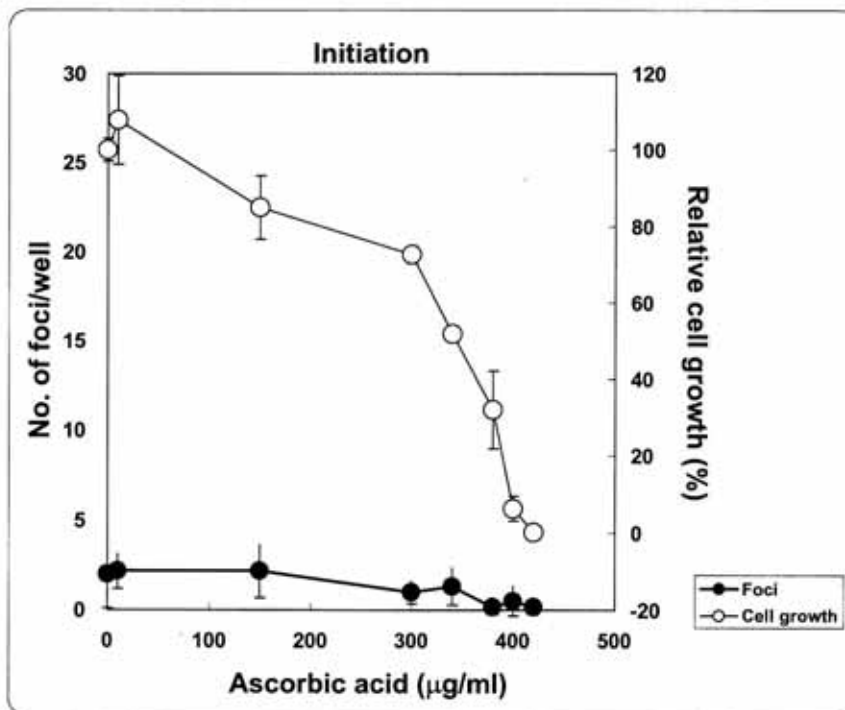
Initiation

Cell Growth Assay		Solvent: Water 5%									
Compound (ug/ml)	Blank	0	10	150	300	340	380	400	420	DMSO 0.1%	MCA 1 ug/mL
OD	0.0818	0.8058	0.7894	0.6374	0.5769	0.4324	0.3673	0.1482	0.0848	0.6681	0.365
	0.0852	0.7641	0.8019	0.6593	0.6011	0.4616	0.2297	0.1034	0.0909	0.6958	0.4466
	0.0854	0.7924	0.9369	0.7478	0.6074	0.4524	0.3323	0.1332	0.0809	0.65	0.4105
Average	0.084	0.787	0.843	0.682	0.595	0.449	0.310	0.128	0.086	0.671	0.407
SD	0.002	0.021	0.082	0.058	0.016	0.015	0.072	0.023	0.005	0.023	0.041
Average-Blank	0.000	0.703	0.759	0.597	0.511	0.365	0.226	0.044	0.001	0.587	0.323
Relative cell growth (%)		100.0	107.9	84.9	72.7	51.9	32.1	6.3	0.2	100.0	55.0
SD of relative cell growth(%)		3.0	11.6	8.3	2.3	2.1	10.2	3.2	0.7	3.9	7.0

Transformation Assay		Solvent: Water 5%									
Compound (ug/ml)	Well No.	0	10	150	300	340	380	400	420	DMSO 0.1%	MCA 1 ug/mL
No of foci	1	2	1	2	1	1	0	0	0	2	13
	2	3	2	3	1	0	0	1	0	1	8
	3	0	3	1	1	3	0	2	1	1	22
	4	2	3	3	0	1	1	0	0	3	14
	5	0	1	4	2	2	0	0	0	0	17
	6	5	3	0	1	1	0	0	0	1	21
Average		2.0	2.2	2.2	1.0	1.3	0.2	0.5	0.2	1.3	15.8
SD		1.9	1.0	1.5	0.6	1.0	0.4	0.8	0.4	1.0	5.3
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)											***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab IV

Compound: Ascorbic acid

Transformation Assay

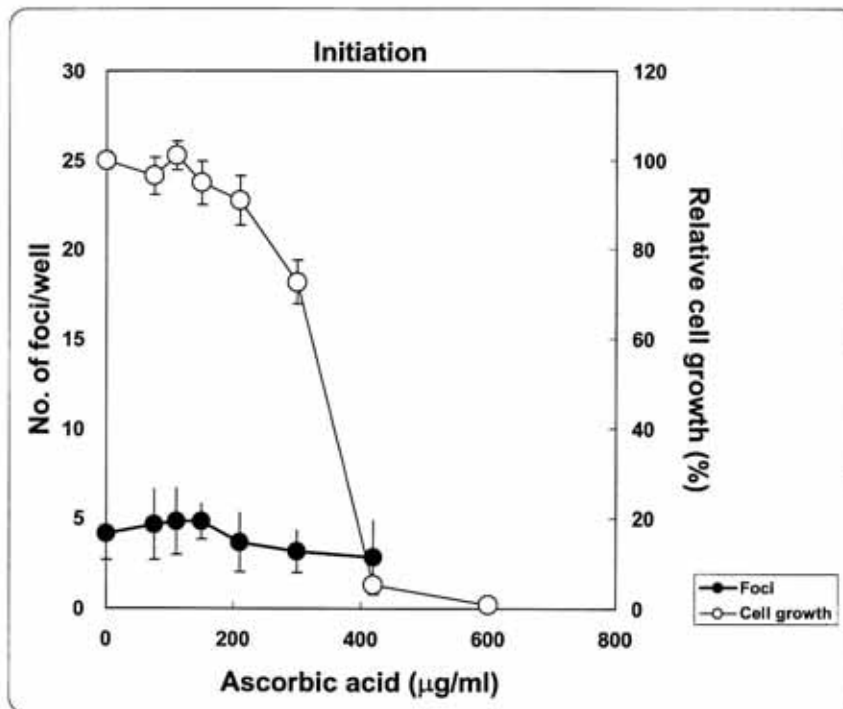
Initiation

Cell Growth Assay		Solvent: DW 5%											
Compound (ug/ml)	Blank	0	75	110	150	210	300	420	600	DMSO 0.1%	MCA 1 ug/mL		
OD	0.039	0.355	0.354	0.37	0.355	0.342	0.291	0.057	0.041				0.114
	0.039	0.36	0.36	0.352	0.326	0.342	0.261	0.06	0.043				0.105
	0.04	0.367	0.334	0.37	0.352	0.311	0.267	0.052	0.042				0.105
Average	0.039	0.361	0.349	0.364	0.344	0.332	0.273	0.056	0.042	#DIV/0!			0.108
SD	0.001	0.006	0.014	0.010	0.016	0.018	0.016	0.004	0.001	#DIV/0!			0.005
Average-Blank	0.000	0.321	0.310	0.325	0.305	0.292	0.234	0.017	0.003	#DIV/0!			0.069
Relative cell growth (%)		100.0	96.5	101.0	94.9	91.0	72.7	5.3	0.8	#DIV/0!			#DIV/0!
SD of relative cell growth(%)		1.9	4.2	3.2	5.0	5.6	4.9	1.3	0.3	#DIV/0!			#DIV/0!

Transformation Assay		Solvent: DW 5%											
Compound (ug/ml)	Well No.	0	75	110	150	210	300	420	600	DMSO 0.1%	MCA 1 ug/mL		
No of foci	1	3	8	3	4	2	3	1	Toxic	3	48		
	2	3	5	5	4	6	4	5	Toxic	7	40		
	3	4	5	7	5	3	2	4	Toxic	4	41		
	4	6	4	3	4	4	5	5	Toxic	6	44		
	5	6	2	4	6	2	3	1	Toxic	8	46		
	6	3	4	7	6	5	2	1	Toxic	5	43		
Average		4.2	4.7	4.8	4.8	3.7	3.2	2.8		5.5	43.7		
SD		1.5	2.0	1.8	1.0	1.6	1.2	2.0		1.9	3.0		
Parametric Dunnett t-Test (Assuming Equal Variances)													

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab I

Compound: Ascorbic acid

Transformation Assay

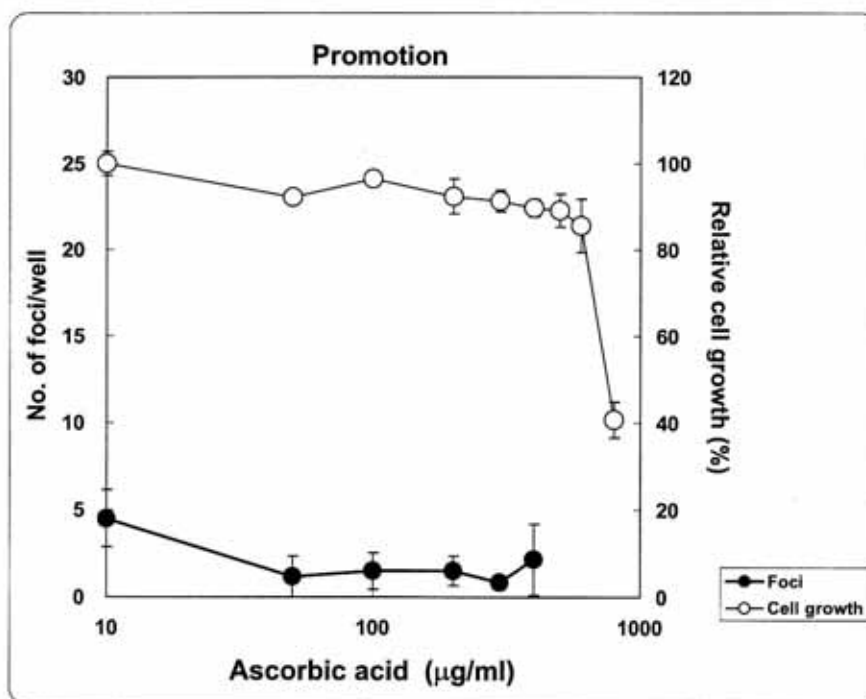
Promotion

Cell Growth Assay	Solvent:		Water 5%								DMSO 0.1%	TPA 50 ng/mL	
	Blank	10	50	100	200	300	400	500	600	800			
Compound (ug/ml)													
OD	0.073	1.148	1.068	1.12	1.04	1.063	1.044	0.99	0.949	0.557	1.108	1.409	
	0.073	1.121	1.049	1.107	1.048	1.027	1.015	1.036	0.965	0.512	1.058	1.298	
	0.077	1.182	1.082	1.109	1.118	1.08	1.059	1.072	1.071	0.469	1.076	1.231	
Average	0.074	1.150	1.066	1.112	1.069	1.057	1.039	1.033	0.995	0.513	1.081	1.313	
SD	0.002	0.031	0.017	0.007	0.043	0.027	0.022	0.041	0.066	0.044	0.025	0.090	
Average-Blank	0.000	1.076	0.992	1.038	0.994	0.982	0.965	0.958	0.921	0.438	1.006	1.238	
Relative cell growth (%)		100.0	92.2	96.4	92.4	91.3	89.7	89.1	85.6	40.7	100.0	123.1	
SD of relative cell growth(%)		2.8	1.5	0.7	4.0	2.5	2.1	3.8	6.2	4.1	2.5	8.9	

Transformation Assay	Solvent:		Water 5%								DMSO 0.1%	TPA 50 ng/mL
	Well No.	10	50	100	200	300	400	500	600	800		
Compound (ug/ml)												
No of foci	1	4	0	2	1	1	6	Tox	ToX	ToX	3	35
	2	5	1	3	1	1	1	Tox	ToX	ToX	9	30
	3	5	1	1	1	1	1	Tox	ToX	ToX	12	43
	4	2	3	2	2	0	1	Tox	ToX	ToX	4	35
	5	4	0	0	1	1	3	Tox	ToX	ToX	5	40
	6	7	2	1	3	1	1	Tox	ToX	ToX	12	40
Average		4.5	1.2	1.5	1.5	0.8	2.2				7.5	37.2
SD		1.6	1.2	1.0	0.8	0.4	2.0				4.0	4.7
Dunnett test												
t-Test (Assuming Equal Variances)												

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Ascorbic acid

Transformation Assay

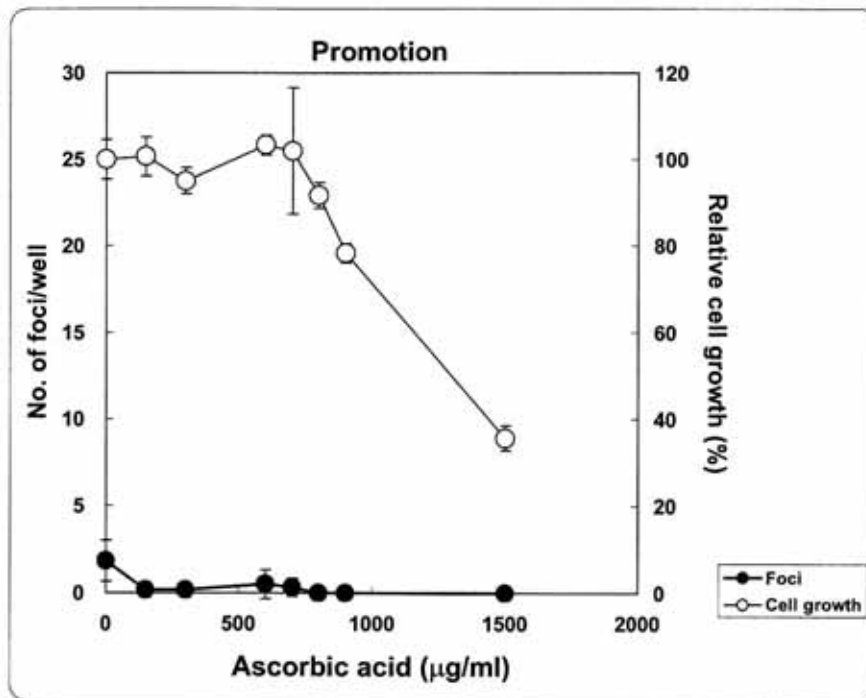
Promotion

Cell Growth Assay	Solvent: Water 5%										DMSO 0.1%	TPA 50 ng/mL
	Blank	0	150	300	600	700	800	900	1500			
Compound (ug/ml)												
OD	0.075	0.5394	0.5639	0.5204	0.5621	0.5078	0.4843	0.4388	0.2523	0.5052	0.7007	
	0.0765	0.5567	0.5306	0.524	0.554	0.6241	0.5111	0.4453	0.24	0.5333	0.7167	
	0.0739	0.5151	0.5263	0.4986	0.5416	0.5067	0.5007	0.4253	0.2259	0.4879	0.68	
平均	0.075	0.537	0.540	0.514	0.553	0.546	0.499	0.436	0.239	0.509	0.699	
SD	0.001	0.021	0.021	0.014	0.010	0.067	0.014	0.010	0.013	0.023	0.018	
Average-Blank	0.000	0.462	0.465	0.439	0.477	0.471	0.424	0.361	0.164	0.434	0.624	
Relative cell growth (%)		100.0	100.7	95.1	103.4	102.0	91.7	78.2	35.6	100.0	143.9	
SD of relative cell growth(%)		4.5	4.5	3.0	2.2	14.6	2.9	2.2	2.9	5.3	4.2	

Transformation Assay	Solvent: Water 5%										DMSO 0.1%	TPA 50 ng/mL
	Well No.	0	150	300	600	700	800	900	1500			
Compound (ug/ml)												
No of foci	1	3	1	0	0	0	0	0	0	3	11	
	2	1	0	0	2	0	0	0	0	4	11	
	3	2	0	0	0	0	0	0	0	1	12	
	4	3	0	0	0	1	0	0	0	6	10	
	5	2	0	1	0	0	0	0	0	4	12	
	6	0	0	0	1	1	0	0	0	1	11	
Average		1.8	0.2	0.2	0.5	0.3	0.0	0.0	0.0	3.2	11.2	
SD		1.2	0.4	0.4	0.8	0.5	0.0	0.0	0.0	1.9	0.8	
Dunnett test												
t-Test for Unequal Variances (Aspin-Welch)											***	

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Assay Laboratory: Lab II

Compound: Ascorbic acid

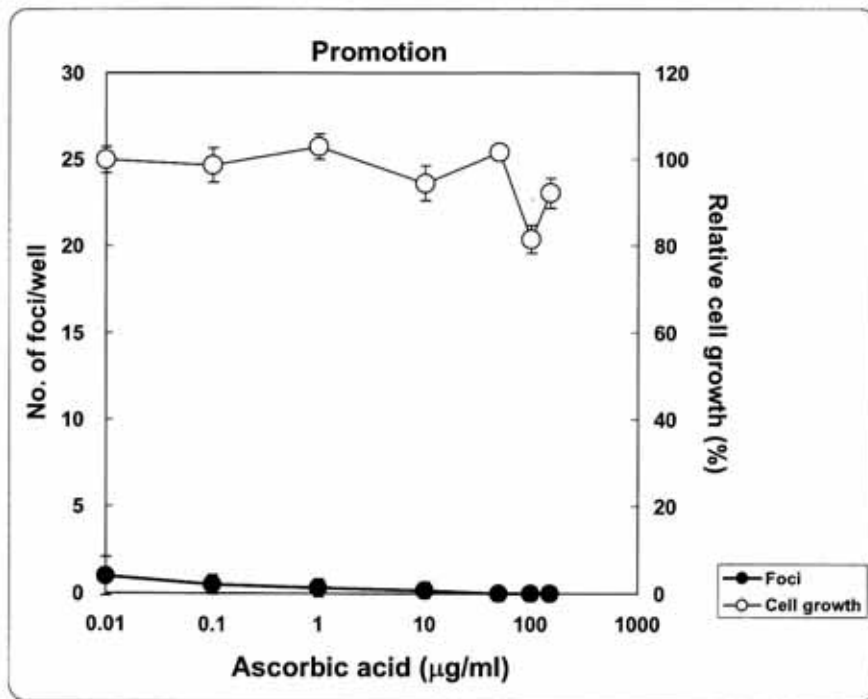
Transformation Assay

Promotion

Cell Growth Assay		Solvent: water 5%									
Compound (ug/ml)	Blank	0.01	0.1	1	10	50	100	150	DMSO 0.1%	TPA 50 ng/mL	
OD	0.0595	0.358	0.3564	0.3605	0.3561	0.361	0.3122	0.3374	0.35	0.4836	
	0.0598	0.3715	0.3695	0.3728	0.3463	0.3669	0.3098	0.3483	0.3514	0.5203	
	0.0597	0.3542	0.3457	0.3774	0.3316	0.3706	0.2944	0.3272	0.3301	0.5025	
Average	0.060	0.361	0.357	0.370	0.345	0.366	0.305	0.338	0.344	0.502	
SD	0.000	0.009	0.012	0.009	0.012	0.005	0.010	0.011	0.012	0.018	
Average-Blank	0.000	0.302	0.298	0.311	0.285	0.307	0.246	0.278	0.284	0.442	
Relative cell growth (%)		100.0	98.7	103.0	94.5	101.6	81.5	92.2	100.0	155.7	
SD of relative cell growth(%)		3.0	4.0	2.9	4.1	1.6	3.2	3.5	4.2	6.5	

Transformation Assay		Solvent: water 5%									
Compound (ug/ml)	Well No.	0.01	0.1	1	10	50	100	150	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	0	1	0	0	0	0	0	1	9	
	2	1	0	1	0	0	0	0	0	12	
	3	1	0	0	0	0	0	0	3	10	
	4	1	1	0	1	0	0	0	2	14	
	5	3	1	1	0	0	0	0	0	11	
	6	0	0	0	-	0	0	0	0	13	
Average		1.0	0.5	0.3	0.2	0.0	0.0	0.0	1.0	11.5	
SD		1.1	0.5	0.5	0.4	0.0	0.0	0.0	1.3	1.9	
Dunnett test											N.S.
t-Test (Assuming Equal Variances)											***

The true value is zero. This value was temporarily put to make logarithm graph.



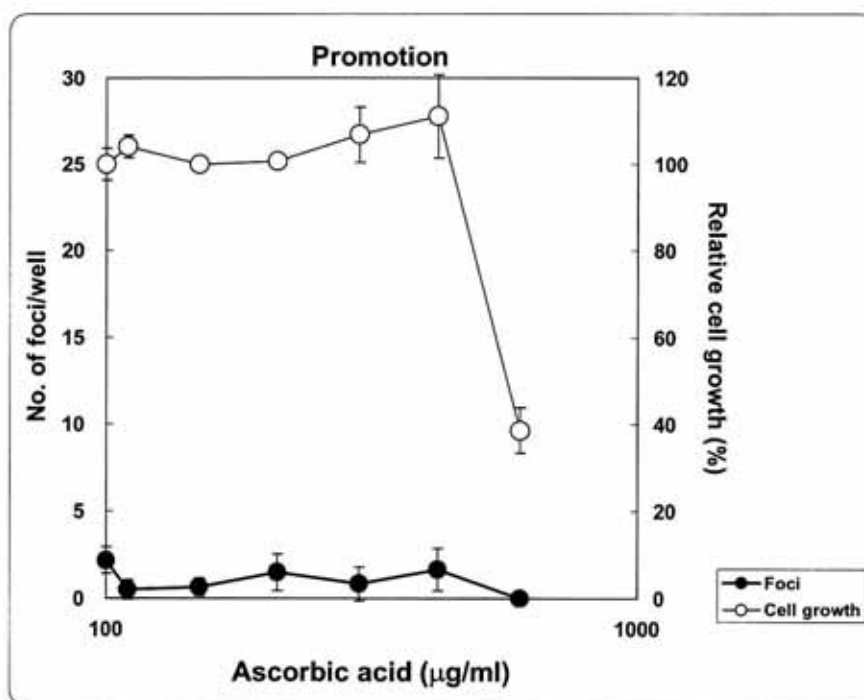
Assay Laboratory: Lab IV
 Compound: Ascorbic acid
 Transformation Assay
 Promotion

Cell Growth Assay	Solvent:	DW 5%									
Compound (ug/ml)	Blank	100	110	150	210	300	420	600	DMSO 0.1%	TPA 50 ng/mL	
OD	0.062	0.312	0.321	0.318	0.322	0.317	0.328	0.158			0.388
	0.061	0.313	0.332	0.315	0.315	0.348	0.337	0.147			0.402
	0.058	0.329	0.333	0.321	0.323	0.342	0.375	0.174			0.457
Average	0.060	0.318	0.329	0.318	0.320	0.336	0.347	0.160	#DIV/0!		0.416
SD	0.002	0.010	0.007	0.003	0.004	0.016	0.025	0.014	#DIV/0!		0.036
Average-Blank	0.000	0.258	0.268	0.258	0.260	0.275	0.286	0.099	#DIV/0!		0.355
Relative cell growth (%)		100.0	104.1	100.0	100.8	106.9	111.1	38.6	#DIV/0!	#DIV/0!	
SD of relative cell growth(%)		3.7	2.6	1.2	1.7	6.4	9.7	5.3	#DIV/0!	#DIV/0!	

Transformation Assay	Solvent:	DW 5%									
Compound (ug/ml)	Well No.	100	110	150	210	300	420	600	DMSO 0.1%	TPA 50 ng/mL	
No of foci	1	1	1	1	2	2	0 TOX		2		26
	2	2	1	1	2	2	2		1		20
	3	3	0	0	1	0	1		2		17
	4	3	1	0	3	0	3		3		17
	5	2	0	1	1	1	1		3		19
	6	2	0	1	0	0	3		5		21
Average		2.2	0.5	0.7	1.5	0.8	1.7	#DIV/0!	2.7		20.0
SD		0.8	0.5	0.5	1.0	1.0	1.2	#DIV/0!	1.4		3.3
Dunnett test											
t-Test for Unequal Variances (Aspin-Welch)											***

N.S.

The true value is zero. This value was temporarily put to make logarithm graph.



Annex 5

Table 55

Supplier, catalog number and lot number of test chemicals in the phase I of 96-well method validation study.

Chemical	CAS no. ^a	Supplier	Catalog no.	Lot no.	
3-Methylcholanthrene	56-49-5	Supelco	442388	LB61087	
Benzo[<i>a</i>]pyrene	50-32-8	Wako	020-13591	TSL8636	
2-Acetylaminofluorene	53-96-3	Ultra Scientific	Rcc-002	NTO1276	
TPA ^b	16561-29-8	Wako	545-00261	UIL0088	
<i>o</i> -Toluidine	95-53-4	Wako	209-01992	PEH9805	
Anthracene	120-12-7	Wako	010-04222	PEG1159	
Phenanthrene	85-01-8	Wako	162-00852	PEN0411	
3-Methylcholanthrene	56-49-5	Aldrich	213942	11303JE	Positive control
TPA	16561-29-8	Sigma	P8139	038K1501	Positive control

^a Chemical Abstract Service registry number.

^b 12-*O*-tetradecanoylphorbol-13-acetate.

Table 56

Supplier, catalog number and lot number of test chemicals in the phase II of 96-well method validation study.

Chemical	CAS no. ^a	Supplier	Catalog no.	Lot no.	
<i>N</i> -Methyl- <i>N</i> '-nitro- <i>N</i> -nitrosoguanidine	70-25-7	TCI	M0527	2VXVI	
Benzo[<i>a</i>]pyrene	50-32-8	Wako	020-13591	KWP9839	
Dibenz[<i>a,h</i>]anthracene	53-70-3	TCI	D0145	JF01	
Sodium arsenite	7784-46-5	Wako	199-01242	CDJ5162	
Cadmium chloride	10108-64-2	Wako	032-00122	PEE3332	
Methapyrilene HCl	135-23-9	Sigma	M9125-25G	037F09291	
Mezerein	34807-41-5	Wako	544-01691	GNM1691	
Lithocholic acid	434-13-9	Wako	127-04982	DPF1761	
Pyrene	129-00-0	Wako	167-05302	KWM0171	
Caprolactam	105-60-2	Wako	036-01122	CDL2706	
Ampicillin sodium	69-52-3	Wako	016-10373	CDM1299	
L-Ascorbic acid	50-81-7	Wako	012-04802	CDG0167	
D-Mannitol	69-65-8	Wako	139-00842	CDK3426	
Caffeine	58-08-2	Wako	031-06792	CDK3686	
Phorbol	17673-25-5	MP Biomedicals	151849	8218F	
Eugenol	97-53-0	Wako	053-03932	KWJ3327	
3-Methylcholanthrene	56-49-5	Aldrich	11303JE		Positive control
TPA ^b	16561-29-8	Sigma	038K1501		Positive control

^a Chemical Abstract Service registry number.

^b 12-*O*-tetradecanoylphorbol-13-acetate.

Annex 6

Protocol of Bhas 42 Cell Transformation Assay (2008.11.18) 96-Well Method Ver. 2

This protocol is described for the international validation study of Bhas 42 cell transformation assay using 96-well plates. Materials and reagents with their catalogue numbers are listed in annex 1.

I. Preparation

1. Materials

1) Cell line

Bhas 42 cells (v-Ha-*ras*-transfected Balb/c 3T3 clone A31-1-1 cells)^{1,2}: Free from bacteria, fungi and mycoplasma, supplied from HRI (Hatano Research Institute, Food and Drug Safety Center, Japan) Cell Bank.

2) Media

MEM: Minimum essential medium with 2.2 g/L NaHCO₃.

DMEM/F12: Dulbecco's modified Eagle's medium/F12 with 1.2 g/L NaHCO₃.

FBS: Fetal bovine serum should be selected showing a low spontaneous focus formation and a high focus formation in the positive control.

PS: Penicillin G sodium (10000 units/mL) and streptomycin sulfate (10 mg/mL).

M10F: MEM + 10% FBS + 1% PS (500 mL MEM + 56 mL FBS + 5 mL PS): Used for the expansion of provided cells, cell storage, and the first culture after thawing.

DF5F: DMEM/F12 + 5% FBS + 1% PS (500 mL DMEM/F12 + 26.5 mL FBS + 5 mL PS): Used for routine passages, cell growth assays and transformation assays.

3) Chemicals

Test chemicals and solvent/vehicle: Test chemicals are dissolved or suspended in an appropriate solvent or vehicle and diluted with the solvent/vehicle to each individual concentration before added to culture media so that all chemical treatment media contain an equal concentration of the solvent/vehicle. The final concentration of the solvent/vehicle in the medium is 5% with distilled water/ultra pure water and 0.1% with DMSO (permissible up to 0.5% when a test chemical does not dissolve).

Negative and positive controls: The solvent/vehicle for a test chemical is used as the negative control. A known initiator, 3-methylcholanthrene (MCA: final concentration of 1 µg/mL), and a promoter, 12-*O*-tetradecanoylphorbol-13- acetate (TPA: final concentration of 50 ng/mL), are used for the positive controls.

4) Fixatives and staining solutions

Formalin (37% formaldehyde): Used for fixing cells.

Methanol: Used for fixing cells.

0.1% crystal violet (CV) solution: Used for staining cells in cell growth assays. First, 1 g of crystal violet is dissolved in 50 mL of ethanol, and the total volume is adjusted to 1 L with distilled water/ultra pure water.

Extraction solution (0.02 mol/L HCl and 50% ethanol): Used for extracting CV in cell growth assays. 480 mL distilled water/ultra pure water + 500 mL ethanol + 20 mL 1 M HCl.

5% Giemsa solution: Used for staining cells in transformation assays.

5) Culture vessels

φ100-mm dishes: Used for routine passage.

96-well plates: Used for cell growth and transformation assays.

6) Micropipettes

Electronic multichannel pipettes are convenient.

2. Cell culture

1) Cell culture and passage

Bhas 42 cells are cultured in a humidified 5% CO₂ incubator at 37°C. The passage should be done at about 70% confluence of cell growth.

2) Preparation of cell stock

The provided cells are expanded in M10F. The regular method of preparation of cell stock is as follows:

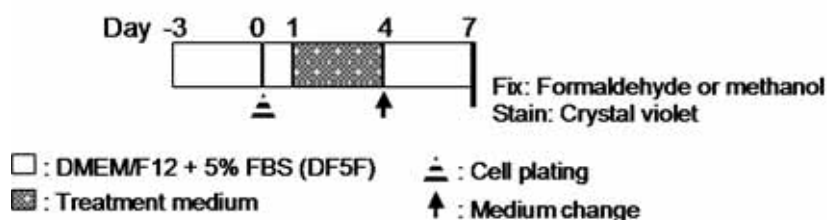
One tube (2.5 x 10⁵ cells) is thawed and suspended in 50 mL of M10F. The cell suspension is distributed into 5 of 100-mm tissue culture dishes at a volume of 10 mL/dish and cultured. Usually the cultures become to be 50-70% confluent in 4 days under these conditions. The cultures at 50-70% confluence are trypsinized with 0.25% trypsin after washed once with 0.02% EDTA-PBS(-). The cells are resuspended at a cell density of 5 x 10⁵ cells/mL in cold fresh M10F containing 5% DMSO, frozen in 0.5 mL aliquots (2.5 x 10⁵ cells/tube) at - 80°C and stored in liquid nitrogen. Every transformation experiment is started from this frozen stock.

For this validation study Bhas 42 cells are provided at passage 17 (2006.12.16, P17) from HRI. In this validation study Bhas 42 cells must be used at passage 18 for the transformation assays. But for cell growth assays the cells at higher passages can be used and should be used to save frozen cell stocks for subsequent transformation assays.

II. Experimental procedures

A. Initiation assay³⁾

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

The highest concentration is 5 mg/mL^{4,5)}. In the case of chemicals difficult to dissolve, the highest concentration may be one or two level higher than the concentration showing the utmost solubility. Five or more concentration levels are set by an appropriate serial dilution factor such as 10, square root 10 or 2. In some chemicals the cell

growth assay should be repeated in a narrower concentration range. The solvent/vehicle for a test chemical is used as the control. Each cell growth assay should include each solvent/vehicle control. Eight wells of 96-well plates are used for each concentration.

Blank control: Wells added with the medium alone are prepared for the blank control in cell growth assays. The blank control can be shared among different experiments conducted at the same time.

2) Procedure of cell growth assay

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cell suspension is transferred at a volume of 10 mL to $\phi 100$ -mm dishes. The cells with a high passage number can be used for the cell growth assay.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 4000 cells/mL. The cell suspension is distributed into each well of 96-well plates at a volume of 0.05 mL (200 cells/well). Because the volume is very small, the cell suspension is spread on the bottom of well by tapping the plates lightly. After keeping the plates at room temperature for 15 min until the cells are attached to the bottom, they are put into a incubator. Eight wells are prepared for each treatment condition.

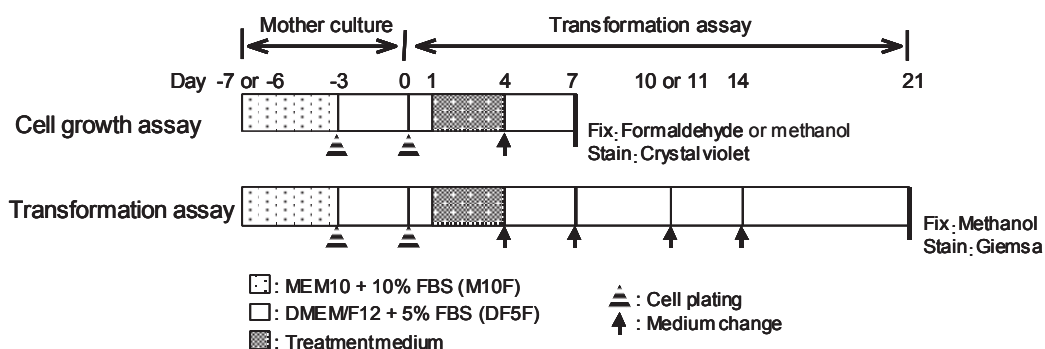
Day 1: Media containing test chemicals at two times the final concentration are prepared, and 0.05 mL of the preparations are added to the wells without medium change. The plate is tapped to mix media in the wells.

Day 4: Medium is changed with 0.1 mL of fresh DF5F.

Day 7: After fixing, the cells are stained with 0.1 mL of CV solution for 15 min or more, rinsed well with water and dried.

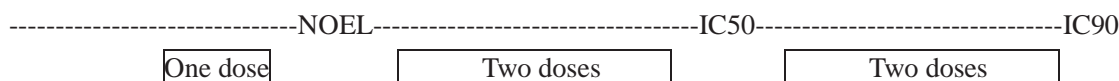
Measurement: The stained dye of each well is extracted with 0.1 mL of extraction solution, gently shaking for 10 min or more, and the optical density of each well is measured at a wavelength between 540 and 570 nm. Before measurement, the plates are shaken for several seconds (a microplate reader with mixing function is recommended). Bubbles, if any, are broken by touching with lightly heated tweezers. Growth rates relative to the solvent/vehicle control culture are calculated.

2. Transformation assay



1) Test concentrations and used vessels

Five to nine concentrations are set up based on the results of cell growth assays. These concentrations cover a range from highest toxicity (less than 20% survival compared to the control culture) to little or no toxicity. Ideally, one dose below NOEL, two doses between NOEL and IC50 and two doses between IC50 and IC90 are assessed in the initiation assay, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For low cytotoxic chemicals, the assay is performed at 0.5, 1, 2, 3, 4 and 5 mg/mL^{4,5)}.

One plate for the transformation assay (96 wells), and eight wells for the concurrent cell growth assay are prepared at each concentration.

Each assay of a chemical should include negative and positive controls (when the solvent of the test chemical is 5% water, DMSO control is also necessary as the control for MCA). The control data cannot be shared among experiments of different chemicals even if they are carried out at the same time.

2) Procedure of transformation assay and concurrent cell growth assay

Day -7 or -6: Frozen stock cells are thawed, and cultured in 10 mL of M10F in ϕ 100-mm dishes. Transformation assays must be started from the frozen stock cells but not from the cells with a high passage number.

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cell suspension is transferred at a volume of 10 mL to ϕ 100-mm dishes.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 4000 cells/mL. The cell suspension is distributed into each well of 96-well plates at a volume of 0.05 mL for the transformation assay and the concurrent cell growth assay (200 cells/well). One plate is prepared for each treatment concentration of transformation assay. Eight wells are prepared for each treatment concentration of concurrent cell growth assay.

Day 1: Media containing test chemicals at two times the final concentration are prepared, and 0.05 mL of the preparations are added to the wells without medium change. The plate is tapped to mix media in wells.

Day 4: The medium is changed with 0.1 mL of fresh DF5F.

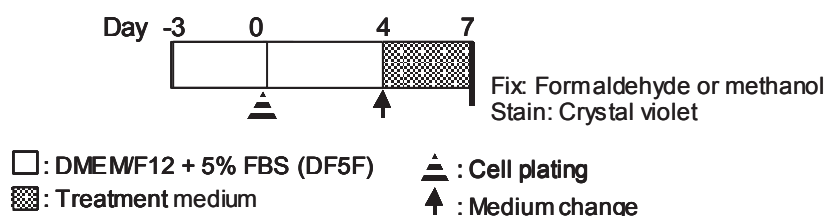
Day 7: The medium of transformation assay is changed with fresh DF5F. The cells for the concurrent growth assay are fixed and processed according to the procedures described above (II.A.1.2).

Day 10 or 11, and 14: The medium of the transformation assay is changed with fresh DF5F.

Day 21: The cells are fixed and stained with 5% Giemsa solution for 30 min or more.

B. Promotion assay^{6,7)}

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

Same as the initiation assay (II.A.1.1).

2) Procedure of cell growth assay

Day -3: The cells are subcultured in the same manner as the initiation assay (II.A.1.2).

Day 0: The cells are replated in the same manner as the initiation assay (II.A.1.2) except that 0.1 mL of the cell

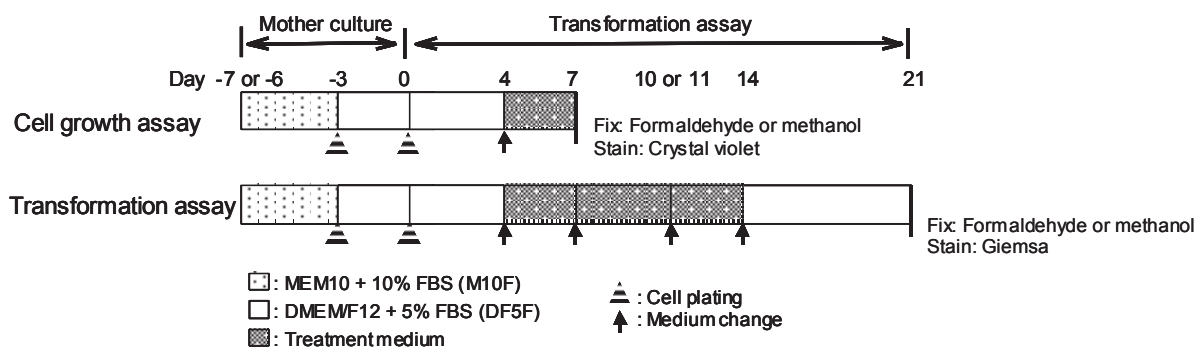
suspension are plated (400 cells/well).

Day 4: Media containing test chemicals at the final concentrations are prepared, and used for medium change (0.1 mL/well).

Day 7: The cells are fixed and processed according to the procedures described above (II.A.1.2).

Measurement: The optical density is measured in the same way as the initiation assay (II.A.1.2). Growth rates relative to the solvent/vehicle control culture are calculated.

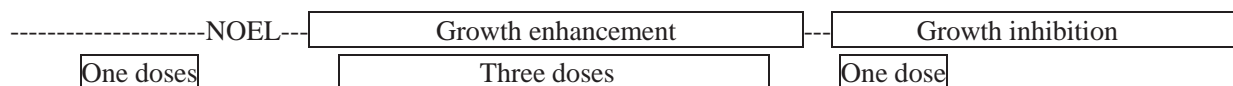
2. Transformation assay



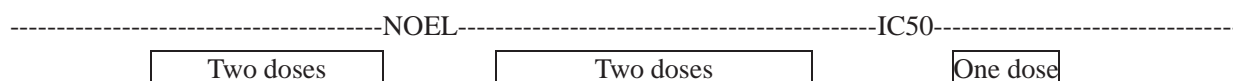
1) Test concentrations and used vessels

Five to nine concentrations are set up based on the results of cell growth assays.

For the chemicals that exhibit marked growth enhancement, test concentrations are selected to cover from growth enhancement to little effect on cell growth. Ideally, one doses below NOEL, three doses in the range of growth enhancement, and one dose in the range of weak growth inhibition are assessed in the promotion assay, as follows;



For the chemicals that do not induce marked growth enhancement, test concentrations are selected ranging from a dose exhibiting below 50% growth level to that two or three levels lower than the non-effective concentration. Ideally, two doses below NOEL, two doses between NOEL and IC50 and one dose above IC50 are assessed, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For low cytotoxic chemicals, the assay is performed at 0.5, 1, 2, 3, 4 and 5 mg/mL^{4,5}).

One plate for the transformation assay (96 wells), and eight wells for the concurrent cell growth assay are prepared at each concentration.

Each assay of a chemical should include negative and positive controls (when the solvent of the test chemical is 5% water, DMSO control is also necessary as the control of TPA). The control data cannot be shared among experiments of different chemicals even if they are carried out at the same time.

2) Procedure of transformation assay and concurrent cell growth assay

Day -7 or -6: The cells are thawed in the same manner as the initiation assay (II.A.2.2).

Day -3: The cells are subcultured in the same manner as the initiation assay (II.A.2.2).

Day 0: The cells are replated in the same manner as the initiation assay (II.A.2.2) except that 0.1 mL of the cell suspension are plated (400 cells/well).

Day 4: Media containing test chemicals at the final concentrations are prepared, and used for medium change (0.1 mL/well).

Day 7: The medium of transformation assay is changed with a media containing a test chemical. The cells for the concurrent growth assay are fixed and processed according to the procedures described above (II.A.2.2).

Day 10 or 11: The medium of transformation assay is changed with a media containing a test chemical.

Day 14: The medium is changed with 0.1 mL of fresh DF5F.

Day 21: The cells are fixed and stained in the same manner as the initiation assay (II.A.2.2).

C. Medium change

The medium is removed by inverting the plates, i.e. paper towels or diapers are laid to catch the medium and the plates are swing and shake three times to drain off the medium completely. The medium is gently dispensed to the cells from tips which are touched to the upper walls of the wells.

D. Cell fixation

The cells are fixed with direct addition of 0.01 mL of formalin to the culture medium, or with 10 % formalin or methanol after the medium is removed. After 30 min or more (when fixed with formalin) or 10 min or more (when fixed with methanol), the cells are washed with tap water and dried.

E. Focus count

Transformed foci are featured by the following morphological characteristics; (a) more than 100 cells, (b) spindle-shaped cells different from the contact-inhibited monolayer cells (spindle-shaped), (c) deep basophilic staining (basophilic), (d) random orientation of cells at the edge of foci (criss-cross), (e) dense multilayering of cells (piling up) and (f) invasive growth into the monolayer of surrounding contact-inhibited cells. There are transformed foci not prominent in some of these characteristics. The number of wells having transformed foci is recorded for each plate: a well having one focus is counted as one and a well having two or more foci is also counted as one.

Transformed foci are counted on the bottom surface but not on the wall of a well.

F. Test acceptance of criteria

The initiation or promotion assay is repeated independently, as needed, to satisfy acceptance criteria.

1. Concurrent cell growth assay

- At least should exist one concentration near NOEL which gives 80-120% cell growth compared to solvent/vehicle control.
- When contamination or technical problems are observed, four undamaged wells are necessary at the minimum for each concentration.

2. Transformation assay

1) Initiation assay and promotion assay

- If the cells are killed and/or not confluent at the end of transformation assay, the concentration is not valid and excluded from focus-counting, statistical analysis and judgment, and “toxicity” is written in the data sheet.

- When contamination or technical problems are observed, if the number of damaged wells are 7 or more the concentration is not valid and excluded from focus-counting, statistical analysis and judgment, and “contamination”, “accident”, “technical error” and so on are written in the data sheet.

2) Initiation assay

- Negative control: The number of wells having transformed foci should be 10 wells/plate or less.
- Positive control: The number of wells having transformed foci should be 30 wells/plate or more.
- Concentrations: Four valid concentrations are necessary at the minimum to accept the transformation assay for evaluating a chemical: at least one concentration near the NOEL and three concentrations in the range between the NOEL and IC90.

3) Promotion assay

- Negative control: The number of wells having transformed foci should be 20 wells/plate or less.
- Positive control: The number of wells having transformed foci should be 30 wells/plate or more. Furthermore, the proportion of wells having transformed foci should be significantly increased in the positive control plate compared to the solvent control plate (chi-square test, $p < 0.05$).
- Concentrations: Four valid concentrations are necessary at the minimum to accept the transformation assay for evaluating a chemical: at least one concentration near the NOEL and two concentrations in the range of growth enhancement when the chemical enhances cell growth (increase cell density). The durations of chemical exposure to the cells are different between the cell growth assay and the transformation assay. Then chemical toxicity is sometimes accumulated during 10 days in the promotion assay and plates may be lost because of chemical toxicity. In such cases four valid plates are also requested in the concentration range where cells are not killed and confluent at the end of transformation assay. It may be required to repeat the assay in a lower concentration range.

(The parts underlined are tentative and may be shifted with change in the batches of FBS from Moregate, Lot no. 7825120 to GIBCO, Lot no. 1391481.)

G. Three independent control data

To evaluate interlaboratory and intralaboratory variability, three independent data, at least, of positive and negative controls are needed in each laboratory. Independent data mean data obtained from the cells thawed on different days.

An example of three independent control data by plating cells on different days

Lab	Chemicals for each lab	Chemicals evaluated at the same time	Thawing frozen cells
Lab-A	a, b, c, d, e, f	a, b	1 Dec 2008
		c, d	8 Dec 2008
		e, f	15 Dec 2008
Lab-B	a, b, c, d, e, f	a	8 Dec 2008
		b, c	25 Dec 2008
		d, e, f	5 Jan 2009
Lab-C	a, b, c, d, e, f	a, b	4 Dec 2008
		c, d, e	18 Dec 2008

H. Statistical analysis

The proportion of wells with transformed foci in the plate treated with a test chemical at each concentration is compared with that in the solvent/vehicle control plate by chi-square test ($p < 0.05$). For multiplicity, the number of concentrations that satisfy “Test acceptance criteria” (**II.F**) is considered by Bonferroni method.

H. Judgment

The result is considered positive if significant increases in the proportion of wells with transformed foci are observed at two serial concentrations when analyzed by chi-square test ($p < 0.05$, multiple comparison analysis). If the increase is significant at only one concentration, the transformation assay should be repeated in a narrower concentration range and judged based on the repeated assay.

Judgment of results

Results		Judgment
----- 1st experiment	Repeated experiment	
Significant at two and more serial concentrations	-	Positive
Significant at one concentration	No significant concentration	Negative
	Significant at one or more concentrations	Positive
No significant concentration	-	Negative

The final judgment is based on a comprehensive evaluation that also takes into account the biological significance.

All results are entered in the data sheet given and sent to Arai (arai.s@fdsc.or.jp, Hadano Research Institute).

Transformation plates must be stored and archived in each lab until the termination of validation study.

III. References

- (1) K. Sasaki, H. Mizusawa and M. Ishidate, Isolation and characterization of ras-transfected BALB/3T3 clone showing morphological transformation by 12-*O*-tetradecanoyl-phorbol-13-acetate, *Jpn. J. Cancer Res.* 79 (1988) 921-930.
- (2) K. Sasaki, H. Mizusawa, M. Ishidate and N. Tanaka, Establishment of a highly reproducible transformation assay of a ras-transfected BALB/3T3 clone by treatment with promoters, *Basic Life Sci.* 52 (1990) 411-416.
- (3) S. Asada, K. Sasaki, N. Tanaka, K. Takeda, M. Hayashi and M. Umeda, Detection of initiating activities of chemicals using v-Ha-ras-transfected BALB/c 3T3 cells (Bhas 42 cells), *Mutat. Res.* 588 (2005) 7-21.
- (4) OECD Guidelines for the Testing of Chemicals. Test No. 473: *In vitro* Mammalian Chromosome Aberration

Test.

- (5) OECD Guidelines for the Testing of Chemicals. Test No. 476: *In vitro* Mammalian Cell Gene Mutation Test.
- (6) K. Ohmori, K. Sasaki, S. Asada, N. Tanaka and M. Umeda, An assay method for the prediction of tumor promoting potential of chemicals by the use of Bhas 42 cells, *Mutat. Res.* 557 (2004) 191-202.
- (7) K. Ohmori *et al.*, Inter-laboratory collaborative study of cell transformation assay for tumor promoters using Bhas 42 cells by non-genotoxic carcinogen study group in Japan, *ATRA* 33 (2005) 1-21.

Annex 1. Materials and Reagents Used and their Catalogue Numbers (and Lot Numbers)

	Supplier	Cat. No.	Lot No.
Positive control			
3-Methylcholanthrene (MCA)	Aldrich	213942-100MG	11303JE
Phorbol 12-myristate 13-acetate (TPA)	SIGMA	P-1585-1MG	038K1501
Culture reagents			
Minimum Essential Medium	GIBCO	11095-080(500mL) (Japan) 11095-098(500mLx10) 31095-029(500mL) (Europe) 31095-052(500mLx10)(Europe)	----
Fetal bovine serum	GIBCO		1391481
Dulbecco's modified Eagle's medium/F12	GIBCO	11330-032(500mL) (Japan) 11330-057(500mLx10) (Japan) 31330-038(500mL) (Europe) 31330-095(500mLx10)(Europe)	----
Penicillin(10000units/mL)-Streptomycin (10 mg/mL)	GIBCO	15140-122	----
Trypsin, 0.25% EDTA(0.02%)-PBS(-)	GIBCO ----	15050-065	
Other reagent			
DMSO	SIGMA	D8418	----
Fixation and staining solutions			
Methanol	----	----	----
Giemsa solution *	MERK	1.09204	----
Giemsa solution (0.4%)*	SIGMA	GS 500	----
Formalin (37% formaldehyde)	SIGMA	F8775	----
Crystal violet	SIGMA	C3886	----
Ethanol	----	----	----
HCl	----	----	----
Culture vessels			
96-well microplates	CORNING (COSTAR) (Do not use FALCON)	3598	----
100-mm tissue culture dish	CORNING	430167	

*: Giemsa solution purchased from Merck is used after $\times 20$ dilution. Either Giemsa solution can be used.

Annex 7

Protocol of Bhas 42 Cell Transformation Assay (2009.1.26) 96-Well Method Ver. 3

This protocol is described for the international validation study of Bhas 42 cell transformation assay using 96-well plates. Materials and reagents with their catalogue numbers are listed in annex 1.

I. Preparation

1. Materials

1) Cell line

Bhas 42 cells (v-Ha-*ras*-transfected Balb/c 3T3 clone A31-1-1 cells)^{1,2}: Free from bacteria, fungi and mycoplasma, supplied from HRI (Hatano Research Institute, Food and Drug Safety Center, Japan) Cell Bank.

2) Media

MEM: Minimum essential medium with 2.2 g/L NaHCO₃ and 0.292 g/L L-glutamine.

DMEM/F12: Dulbecco's modified Eagle's medium/F12 with 1.2 g/L NaHCO₃.

FBS: Fetal bovine serum should be selected showing a low spontaneous focus formation and a high focus formation in the positive control.

PS: Penicillin G sodium (10000 units/mL) and streptomycin sulfate (10 mg/mL).

M10F: MEM + 10% FBS + 1% PS (500 mL MEM + 56 mL FBS + 5 mL PS): Used for the expansion of provided cells, cell storage, and the first culture after thawing.

DF5F: DMEM/F12 + 5% FBS + 1% PS (500 mL DMEM/F12 + 26.5 mL FBS + 5 mL PS): Used for routine passages, cell growth assays and transformation assays.

3) Chemicals

Test chemicals and solvent/vehicle: Test chemicals are dissolved or suspended in an appropriate solvent or vehicle and diluted with the solvent/vehicle to each individual concentration before added to culture media so that all chemical treatment media contain an equal concentration of the solvent/vehicle. The final concentration of the solvent/vehicle in the medium is 5% with distilled water/ultra pure water and 0.1% with DMSO (permissible up to 0.5% when a test chemical does not dissolve).

Negative and positive controls: The solvent/vehicle for a test chemical is used as the negative control. A known initiator, 3-methylcholanthrene (MCA: final concentration of 1 µg/mL), and a promoter, 12-*O*-tetradecanoylphorbol-13- acetate (TPA: final concentration of 50 ng/mL), are used for the positive controls.

Preparation and storage of test-chemical solutions: Test-chemical solutions are prepared before use as a general rule. Working solutions may be preserved in aliquots at -20°C for less than 10 days and thawed before use, but must not be re-frozen. When the test chemical supplied is too little to be weighed at every assay, a stock solution is prepared at a concentration as high as possible or at an appropriate concentration and stored in aliquots at -20°C for a year and thawed before use. The stock solution is never re-frozen. The suspensions of test chemicals must be prepared before use and cannot be stored for the future use. The stock solutions of MCA and TPA in DMSO can be stored at -20 °C for two years at least, if they are not thawed.

4) Fixatives and staining solutions

Formalin (37% formaldehyde): Used for fixing cells.

Methanol: Used for fixing cells.

0.1% crystal violet (CV) solution: Used for staining cells in cell growth assays. First, 1 g of crystal violet is dissolved in 50 mL of ethanol, and the total volume is adjusted to 1 L with distilled water/ultra pure water.

Extraction solution (0.02 mol/L HCl and 50% ethanol): Used for extracting CV in cell growth assays. 480 mL distilled water/ultra pure water + 500 mL ethanol + 20 mL 1 M HCl.

5% Giemsa solution: Used for staining cells in transformation assays.

5) Culture vessels

φ100-mm dishes: Used for routine passage.

96-well plates: Used for cell growth and transformation assays.

6) Micropipettes

Electronic multichannel pipettes are convenient.

2. Cell culture

1) Cell culture and passage

Bhas 42 cells are cultured in a humidified 5% CO₂ incubator at 37°C. The passage should be done at about 70% confluence of cell growth.

2) Preparation of cell stock

The provided cells are expanded in M10F. The regular method of preparation of cell stock is as follows:

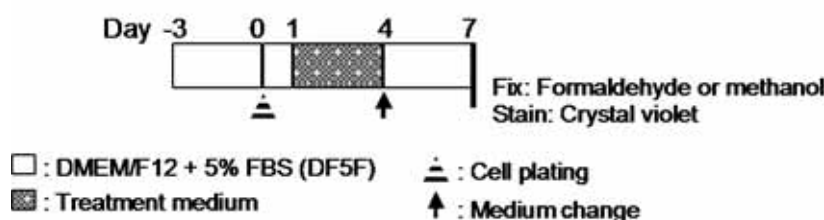
One tube (2.5 x 10⁵ cells) is thawed and suspended in 50 mL of M10F. The cell suspension is distributed into 5 of 100-mm tissue culture dishes at a volume of 10 mL/dish and cultured. Usually the cultures become to be 50-70% confluent in 4 days under these conditions. The cultures at 50-70% confluence are trypsinized with 0.25% trypsin after washed once with 0.02% EDTA-PBS(-). The cells are resuspended at a cell density of 5 x 10⁵ cells/mL in cold fresh M10F containing 5% DMSO, frozen in 0.5 mL aliquots (2.5 x 10⁵ cells/tube) at - 80°C and stored in liquid nitrogen. Every transformation experiment is started from this frozen stock.

For this validation study Bhas 42 cells are provided at passage 17 (2006.12.16, P17) from HRI. In this validation study Bhas 42 cells must be used at passage 18 for the transformation assays. But for cell growth assays the cells at higher passages can be used and should be used to save frozen cell stocks for subsequent transformation assays.

II. Experimental procedures

A. Initiation assay³⁾

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

The highest concentration is 5 mg/mL^{4,5}. In the case of chemicals difficult to dissolve, the highest concentration may be one or two level higher than the concentration showing the utmost solubility. Five or more concentration levels are set by an appropriate serial dilution factor such as 10, square root 10 or 2. In some chemicals the cell growth assay should be repeated in a narrower concentration range. The solvent/vehicle for a test chemical is used as the control. Each cell growth assay should include each solvent/vehicle control. Eight wells of 96-well plates are used for each concentration.

Blank control: Wells added with the medium alone are prepared for the blank control in cell growth assays. The blank control can be shared among different experiments conducted at the same time.

2) Procedure of cell growth assay

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1 x 10⁴ cells/mL. The cell suspension is transferred at a volume of 10 mL to φ100-mm dishes. The cells with a high passage number can be used for the cell growth assay.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 4000 cells/mL. The cell suspension is distributed into each well of 96-well plates at a volume of 0.05 mL (200 cells/well). Because the volume is very small, the cell suspension is spread on the bottom of well by tapping the plates lightly. After keeping the plates at room temperature for 15 min until the cells are attached to the bottom, they are put into a incubator. Eight wells are prepared for each treatment condition.

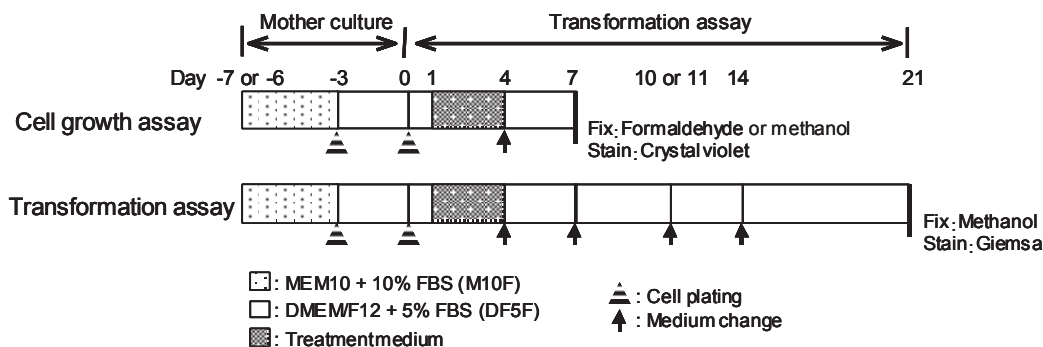
Day 1: Media containing test chemicals at two times the final concentration are prepared, and 0.05 mL of the preparations are added to the wells without medium change. The plate is tapped to mix media in the wells.

Day 4: Medium is changed with 0.1 mL of fresh DF5F.

Day 7: After fixing, the cells are stained with 0.1 mL of CV solution for 15 min or more, rinsed well with water and dried.

Measurement: The stained dye of each well is extracted with 0.1 mL of extraction solution, occasionally gently shaking for 10 min or more, and the optical density of each well is measured at a wavelength between 540 and 570 nm. Before measurement, the plates are shaken for several seconds (a microplate reader with mixing function is recommended). Bubbles, if any, are broken by touching with lightly heated tweezers. Growth rates relative to the solvent/vehicle control culture are calculated.

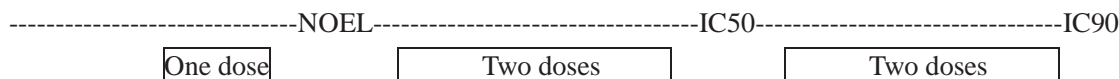
2. Transformation assay



1) Test concentrations and used vessels

Five to nine concentrations are set up based on the results of cell growth assays. These concentrations cover a range from highest toxicity (less than 20% survival compared to the control culture) to little or no toxicity. Ideally,

one dose below NOEL, two doses between NOEL and IC50 and two doses between IC50 and IC90 are assessed in the initiation assay, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For low cytotoxic chemicals, the assay is performed at 0.5, 1, 2, 3, 4 and 5 mg/mL^{4,5)}.

One plate for the transformation assay (96 wells), and eight wells for the concurrent cell growth assay are prepared at each concentration.

Each assay of a chemical should include negative and positive controls (when the solvent of the test chemical is 5% water, DMSO control is also necessary as the control for MCA). The control data cannot be shared among experiments of different chemicals even if they are carried out at the same time.

2) Procedure of transformation assay and concurrent cell growth assay

Day -6 or -7: Frozen stock cells (2.5×10^5 cells) are thawed, and cultured in 20 to 50 mL of M10F in 2 to 5 of ϕ 100-mm dishes. Transformation assays must be started from the frozen stock cells but not from the cells with a high passage number.

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cell suspension is transferred at a volume of 10 mL to ϕ 100-mm dishes.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 4000 cells/mL. The cell suspension is distributed into each well of 96-well plates at a volume of 0.05 mL for the transformation assay and the concurrent cell growth assay (200 cells/well). One plate is prepared for each treatment concentration of transformation assay. Eight wells are prepared for each treatment concentration of concurrent cell growth assay.

Day 1: Media containing test chemicals at two times the final concentration are prepared, and 0.05 mL of the preparations are added to the wells without medium change. The plate is tapped to mix media in wells.

Day 4: The medium is changed with 0.1 mL of fresh DF5F.

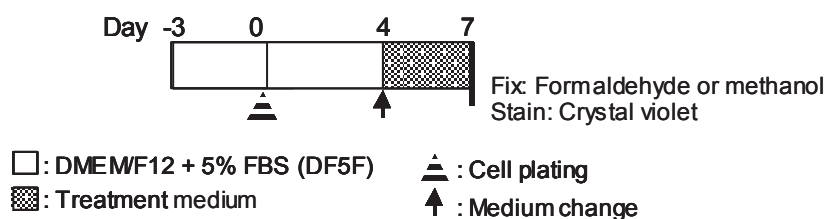
Day 7: The medium of transformation assay is changed with fresh DF5F. The cells for the concurrent growth assay are fixed and processed according to the procedures described above (II.A.1.2).

Day 10 or 11, and 14: The medium of the transformation assay is changed with fresh DF5F.

Day 21: The cells are fixed and stained with 5% Giemsa solution for 30 min or more.

B. Promotion assay^{6,7)}

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

Same as the initiation assay (II.A.1.1).

2) Procedure of cell growth assay

Day -3: The cells are subcultured in the same manner as the initiation assay (II.A.1.2).

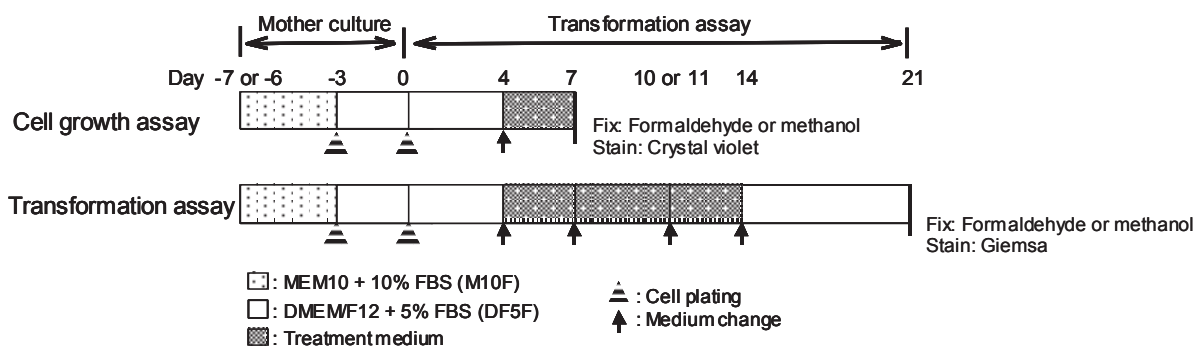
Day 0: The cells are replated in the same manner as the initiation assay (II.A.1.2) except that 0.1 mL of the cell suspension are plated (400 cells/well).

Day 4: Media containing test chemicals at the final concentrations are prepared, and used for medium change (0.1 mL/well).

Day 7: The cells are fixed and processed according to the procedures described above (II.A.1.2).

Measurement: The optical density is measured in the same way as the initiation assay (II.A.1.2). Growth rates relative to the solvent/vehicle control culture are calculated.

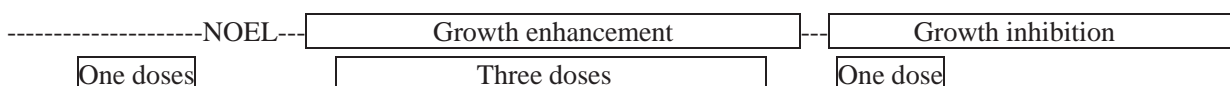
2. Transformation assay



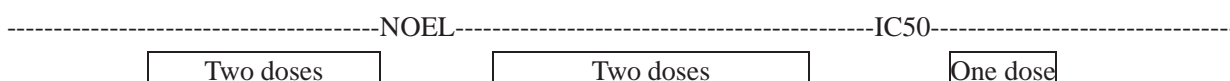
1) Test concentrations and used vessels

Five to nine concentrations are set up based on the results of cell growth assays.

For the chemicals that exhibit marked growth enhancement, test concentrations are selected to cover from growth enhancement to little effect on cell growth. Ideally, one dose below NOEL, three doses in the range of growth enhancement, and one dose in the range of weak growth inhibition are assessed in the promotion assay, as follows;



For the chemicals that do not induce marked growth enhancement, test concentrations are selected ranging from a dose exhibiting below 50% growth level to that two or three levels lower than the non-effective concentration. Ideally, two doses below NOEL, two doses between NOEL and IC50 and one dose above IC50 are assessed, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For low cytotoxic chemicals, the assay is performed at 0.5, 1, 2, 3, 4 and 5 mg/mL^{4,5)}.

One plate for the transformation assay (96 wells), and eight wells for the concurrent cell growth assay are prepared at each concentration.

Each assay of a chemical should include negative and positive controls (when the solvent of the test chemical is 5% water, DMSO control is also necessary as the control of TPA). The control data cannot be shared among experiments of different chemicals even if they are carried out at the same time.

2) Procedure of transformation assay and concurrent cell growth assay

Day -7 or -6: The cells are thawed in the same manner as the initiation assay (II.A.2.2).

Day -3: The cells are subcultured in the same manner as the initiation assay (II.A.2.2).

Day 0: The cells are replated in the same manner as the initiation assay (II.A.2.2) except that 0.1 mL of the cell suspension are plated (400 cells/well).

Day 4: Media containing test chemicals at the final concentrations are prepared, and used for medium change (0.1 mL/well).

Day 7: The medium of transformation assay is changed with a media containing a test chemical. The cells for the concurrent growth assay are fixed and processed according to the procedures described above (II.A.2.2).

Day 10 or 11: The medium of transformation assay is changed with a media containing a test chemical.

Day 14: The medium is changed with 0.1 mL of fresh DF5F.

Day 21: The cells are fixed and stained in the same manner as the initiation assay (II.A.2.2).

C. Medium change

The medium is removed by inverting the plates, i.e. paper towels or diapers are laid to catch the medium and the plates are swing and shake three times to drain off the medium completely. The medium is gently dispensed to the cells from tips which are touched to the upper walls of the wells.

D. Cell fixation

The cells are fixed with direct addition of 0.01 mL of formalin to the culture medium, or with 10 % formalin or methanol after the medium is removed. After 30 min or more (when fixed with formalin) or 10 min or more (when fixed with methanol), the cells are washed with tap water and dried.

E. Focus count

Transformed foci are featured by the following morphological characteristics; (a) more than 100 cells, (b) spindle-shaped cells different from the contact-inhibited monolayer cells (spindle-shaped), (c) deep basophilic staining (basophilic), (d) random orientation of cells at the edge of foci (criss-cross), (e) dense multilayering of cells (piling up) and (f) invasive growth into the monolayer of surrounding contact-inhibited cells. There are transformed foci not prominent in some of these characteristics. The number of wells having transformed foci is recorded for each plate: a well having one focus is counted as one and a well having two or more foci is also counted as one.

Transformed foci are counted on the bottom surface but not on the wall of a well.

F. Test acceptance of criteria

The initiation or promotion assay is repeated independently, as needed, to satisfy acceptance criteria.

1. Concurrent cell growth assay

- When contamination or technical problems are observed, four undamaged wells are necessary at the minimum for each concentration.

2. Transformation assay

1) Initiation assay and promotion assay

- If the cells are killed and/or not confluent at the end of transformation assay, the concentration is not valid and excluded from focus-counting, statistical analysis and judgment, and “toxicity” is written in the data sheet..
- When contamination or technical problems are observed, if the number of damaged wells are 7 or more the concentration is not valid and excluded from focus-counting, statistical analysis and judgment, and “contamination”, “accident”, “technical error” and so on are written in the data sheet.

2) Initiation assay

- Negative control: The number of wells having transformed foci should be 15 wells/plate or less.
- Positive control: The number of wells having transformed foci should be 40 wells/plate or more.
- Concentrations: Four valid concentrations are necessary at the minimum to accept the transformation assay for evaluating a chemical: at least one concentration near the NOEL (80 – 120 %) and three concentrations in the range between the NOEL and IC90 in the concurrent cell growth assay.

3) Promotion assay

- Negative control: The number of wells having transformed foci should be 20 wells/plate or less.
- Positive control: The number of wells having transformed foci should be 40 wells/plate or more.
- Concentrations: Four valid concentrations are necessary at the minimum to accept the transformation assay for evaluating a chemical: in the concurrent assay, at least one concentration near the NOEL and two concentrations in the range of growth enhancement, when the chemical enhances cell growth (increase cell density). The durations of chemical exposure to the cells are different between the cell growth assay and the transformation assay. Then chemical toxicity is sometimes accumulated during 10 days in the promotion assay and plates may be lost because of chemical toxicity. In such cases four valid plates are also requested in the concentration range where cells are confluent and not killed at the end of transformation assay. It may be required to repeat the assay in a lower concentration range.

(The parts underlined were tentative in the protocol for the pre-validation phase and have been shifted with change in the batches of FBS from Moregate, Lot no. 7825120 to GIBCO, Lot no. 1391481, based on the results of the pre-validation study.)

G. Three independent control data

To evaluate interlaboratory and intralaboratory variability, three independent data, at least, of positive and negative controls are needed in each laboratory. Independent data mean data obtained from the cells thawed on different days.

An example of three independent control data by plating cells on different days

Lab	Chemicals for each lab	Chemicals evaluated at the same time	Thawing frozen cells
Lab-A	a, b, c, d, e, f	a, b	1 Dec 2008
		c, d	8 Dec 2008
		e, f	15 Dec 2008
Lab-B	a, b, c, d, e, f	a	8 Dec 2008
		b, c	25 Dec 2008
		d, e, f	5 Jan 2009

Lab-C	a, b, c, d, e, f	a, b	4 Dec 2008
		c, d, e	18 Dec 2008
		f, (repeated a)	8 Jan 2009

H. Statistical analysis

The proportion of wells with transformed foci in the plate treated with a test chemical at each concentration is compared with that in the solvent/vehicle control plate by chi-square test with Bonferroni adjustment (p-value<0.025, upper-sided)⁸⁾. For multiplicity, the number of concentrations that satisfy “TEST acceptance criteria” (II.F) is considered by this methodology.

H. Judgment

The result is considered positive if significant increases in the proportion of wells with transformed foci are observed at two serial concentrations when analyzed by chi-square test (p-value<0.025, upper-sided, multiple comparison analysis). If the increase is significant at only one or non-serial concentrations, the transformation assay should be repeated in a narrower concentration range and judged based on the repeated assay.

Judgment of results

Results		Judgment
1st experiment	Repeated experiment	
Significant at two and more serial concentrations	-	Positive
Significant at one or non-serial concentrations	No significant concentration	Negative
	Significant at one or more concentrations	Positive
No significant concentration	-	Negative

The final judgment is based on a comprehensive evaluation that also takes into account the biological significance.

All results are entered in the data sheet given and sent to both Shoko Arai (arai.s@fdsc.or.jp, Hadano Research Institute) and Masaya Suzuki (masaya@anpyo.or.jp, Biosafety Research Center, Foods, Drugs and Pesticides).

Transformation plates must be stored and archived in each lab until the termination of validation study.

III. References

- (1) K. Sasaki, H. Mizusawa and M. Ishidate, Isolation and characterization of ras-transfected BALB/3T3 clone showing morphological transformation by 12-O-tetradecanoyl-phorbol-13-acetate, Jpn. J. Cancer Res. 79 (1988) 921-930.
- (2) K. Sasaki, H. Mizusawa, M. Ishidate and N. Tanaka, Establishment of a highly reproducible transformation

assay of a ras-transfected BALB/3T3 clone by treatment with promoters, Basic Life Sci. 52 (1990) 411-416.

- (3) S. Asada, K. Sasaki, N. Tanaka, K. Takeda, M. Hayashi and M. Umeda, Detection of initiating activities of chemicals using v-Ha-ras-transfected BALB/c 3T3 cells (Bhas 42 cells), Mutat. Res. 588 (2005) 7-21.
- (4) OECD Guidelines for the Testing of Chemicals. Test No. 473: *In vitro* Mammalian Chromosome Aberration Test.
- (5) OECD Guidelines for the Testing of Chemicals. Test No. 476: *In vitro* Mammalian Cell Gene Mutation Test.
- (6) K. Ohmori, K. Sasaki, S. Asada, N. Tanaka and M. Umeda, An assay method for the prediction of tumor promoting potential of chemicals by the use of Bhas 42 cells, Mutat. Res. 557 (2004) 191-202.
- (7) K. Ohmori *et al.*, Inter-laboratory collaborative study of cell transformation assay for tumor promoters using Bhas 42 cells by non-genotoxic carcinogen study group in Japan, ATLA 33 (2005) 1-21.
- (8) G. W. Snedecor and W. G. Cochran, Statistical Methods (8th Edition), Iowa State University Press, Ames, 1989, pp.125-128.

Annex 1. Materials and Reagents Used and their Catalogue Numbers (and Lot Numbers)

	Supplier	Cat. No.	Lot No.
Positive control			
3-Methylcholanthrene (MCA)	Aldrich	213942-100MG	11303JE
Phorbol 12-myristate 13-acetate (TPA)	SIGMA	P-1585-1MG	038K1501
Culture reagents			
Minimum Essential Medium	GIBCO	11095-080(500mL) (Japan) 11095-098(500mLx10) 31095-029(500mL) (Europe) 31095-052(500mLx10)(Europe)	----
Fetal bovine serum	GIBCO		1391481
Dulbecco's modified Eagle's medium/F12	GIBCO	11330-032(500mL) (Japan) 11330-057(500mLx10) (Japan) 31330-038(500mL) (Europe) 31330-095(500mLx10)(Europe)	----
Penicillin(10000units/mL)-Streptomycin (10 mg/mL)	GIBCO	15140-122	----
Trypsin, 0.25% EDTA(0.02%)-PBS(-)	GIBCO ----	15050-065	
Other reagent			
DMSO	SIGMA	D8418	----
Fixation and staining solutions			
Methanol	----	----	----
Giemsa solution *	MERK	1.09204	----
Giemsa solution (0.4%)*	SIGMA	GS 500	----
Formalin (37% formaldehyde)	SIGMA	F8775	----
Crystal violet	SIGMA	C3886	----
Ethanol	----	----	----
HCl	----	----	----
Culture vessels			
96-well microplates	CORNING (COSTAR) (Do not use FALCON)	3598	----
100-mm tissue culture dish	CORNING	430167	

*: Giemsa solution purchased from Merck is used after $\times 20$ dilution. Either Giemsa solution can be used.

Annex 8

Protocol of Bhas 42 Cell Transformation Assay (2009.10.28) 96-Well Method Ver. 4

This protocol is described for the international validation study of Bhas 42 cell transformation assay using 96-well plates. Materials and reagents with their catalogue numbers are listed in annex 1.

I. Preparation

1. Materials

1) Cell line

Bhas 42 cells (v-Ha-*ras*-transfected Balb/c 3T3 clone A31-1-1 cells)^{1,2}: Free from bacteria, fungi and mycoplasma, supplied from HRI (Hatano Research Institute, Food and Drug Safety Center, Japan) Cell Bank.

2) Media

MEM: Minimum essential medium with 2.2 g/L NaHCO₃ and 0.292 g/L L-glutamine.

DMEM/F12: Dulbecco's modified Eagle's medium/F12 with 1.2 g/L NaHCO₃.

FBS: Fetal bovine serum should be selected showing a low spontaneous focus formation and a high focus formation in the positive control.

PS: Penicillin G sodium (10000 units/mL) and streptomycin sulfate (10 mg/mL).

M10F: MEM + 10% FBS + 1% PS (500 mL MEM + 56 mL FBS + 5 mL PS): Used for the expansion of provided cells, cell storage, and the first culture after thawing.

DF5F: DMEM/F12 + 5% FBS + 1% PS (500 mL DMEM/F12 + 26.5 mL FBS + 5 mL PS): Used for routine passages, cell growth assays and transformation assays.

3) Chemicals

Test chemicals and solvent/vehicle: Test chemicals are dissolved or suspended in an appropriate solvent or vehicle and diluted with the solvent/vehicle to each individual concentration before added to culture media so that all chemical treatment media contain an equal concentration of the solvent/vehicle. The final concentration of the solvent/vehicle in the medium is 5% with distilled water/ultra pure water and 0.1% with DMSO (permissible up to 0.5% when a test chemical does not dissolve).

Negative and positive controls: The solvent/vehicle for a test chemical is used as the negative control. A known initiator, 3-methylcholanthrene (MCA: final concentration of 1 µg/mL), and a promoter, 12-*O*-tetradecanoylphorbol-13- acetate (TPA: final concentration of 50 ng/mL), are used for the positive controls.

Preparation and storage of test-chemical solutions: Test-chemical solutions are prepared before use as a general rule. Working solutions may be preserved in aliquots at -20°C for less than 10 days and thawed before use, but must not be re-frozen. When the test chemical supplied is too little to be weighed at every assay, a stock solution is prepared at a concentration as high as possible or at an appropriate concentration and stored in aliquots at -20°C for a year and thawed before use. The stock solution is never re-frozen. The suspensions of test chemicals must be prepared before use and cannot be stored for the future use. The stock solutions of MCA and TPA in DMSO can be stored at -20 °C for two years at least, if they are not thawed.

4) Fixatives and staining solutions

Formalin (37% formaldehyde): Used for fixing cells.

Methanol: Used for fixing cells.

0.1% crystal violet (CV) solution: Used for staining cells in cell growth assays. First, 1 g of crystal violet is dissolved in 50 mL of ethanol, and the total volume is adjusted to 1 L with distilled water/ultra pure water.

Extraction solution (0.02 mol/L HCl and 50% ethanol): Used for extracting CV in cell growth assays. 480 mL distilled water/ultra pure water + 500 mL ethanol + 20 mL 1 M HCl.

5% Giemsa solution: Used for staining cells in transformation assays.

5) Culture vessels

φ100-mm dishes: Used for routine passage.

96-well plates: Used for cell growth and transformation assays.

6) Micropipettes

Electronic multichannel pipettes are convenient.

2. Cell culture

1) Cell culture and passage

Bhas 42 cells are cultured in a humidified 5% CO₂ incubator at 37°C. The passage should be done at about 70% confluence of cell growth.

2) Preparation of cell stock

The provided cells are expanded in M10F. The regular method of preparation of cell stock is as follows:

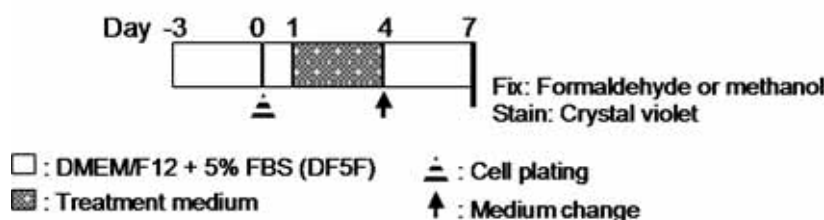
One tube (2.5 x 10⁵ cells) is thawed and suspended in 50 mL of M10F. The cell suspension is distributed into 5 of 100-mm tissue culture dishes at a volume of 10 mL/dish and cultured. Usually the cultures become to be 50-70% confluent in 4 days under these conditions. The cultures at 50-70% confluence are trypsinized with 0.25% trypsin after washed once with 0.02% EDTA-PBS(-). The cells are resuspended at a cell density of 5 x 10⁵ cells/mL in cold fresh M10F containing 5% DMSO, frozen in 0.5 mL aliquots (2.5 x 10⁵ cells/tube) at - 80°C and stored in liquid nitrogen. Every transformation experiment is started from this frozen stock.

For this validation study Bhas 42 cells are provided at passage 17 (2006.12.16, P17) from HRI. In this validation study Bhas 42 cells must be used at passage 18 for the transformation assays. But for cell growth assays the cells at higher passages can be used and should be used to save frozen cell stocks for subsequent transformation assays.

II. Experimental procedures

A. Initiation assay³⁾

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

The highest concentration is 5 mg/mL in medium^{4,5}). In the case of chemicals difficult to dissolve in a solvent/vehicle, the highest concentration may be one or two level higher than the concentration showing the utmost solubility. Five or more concentration levels are set by an appropriate serial dilution factor such as 10, square root 10 or 2. In some chemicals the cell growth assay should be repeated in a narrower concentration range. The solvent/vehicle for a test chemical is used as the control. Each cell growth assay should include each solvent/vehicle control. Eight wells of 96-well plates are used for each concentration.

Blank control: Wells added with the medium alone are prepared for the blank control in cell growth assays. The blank control can be shared among different experiments conducted at the same time.

2) Procedure of cell growth assay

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cell suspension is transferred at a volume of 10 mL to $\phi 100$ -mm dishes. The cells with a high passage number can be used for the cell growth assay.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 4000 cells/mL. The cell suspension is distributed into each well of 96-well plates at a volume of 0.05 mL (200 cells/well). Because the volume is very small, the cell suspension is spread on the bottom of well by tapping the plates lightly. After keeping the plates at room temperature for 15 min until the cells are attached to the bottom, they are put into a incubator. Eight wells are prepared for each treatment condition.

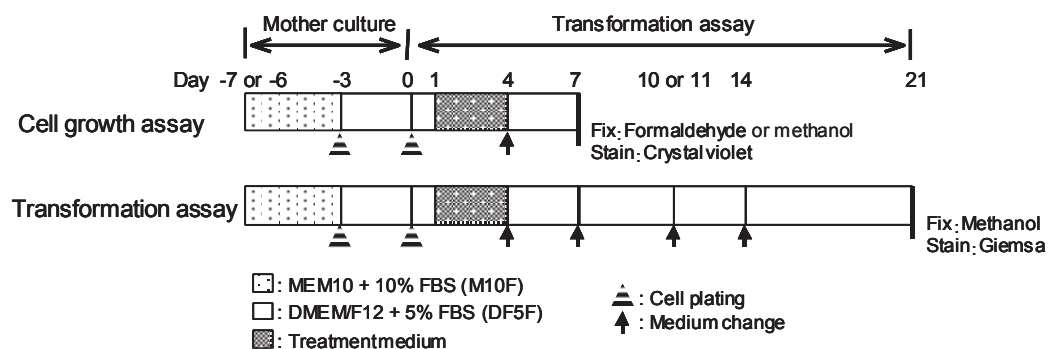
Day 1: Media containing test chemicals at two times the final concentration are prepared, and 0.05 mL of the preparations are added to the wells without medium change. The plate is tapped to mix media in the wells.

Day 4: Medium is changed with 0.1 mL of fresh DF5F.

Day 7: After fixing, the cells are stained with 0.1 mL of CV solution for 15 min or more, rinsed well with water and dried.

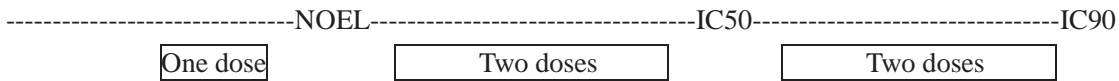
Measurement: The stained dye of each well is extracted with 0.1 mL of extraction solution, occasionally gently shaking for 10 min or more, and the optical density of each well is measured at a wavelength between 540 and 570 nm. Before measurement, the plates are shaken for several seconds (a microplate reader with mixing function is recommended). Bubbles, if any, are broken by touching with lightly heated tweezers. Growth rates relative to the solvent/vehicle control culture are calculated.

2. Transformation assay



1) Test concentrations and used vessels

Five to nine concentrations are set up based on the results of cell growth assays. These concentrations cover a range from highest toxicity (less than 20% survival compared to the control culture) to little or no toxicity. Ideally, one dose below NOEL, two doses between NOEL and IC50 and two doses between IC50 and IC90 are assessed in the initiation assay, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For low cytotoxic chemicals, the assay is performed at 0.5, 1, 2, 3, 4 and 5 mg/mL^{4,5)}.

One plate for the transformation assay (96 wells), and eight wells for the concurrent cell growth assay are prepared at each concentration.

Each assay of a chemical should include negative and positive controls (when the solvent of the test chemical is 5% water, DMSO control is also necessary as the control for MCA). The control data cannot be shared among experiments of different chemicals even if they are carried out at the same time.

2) Procedure of transformation assay and concurrent cell growth assay

Day -6 or -7: Frozen stock cells (2.5×10^5 cells) are thawed, and cultured in 20 to 50 mL of M10F in 2 to 5 of ϕ 100-mm dishes. Transformation assays must be started from the frozen stock cells but not from the cells with a high passage number.

Day -3: The cells at about 70% confluence are trypsinized and suspended in DF5F at 0.7 to 1×10^4 cells/mL. The cell suspension is transferred at a volume of 10 mL to ϕ 100-mm dishes.

Day 0: The cells at about 70% confluence are trypsinized and suspended in DF5F at 4000 cells/mL. The cell suspension is distributed into each well of 96-well plates at a volume of 0.05 mL for the transformation assay and the concurrent cell growth assay (200 cells/well). One plate is prepared for each treatment concentration of transformation assay. Eight wells are prepared for each treatment concentration of concurrent cell growth assay.

Day 1: Media containing test chemicals at two times the final concentration are prepared, and 0.05 mL of the preparations are added to the wells without medium change. The plate is tapped to mix media in wells.

Day 4: The medium is changed with 0.1 mL of fresh DF5F.

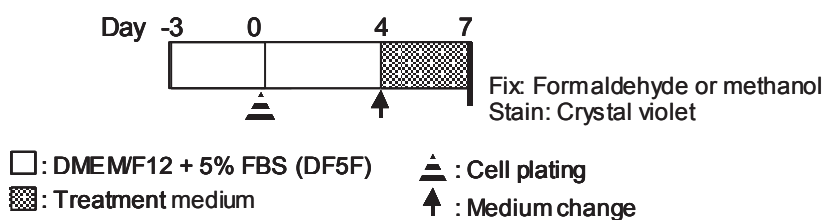
Day 7: The medium of transformation assay is changed with fresh DF5F. The cells for the concurrent growth assay are fixed and processed according to the procedures described above (II.A.1.2).

Day 10 or 11, and 14: The medium of the transformation assay is changed with fresh DF5F.

Day 21: The cells are fixed and stained with 5% Giemsa solution for 30 min or more.

B. Promotion assay^{6,7)}

1. Cell growth assay for determination of test concentrations (crystal violet method)



1) Test concentrations and used vessels

Same as the initiation assay (II.A.1.1).

2) Procedure of cell growth assay

Day -3: The cells are subcultured in the same manner as the initiation assay (II.A.1.2).

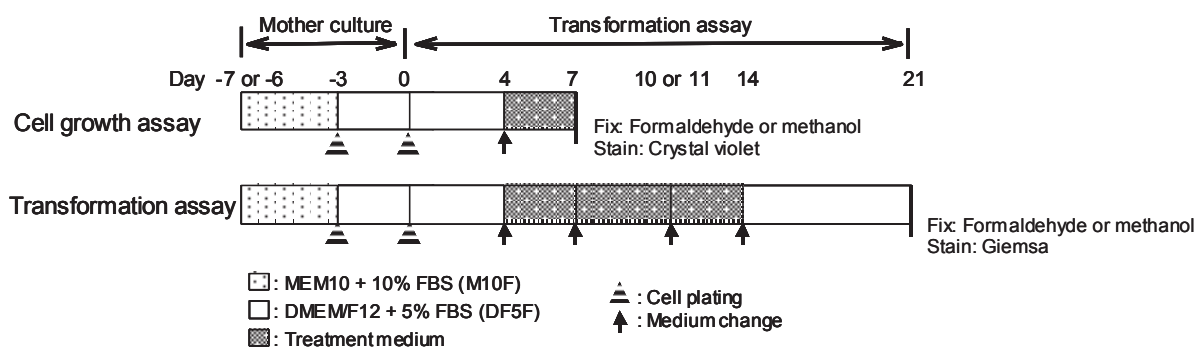
Day 0: The cells are replated in the same manner as the initiation assay (II.A.1.2) except that 0.1 mL of the cell suspension are plated (400 cells/well).

Day 4: Media containing test chemicals at the final concentrations are prepared, and used for medium change (0.1 mL/well).

Day 7: The cells are fixed and processed according to the procedures described above (II.A.1.2).

Measurement: The optical density is measured in the same way as the initiation assay (II.A.1.2). Growth rates relative to the solvent/vehicle control culture are calculated.

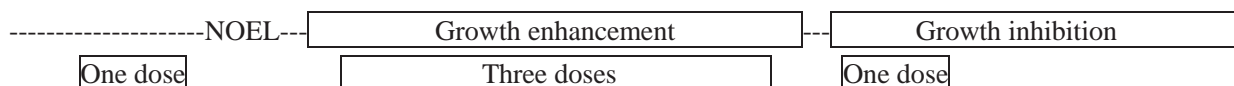
2. Transformation assay



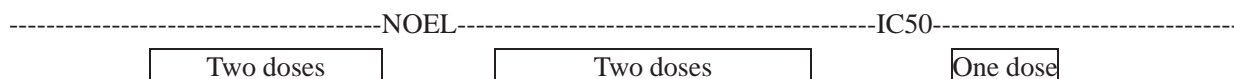
1) Test concentrations and used vessels

Five to nine concentrations are set up based on the results of cell growth assays.

For the chemicals that exhibit marked growth enhancement, test concentrations are selected to cover from growth enhancement to little effect on cell growth. Ideally, one doses bellow NOEL, three doses in the range of growth enhancement, and one dose in the range of weak growth inhibition are assessed in the promotion assay, as follows;



For the chemicals that do not induce marked growth enhancement, test concentrations are selected ranging from a dose exhibiting below 50% growth level to that two or three levels lower than the non-effective concentration. Ideally, two doses below NOEL, two doses between NOEL and IC50 and one dose above IC50 are assessed, as follows;



For a chemical which gives the sharp decline of cell growth within a narrow concentration range, one or two more doses above or below the predicted concentration range may be set up as a precaution against the fluctuation of cell response among experiments.

For low cytotoxic chemicals, the assay is performed at 0.5, 1, 2, 3, 4 and 5 mg/mL^{4,5)}.

One plate for the transformation assay (96 wells), and eight wells for the concurrent cell growth assay are prepared at each concentration.

Each assay of a chemical should include negative and positive controls (when the solvent of the test chemical is 5% water, DMSO control is also necessary as the control of TPA). The control data cannot be shared among experiments of different chemicals even if they are carried out at the same time.

2) Procedure of transformation assay and concurrent cell growth assay

Day -7 or -6: The cells are thawed in the same manner as the initiation assay (II.A.2.2).

Day -3: The cells are subcultured in the same manner as the initiation assay (II.A.2.2).

Day 0: The cells are replated in the same manner as the initiation assay (II.A.2.2) except that 0.1 mL of the cell suspension are plated (400 cells/well).

Day 4: Media containing test chemicals at the final concentrations are prepared, and used for medium change (0.1 mL/well).

Day 7: The medium of transformation assay is changed with a media containing a test chemical. The cells for the concurrent growth assay are fixed and processed according to the procedures described above (II.A.2.2).

Day 10 or 11: The medium of transformation assay is changed with a media containing a test chemical.

Day 14: The medium is changed with 0.1 mL of fresh DF5F.

Day 21: The cells are fixed and stained in the same manner as the initiation assay (II.A.2.2).

C. Medium change

The medium is removed by inverting the plates, i.e. paper towels or diapers are laid to catch the medium and the plates are swing and shake three times to drain off the medium completely. The medium is gently dispensed to the cells from tips which are touched to the upper walls of the wells.

D. Cell fixation

The cells are fixed with direct addition of 0.01 mL of formalin to the culture medium, or with 10 % formalin or methanol after the medium is removed. After 30 min or more (when fixed with formalin) or 10 min or more (when fixed with methanol), the cells are washed with tap water and dried.

E. Focus count

Transformed foci are featured by the following morphological characteristics; (a) more than 100 cells, (b) spindle-shaped cells different from the contact-inhibited monolayer cells (spindle-shaped), (c) deep basophilic staining (basophilic), (d) random orientation of cells at the edge of foci (criss-cross), (e) dense multilayering of cells (piling up) and (f) invasive growth into the monolayer of surrounding contact-inhibited cells. There are transformed foci not prominent in some of these characteristics. The number of wells having transformed foci is recorded for each plate: a well having one focus is counted as one and a well having two or more foci is also counted as one.

Transformed foci are counted on the bottom surface but not on the wall of a well.

F. Test acceptance of criteria

The initiation or promotion assay is repeated independently, as needed, to satisfy acceptance criteria.

1. Concurrent cell growth assay

- When contamination or technical problems are observed, four undamaged wells are necessary at the minimum for each concentration.

2. Transformation assay

1) Initiation assay and promotion assay

- If the cells are killed and/or not confluent at the end of transformation assay, the concentration is not valid and excluded from focus-counting, statistical analysis and judgment, and “toxicity” is written in the data sheet..
- When contamination or technical problems are observed, if the number of damaged wells are 7 or more the concentration is not valid and excluded from focus-counting, statistical analysis and judgment, and “contamination”, “accident”, “technical error” and so on are written in the data sheet.

2) Initiation assay

- Negative control: The number of wells having transformed foci should be 15 wells/plate or less.
- Positive control: The number of wells having transformed foci should be 40 wells/plate or more.
- Concentrations: Four valid concentrations are necessary at the minimum to accept the transformation assay for evaluating a chemical: at least one concentration near the NOEL (80 – 120 %) and three concentrations in the range between the NOEL and IC90 in the concurrent cell growth assay.

3) Promotion assay

- Negative control: The number of wells having transformed foci should be 20 wells/plate or less.
- Positive control: The number of wells having transformed foci should be 40 wells/plate or more.
- Concentrations: Four valid concentrations are necessary at the minimum to accept the transformation assay for evaluating a chemical: in the concurrent assay, at least one concentration near the NOEL and two concentrations in the range of growth enhancement, when the chemical enhances cell growth (increase cell density). The durations of chemical exposure to the cells are different between the cell growth assay and the transformation assay. Then chemical toxicity is sometimes accumulated during 10 days in the promotion assay and plates may be lost because of chemical toxicity. In such cases four valid plates are also requested in the concentration range where cells are confluent and not killed at the end of transformation assay. It may be required to repeat the assay in a lower concentration range.

(The parts underlined were tentative in the protocol for the pre-validation phase and have been shifted with change in the batches of FBS from Moregate, Lot no. 7825120 to GIBCO, Lot no. 1391481, based on the results of the pre-validation study.)

G. Three independent control data

To evaluate interlaboratory and intralaboratory variability, three independent data, at least, of positive and negative controls are needed in each laboratory. Independent data mean data obtained from the cells thawed on different days.

An example of three independent control data by plating cells on different days

Lab	Chemicals for each lab	Chemicals evaluated at the same time	Thawing frozen cells
Lab-A	a, b, c, d, e, f	a, b	1 Dec 2009
		c, d	8 Dec 2009
		e, f	15 Dec 2009
Lab-B	a, b, c, d, e, f	a	8 Dec 2009
		b, c	25 Dec 2009
		d, e, f	5 Jan 2010

Lab-C	a, b, c, d, e, f	a, b	4 Dec 2009
		c, d, e	18 Dec 2009
		f, (repeated a)	8 Jan 2010

H. Statistical analysis

The proportion of wells with transformed foci in the plate treated with a test chemical at each concentration is compared with that in the solvent/vehicle control plate by chi-square test with Bonferroni adjustment (p-value<0.05, upper-sided)⁸⁾. For multiplicity, the number of concentrations that satisfy “TEST acceptance criteria” (II.F) is considered by this methodology.

I. Judgment

The result is considered positive if significant increases in the proportion of wells with transformed foci are observed at two serial concentrations when analyzed by chi-square test (p-value<0.05, upper-sided, multiple comparison analysis). If the increase is significant at only one or non-serial concentrations, the result should be judged as “equivocal” when the assay is not repeated, and when the assay is repeated in a narrower concentration range the judgment should be based on the repeated assay as follows.

The judgment of results

	Results		Judgment
	1st experiment	Repeated experiment	
Significant at two and more serial concentrations	-		Positive
Significant at one or non-serial concentrations	-	No significant concentration	equivocal
		Significant at one or more concentrations	Negative
No significant concentration	-		Positive
			Negative

The final judgment is based on a comprehensive evaluation that also takes into account the biological significance.

All results are entered in the data sheet given and sent to both Shoko Arai (arai.s@fdsc.or.jp, Hadano Research Institute) and Masaya Suzuki (masaya@anpyo.or.jp, Biosafety Research Center, Foods, Drugs and Pesticides).

Transformation plates must be stored and archived in each lab until the termination of validation study.

III. References

- (1) K. Sasaki, H. Mizusawa and M. Ishidate, Isolation and characterization of ras-transfected BALB/3T3 clone showing morphological transformation by 12-*O*-tetradecanoyl-phorbol-13-acetate, Jpn. J. Cancer Res. 79 (1988) 921-930.
- (2) K. Sasaki, H. Mizusawa, M. Ishidate and N. Tanaka, Establishment of a highly reproducible transformation assay of a ras-transfected BALB/3T3 clone by treatment with promoters, Basic Life Sci. 52 (1990) 411-416.

- (3) S. Asada, K. Sasaki, N. Tanaka, K. Takeda, M. Hayashi and M. Umeda, Detection of initiating activities of chemicals using v-Ha-ras-transfected BALB/c 3T3 cells (Bhas 42 cells), *Mutat. Res.* 588 (2005) 7-21.
- (4) OECD Guidelines for the Testing of Chemicals. Test No. 473: *In vitro* Mammalian Chromosome Aberration Test.
- (5) OECD Guidelines for the Testing of Chemicals. Test No. 476: *In vitro* Mammalian Cell Gene Mutation Test.
- (6) K. Ohmori, K. Sasaki, S. Asada, N. Tanaka and M. Umeda, An assay method for the prediction of tumor promoting potential of chemicals by the use of Bhas 42 cells, *Mutat. Res.* 557 (2004) 191-202.
- (7) K. Ohmori *et al.*, Inter-laboratory collaborative study of cell transformation assay for tumor promoters using Bhas 42 cells by non-genotoxic carcinogen study group in Japan, *ATLA* 33 (2005) 1-21.
- (8) G. W. Snedecor and W. G. Cochran, *Statistical Methods* (8th Edition), Iowa State University Press, Ames, 1989, pp.125-128.

Annex 1. Materials and Reagents Used and their Catalogue Numbers (and Lot Numbers)

	Supplier	Cat. No.	Lot No.
Positive control			
3-Methylcholanthrene (MCA)	Aldrich	213942-100MG	11303JE
Phorbol 12-myristate 13-acetate (TPA)	SIGMA	P-1585-1MG	038K1501
Culture reagents			
Minimum Essential Medium	GIBCO	11095-080(500mL) (Japan) 11095-098(500mLx10) 31095-029(500mL) (Europe) 31095-052(500mLx10)(Europe)	----
Fetal bovine serum	GIBCO		1391481
Dulbecco's modified Eagle's medium/F12	GIBCO	11330-032(500mL) (Japan) 11330-057(500mLx10) (Japan) 31330-038(500mL) (Europe) 31330-095(500mLx10)(Europe)	----
Penicillin(10000units/mL)-Streptomycin (10 mg/mL)	GIBCO	15140-122	----
Trypsin, 0.25% EDTA(0.02%)-PBS(-)	GIBCO ----	15050-065	
Other reagent			
DMSO	SIGMA	D8418	----
Fixation and staining solutions			
Methanol	----	----	----
Giemsa solution *	MERK	1.09204	----
Giemsa solution (0.4%)*	SIGMA	GS 500	----
Formalin (37% formaldehyde)	SIGMA	F8775	----
Crystal violet	SIGMA	C3886	----
Ethanol	----	----	----
HCl	----	----	----
Culture vessels			
96-well microplates	CORNING (COSTAR) (Do not use FALCON)	3598	----
100-mm tissue culture dish	CORNING	430167	

*: Giemsa solution purchased from Merck is used after $\times 20$ dilution. Either Giemsa solution can be used.

Annex 9

**Results Submitted from Laboratories
in
the Pre-validation Phase
of
Validation Study
of
96-Well Method**

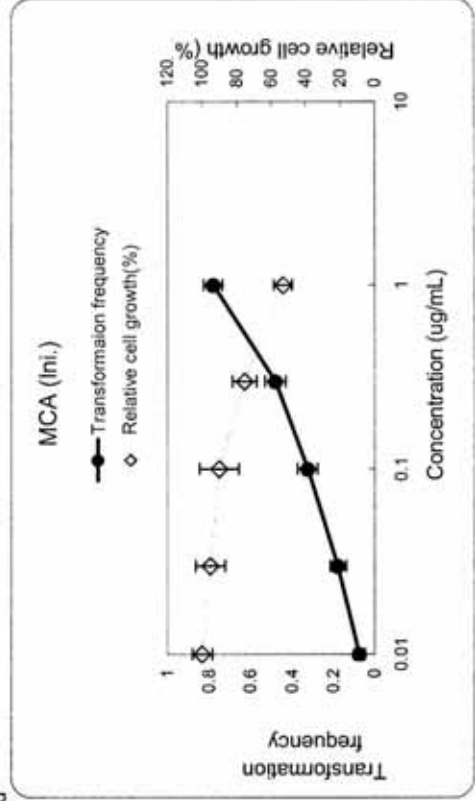
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	MCA
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Solvent		Concentrations (ug/ml)								
			0.01	0.03	0.1	0.3	1	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!		
1	0.239	1.566	1.555	1.526	1.366	0.921							
2	0.211	1.463	1.666	1.630	1.298	1.081							
3	0.217	1.591	1.481	1.608	1.278	0.883							
4	0.218	1.496	1.518	1.350	1.179	0.877							
5	0.191	1.475	1.538	1.320	1.081	0.919							
6	0.199	1.532	1.307	1.374	1.207	0.868							
7	0.198	1.682	1.352	1.365	1.261	0.962							
8	0.199	1.626	1.499	1.187	1.116	0.883							
Average	0.209	1.554	1.490	1.420	1.223	0.924	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!
SD	0.016	0.077	0.114	0.154	0.096	0.071	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!
Average - Blank	0.000	1.345	1.281	1.211	1.014	0.715	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!
Relative cell growth (%)		100.0	95.2	90.0	75.4	53.2	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!
SD of relative cell growth		5.7	8.5	11.4	7.1	5.2	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!

Transformation Assay	Solvent		Concentrations (ug/ml)											
	0.01	0.03	0.1	0.3	1	5	96	96	96	96	96	0	0	0
Giemsa Staining	0	96	96	96	96	91	96	96	96	96	96	0	0	0
No. of wells excluded	0	0	0	0	0	5								
No. of wells counted	96	96	96	96	96	91	96	96	96	96	96	0	0	0
No. of wells having foci	7	17	31	46	71									
Transformation frequency*	0.07291667	0.17708333	0.32291667	0.47916667	0.78021978									
SE	0.02653610	0.03896110	0.04772332	0.05098672	0.04340923	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



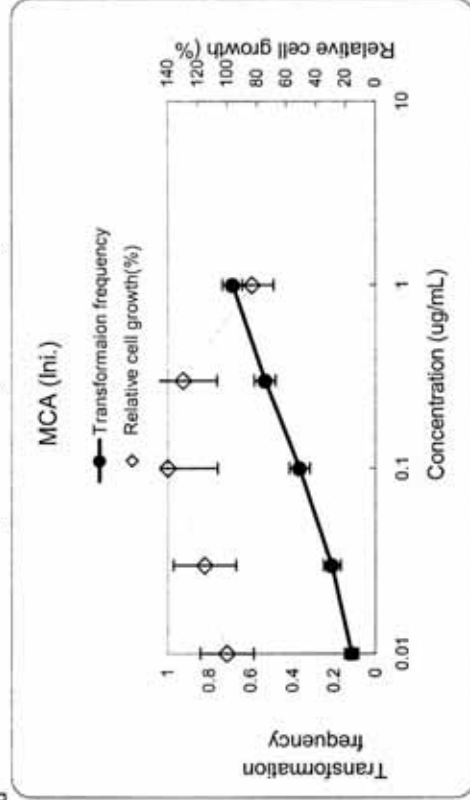
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	MCA
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining	Blank	Solvent	0.03	0.1	0.3	1	
OD540nm		0.01	0.491	0.449	0.692	0.376	
1	0.072	0.376	0.491	0.449	0.692	0.376	
2	0.080	0.424	0.458	0.480	0.677	0.462	
3	0.077	0.427	0.460	0.569	0.603	0.531	
4	0.082	0.494	0.592	0.677	0.444	0.444	
5	0.077	0.540	0.537	0.779	0.423	0.386	
6	0.081	0.603	0.689	0.831	0.665	0.377	
7	0.076	0.531	0.511	0.689	0.557	0.395	
8	0.070	0.489	0.643	0.716	0.558	0.361	
Average	0.077	0.486	0.548	0.649	0.607	0.417	#DIV/0!
SD	0.004	0.074	0.086	0.137	0.092	0.058	#DIV/0!
Average - Blank	0.000	0.409	0.471	0.572	0.530	0.340	#DIV/0!
Relative cell growth (%)		100.0	115.2	140.0	129.6	83.1	#DIV/0!
SD of relative cell growth		18.1	21.0	33.6	22.5	14.2	#DIV/0!

Transformation Assay		Concentrations (ug/ml)					
Giemsa Staining	Solvent	0.03	0.1	0.3	1	0	0
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	11	20	35	51	66	0	0
Transformation frequency*	0.11458333	0.20833333	0.36458333	0.53125	0.6875	0.00000000	0.00000000
SE	0.03250865	0.04144908	0.04912382	0.05093127	0.04730704	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



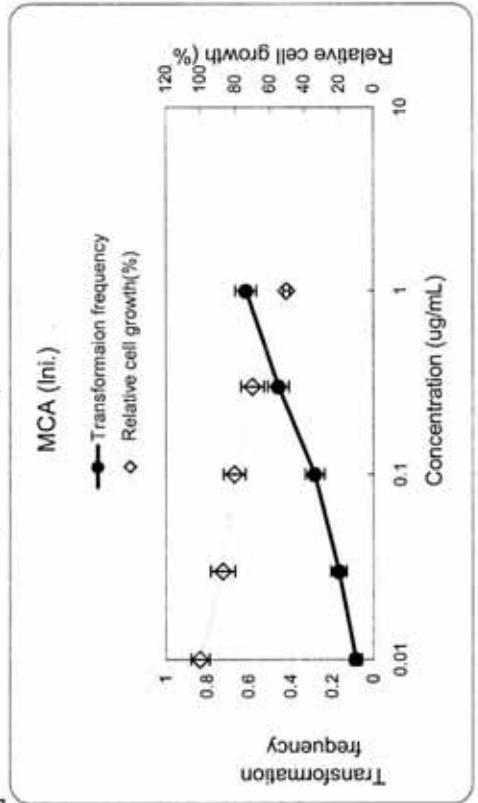
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	MCA
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining	Blank	0.1% DMSO	0.03	0.1	0.3	1	
OD540nm							
1	0.154	1.198	1.015	0.963	0.938	0.705	#DIV/0!
2	0.176	1.177	1.158	1.078	0.943	0.735	#DIV/0!
3	0.166	1.321	1.156	0.985	0.864	0.729	#DIV/0!
4	0.199	1.216	1.040	1.085	0.884	0.705	#DIV/0!
5	0.221	1.323	1.134	1.045	0.902	0.742	#DIV/0!
6	0.190	1.240	1.148	0.969	0.968	0.747	#DIV/0!
7	0.195	1.311	1.047	1.149	0.891	0.715	#DIV/0!
8	0.250	1.246	1.231	1.072	1.084	0.752	#DIV/0!
Average	0.194	1.254	1.116	1.043	0.934	0.729	#DIV/0!
SD	0.031	0.058	0.074	0.066	0.070	0.019	#DIV/0!
Average - Blank	0.000	1.060	0.922	0.849	0.740	0.535	#DIV/0!
Relative cell growth (%)	100.0	87.0	80.1	69.8	50.5	1.7	#DIV/0!
SD of relative cell growth	5.4	7.0	6.2	6.6	6.6	1.7	#DIV/0!

Transformation Assay		Concentrations (ug/ml)					
Giemsa Staining	Blank	0.1% DMSO	0.03	0.1	0.3	1	
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	8	16	27	44	59	59	96
Transformation frequency*	0.08333333	0.16666667	0.28125	0.45833333	0.61458333	0.61458333	0
SE	0.02820847	0.03803629	0.04588805	0.05085354	0.04967296	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



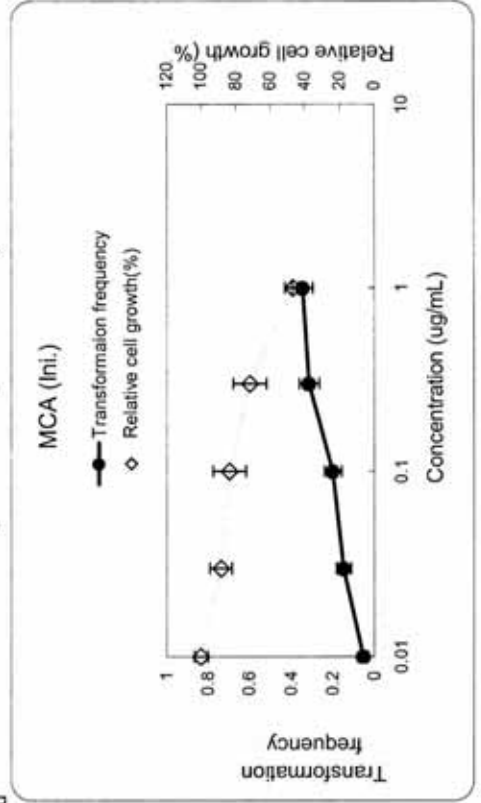
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	MCA
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)							
Crystal Violet Staining OD540nm	Blank	Solvent	0.01	0.03	0.1	0.3	1		
1	0.065	0.300	0.284	0.267	0.270	0.230	0.175		
2	0.065	0.303	0.283	0.270	0.254	0.177			
3	0.065	0.295	0.279	0.241	0.218	0.182			
4	0.066	0.294	0.257	0.239	0.254	0.161			
5	0.066	0.303	0.275	0.251	0.249	0.155			
6	0.067	0.314	0.242	0.250	0.207	0.192			
7	0.065	0.294	0.286	0.308	0.199	0.177			
8	0.067	0.284	0.269	0.255	0.251	0.175			
Average	0.066	0.298	0.272	0.260	0.233	0.174	#DIV/0!	#DIV/0!	#DIV/0!
SD	0.001	0.009	0.015	0.022	0.023	0.012	#DIV/0!	#DIV/0!	#DIV/0!
Average - Blank	0.000	0.233	0.206	0.194	0.167	0.109	#DIV/0!	#DIV/0!	#DIV/0!
Relative cell growth (%)		100.0	88.5	83.5	71.8	46.6	#DIV/0!	#DIV/0!	#DIV/0!
SD of relative cell growth		3.9	6.5	9.6	9.7	5.0	#DIV/0!	#DIV/0!	#DIV/0!

Transformation Assay		Concentrations (ug/ml)							
Giemsa Staining	Blank	Solvent	0.01	0.03	0.1	0.3	1		
No. of wells excluded	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96
No. of wells having foci	5	14	19	30	33				
Transformation frequency*	0.05208333	0.14583333	0.19791667	0.3125	0.34375	0.00000000	0.00000000	0.00000000	0.00000000
SE	0.02267769	0.03602169	0.04066449	0.04730704	0.04847529	0.00000000	0.00000000	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



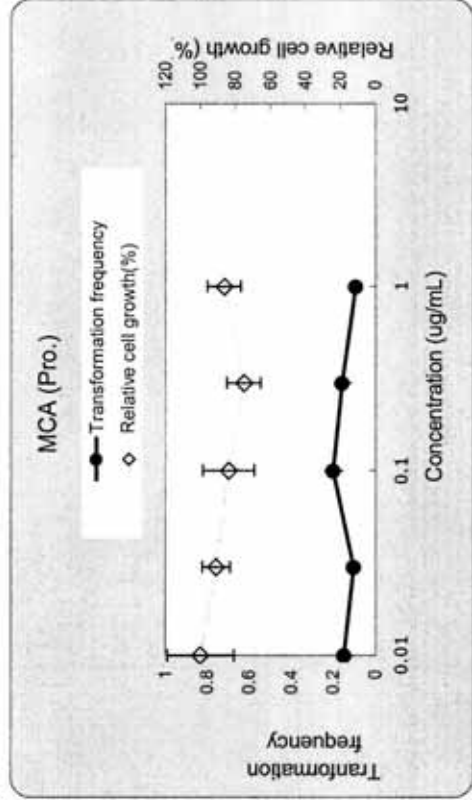
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	MCA
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining	Blank	Solvent	0.03	0.1	0.3	1	
OD540nm		0.01					
1	0.241	1.674	1.399	1.271	1.157	1.237	#DIV/0!
2	0.245	1.439	1.411	1.534	0.987	1.195	#DIV/0!
3	0.230	1.617	1.592	1.266	1.304	1.375	#DIV/0!
4	0.250	1.062	1.472	1.221	1.249	1.364	#DIV/0!
5	0.280	1.397	1.583	1.409	1.262	1.579	#DIV/0!
6	0.285	1.651	1.325	0.999	1.403	1.501	#DIV/0!
7	0.219	1.837	1.320	1.474	1.170	1.401	#DIV/0!
8	0.268	1.754	1.401	1.574	1.311	1.359	#DIV/0!
Average	0.252	1.554	1.438	1.344	1.230	1.376	#DIV/0!
SD	0.024	0.247	0.104	0.191	0.126	0.125	#DIV/0!
Average - Blank	0.000	1.302	1.186	1.091	0.978	1.124	#DIV/0!
Relative cell growth (%)	100.0	91.1	83.8	75.1	86.4	86.4	#DIV/0!
SD of relative cell growth	19.0	8.0	14.7	9.7	9.6	9.6	#DIV/0!

Transformation Assay		Concentrations (ug/ml)					
Glensa Staining	Solvent	0.01	0.03	0.1	0.3	1	
No. of wells excluded	2	0	0	0	0	0	0
No. of wells counted	94	96	96	96	96	96	96
No. of wells having foci	14	10	19	15	9	9	96
Transformation frequency*	0.14893617	0.10416667	0.19791667	0.15625	0.09375	0.09375	0
SE	0.03672123	0.03117758	0.04066449	0.03705794	0.02974911	0.02974911	0.00000000
							0.00000000
							0.00000000
							0.00000000
							0.00000000

* No. of wells having foci/ No. of wells counted



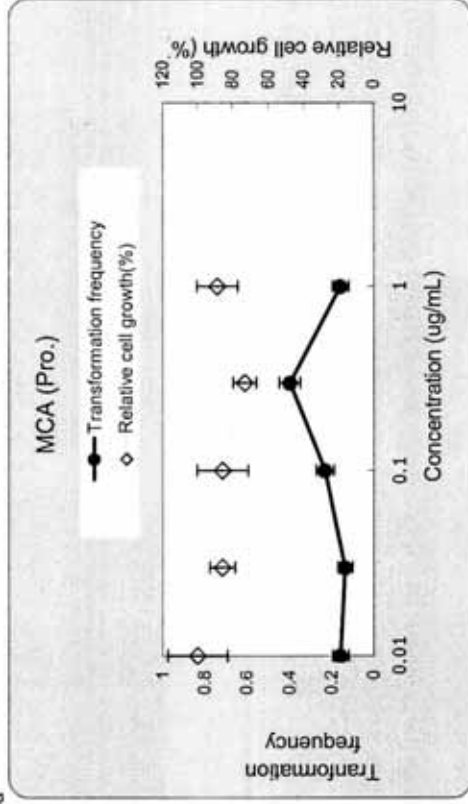
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	MCA
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)							
Crystal Violet Staining	Blank	Solvent	0.01	0.03	0.1	0.3	1		
OD540nm									
1	0.072	0.221	0.260	0.222	0.217	0.228			
2	0.080	0.255	0.252	0.268	0.239	0.293			
3	0.077	0.289	0.265	0.298	0.247	0.266			
4	0.082	0.296	0.258	0.264	0.244	0.281			
5	0.077	0.287	0.287	0.256	0.232	0.285			
6	0.081	0.327	0.244	0.305	0.221	0.285			
7	0.076	0.315	0.239	0.241	0.214	0.251			
8	0.070	0.325	0.272	0.223	0.247	0.237			
Average	0.077	0.289	0.260	0.260	0.233	0.266	#DIV/0!	#DIV/0!	#DIV/0!
SD	0.004	0.036	0.015	0.031	0.014	0.024	#DIV/0!	#DIV/0!	#DIV/0!
Average - Blank	0.000	0.213	0.183	0.183	0.156	0.189	#DIV/0!	#DIV/0!	#DIV/0!
Relative cell growth (%)		100.0	86.0	86.0	73.3	88.9	#DIV/0!	#DIV/0!	#DIV/0!
SD of relative cell growth		17.1	7.3	14.6	6.4	11.5	#DIV/0!	#DIV/0!	#DIV/0!

Transformation Assay		Concentrations (ug/ml)							
Giemsa Staining	Solvent	0.01	0.03	0.1	0.3	1	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96
No. of wells having foci	15	13	22	38	15				
Transformation frequency*	0.15625	0.13541667	0.22916667	0.39583333	0.15625				
SE	0.03705794	0.03492238	0.04289635	0.04991131	0.03705794	0.00000000	0.00000000	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



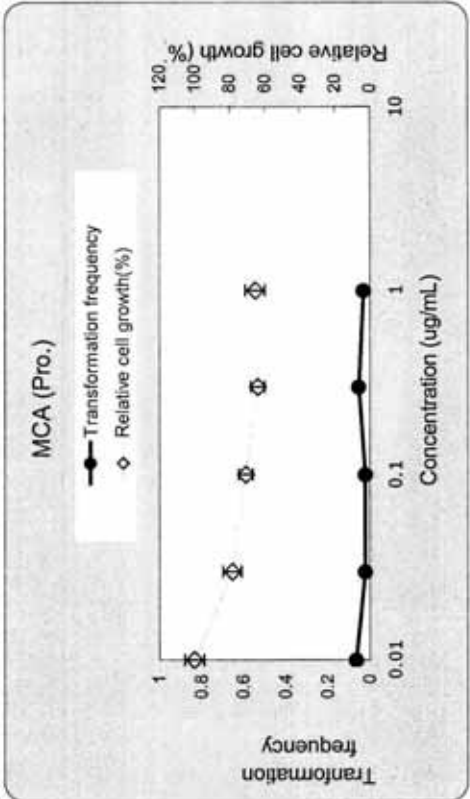
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	MCA
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)				
Crystal Violet Staining OD540nm	Blank	0.1% DMSO	0.03	0.1	0.3	1
1	0.154	1.166	0.904	0.864	0.765	0.772
2	0.183	1.190	0.942	0.869	0.798	0.829
3	0.182	1.163	0.988	0.924	0.869	0.834
4	0.185	1.239	1.066	0.913	0.884	0.827
5	0.234	1.240	1.022	0.909	0.853	0.926
6	0.193	1.192	1.031	0.903	0.855	0.937
7	0.210	1.181	0.961	0.950	0.875	0.907
8	0.210	1.329	1.021	0.985	0.856	0.850
Average	0.194	1.213	0.992	0.915	0.844	0.860
SD	0.024	0.056	0.053	0.040	0.041	0.057
Average - Blank	0.000	1.019	0.798	0.721	0.651	0.666
Relative cell growth (%)		100.0	78.3	70.8	63.9	65.4
SD of relative cell growth		5.4	5.2	3.9	4.0	5.6
					#DIV/0!	#DIV/0!
					#DIV/0!	#DIV/0!
					#DIV/0!	#DIV/0!
					#DIV/0!	#DIV/0!
					#DIV/0!	#DIV/0!
					#DIV/0!	#DIV/0!
					#DIV/0!	#DIV/0!

Transformation Assay		Concentrations (ug/ml)				
Giemsa Staining	0.1% DMSO	0.03	0.1	0.3	1	0
No. of wells excluded	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96
No. of wells having foci	6	2	2	5	3	96
Transformation frequency*	0.0625	0.02083333	0.02083333	0.05208333	0.03125	0
SE	0.02470529	0.01457713	0.01457713	0.02267769	0.01775805	0.00000000
						0.00000000
						0.00000000
						0.00000000
						0.00000000
						0.00000000

* No. of wells having foci/ No. of wells counted



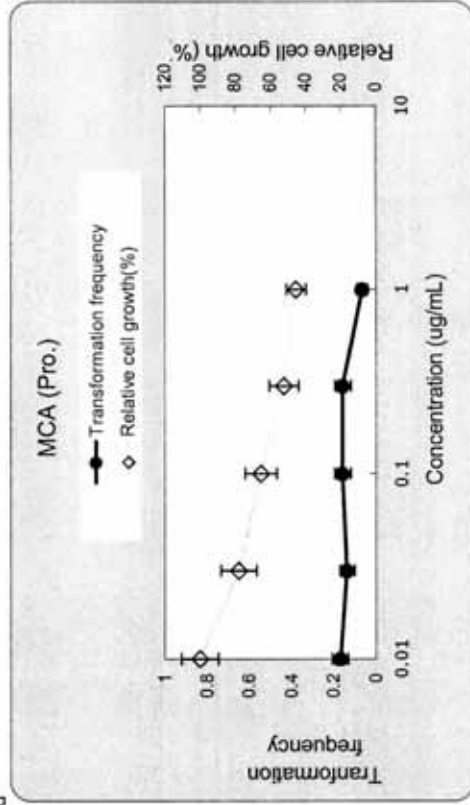
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	MCA
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining	Blank	Solvent	0.03	0.1	0.3	1	
OD540nm		0.01	0.03	0.1	0.3	1	
1	0.065	0.303	0.269	0.232	0.215	0.186	
2	0.065	0.294	0.223	0.239	0.183	0.173	
3	0.065	0.282	0.244	0.194	0.162	0.157	
4	0.066	0.280	0.226	0.207	0.172	0.170	
5	0.066	0.259	0.243	0.189	0.178	0.162	
6	0.067	0.311	0.246	0.200	0.187	0.150	
7	0.065	0.276	0.218	0.204	0.166	0.151	
8	0.067	0.242	0.197	0.185	0.162	0.155	
Average	0.066	0.281	0.233	0.206	0.178	0.163	#DIV/0!
SD	0.001	0.023	0.022	0.020	0.018	0.012	#DIV/0!
Average - Blank	0.000	0.215	0.168	0.141	0.112	0.097	#DIV/0!
Relative cell growth (%)		100.0	77.9	65.3	52.3	45.2	#DIV/0!
SD of relative cell growth		10.5	10.1	9.1	8.2	5.7	#DIV/0!

Transformation Assay		Concentrations (ug/ml)					
Glensa Staining	Solvent	0.03	0.1	0.3	1	0	0
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	16	13	15	15	6	0	0
Transformation frequency*	0.16666667	0.13541667	0.15625	0.15625	0.0625	0.00000000	0.00000000
SE	0.03803629	0.03492238	0.03705794	0.03705794	0.02470529	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



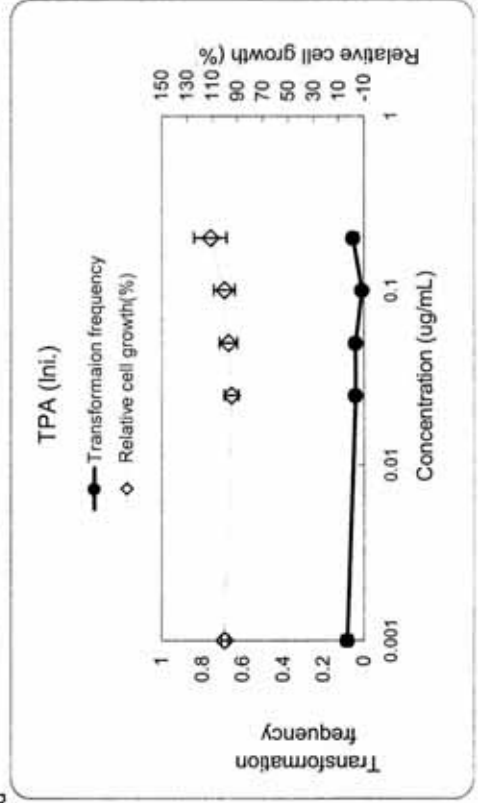
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	TPA
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining	Blank	Solvent	0.025	0.05	0.1	0.2	
OD540nm							
1	0.210	1.732	1.598	1.889	1.925	2.426	
2	0.316	1.751	1.878	1.805	1.775	1.987	
3	0.212	1.788	1.747	1.603	1.600	1.836	
4	0.202	1.848	1.669	1.605	1.936	1.842	
5	0.201	1.805	1.633	1.824	1.640	1.949	
6	0.218	1.835	1.761	1.809	1.768	1.942	
7	0.214	1.817	1.699	1.653	1.780	1.776	
8	0.233	1.751	1.700	1.796	1.939	1.993	
Average	0.226	1.791	1.711	1.748	1.795	1.989	#DIV/0!
SD	0.038	0.043	0.087	0.110	0.131	0.201	#DIV/0!
Average - Blank	0.000	1.565	1.485	1.522	1.570	1.743	#DIV/0!
Relative cell growth (%)		100.0	94.9	97.3	100.3	111.4	#DIV/0!
SD of relative cell growth		2.7	5.5	7.1	8.4	12.8	#DIV/0!

Transformation Assay		Concentrations (ug/ml)					
Giemsa Staining	Solvent	0.025	0.05	0.1	0.2		
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	8	4	4	1	5		
Transformation frequency*	0.08333333	0.04166667	0.04166667	0.01041667	0.05208333	0	0
SE	0.02820847	0.02039469	0.02039469	0.01036227	0.02267769	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



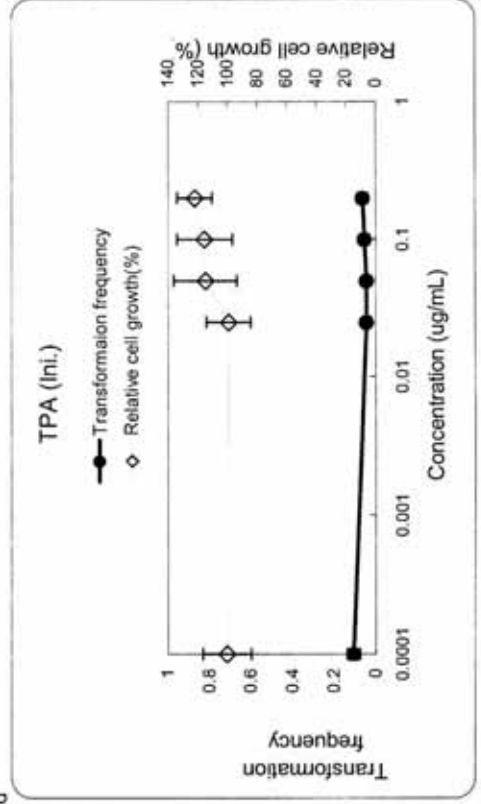
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	TPA
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining OD540nm	Blank	0.0001	0.025	0.05	0.1	0.2	
1	0.072	0.291	0.351	0.322	0.391	0.430	#DIV/0!
2	0.080	0.362	0.334	0.421	0.408	0.517	#DIV/0!
3	0.077	0.366	0.332	0.352	0.371	0.454	#DIV/0!
4	0.082	0.419	0.332	0.425	0.354	0.457	#DIV/0!
5	0.077	0.456	0.423	0.453	0.440	0.469	#DIV/0!
6	0.081	0.388	0.399	0.511	0.456	0.435	#DIV/0!
7	0.076	0.407	0.418	0.494	0.524	0.397	#DIV/0!
8	0.070	0.359	0.439	0.431	0.477	0.426	#DIV/0!
Average	0.077	0.381	0.379	0.426	0.428	0.448	#DIV/0!
SD	0.004	0.049	0.046	0.064	0.057	0.036	#DIV/0!
Average - Blank	0.000	0.304	0.302	0.349	0.351	0.371	#DIV/0!
Relative cell growth (%)	100.0	99.2	114.8	115.3	122.1	117.7	#DIV/0!
SD of relative cell growth	16.2	15.1	21.1	18.9	11.7	11.7	#DIV/0!

Transformation Assay		Concentrations (ug/ml)					
Glomse Staining	Solvent	0.0001	0.025	0.05	0.1	0.2	
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	10	4	4	5	6	6	96
Transformation frequency*	0.10416667	0.04166667	0.04166667	0.05208333	0.0625	0	0
SE	0.03117758	0.02039469	0.02039469	0.02267769	0.02470529	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



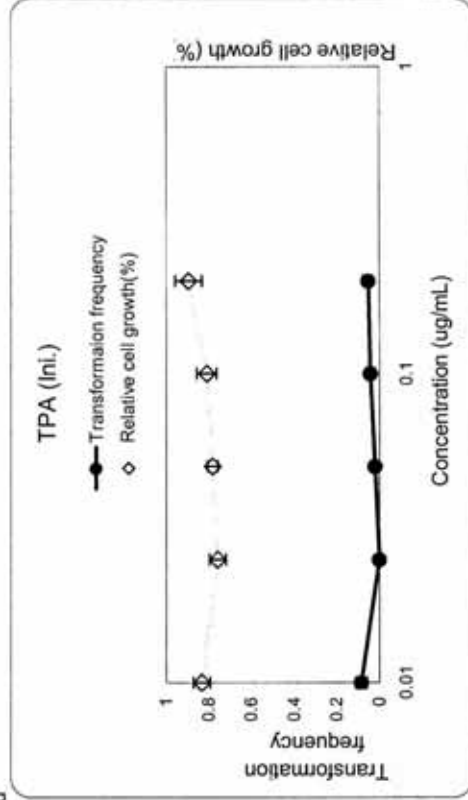
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	TPA
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)							
Crystal Violet Staining OD540nm	Blank	0.1% DMSO	0.025	0.05	0.1	0.2	0.1	0.2	#DIV/0!
1	0.154	1.171	1.073	1.138	1.181	1.182	1.181	1.182	#DIV/0!
2	0.176	1.158	1.059	1.090	1.071	1.161	1.071	1.161	#DIV/0!
3	0.166	1.120	1.125	1.114	1.162	1.227	1.162	1.227	#DIV/0!
4	0.199	1.184	1.043	1.124	1.117	1.300	1.117	1.300	#DIV/0!
5	0.221	1.276	1.095	1.121	1.214	1.282	1.214	1.282	#DIV/0!
6	0.190	1.225	1.158	1.152	1.145	1.299	1.145	1.299	#DIV/0!
7	0.195	1.219	1.140	1.152	1.206	1.315	1.206	1.315	#DIV/0!
8	0.250	1.210	1.160	1.185	1.241	1.393	1.241	1.393	#DIV/0!
Average	0.194	1.195	1.107	1.135	1.167	1.270	1.167	1.270	#DIV/0!
SD	0.031	0.048	0.046	0.029	0.056	0.076	0.056	0.076	#DIV/0!
Average - Blank	0.000	1.002	0.913	0.941	0.973	1.076	0.973	1.076	#DIV/0!
Relative cell growth (%)	100.0	91.1	93.9	97.2	97.2	107.4	97.2	107.4	#DIV/0!
SD of relative cell growth	4.8	4.5	2.9	5.6	5.6	7.6	5.6	7.6	#DIV/0!

Transformation Assay		Concentrations (ug/ml)							
Giemsa Staining	Blank	0.1% DMSO	0.025	0.05	0.1	0.2	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96
No. of wells having foci	8	0	2	4	5	5	5	5	5
Transformation frequency*	0.08333333	0	0.02083333	0.04166667	0.05208333	0.05208333	0	0	0
SE	0.02820847	0.00000000	0.01457713	0.02039469	0.02267769	0.00000000	0.00000000	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



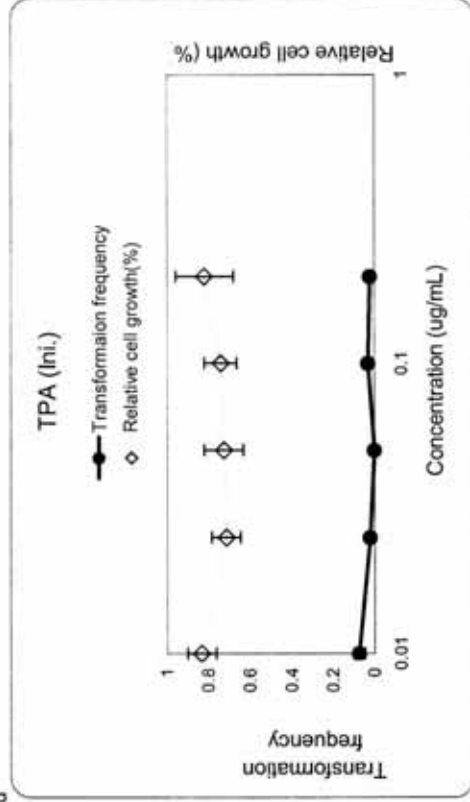
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical:	TPA
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining	Blank	Solvent	0.01	0.025	0.05	0.1	0.2
1	0.065	0.352	0.307	0.339	0.331	0.331	0.392
2	0.065	0.305	0.284	0.256	0.282	0.288	0.339
3	0.065	0.352	0.278	0.256	0.282	0.288	0.339
4	0.066	0.317	0.295	0.307	0.259	0.361	0.288
5	0.066	0.329	0.289	0.305	0.266	0.281	0.361
6	0.067	0.328	0.309	0.255	0.300	0.309	0.281
7	0.065	0.301	0.265	0.274	0.306	0.265	0.309
8	0.067	0.299	0.248	0.303	0.307	0.265	0.316
Average	0.066	0.323	0.287	0.290	0.295	0.295	0.319
SD	0.001	0.021	0.022	0.029	0.024	0.043	0.043
Average - Blank	0.000	0.257	0.221	0.225	0.229	0.253	0.253
Relative cell growth (%)		100.0	85.9	87.4	89.1	98.5	98.5
SD of relative cell growth		8.3	8.4	11.1	9.4	16.7	16.7

Transformation Assay		Concentrations (ug/ml)					
Giemsa Staining	Solvent	0.01	0.025	0.05	0.1	0.2	0
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	7	2	0	3	2	0	0
Transformation frequency*	0.07291667	0.02083333	0	0.03125	0.02083333	0	0
SE	0.02853610	0.01457713	0.00000000	0.01775805	0.01457713	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



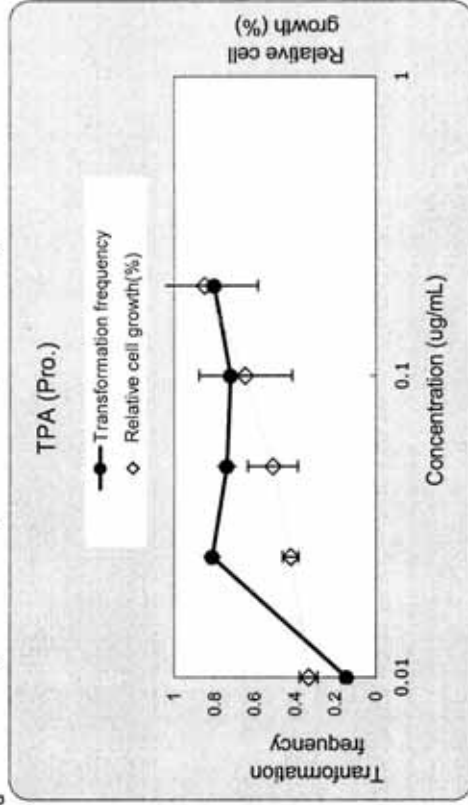
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	TPA
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)												
Crystal Violet Staining	Blank	Solvent	0.01	0.025	0.05	0.1	0.2	0.367	100	254.6	79.6	#DIV/0!	#DIV/0!	#DIV/0!
OD540nm														
1	0.047	0.164	0.202	0.207	0.239	0.565								
2	0.051	0.160	0.211	0.217	0.249	0.389								
3	0.043	0.177	0.235	0.266	0.439	0.437								
4	0.047	0.167	0.197	0.197	0.408	0.363								
5	0.043	0.155	0.218	0.203	0.214	0.259								
6	0.047	0.192	0.197	0.215	0.306	0.344								
7	0.049	0.163	0.193	0.321	0.220	0.268								
8	0.048	0.202	0.197	0.289	0.251	0.309								
Average	0.047	0.173	0.206	0.239	0.291	0.367								
SD	0.003	0.017	0.014	0.046	0.087	0.100								
Average - Blank	0.000	0.126	0.159	0.193	0.244	0.320								
Relative cell growth (%)		100.0	126.9	153.2	194.1	254.6								
SD of relative cell growth		13.2	11.4	37.0	69.2	79.6								

Transformation Assay		Concentrations (ug/ml)											
Giensa Staining	Solvent	0.01	0.025	0.05	0.1	0.2	0	0	0	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	14	78	71	69	77								
Transformation frequency*	0.14583333	0.8125	0.73958333	0.71875	0.80208333								
SE	0.03602169	0.03983609	0.04479116	0.04588805	0.04066449								

* No. of wells having foci/ No. of wells counted



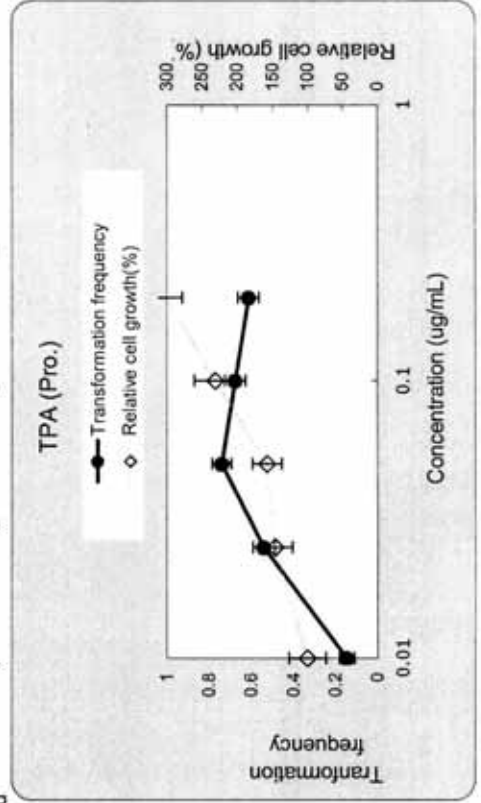
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	TPA
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)													
Crystal Violet Staining	Blank	Solvent	0.01	0.025	0.05	0.1	0.2	0.695	0.745	0.750	0.811	0.805	0.764	0.724	0.849
OD540nm			0.072	0.356	0.365	0.465	0.465	0.365	0.473	0.580	0.633	0.670	0.664	0.578	0.675
1	0.080	0.225	0.077	0.409	0.439	0.597	0.608	0.409	0.597	0.745	0.811	0.805	0.764	0.724	0.849
2	0.077	0.322	0.004	0.508	0.430	0.580	0.070	0.430	0.580	0.750	0.811	0.805	0.764	0.724	0.849
3	0.082	0.341	0.000	0.385	0.435	0.633	0.531	0.435	0.633	0.811	0.805	0.764	0.724	0.849	0.849
4	0.077	0.325	0.000	0.385	0.499	0.670	0.691	0.499	0.670	0.805	0.805	0.764	0.724	0.849	0.849
5	0.081	0.360	0.000	0.484	0.478	0.664	0.664	0.478	0.664	0.764	0.764	0.764	0.764	0.764	0.764
6	0.076	0.390	0.000	0.406	0.453	0.578	0.578	0.453	0.578	0.724	0.724	0.724	0.724	0.724	0.724
7	0.070	0.246	0.000	0.361	0.377	0.675	0.675	0.377	0.675	0.849	0.849	0.849	0.849	0.849	0.849
8	0.077	0.307	0.000	0.412	0.439	0.608	0.608	0.439	0.608	0.768	0.768	0.768	0.768	0.768	0.768
Average	0.004	0.061	0.000	0.056	0.048	0.070	0.051	0.056	0.070	0.051	0.051	0.051	0.051	0.051	0.051
SD	0.000	0.230	0.000	0.335	0.362	0.531	0.691	0.362	0.531	0.691	0.691	0.691	0.691	0.691	0.691
Relative cell growth (%)	100.0	145.7	100.0	157.4	157.4	230.9	300.6	157.4	230.9	300.6	300.6	300.6	300.6	300.6	300.6
SD of relative cell growth	26.4	24.2	26.4	20.7	20.7	30.5	22.0	20.7	30.5	22.0	22.0	22.0	22.0	22.0	22.0

Transformation Assay		Concentrations (ug/ml)									
Giemsa Staining	Solvent	0.01	0.025	0.05	0.1	0.2	0	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	14	52	71	65	59	59	59	59	59	59	59
Transformation frequency*	0.14583333	0.54166667	0.73958333	0.67708333	0.61458333	0.61458333	0.61458333	0.61458333	0.61458333	0.61458333	0.61458333
SE	0.03602169	0.05085354	0.04479116	0.04772332	0.04967296	0.04967296	0.04967296	0.04967296	0.04967296	0.04967296	0.04967296

* No. of wells having foci/ No. of wells counted



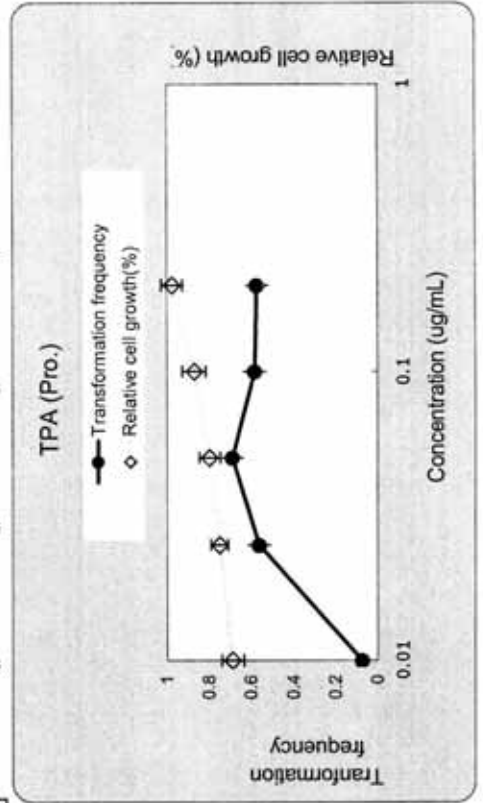
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	TPA
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)					
Crystal Violet Staining OD540nm	Blank	0.1% DMSO	0.025	0.05	0.1	0.2	
1	0.154	1.067	1.242	1.253	1.424	1.744	
2	0.183	1.308	1.300	1.395	1.558	1.673	
3	0.182	1.239	1.342	1.444	1.449	1.660	
4	0.185	1.206	1.360	1.527	1.519	1.757	
5	0.234	1.334	1.463	1.432	1.649	1.635	
6	0.193	1.351	1.422	1.505	1.695	1.757	
7	0.210	1.281	1.394	1.451	1.591	1.838	
8	0.210	1.231	1.345	1.482	1.622	1.869	
Average	0.194	1.252	1.359	1.436	1.563	1.742	#DIV/0!
SD	0.024	0.091	0.089	0.085	0.095	0.083	#DIV/0!
Average - Blank	0.000	1.058	1.165	1.242	1.370	1.548	#DIV/0!
Relative cell growth (%)		100.0	110.1	117.4	129.4	146.3	#DIV/0!
SD of relative cell growth		8.6	6.6	8.0	9.0	7.9	#DIV/0!

Transformation Assay		Concentrations (ug/ml)					
Giemsa Staining	0.1% DMSO	0.025	0.05	0.1	0.2		
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	7	54	66	56	55		
Transformation frequency*	0.07291667	0.5625	0.6875	0.58333333	0.57291667		
SE	0.02653610	0.05063079	0.04730704	0.05031728	0.05048547		

* No. of wells having foci/ No. of wells counted



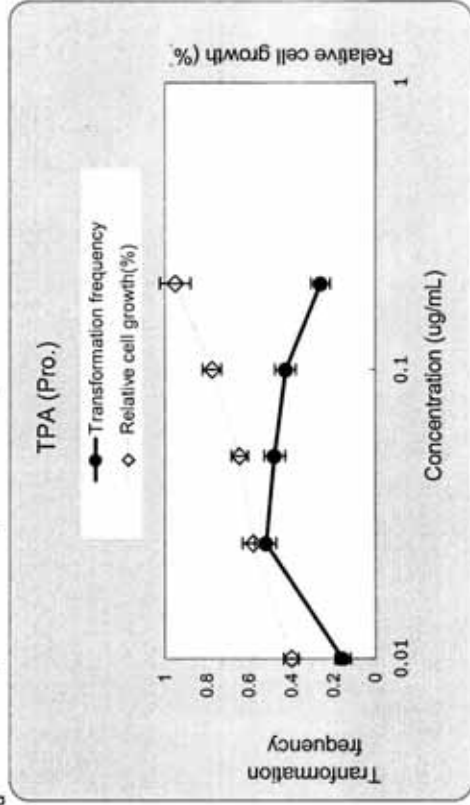
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical:	TPA
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)												
Crystal Violet Staining	Blank	Solvent	0.01	0.025	0.05	0.1	0.2	0.615	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!
OD540nm	0.065	0.310	0.419	0.440	0.490	0.520	0.552	0.634	0.577	0.554	0.529	0.527	0.562	
1	0.065	0.286	0.374	0.398	0.472	0.486	0.554	0.529	0.527	0.562	0.569	0.477	0.407	0.477
2	0.065	0.284	0.389	0.375	0.480	0.486	0.554	0.529	0.527	0.562	0.407	0.407	0.407	0.407
3	0.066	0.271	0.369	0.402	0.486	0.445	0.529	0.527	0.562	0.562	0.407	0.407	0.407	0.407
4	0.066	0.284	0.381	0.434	0.486	0.445	0.529	0.527	0.562	0.562	0.407	0.407	0.407	0.407
5	0.067	0.272	0.371	0.408	0.445	0.463	0.460	0.460	0.460	0.460	0.460	0.460	0.460	0.460
6	0.065	0.248	0.340	0.403	0.395	0.395	0.395	0.395	0.395	0.395	0.395	0.395	0.395	0.395
7	0.067	0.262	0.332	0.372	0.407	0.477	0.569	0.569	0.569	0.569	0.569	0.569	0.569	0.569
8	0.066	0.277	0.372	0.407	0.477	0.569	0.569	0.569	0.569	0.569	0.569	0.569	0.569	0.569
Average	0.001	0.019	0.027	0.021	0.023	0.038	0.038	0.038	0.038	0.038	0.038	0.038	0.038	0.038
SD	0.000	0.211	0.306	0.341	0.411	0.503	0.503	0.503	0.503	0.503	0.503	0.503	0.503	0.503
Average - Blank		100.0	144.9	161.4	194.4	237.9	237.9	237.9	237.9	237.9	237.9	237.9	237.9	237.9
Relative cell growth (%)		8.8	12.9	10.0	10.8	18.2	18.2	18.2	18.2	18.2	18.2	18.2	18.2	18.2
SD of relative cell growth														

Transformation Assay		Concentrations (ug/ml)										
Giemsa Staining	Solvent	0.01	0.025	0.05	0.1	0.2	0	0	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	15	50	46	41	25	25	25	25	25	25	25	25
Transformation frequency*	0.15625	0.520833333	0.47916667	0.42708333	0.26041667	0.26041667	0.26041667	0.26041667	0.26041667	0.26041667	0.26041667	0.26041667
SE	0.03705794	0.05098672	0.05098672	0.05048547	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116

* No. of wells having foci/ No. of wells counted



Annex 10

Results Submitted from Laboratories in the Phase I of Validation Study of 96-Well Method

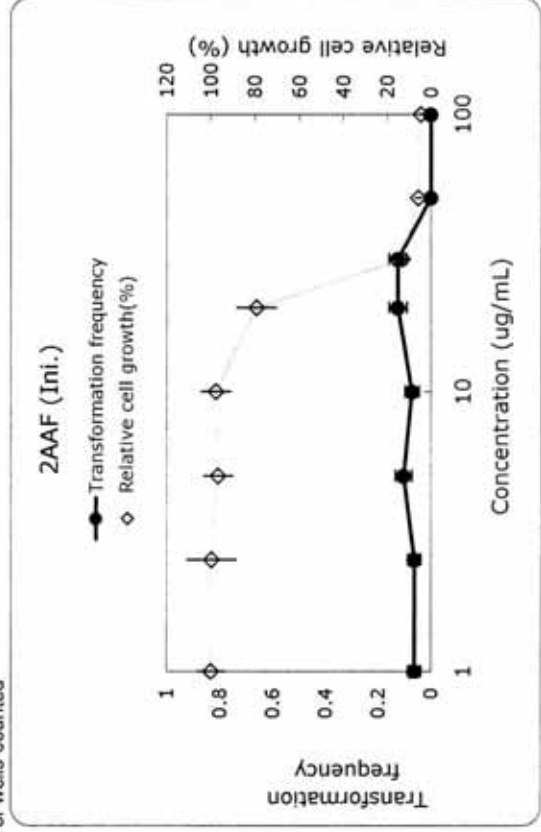
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	2AAF
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)								Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
		Blank	1	2.5	5	10	20	30	50		
1	0.118	0.678	0.720	0.618	0.659	0.530	0.209	0.161	0.148	0.678	0.224
2	0.124	0.701	0.711	0.631	0.625	0.538	0.191	0.148	0.139	0.701	0.275
3	0.128	0.696	0.676	0.677	0.621	0.634	0.188	0.145	0.141	0.696	0.306
4	0.114	0.661	0.676	0.663	0.648	0.511	0.164	0.147	0.138	0.661	0.288
5	0.104	0.609	0.733	0.729	0.730	0.526	0.166	0.150	0.138	0.609	0.258
6	0.123	0.697	0.534	0.634	0.701	0.590	0.202	0.138	0.146	0.697	0.287
7	0.096	0.708	0.678	0.634	0.661	0.519	0.188	0.135	0.124	0.708	0.236
8	0.102	0.651	0.663	0.668	0.654	0.623	0.185	0.140	0.137	0.651	0.220
Average	0.114	0.675	0.674	0.657	0.662	0.559	0.187	0.146	0.139	0.675	0.262
SD	0.012	0.033	0.062	0.036	0.037	0.049	0.016	0.008	0.007	0.033	0.032
Average - Blank	0.000	0.562	0.560	0.543	0.549	0.445	0.073	0.032	0.025	0.562	0.148
Relative Cell Growth (%)		100.0	99.8	96.7	97.7	79.3	13.0	5.7	4.5	100.0	26.4
SD of Relative Cell Growth		6.0	11.0	6.4	6.5	8.8	2.8	1.5	1.3	6.0	5.8

Transformation Assay	Giemsa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	Transformation frequency* SE	Concentrations (ug/ml)								Negative Control	Positive Control	
						1	2.5	5	10	20	30	50	100			0
		0	96	6	0.0625	0.0625	0.104166667	0.072916667	0.125	0.125	0.125	0.125	0.125	0.125	0.0625	0.458333333
		0	96	6	0.02470529	0.02470529	0.03117758	0.02653610	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.02470529	0.05085354
		0	96	6	0.0625	0.0625	0.104166667	0.072916667	0.125	0.125	0.125	0.125	0.125	0.125	0.0625	0.458333333
		0	96	6	0.02470529	0.02470529	0.03117758	0.02653610	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.02470529	0.05085354

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

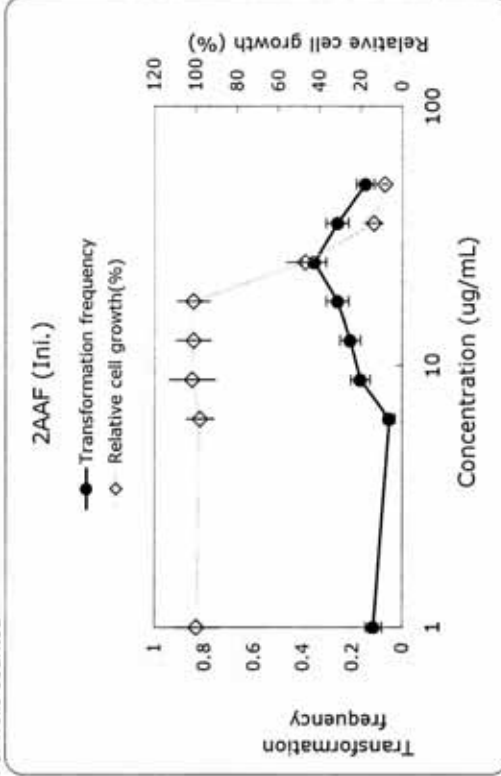
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	ZAAF
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (ug/mL)
	Blank	6.25	8.84	12.5	17.7	25	35.4	50	70.7	100.0		
Crystal Violet Staining OD _{540nm}	Blank	1	0.246	0.301	0.288	0.295	0.182	0.064	0.041	0.246	0.146	
1	0.018	0.282	0.301	0.288	0.295	0.182	0.064	0.041	0.246	0.146		
2	0.023	0.322	0.385	0.327	0.328	0.194	0.060	0.051	0.341	0.186		
3	0.021	0.299	0.279	0.322	0.358	0.135	0.060	0.046	0.316	0.189		
4	0.020	0.345	0.338	0.345	0.339	0.168	0.053	0.051	0.341	0.189		
5	0.019	0.327	0.321	0.356	0.305	0.119	0.045	0.039	0.318	0.134		
6	0.018	0.315	0.350	0.296	0.315	0.146	0.047	0.043	0.339	0.152		
7	0.016	0.295	0.300	0.341	0.339	0.188	0.064	0.039	0.317	0.155		
8	0.017	0.312	0.314	0.298	0.292	0.142	0.087	0.043	0.327	0.156		
Average	0.019	0.312	0.324	0.322	0.321	0.159	0.060	0.044	#DIV/0!	#DIV/0!	0.163	
SD	0.002	0.020	0.033	0.025	0.024	0.028	0.013	0.005	#DIV/0!	#DIV/0!	0.022	
Average - Blank	0.000	0.293	0.305	0.303	0.302	0.140	0.041	0.025	#DIV/0!	#DIV/0!	0.144	
Relative Cell Growth (%)	100.0	98.0	101.8	101.2	101.1	46.9	13.7	8.4	#DIV/0!	#DIV/0!	48.3	
SD of Relative Cell Growth	10.4	6.7	11.2	8.5	7.9	9.2	4.4	1.6	#DIV/0!	#DIV/0!	7.2	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	1	6.25	8.84	12.5	17.7	25	35.4	50	70.7	100.0		
No. of wells excluded	1	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	11	5	16	20	25	34	25	14	11	11	11	68
Transformation frequency*	0.11458333	0.05208333	0.16666667	0.20833333	0.26041667	0.35416667	0.26041667	0.14583333	0	0	0.11458333	0.70833333
SE	0.03250865	0.02267769	0.03803629	0.04144908	0.04479116	0.04881221	0.04479116	0.03602169	0.00000000	0.00000000	0.03250865	0.04639024

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

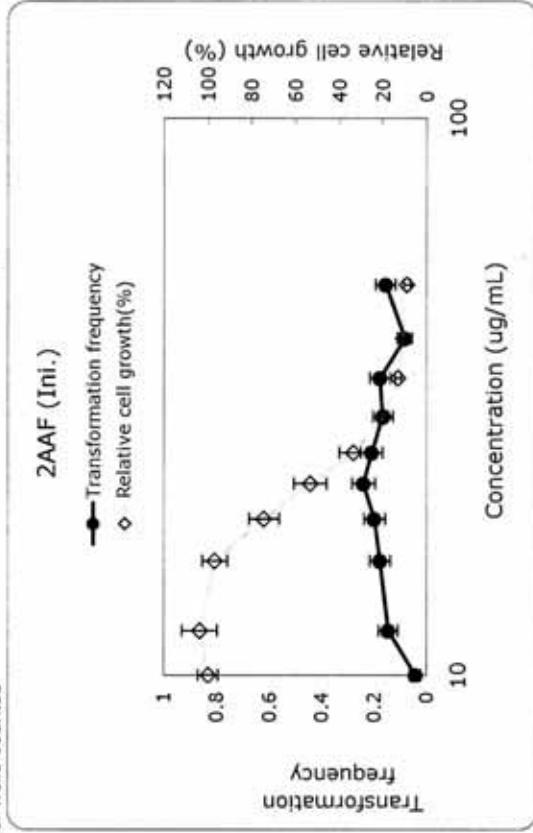
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	2AAF
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control		Positive Control	
			10	12	16	19	22	25	29	34	40	50	0.1% DMSO	MCA (1ug/mL)		
1	0.101	0.961	0.992	0.917	0.812	0.616	0.299	0.252	0.233	0.191	0.190	0.961	0.495			
2	0.111	0.885	0.973	0.890	0.724	0.528	0.390	0.257	0.209	0.188	0.197	0.885	0.520			
3	0.111	0.957	0.839	0.863	0.688	0.476	0.387	0.266	0.229	0.225	0.186	0.957	0.532			
4	0.112	0.897	0.904	0.934	0.721	0.495	0.414	0.287	0.217	0.212	0.193	0.897	0.523			
5	0.111	0.866	1.036	0.802	0.660	0.552	0.436	0.257	0.205	0.206	0.184	0.866	0.457			
6	0.122	0.875	0.879	0.867	0.638	0.630	0.431	0.299	0.210	0.194	0.177	0.875	0.535			
7	0.112	0.911	0.925	0.864	0.667	0.506	0.331	0.263	0.217	0.189	0.173	0.911	0.483			
8	0.113	0.906	0.954	0.934	0.727	0.483	0.332	0.255	0.199	0.181	0.159	0.906	0.475			
Average	0.112	0.907	0.938	0.883	0.705	0.536	0.378	0.267	0.215	0.198	0.182	0.907	0.503			
SD	0.006	0.035	0.064	0.045	0.055	0.059	0.051	0.017	0.012	0.015	0.012	0.035	0.029			
Average - Blank	0.000	0.796	0.826	0.771	0.593	0.424	0.266	0.155	0.103	0.087	0.071	0.796	0.391			
Relative Cell Growth (%)	100.0	103.8	103.8	96.9	74.5	53.3	33.4	19.5	13.0	10.9	8.9	100.0	49.1			
SD of Relative Cell Growth	4.4	8.0	8.0	5.6	6.9	7.4	6.4	2.1	1.5	1.9	1.5	4.4	3.7			

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
	10	12	16	19	22	25	29	34	40	50	0.1% DMSO	MCA (1ug/mL)		
Giemsa Staining	10	12	16	19	22	25	29	34	40	50	0	0		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0		
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96		
No. of wells having foci	4	14	17	19	23	20	16	17	8	15	4	83		
Transformation frequency*	0.041666667	0.145833333	0.177083333	0.197916667	0.239583333	0.208333333	0.166666667	0.177083333	0.083333333	0.15625	0.041666667	0.864583333		
SE	0.02039469	0.03602169	0.03896110	0.04066449	0.04356307	0.04144908	0.03803629	0.03896110	0.02820847	0.03705794	0.02039469	0.03492238		

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

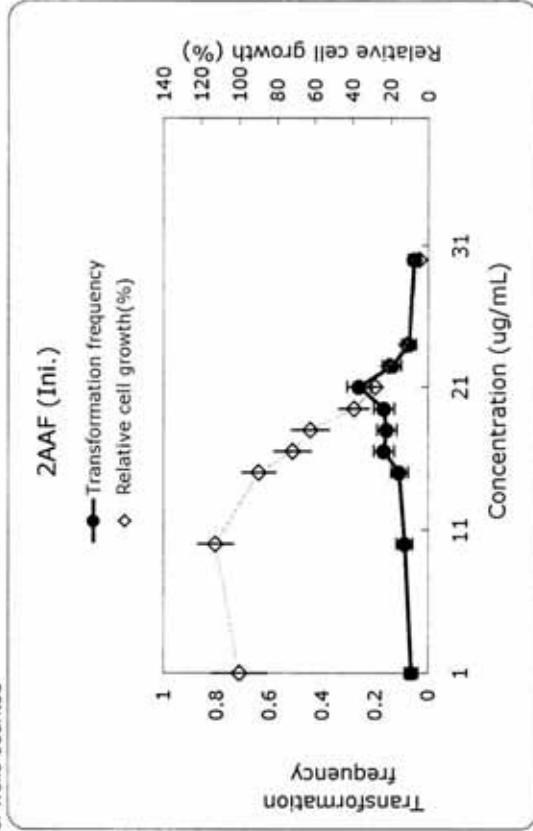
Chemical Code:	ZAAF
Institution:	Lab 4
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control		Positive Control	
			1	10	15	16.5	18	19.5	21	22.5	24	30	0.1% DMSO	MCA (1ug/mL)		
1	0.072	0.268	0.341	0.263	0.242	0.210	0.156	0.128	0.124	0.108	0.085	0.268	0.221			
2	0.071	0.281	0.291	0.280	0.244	0.200	0.147	0.136	0.109	0.100	0.086	0.281	0.192			
3	0.074	0.271	0.285	0.292	0.234	0.254	0.175	0.130	0.124	0.099	0.086	0.271	0.184			
4	0.075	0.306	0.293	0.261	0.227	0.202	0.152	0.128	0.114	0.097	0.083	0.306	0.205			
5	0.073	0.345	0.314	0.258	0.222	0.206	0.190	0.135	0.122	0.097	0.087	0.345	0.192			
6	0.074	0.297	0.327	0.248	0.214	0.184	0.150	0.150	0.119	0.091	0.081	0.297	0.173			
7	0.075	0.255	0.321	0.232	0.238	0.197	0.145	0.128	0.114	0.096	0.079	0.255	0.180			
8	0.076	0.255	0.318	0.270	0.181	0.191	0.139	0.128	0.116	0.097	0.081	0.255	0.183			
Average	0.074	0.285	0.311	0.263	0.225	0.206	0.157	0.133	0.118	0.098	0.084	0.285	0.191			
SD	0.001	0.030	0.020	0.018	0.021	0.021	0.017	0.008	0.005	0.005	0.003	0.030	0.015			
Average - Blank	0.000	0.211	0.237	0.189	0.152	0.132	0.083	0.059	0.044	0.024	0.010	0.211	0.117			
Relative Cell Growth (%)		100.0	112.6	89.7	71.9	62.5	39.3	28.0	20.9	11.5	4.6	100.0	55.6			
SD of Relative Cell Growth		14.4	9.3	8.7	9.7	10.0	8.0	3.7	2.5	2.3	1.3	14.4	7.3			

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining	1	10	15	16.5	18	19.5	21	22.5	24	30	0.1% DMSO	MCA (1ug/mL)		
No. of wells excluded	0	5	3	0	0	0	0	0	0	0	0	0		
No. of wells counted	96	91	93	96	96	96	96	96	96	96	96	96		
No. of wells having foci	6	8	10	16	15	16	25	13	7	5	6	41		
Transformation frequency*	0.0625	0.087912088	0.107526882	0.166666667	0.15625	0.166666667	0.260416667	0.135416667	0.072916667	0.052083333	0.0625	0.427083333		
SE	0.02470529	0.02968397	0.03212289	0.03803629	0.03705794	0.03803629	0.04479116	0.03492238	0.02653610	0.02267769	0.02470529	0.05048547		

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

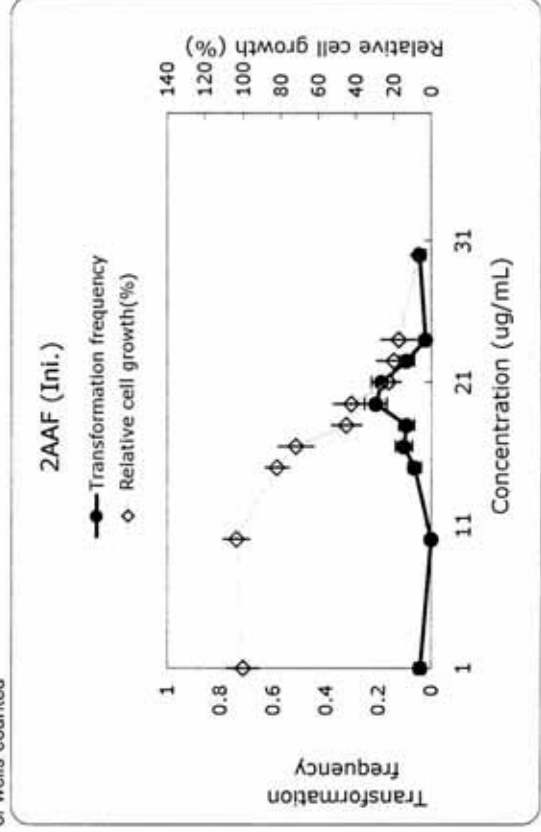
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	2AAF
Institution:	Lab 4
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)											Negative Control	Positive Control				
		0.1% DMSO	10	15	16.5	18	19.5	21	22.5	24	30	30						
Crystal Violet Staining																		
OD _{550nm}																		
1	0.062	0.320	0.264	0.287	0.232	0.208	0.163	0.160	0.167	0.084	0.320	0.165						
2	0.065	0.318	0.281	0.289	0.202	0.199	0.143	0.151	0.133	0.082	0.318	0.196						
3	0.065	0.361	0.282	0.288	0.172	0.188	0.116	0.128	0.097	0.081	0.361	0.201						
4	0.063	0.321	0.303	0.274	0.200	0.208	0.119	0.112	0.101	0.087	0.321	0.203						
5	0.066	0.364	0.318	0.243	0.184	0.192	0.116	0.103	0.097	0.083	0.364	0.212						
6	0.071	0.374	0.292	0.227	0.170	0.141	0.124	0.101	0.093	0.081	0.374	0.203						
7	0.063	0.335	0.277	0.234	0.180	0.152	0.105	0.097	0.097	0.082	0.335	0.178						
8	0.061	0.321	0.298	0.251	0.166	0.160	0.130	0.105	0.108	0.081	0.321	0.185						
Average	0.065	0.349	0.289	0.262	0.188	0.181	0.127	0.120	0.112	0.083	0.339	0.193						
SD	0.003	0.023	0.017	0.026	0.022	0.026	0.018	0.024	0.026	0.002	0.023	0.016						
Average - Blank	0.000	0.284	0.225	0.197	0.124	0.117	0.063	0.055	0.047	0.018	0.275	0.128						
Relative Cell Growth (%)		103.4	81.8	71.7	45.0	42.4	22.7	20.1	17.2	6.6	100.0	46.7						
SD of Relative Cell Growth		6.8	6.2	9.4	8.0	9.6	6.7	8.8	9.4	0.8	8.5	5.7						

Transformation Assay	1	10	15	16.5	18	19.5	21	22.5	24	30	Negative Control	Positive Control
No. of wells excluded	0	96	0	0	0	1	0	0	0	2	0	0
No. of wells counted	96	0	96	96	96	95	96	96	96	94	96	96
No. of wells having foci	4	6	10	10	9	20	18	9	2	4	4	52
Transformation frequency*	0.041666667	#DIV/0!	0.0625	0.104166667	0.09375	0.210526316	0.1875	0.09375	0.020833333	0.042553191	0.041666667	0.541666667
SE	0.02039469	#DIV/0!	0.02470529	0.03117758	0.02974911	0.04182734	0.03983609	0.02974911	0.01457713	0.02081898	0.02039469	0.05085354

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

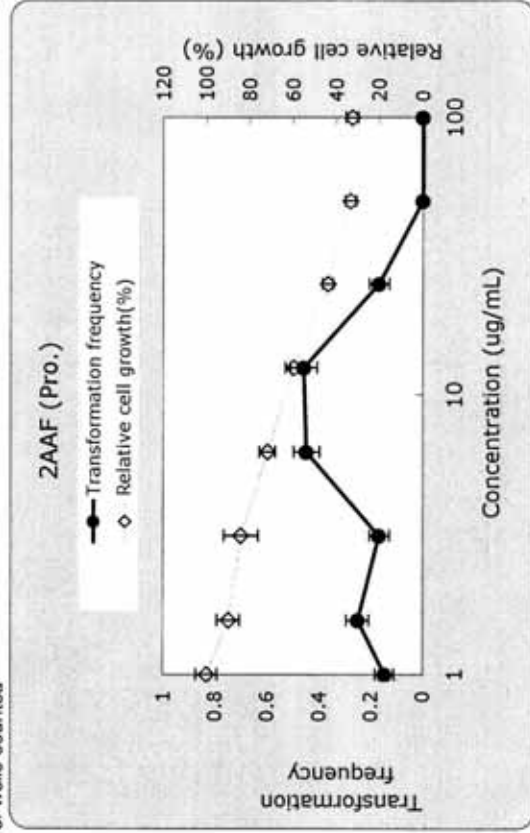
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	2AAF
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control	Positive Control
			0.1% DMSO	1.56	3.13	6.25	12.5	25	50	100		
1	0.112	0.905	0.757	0.688	0.666	0.551	0.456	0.372	0.352	0.905	0.940	
2	0.121	0.882	0.807	0.725	0.684	0.569	0.464	0.369	0.359	0.882	0.971	
3	0.120	0.870	0.881	0.758	0.682	0.576	0.441	0.378	0.379	0.870	0.980	
4	0.128	0.981	0.849	0.767	0.689	0.589	0.452	0.374	0.362	0.981	1.053	
5	0.136	0.919	0.797	0.800	0.644	0.575	0.467	0.375	0.393	0.919	0.992	
6	0.134	0.896	0.852	0.791	0.696	0.624	0.502	0.408	0.410	0.896	0.967	
7	0.135	0.862	0.843	0.889	0.742	0.634	0.477	0.416	0.405	0.862	1.017	
8	0.153	0.897	0.808	0.816	0.673	0.613	0.486	0.407	0.387	0.897	0.985	
Average	0.130	0.902	0.824	0.779	0.685	0.591	0.468	0.387	0.381	0.902	0.988	
SD	0.013	0.037	0.039	0.061	0.028	0.029	0.020	0.019	0.022	0.037	0.034	
Average - Blank	0.000	0.772	0.694	0.649	0.555	0.462	0.338	0.258	0.251	0.772	0.858	
Relative Cell Growth (%)		100.0	90.0	84.2	71.9	59.8	43.8	33.4	32.5	100.0	111.2	
SD of Relative Cell Growth		4.8	5.1	7.9	3.7	3.8	2.5	2.5	2.8	4.8	4.4	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	1	1.56	3.13	6.25	12.5	25	50	100	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	14	24	16	43	44	16	0	0	0	14	76	
Transformation frequency*	0.145833333	0.25	0.166666667	0.447916667	0.458333333	0.166666667	0	#VALUE!	0	0.145833333	0.791666667	
SE	0.03602169	0.04419417	0.03803629	0.05075342	0.05085354	0.03803629	0.00000000	0.00000000	0.00000000	0.03602169	0.04144908	

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

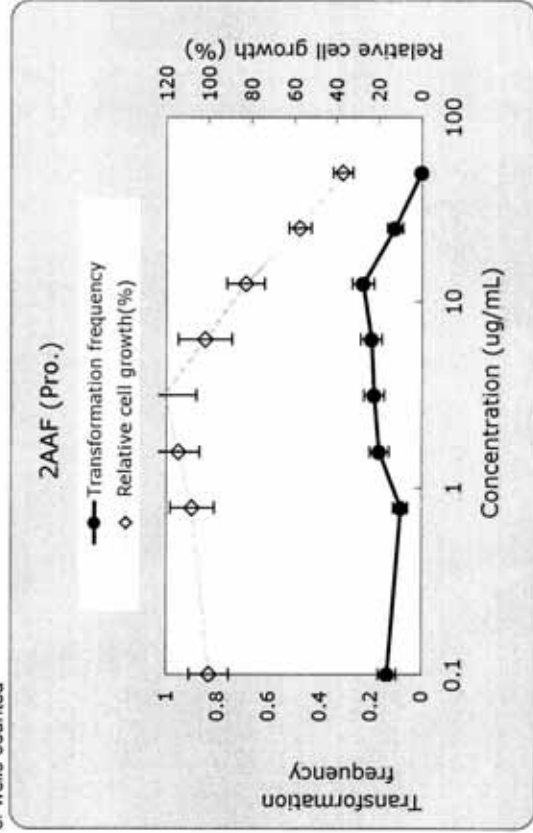
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	2AAF
Institution:	Lab 2
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)								Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)	
		0.1	0.78	1.56	3.13	6.25	12.5	25	50			
Crystal Violet Staining OD _{540nm}												
1	0.027	0.312	0.351	0.377	0.346	0.265	0.184	0.132	0.312	0.432		
2	0.027	0.369	0.362	0.336	0.363	0.313	0.195	0.157	0.369	0.525		
3	0.023	0.289	0.328	0.388	0.268	0.281	0.187	0.136	0.289	0.385		
4	0.027	0.280	0.352	0.351	0.355	0.253	0.226	0.126	0.280	0.439		
5	0.032	0.327	0.372	0.385	0.329	0.247	0.190	0.135	0.327	0.444		
6	0.024	0.317	0.309	0.371	0.306	0.242	0.174	0.136	0.317	0.368		
7	0.023	0.306	0.343	0.323	0.269	0.277	0.183	0.110	0.306	0.447		
8	0.022	0.315	0.282	0.315	0.319	0.236	0.190	0.127	0.315	0.330		
Average	0.026	0.314	0.337	0.356	0.319	0.264	0.191	0.132	0.314	0.421		
SD	0.003	0.027	0.030	0.029	0.037	0.025	0.015	0.013	0.027	0.060		
Average - Blank	0.000	0.289	0.312	0.330	0.294	0.239	0.166	0.107	0.289	0.396		
Relative Cell Growth (%)	100.0	114.3	120.8	101.7	82.6	57.3	37.0	4.6	100.0	137.0		
SD of Relative Cell Growth	9.3	10.3	15.0	12.6	8.8	5.3	4.6	9.3	9.3	20.6		

Transformation Assay	Concentrations (ug/ml)								Negative Control	Positive Control
Giemsa Staining	0.1	0.78	1.56	3.13	6.25	12.5	25	50		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	13	8	16	18	19	22	10	0	13	46
Transformation frequency*	0.135416667	0.083333333	0.166666667	0.1875	0.197916667	0.229166667	0.104166667	0	0.135416667	0.479166667
SE	0.03492238	0.02820847	0.03803629	0.03983609	0.04066449	0.04289635	0.03117758	0.00000000	0.03492238	0.05098672

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

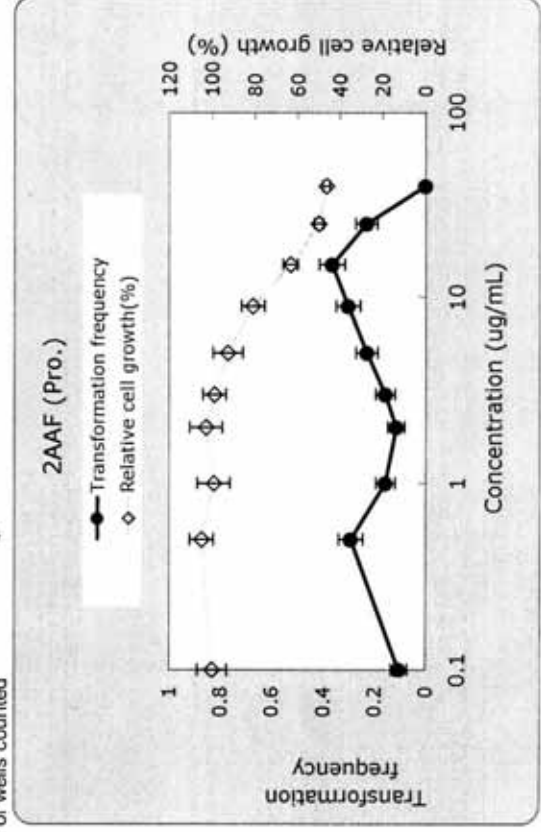
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	2AAF
Institution:	Lab 3
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)		
		0.1	0.5	1	2	3	5	9	15	25	40				
Crystal Violet Staining															
OD _{540nm}															
1	0.101	0.902	0.951	1.033	0.894	0.793	0.761	0.589	0.485	0.471	0.945	1.280			
2	0.111	1.030	0.910	0.975	0.909	0.963	0.838	0.650	0.523	0.476	0.875	1.208			
3	0.111	0.974	0.964	0.857	0.902	0.854	0.722	0.616	0.521	0.484	1.024	1.382			
4	0.112	0.963	0.833	0.977	0.923	0.859	0.750	0.629	0.530	0.488	0.861	1.252			
5	0.111	0.899	1.002	0.888	0.953	0.890	0.778	0.653	0.515	0.469	0.851	1.346			
6	0.122	0.976	0.879	0.966	0.947	0.832	0.774	0.628	0.496	0.480	0.925	1.209			
7	0.112	0.927	0.870	0.905	0.881	0.813	0.760	0.602	0.494	0.511	0.910	1.302			
8	0.113	0.942	0.839	0.876	0.813	0.807	0.688	0.581	0.526	0.494	0.910	1.199			
Average	0.112	0.952	0.906	0.935	0.903	0.851	0.759	0.619	0.511	0.484	0.913	1.272			
SD	0.006	0.044	0.061	0.062	0.044	0.055	0.044	0.027	0.017	0.014	0.055	0.068			
Average - Blank	0.000	0.840	0.794	0.823	0.791	0.740	0.647	0.507	0.400	0.373	0.801	1.161			
Relative Cell Growth (%)		104.9	99.2	102.7	98.8	92.4	80.8	63.3	49.9	46.5	100.0	144.9			
SD of Relative Cell Growth		5.4	7.7	7.7	5.5	6.9	5.4	3.3	2.1	1.7	6.9	8.5			

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	0.1	0.5	1	2	3	5	9	15	25	40			
Giemsa Staining													
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	10	28	15	11	15	22	29	35	22	22	10	88	88
Transformation frequency*	0.104166667	0.291666667	0.15625	0.114583333	0.15625	0.229166667	0.302083333	0.364583333	0.229166667	0.229166667	0.104166667	0.916666667	0.916666667
SE	0.03117758	0.04639024	0.03705794	0.03250865	0.03705794	0.04289635	0.04686294	0.04912382	0.04289635	0.04289635	0.03117758	0.02820847	0.02820847

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

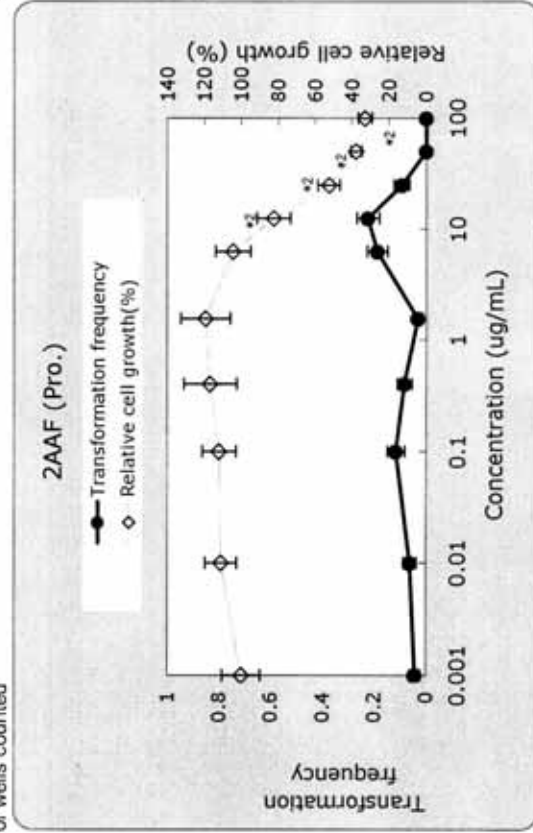
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	2AAF
Institution:	Lab 4
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)		
		0.001	0.01	0.1	0.4	1.56	6.25	12.5	25	50	100				
Crystal Violet Staining															
OD _{540nm}															
1	0.072	0.264	0.278	0.252	0.287	0.250	0.210	0.150	0.133	0.126	0.245	0.319			
2	0.073	0.250	0.257	0.286	0.290	0.250	0.219	0.167	0.142	0.128	0.256	0.367			
3	0.073	0.266	0.278	0.321	0.289	0.260	0.249	0.179	0.139	0.124	0.244	0.376			
4	0.074	0.297	0.276	0.286	0.330	0.247	0.208	0.166	0.138	0.130	0.275	0.354			
5	0.074	0.264	0.298	0.289	0.285	0.247	0.234	0.180	0.144	0.144	0.248	0.354			
6	0.072	0.253	0.250	0.280	0.271	0.285	0.209	0.163	0.146	0.136	0.248	0.338			
7	0.071	0.282	0.263	0.269	0.274	0.274	0.203	0.161	0.140	0.131	0.259	0.324			
8	0.074	0.267	0.254	0.239	0.249	0.234	0.209	0.153	0.131	0.129	0.210	0.320			
Average	0.073	0.268	0.269	0.278	0.282	0.256	0.218	0.165	0.139	0.131	0.248	0.344			
SD	0.001	0.015	0.016	0.025	0.024	0.016	0.016	0.011	0.005	0.006	0.018	0.022			
Average - Blank	0.000	0.195	0.197	0.205	0.209	0.183	0.145	0.092	0.066	0.058	0.176	0.271			
Relative Cell Growth (%)															
SD of Relative Cell Growth															
		100.0	112.0	116.7	119.1	104.4	82.5	52.6	37.8	33.1	100.0	154.6			
		10.5	8.7	14.3	13.4	9.4	8.9	6.1	2.9	3.6	10.5	12.5			

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	0.001	0.01	0.1	0.4	1.56	6.25	12.5	25	50	100		
No. of wells excluded	0	0	0	0	0	0	2	0	0	96	0	0
No. of wells counted	96	96	96	96	96	96	94	96	96	96	96	96
No. of wells having foci	4	6	11	8	3	18	21	9	0	0	4	43
Transformation frequency*	0.041666667	0.0625	0.114583333	0.083333333	0.03125	0.1875	0.223404255	0.09375	0	#DIV/0!	0.041666667	0.447916667
SE	0.02039469	0.02470529	0.03250865	0.02820847	0.01775805	0.03983609	0.04296149	0.02974911	0.00000000	#DIV/0!	0.02039469	0.05075342

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

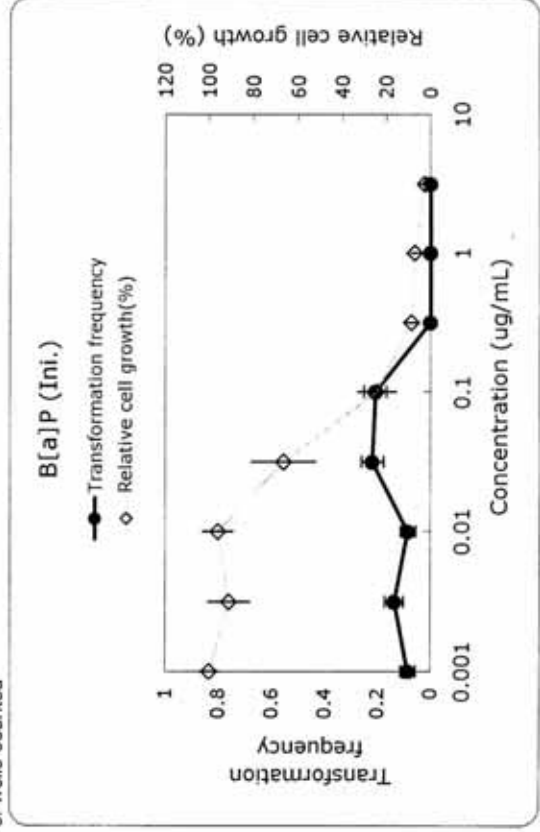
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	B[a]P
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										
Crystal Violet Staining	Blank	0.00316	0.01	0.0316	0.1	0.316	1	3.16	Negative Control	Positive Control		
OD _{540nm}		0.001	0.00316	0.01	0.0316	0.1	0.316	1	0.1% DMSO	MCA (1ug/mL)		
1	0.089	0.588	0.581	0.394	0.221	0.123	0.130	0.094	0.619	0.200		
2	0.095	0.624	0.607	0.472	0.229	0.167	0.126	0.122	0.651	0.256		
3	0.085	0.596	0.584	0.506	0.268	0.152	0.139	0.102	0.644	0.262		
4	0.098	0.551	0.545	0.516	0.253	0.143	0.138	0.102	0.597	0.245		
5	0.092	0.500	0.642	0.410	0.251	0.139	0.124	0.112	0.669	0.285		
6	0.089	0.541	0.650	0.548	0.240	0.126	0.144	0.119	0.628	0.249		
7	0.083	0.656	0.628	0.446	0.121	0.130	0.115	0.097	0.621	0.242		
8	0.095	0.601	0.637	0.308	0.190	0.120	0.119	0.087	0.614	0.182		
Average	0.091	0.582	0.609	0.450	0.222	0.138	0.129	0.104	0.630	0.240		
SD	0.005	0.050	0.037	0.078	0.047	0.016	0.010	0.012	0.023	0.033		
Average - Blank	0.000	0.491	0.519	0.359	0.131	0.047	0.039	0.014	0.540	0.149		
Relative Cell Growth (%)		91.1	96.1	66.6	24.3	8.7	7.2	2.5	100.0	27.7		
SD of Relative Cell Growth		9.2	6.8	14.4	8.7	3.0	1.9	2.3	4.3	6.2		

Transformation Assay		Concentrations (ug/ml)										
Giemsa Staining	Blank	0.00316	0.01	0.0316	0.1	0.316	1	3.16	Negative Control	Positive Control		
No. of wells excluded		0	0	0	0	33	76	96	0	0		
No. of wells counted		96	96	96	96	63	20	0	96	96		
No. of wells having foci		8	13	8	21	TOX	TOX	TOX	8	41		
Transformation frequency*		0.083333333	0.135416667	0.083333333	0.21875	0.208333333	#VALUE!	#VALUE!	0.083333333	0.427083333		
SE		0.02820847	0.03492238	0.02820847	0.04219232	0.04144908	#VALUE!	#VALUE!	0.02820847	0.05048547		

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

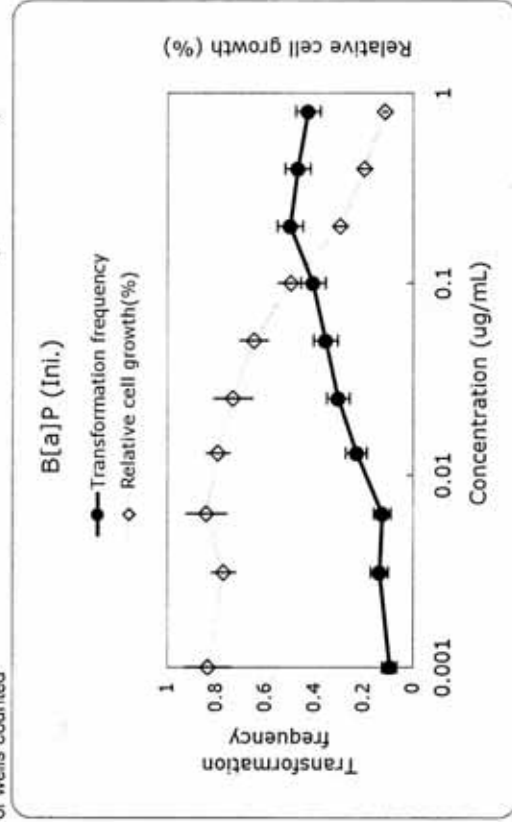
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	B[a]P
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control	Positive Control
			0.1% DMSO	0.0031	0.0063	0.013	0.025	0.05	0.1	0.2	0.4	0.8		
1	0.049	0.496	0.502	0.494	0.486	0.491	0.417	0.336	0.260	0.177	0.138	0.496	0.228	
2	0.050	0.579	0.517	0.581	0.499	0.527	0.478	0.328	0.254	0.172	0.133	0.579	0.273	
3	0.049	0.594	0.484	0.532	0.522	0.391	0.395	0.357	0.231	0.180	0.133	0.594	0.210	
4	0.052	0.634	0.519	0.627	0.545	0.523	0.449	0.364	0.211	0.201	0.126	0.634	0.218	
5	0.070	0.527	0.482	0.565	0.510	0.487	0.410	0.321	0.232	0.173	0.130	0.527	0.238	
6	0.080	0.514	0.561	0.518	0.567	0.462	0.438	0.375	0.225	0.161	0.129	0.514	0.244	
7	0.080	0.501	0.476	0.561	0.523	0.464	0.409	0.303	0.225	0.180	0.123	0.501	0.201	
8	0.074	0.481	0.498	0.488	0.495	0.520	0.480	0.396	0.224	0.167	0.120	0.481	0.224	
Average	0.063	0.541	0.505	0.546	0.518	0.483	0.435	0.348	0.233	0.176	0.129	0.541	0.230	
SD	0.014	0.055	0.028	0.047	0.027	0.045	0.032	0.031	0.016	0.012	0.006	0.055	0.022	
Average - Blank	0.000	0.478	0.442	0.483	0.455	0.420	0.372	0.285	0.170	0.113	0.066	0.478	0.167	
Relative Cell Growth (%)		100.0	92.5	101.0	95.3	87.9	77.8	59.5	35.5	23.7	13.8	100.0	34.9	
SD of Relative Cell Growth		11.5	5.8	9.8	5.7	9.4	6.8	6.5	3.4	2.5	1.2	11.5	4.7	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giensa Staining	0.001	0.0031	0.0063	0.013	0.025	0.05	0.1	0.2	0.4	0.8		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	9	13	12	22	29	34	39	48	45	41	9	70
Transformation frequency*	0.09375	0.135416667	0.125	0.229166667	0.302083333	0.354166667	0.40625	0.5	0.46875	0.427083333	0.09375	0.729166667
SE	0.02974911	0.03492238	0.03375386	0.04289635	0.04686294	0.04881221	0.05012598	0.05103104	0.05093127	0.05048547	0.02974911	0.04535538

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

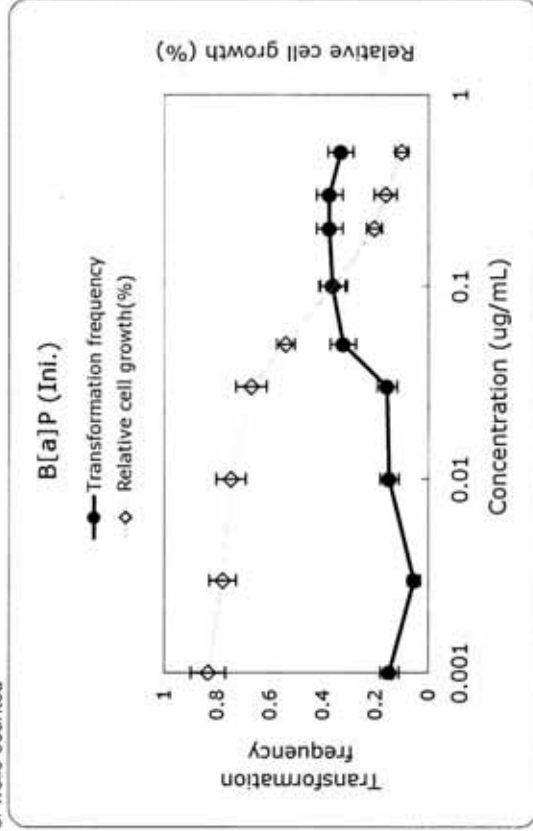
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	B[a]P
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control		
Crystal Violet Staining		0.1% DMSO										0.1% DMSO		MCA (1ug/mL)		
OD _{540nm}		Blank	0.001	0.003	0.01	0.03	0.05	0.1	0.2	0.3	0.5	0.5	0.5	0.5	0.5	0.5
1	0.096	0.096	1.094	0.954	1.043	1.022	0.838	0.632	0.374	0.357	0.238	1.094	0.551			
2	0.111	0.111	1.238	1.167	1.040	0.942	0.836	0.644	0.408	0.310	0.260	1.238	0.541			
3	0.106	0.106	1.119	1.086	1.017	0.936	0.766	0.550	0.379	0.284	0.222	1.119	0.619			
4	0.106	0.106	1.059	1.078	1.003	0.844	0.746	0.484	0.377	0.296	0.270	1.059	0.620			
5	0.099	0.099	1.144	1.069	0.969	0.874	0.764	0.550	0.344	0.407	0.227	1.144	0.586			
6	0.097	0.097	1.314	1.125	1.010	0.924	0.726	0.470	0.371	0.301	0.220	1.314	0.536			
7	0.097	0.097	1.135	1.138	1.081	0.924	0.767	0.523	0.331	0.233	0.236	1.135	0.551			
8	0.099	0.099	1.136	1.064	1.197	0.981	0.822	0.587	0.291	0.266	0.169	1.136	0.432			
Average	0.101	0.101	1.155	1.085	1.045	0.948	0.783	0.555	0.359	0.307	0.230	1.155	0.555			
SD	0.006	0.006	0.082	0.064	0.070	0.072	0.043	0.063	0.036	0.054	0.030	0.082	0.060			
Average - Blank	0.000	0.000	1.054	0.984	0.944	0.847	0.682	0.454	0.258	0.205	0.129	1.054	0.453			
Relative Cell Growth (%)	100.0	93.4	89.6	80.4	64.7	43.1	24.5	19.5	12.2	43.0	7.8	100.0	43.0			
SD of Relative Cell Growth	7.8	6.1	6.6	6.8	4.1	6.0	3.4	5.1	2.9	5.7	7.8	100.0	5.7			

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control		
Giemsa Staining		0.1% DMSO										0.1% DMSO		MCA (1ug/mL)		
No. of wells excluded		0.001	0.003	0.01	0.03	0.05	0.1	0.2	0.3	0.5	0.5	0.5	0.5	0.5	0.5	0.5
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	14	5	14	15	31	35	36	36	36	32	32	14	73			
Transformation frequency* SE	0.145833333	0.052083333	0.145833333	0.15625	0.322916667	0.364583333	0.375	0.375	0.375	0.333333333	0	0.145833333	0.760416667			
	0.03602169	0.02267769	0.03602169	0.03705794	0.04772332	0.04912382	0.04941059	0.04941059	0.04941059	0.04811252	0.00000000	0.03602169	0.04356307			

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

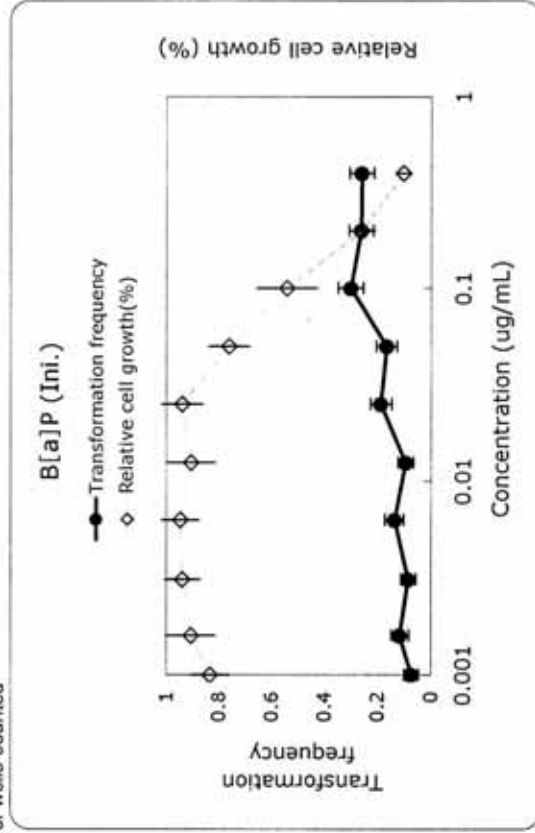
Chemical Code:	B[a]P
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining	Blank	0.1% DMSO	0.0016	0.0031	0.0063	0.0125	0.025	0.05	0.1	0.2	0.4	0.1% DMSO	MCA (1ug/mL)		
1	0.078	0.270	0.266	0.294	0.286	0.299	0.303	0.278	0.212	0.134	0.104	0.270	0.203		
2	0.074	0.267	0.285	0.340	0.328	0.333	0.305	0.286	0.204	0.154	0.102	0.267	0.173		
3	0.073	0.279	0.291	0.300	0.304	0.306	0.313	0.279	0.202	0.144	0.115	0.279	0.182		
4	0.076	0.267	0.323	0.320	0.338	0.308	0.318	0.275	0.279	0.150	0.102	0.267	0.215		
5	0.072	0.288	0.306	0.315	0.302	0.299	0.334	0.259	0.204	0.141	0.098	0.288	0.170		
6	0.074	0.301	0.302	0.330	0.330	0.336	0.336	0.276	0.200	0.147	0.098	0.301	0.181		
7	0.074	0.320	0.339	0.296	0.320	0.267	0.318	0.262	0.214	0.120	0.095	0.320	0.172		
8	0.079	0.293	0.322	0.303	0.305	0.283	0.274	0.225	0.182	0.129	0.096	0.293	0.163		
Average	0.075	0.286	0.304	0.312	0.314	0.304	0.313	0.267	0.212	0.140	0.101	0.286	0.182		
SD	0.002	0.019	0.024	0.017	0.018	0.023	0.019	0.019	0.029	0.012	0.006	0.019	0.018		
Average - Blank	0.000	0.211	0.229	0.237	0.239	0.229	0.238	0.192	0.137	0.065	0.026	0.211	0.107		
Relative Cell Growth (%)		100.0	108.8	112.7	113.6	108.7	112.9	91.4	65.0	30.9	12.4	100.0	51.0		
SD of Relative Cell Growth		9.0	11.2	7.9	8.4	10.9	9.2	9.2	13.7	5.5	3.0	9.0	8.4		

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining	Blank	0.001	0.0016	0.0031	0.0063	0.0125	0.025	0.05	0.1	0.2	0.4	0.1% DMSO	MCA (1ug/mL)		
No. of wells excluded		0	0	0	0	0	0	0	0	1	0	0	0		
No. of wells counted		96	96	96	96	96	96	96	96	95	96	96	96		
No. of wells having foci		7	11	8	13	9	18	16	29	25	25	7	54		
Transformation frequency*		0.072916667	0.114583333	0.083333333	0.135416667	0.09375	0.1875	0.166666667	0.302083333	0.263157895	0.260416667	0.072916667	0.5625		
SE		0.02653610	0.03250885	0.02820847	0.03492238	0.02974911	0.03983609	0.03803629	0.04686294	0.04517869	0.04479116	0.02653610	0.05063079		

* No. of wells having foci/ No. of wells counted

Remarks	



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

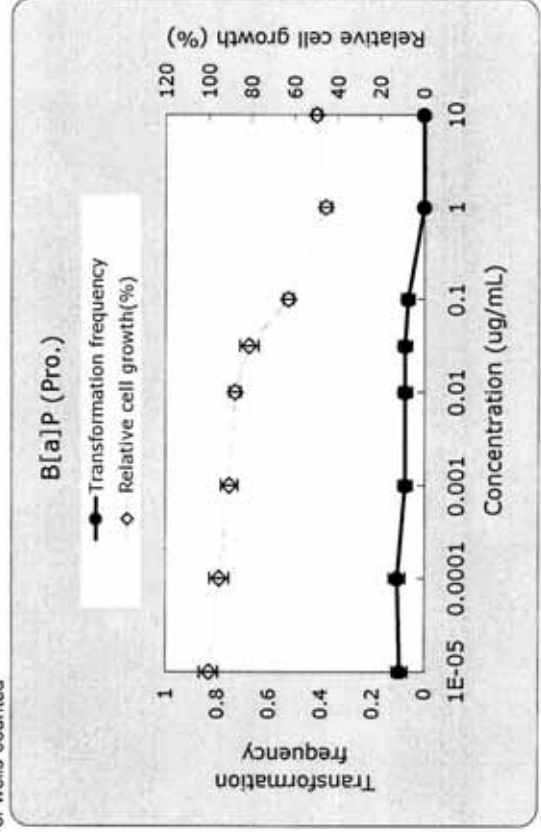
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	B[a]P
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Blank	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)			
			0.00001	0.0001	0.001	0.01	0.0316	0.1	1	10	10	10					
1	0.124	0.124	0.640	0.580	0.513	0.461	0.374	0.395	0.630	0.632	0.670	0.611	0.674	0.659	0.627	0.641	1.011
2	0.125	0.125	0.608	0.592	0.551	0.456	0.337	0.378	0.632	0.632	0.670	0.611	0.674	0.659	0.627	0.641	1.011
3	0.116	0.116	0.592	0.578	0.545	0.448	0.363	0.359	0.670	0.670	0.611	0.674	0.659	0.627	0.641	1.011	1.042
4	0.117	0.117	0.591	0.578	0.524	0.452	0.334	0.391	0.632	0.632	0.670	0.611	0.674	0.659	0.627	0.641	1.042
5	0.106	0.106	0.632	0.594	0.585	0.461	0.369	0.372	0.632	0.632	0.670	0.611	0.674	0.659	0.627	0.641	1.042
6	0.122	0.122	0.644	0.576	0.533	0.435	0.358	0.379	0.659	0.659	0.670	0.611	0.674	0.659	0.627	0.641	1.042
7	0.107	0.107	0.597	0.635	0.563	0.426	0.350	0.377	0.627	0.627	0.670	0.611	0.674	0.659	0.627	0.641	1.042
8	0.112	0.112	0.640	0.582	0.540	0.449	0.360	0.385	0.641	0.641	0.670	0.611	0.674	0.659	0.627	0.641	1.042
Average	0.116	0.116	0.618	0.593	0.544	0.449	0.356	0.380	0.643	0.643	0.670	0.611	0.674	0.659	0.627	0.641	1.042
SD	0.007	0.007	0.023	0.020	0.023	0.012	0.014	0.011	0.022	0.022	0.022	0.011	0.011	0.022	0.022	0.022	0.058
Average - Blank	0.000	0.000	0.502	0.477	0.428	0.332	0.240	0.263	0.527	0.527	0.527	0.263	0.263	0.527	0.527	0.527	0.863
Relative Cell Growth (%)			95.3	90.6	81.3	63.1	45.5	50.0	100.0	100.0	100.0	50.0	50.0	100.0	100.0	100.0	163.7
SD of Relative Cell Growth			4.4	3.8	4.3	2.3	2.7	2.1	4.3	4.3	4.3	2.1	2.1	4.3	4.3	4.3	10.9

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Giemsa Staining	0.00001	0.0001	0.001	0.01	0.0316	0.1	1	10	10	10			0
No. of wells excluded	0	0	0	0	0	0	96	96	96	96	96	96	0
No. of wells counted	96	96	96	96	96	96	0	0	0	0	96	96	96
No. of wells having foci	9	10	7	7	7	6	TOX	TOX	TOX	TOX	9	9	54
Transformation frequency*	0.09375	0.104166667	0.072916667	0.072916667	0.072916667	0.0625	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.09375	0.09375	0.5625
SE	0.02974911	0.03117758	0.02653610	0.02653610	0.02653610	0.02470529	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.02974911	0.02974911	0.05063079

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

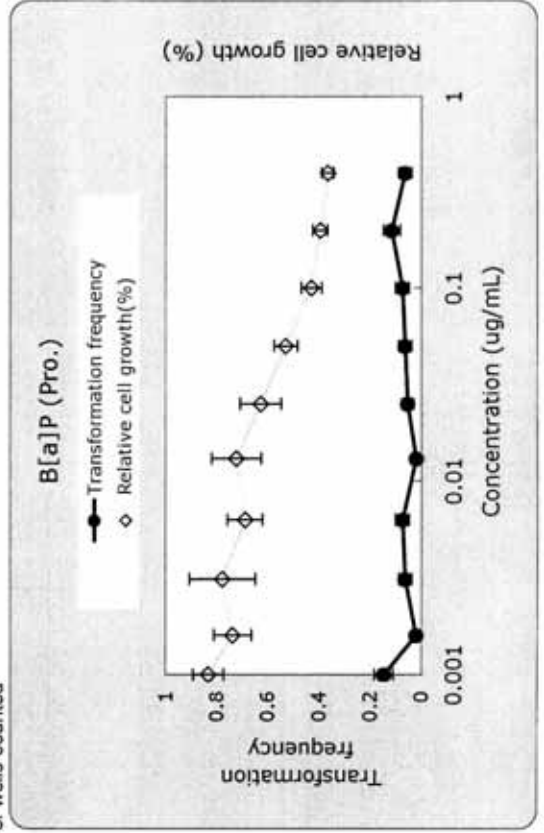
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	B[a]P
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)												
	Blank	0.001	0.0016	0.0031	0.0063	0.013	0.025	0.05	0.1	0.2	0.4	Negative Control	Positive Control
Crystal Violet Staining												0.1% DMSO	TPA (50ng/mL)
OD _{540nm}													
1	0.048	0.547	0.505	0.457	0.457	0.439	0.368	0.333	0.282	0.263	0.246	0.547	0.522
2	0.052	0.573	0.478	0.506	0.490	0.493	0.393	0.353	0.286	0.272	0.273	0.573	0.556
3	0.051	0.522	0.480	0.478	0.402	0.417	0.424	0.336	0.291	0.307	0.280	0.522	0.610
4	0.052	0.585	0.498	0.684	0.508	0.482	0.465	0.389	0.314	0.302	0.271	0.585	0.581
5	0.067	0.491	0.433	0.522	0.503	0.549	0.425	0.404	0.326	0.289	0.275	0.491	0.626
6	0.080	0.567	0.576	0.537	0.477	0.576	0.423	0.395	0.333	0.304	0.293	0.567	0.602
7	0.079	0.573	0.496	0.476	0.451	0.464	0.516	0.376	0.347	0.302	0.277	0.573	0.564
8	0.078	0.507	0.453	0.451	0.411	0.435	0.403	0.389	0.312	0.297	0.282	0.507	0.580
Average	0.063	0.546	0.490	0.514	0.462	0.482	0.427	0.371	0.311	0.292	0.275	0.546	0.580
SD	0.014	0.035	0.042	0.075	0.040	0.056	0.046	0.027	0.024	0.016	0.013	0.035	0.033
Average - Blank	0.000	0.482	0.427	0.451	0.399	0.419	0.364	0.307	0.248	0.229	0.211	0.482	0.517
Relative Cell Growth (%)		100.0	88.4	93.4	82.7	86.8	75.4	63.7	51.4	47.4	43.8	100.0	107.2
SD of Relative Cell Growth		7.2	8.8	15.6	8.3	11.6	9.5	5.5	4.9	3.4	2.8	7.2	6.9

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	0.001	0.0016	0.0031	0.0063	0.013	0.025	0.05	0.1	0.2	0.4	0.1% DMSO	TPA (50ng/mL)
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	14	2	6	7	2	5	6	7	11	6	14	55
Transformation frequency*	0.145833333	0.020833333	0.0625	0.072916667	0.020833333	0.052083333	0.0625	0.072916667	0.114583333	0.0625	0.145833333	0.572916667
SE	0.03602169	0.01457713	0.02470529	0.02653610	0.01457713	0.02267769	0.02470529	0.02653610	0.03250965	0.02470529	0.03602169	0.05048547

*No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

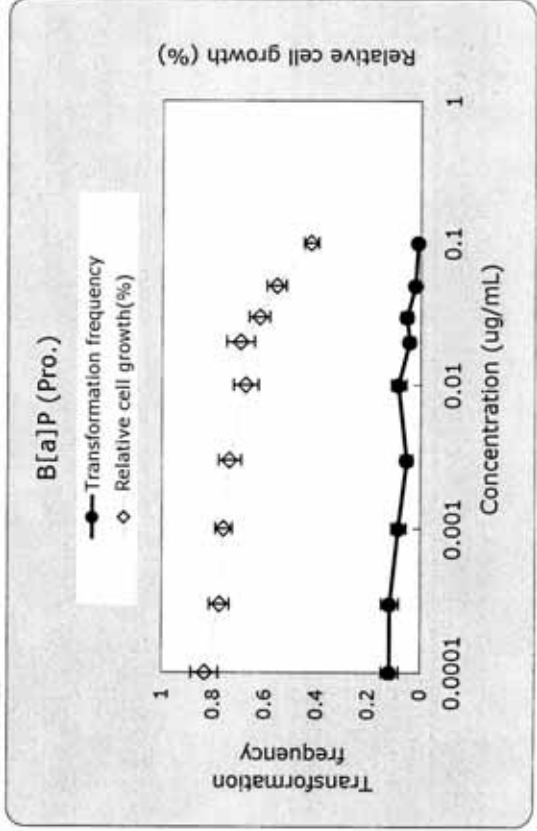
Chemical Code:	B[a]P
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)								Negative Control	Positive Control	
Crystal Violet Staining	Blank	0.0001	0.0003	0.001	0.003	0.01	0.02	0.03	0.05	0.1	0.1% DMSO	TPA (50ng/mL)
1	0.091	1.044	0.914	0.985	0.873	0.863	0.927	0.800	0.761	0.548	1.044	1.245
2	0.092	1.017	1.020	0.983	0.913	0.898	0.930	0.825	0.763	0.599	1.017	1.580
3	0.122	1.195	1.004	0.952	0.975	0.912	1.021	0.913	0.755	0.636	1.195	1.469
4	0.099	1.092	1.011	0.960	0.932	0.785	0.872	0.762	0.777	0.568	1.092	1.385
5	0.080	1.036	1.018	1.071	1.024	0.891	0.846	0.809	0.649	0.557	1.036	1.473
6	0.095	1.064	0.986	0.981	1.000	0.936	0.979	0.871	0.809	0.601	1.064	1.409
7	0.107	1.019	1.070	0.977	0.987	0.971	0.838	0.844	0.744	0.622	1.019	1.496
8	0.120	1.123	1.031	1.015	0.994	0.870	0.901	0.811	0.759	0.622	1.123	1.331
Average	0.101	1.074	1.007	0.991	0.962	0.891	0.914	0.829	0.752	0.594	1.074	1.424
SD	0.015	0.061	0.045	0.038	0.051	0.055	0.064	0.046	0.046	0.033	0.061	0.104
Average - Blank	0.000	0.973	0.906	0.890	0.862	0.790	0.814	0.729	0.651	0.493	0.973	1.323
Relative Cell Growth (%)	100.0	93.1	91.4	88.5	88.5	81.2	83.6	74.9	66.9	50.7	100.0	135.9
SD of Relative Cell Growth	6.3	4.6	3.9	5.3	5.3	5.7	6.5	4.8	4.7	3.4	6.3	10.7

Transformation Assay		Concentrations (ug/ml)								Negative Control	Positive Control	
Giemsa Staining	Blank	0.0001	0.0003	0.001	0.003	0.01	0.02	0.03	0.05	0.1	0.1% DMSO	TPA (50ng/mL)
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	11	11	8	5	8	4	5	5	2	1	11	80
Transformation frequency*	0.114583333	0.114583333	0.083333333	0.052083333	0.083333333	0.0666667	0.041666667	0.052083333	0.020833333	0.010416667	0.114583333	0.833333333
SE	0.03250865	0.03250865	0.02820847	0.02267769	0.02820847	0.02039469	0.02267769	0.02267769	0.01457713	0.01036227	0.03250865	0.03803629

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

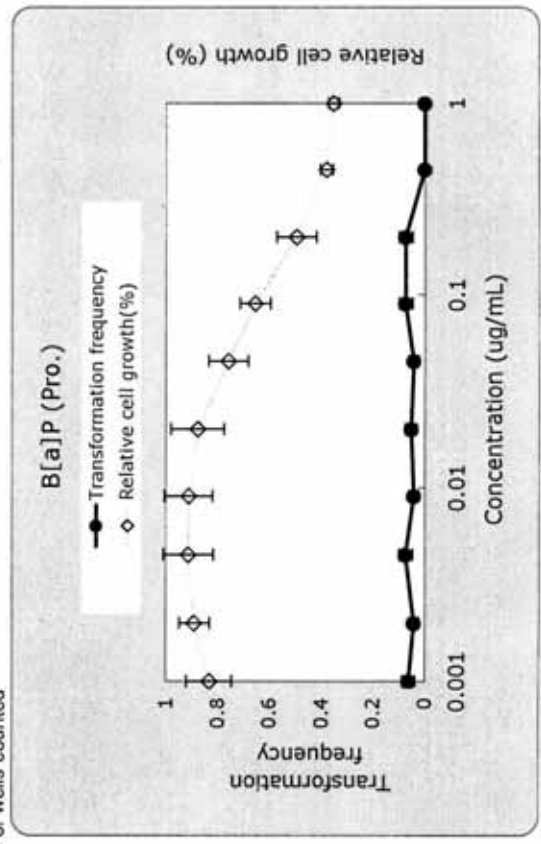
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	B[a]P
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)											
Crystal Violet Staining OD _{550nm}	Blank	0.1% DMSO											
		0.001	0.002	0.0045	0.009	0.02	0.045	0.09	0.2	0.45	1		
1	0.078	0.207	0.241	0.225	0.229	0.209	0.188	0.182	0.147	0.141	0.140	0.207	0.285
2	0.074	0.215	0.250	0.263	0.249	0.219	0.226	0.193	0.152	0.144	0.141	0.215	0.325
3	0.073	0.243	0.239	0.254	0.257	0.257	0.221	0.208	0.165	0.147	0.148	0.243	0.337
4	0.076	0.251	0.226	0.273	0.250	0.259	0.221	0.191	0.179	0.151	0.147	0.251	0.365
5	0.072	0.244	0.246	0.263	0.256	0.252	0.238	0.210	0.182	0.144	0.144	0.244	0.330
6	0.074	0.250	0.263	0.243	0.232	0.262	0.228	0.214	0.190	0.152	0.139	0.250	0.318
7	0.074	0.238	0.241	0.251	0.283	0.245	0.223	0.209	0.175	0.154	0.139	0.238	0.357
8	0.079	0.223	0.252	0.223	0.236	0.237	0.213	0.194	0.167	0.146	0.139	0.223	0.335
Average	0.075	0.234	0.245	0.249	0.249	0.242	0.220	0.200	0.169	0.147	0.142	0.234	0.332
SD	0.002	0.017	0.011	0.018	0.017	0.020	0.015	0.011	0.015	0.005	0.004	0.017	0.025
Average - Blank	0.000	0.159	0.170	0.174	0.174	0.167	0.145	0.125	0.094	0.072	0.067	0.159	0.257
Relative Cell Growth (%)		100.0	106.9	109.7	109.4	105.3	91.1	78.6	59.4	45.6	42.3	100.0	161.5
SD of Relative Cell Growth		10.5	6.9	11.4	11.0	12.3	9.2	7.2	9.2	2.9	2.4	10.5	15.5

Transformation Assay		Concentrations (ug/ml)												
Giemsa Staining	No. of wells excluded	0.1% DMSO												
		0.001	0.002	0.0045	0.009	0.02	0.045	0.09	0.2	0.45	1			
No. of wells counted	0	0	0	0	0	0	0	0	0	0	0	0	0	
No. of wells having foci	96	96	96	96	96	96	96	96	96	96	96	96	96	
Transformation frequency* SE	0.0625 0.02470529	0.041666667 0.02039469	0.072916667 0.02653610	0.041666667 0.02039469	0.052083333 0.02267769	0.041666667 0.02039469	0.072916667 0.02653610	0.041666667 0.02039469	0.072916667 0.02653610	0.041666667 0.02039469	0.072916667 0.02653610	0.041666667 0.02039469	0.0625 0.02470529	0.447916667 0.05075342

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

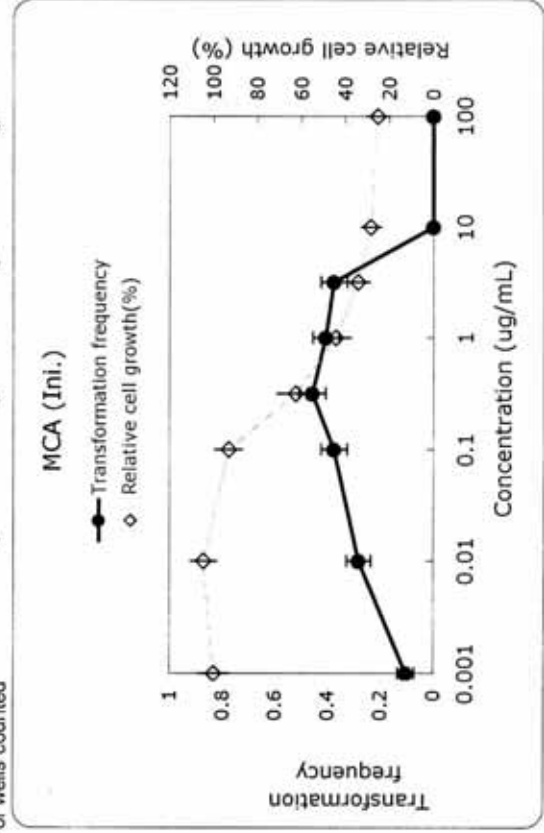
The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	MCA
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)							Negative Control	Positive Control	
Crystal Violet Staining		Blank	0.01	0.1	0.316	1	3.16	10	100	0.1% DMSO	MCA (1ug/mL)
1	OD _{540nm}	0.100	0.662	0.604	0.387	0.387	0.387	0.223	0.227	0.633	0.229
2		0.106	0.674	0.668	0.493	0.356	0.311	0.288	0.257	0.630	0.298
3		0.106	0.686	0.638	0.418	0.367	0.333	0.280	0.226	0.633	0.304
4		0.111	0.644	0.620	0.466	0.377	0.314	0.270	0.280	0.642	0.312
5		0.110	0.709	0.558	0.490	0.355	0.273	0.282	0.236	0.583	0.289
6		0.107	0.693	0.586	0.473	0.337	0.306	0.243	0.275	0.614	0.300
7		0.107	0.685	0.601	0.453	0.314	0.265	0.246	0.238	0.598	0.280
8		0.107	0.610	0.598	0.367	0.270	0.263	0.239	0.196	0.555	0.217
Average		0.107	0.670	0.609	0.443	0.345	0.291	0.259	0.242	0.611	0.279
SD		0.003	0.031	0.033	0.047	0.038	0.028	0.024	0.028	0.030	0.036
Average - Blank		0.000	0.564	0.502	0.337	0.239	0.184	0.152	0.135	0.504	0.172
Relative Cell Growth (%)			104.5	93.2	62.4	44.3	34.1	28.2	25.1	100.0	34.1
SD of Relative Cell Growth			7.0	6.2	8.8	7.1	5.2	4.5	5.2	5.6	6.6
										#DIV/0!	#DIV/0!
										#DIV/0!	#DIV/0!
										#DIV/0!	#DIV/0!
										#DIV/0!	#DIV/0!
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										#DIV/0!	#DIV/0!
										#DIV/0!	#DIV/0!

Transformation Assay		Concentrations (ug/ml)							Negative Control	Positive Control	
Giemsa Staining		0.001	0.01	0.1	0.316	1	3.16	10	100	0	0
No. of wells excluded		0	0	0	0	0	0	52	96	0	0
No. of wells counted		96	96	96	96	96	96	44	96	96	96
No. of wells having foci		10	27	36	44	39	36	TOX	TOX	6	43
Transformation frequency*		0.104166667	0.28125	0.375	0.458333333	0.40625	0.375	#VALUE!	#VALUE!	0	0
SE		0.03117758	0.04588805	0.04941059	0.05085354	0.05012598	0.04941059	#VALUE!	#VALUE!	0.00000000	0.00000000
* No. of wells having foci/ No. of wells counted										0.02470529	0.05075342



Acceptance Criteria (1)	
Negative Control < 15 and 40 < Positive Control	Pass
Acceptance Criteria (2)	
Concurrent Cell Growth Assay	Pass
The Final Data	

Remarks	
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Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	MCA
Institution:	Lab 2
Test Number	

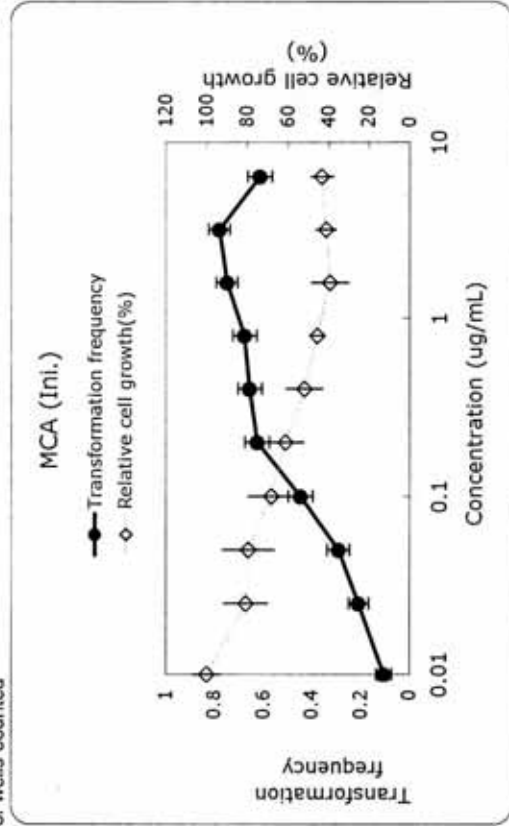
Cell Growth Assay

Crystal Violet Staining OD _{540nm}	Blank	Concentrations (ug/ml)										Negative Control		Positive Control	
		0.01	0.025	0.05	0.1	0.2	0.4	0.8	1.6	3.2	6.4	0.1% DMSO	MCA (1ug/mL)		
1	0.042	0.505	0.523	0.407	0.366	0.325	0.260	0.261	0.246	0.229	0.498	0.269			
2	0.046	0.439	0.420	0.432	0.347	0.308	0.270	0.220	0.223	0.245	0.524	0.299			
3	0.050	0.416	0.382	0.359	0.286	0.311	0.282	0.175	0.242	0.236	0.464	0.231			
4	0.041	0.441	0.409	0.273	0.384	0.250	0.256	0.302	0.247	0.245	0.519	0.294			
5	0.041	0.343	0.355	0.344	0.271	0.268	0.264	0.193	0.201	0.237	0.573	0.289			
6	0.048	0.385	0.368	0.334	0.339	0.219	0.240	0.196	0.278	0.244	0.532	0.301			
7	0.051	0.425	0.434	0.393	0.373	0.314	0.264	0.268	0.232	0.259	0.512	0.264			
8	0.046	0.475	0.495	0.415	0.319	0.342	0.258	0.241	0.259	0.310	0.538	0.291			
Average	0.046	0.429	0.423	0.370	0.336	0.292	0.262	0.232	0.241	0.251	0.520	0.280			
SD	0.004	0.050	0.060	0.052	0.041	0.042	0.012	0.044	0.023	0.026	0.032	0.024			
Average - Blank	0.000	0.383	0.378	0.324	0.290	0.247	0.216	0.186	0.195	0.205	0.474	0.234			
Relative Cell Growth (%)	100.0	80.7	79.6	68.3	61.1	52.0	45.6	39.3	41.2	43.2	100.0	49.4			
SD of Relative Cell Growth	6.7	10.6	12.6	11.1	8.6	8.9	2.5	9.2	4.9	5.4	6.7	5.0			

Transformation Assay

Giemsa Staining	Concentrations (ug/ml)										Negative Control		Positive Control	
No. of wells excluded	0.01	0.025	0.05	0.1	0.2	0.4	0.8	1.6	3.2	6.4	0.1% DMSO	MCA (1ug/mL)		
No. of wells counted	0	0	0	0	0	0	0	0	0	0	0	0		
No. of wells having foci	96	96	96	96	96	96	96	96	96	96	96	96		
Transformation frequency*	0.03117758	0.04144908	0.04639024	0.05075342	0.04941059	0.04847529	0.04772332	0.04419417	0.04219232	0.04967296	0.104166667	0.677083333		
SE	0.104166667	0.208333333	0.291666667	0.447916667	0.625	0.65625	0.677083333	0.75	0.78125	0.614583333	0.03117758	0.04772332		

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

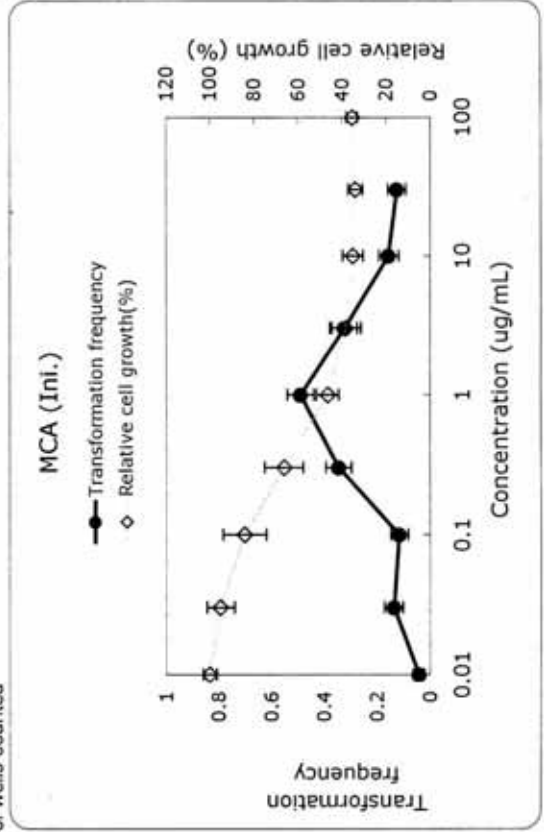
Chemical Code:	MCA
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control	Positive Control
			0.5% DMSO	0.03	0.1	0.3	1	3	10	30		
1	0.120	1.384	1.234	1.273	1.146	0.673	0.676	0.557	0.539	0.543	1.185	0.602
2	0.123	1.342	1.257	1.040	0.922	0.753	0.526	0.484	0.526	0.562	1.320	0.610
3	0.119	1.338	1.374	0.982	0.929	0.604	0.514	0.536	0.517	0.535	1.262	0.648
4	0.118	1.288	1.253	0.990	0.788	0.638	0.510	0.602	0.582	0.561	1.382	0.570
5	0.115	1.289	1.139	1.204	0.825	0.737	0.627	0.547	0.544	0.580	1.411	0.585
6	0.115	1.390	1.290	1.204	0.889	0.619	0.738	0.642	0.526	0.595	1.282	0.627
7	0.119	1.336	1.348	1.235	0.971	0.761	0.548	0.474	0.561	0.502	1.222	0.615
8	0.111	1.341	1.324	1.236	0.944	0.688	0.538	0.511	0.446	0.519	1.326	0.655
Average	0.118	1.339	1.277	1.146	0.927	0.684	0.585	0.544	0.530	0.550	1.299	0.614
SD	0.004	0.037	0.074	0.120	0.108	0.061	0.086	0.057	0.040	0.031	0.077	0.029
Average - Blank	0.000	1.221	1.160	1.028	0.809	0.567	0.467	0.427	0.413	0.432	1.181	0.497
Relative Cell Growth (%)		100.0	95.0	84.2	66.3	46.4	38.3	34.9	33.8	35.4	100.0	42.0
SD of Relative Cell Growth		3.1	6.1	9.9	8.8	5.0	7.0	4.7	3.3	2.5	6.3	2.4

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	0.01	0.03	0.1	0.3	1	3	10	30	100	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	4	13	11	33	47	31	15	12	tox	2	2	41
Transformation frequency*	0.041666667	0.135416667	0.114583333	0.34375	0.489583333	0.322916667	0.15625	0.125	#VALUE!	0	0.020833333	0.427083333
SE	0.02039469	0.03492238	0.03250965	0.04847529	0.05101996	0.04772332	0.03705794	0.03375386	#VALUE!	0.00000000	0.01457713	0.05048547

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

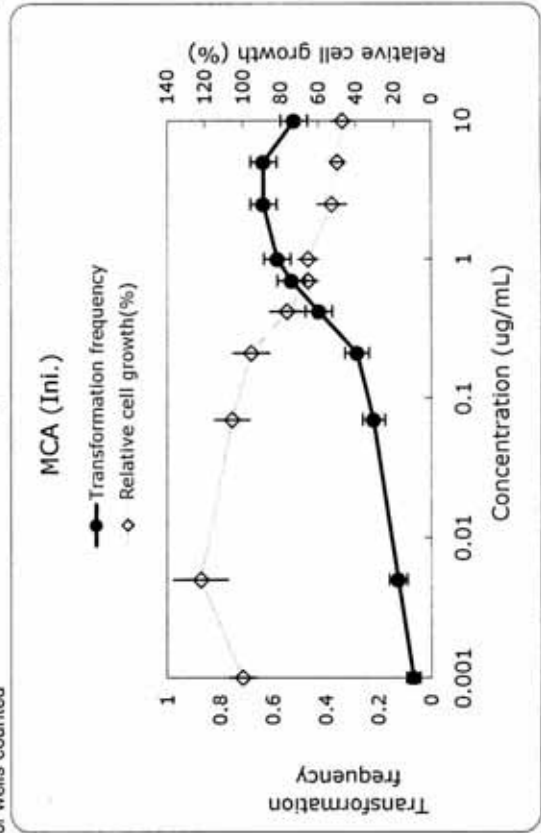
Chemical Code:	MCA
Institution:	Lab 4
Test Number	

Cell Growth Assay	Concentrations (ug/ml)											
	Blank	0.005	0.07	0.21	0.42	0.7	1	2.5	5	10	Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
Crystal Violet Staining												
OD _{540nm}												
1	0.072	0.265	0.272	0.259	0.270	0.207	0.206	0.180	0.174	0.187	0.293	0.203
2	0.071	0.262	0.278	0.272	0.286	0.210	0.208	0.184	0.180	0.164	0.331	0.199
3	0.074	0.279	0.324	0.252	0.283	0.206	0.206	0.193	0.167	0.156	0.333	0.201
4	0.075	0.283	0.335	0.288	0.258	0.183	0.181	0.167	0.170	0.173	0.300	0.183
5	0.073	0.273	0.315	0.276	0.248	0.209	0.205	0.186	0.167	0.163	0.310	0.206
6	0.074	0.272	0.288	0.303	0.250	0.207	0.207	0.173	0.171	0.148	0.287	0.190
7	0.075	0.237	0.346	0.297	0.236	0.195	0.201	0.181	0.165	0.169	0.269	0.178
8	0.076	0.272	0.329	0.283	0.242	0.205	0.190	0.146	0.171	0.160	0.293	0.170
Average	0.074	0.268	0.311	0.279	0.259	0.201	0.201	0.176	0.171	0.165	0.302	0.191
SD	0.001	0.014	0.028	0.018	0.019	0.010	0.010	0.015	0.005	0.012	0.022	0.013
Average - Blank	0.000	0.194	0.237	0.205	0.185	0.148	0.127	0.102	0.097	0.091	0.228	0.117
Relative Cell Growth (%)												
SD of Relative Cell Growth		100.0	122.2	105.6	95.5	65.4	65.3	52.8	50.0	47.1	100.0	51.4
		7.4	14.4	9.2	9.6	5.1	5.0	7.5	2.5	6.1	11.2	6.8

Transformation Assay	Concentrations (ug/ml)											
Giemsa Staining	0.001	0.005	0.07	0.21	0.42	0.7	1	2.5	5	10	Negative Control	Positive Control
No. of wells excluded	6	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	90	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	6	12	21	27	41	51	56	61	61	50	13	48
Transformation frequency*	0.066666667	0.125	0.21875	0.28125	0.427083333	0.53125	0.583333333	0.635416667	0.635416667	0.520833333	0.135416667	0.5
SE	0.02629369	0.03375386	0.04219232	0.04588805	0.05048547	0.05093127	0.05031728	0.04912382	0.04912382	0.05098672	0.03492238	0.05103104

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

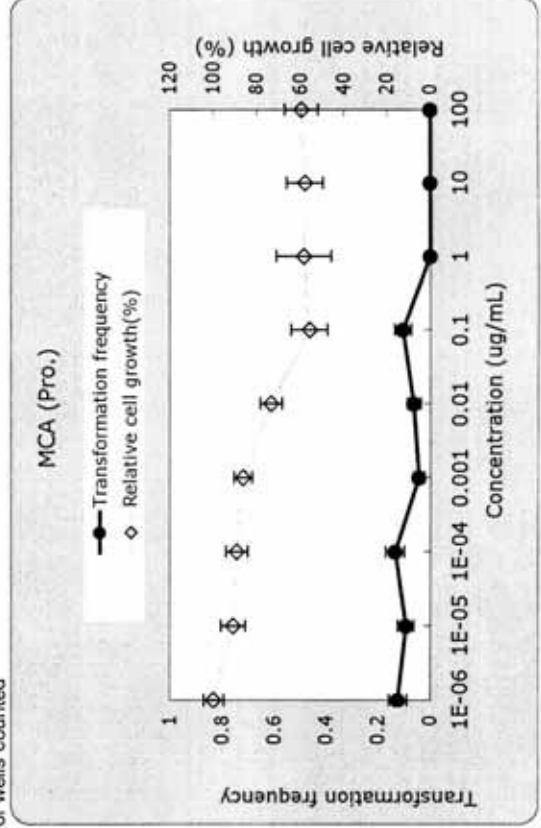
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	MCA
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)											Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)			
	Blank	0.000001	0.00001	0.0001	0.001	0.01	0.1	1	10	100	100					
Crystal Violet Staining OD _{540nm}																
1	0.122	0.624	0.591	0.547	0.577	0.514	0.302	0.560	0.508	0.502	0.675	0.991				
2	0.130	0.648	0.628	0.619	0.574	0.512	0.421	0.400	0.406	0.411	0.721	0.978				
3	0.121	0.587	0.568	0.546	0.513	0.483	0.413	0.390	0.379	0.404	0.660	0.982				
4	0.129	0.602	0.546	0.567	0.548	0.452	0.419	0.421	0.393	0.436	0.605	0.986				
5	0.127	0.586	0.553	0.565	0.541	0.479	0.385	0.371	0.403	0.391	0.643	0.986				
6	0.130	0.637	0.547	0.549	0.542	0.449	0.425	0.389	0.400	0.389	0.590	0.897				
7	0.124	0.620	0.557	0.555	0.544	0.499	0.420	0.373	0.399	0.392	0.644	0.908				
8	0.123	0.620	0.579	0.551	0.545	0.487	0.410	0.387	0.384	0.417	0.638	0.954				
Average	0.126	0.616	0.571	0.562	0.548	0.484	0.399	0.411	0.409	0.418	0.647	0.960				
SD	0.004	0.022	0.028	0.024	0.020	0.024	0.041	0.062	0.041	0.038	0.041	0.037				
Average - Blank	0.000	0.490	0.445	0.437	0.422	0.359	0.274	0.286	0.283	0.292	0.521	0.835				
Relative Cell Growth (%)																
SD of Relative Cell Growth		100.0	90.9	89.2	86.2	73.2	55.9	58.3	57.8	59.6	100.0	160.1				
		4.6	5.7	4.9	4.1	5.0	8.4	12.7	8.4	7.7	8.3	7.7				

Transformation Assay	Concentrations (ug/ml)											Negative Control	Positive Control
Giemsa Staining	0.000001	0.00001	0.0001	0.001	0.01	0.1	1	10	100	100	TOX		
No. of wells excluded	0	0	0	3	0	1	96	0	96	96	0	96	0
No. of wells counted	96	96	96	93	96	95	0	0	0	0	96	96	96
No. of wells having foci	12	9	13	4	6	10	TOX	TOX	TOX	TOX	11	56	56
Transformation frequency*	0.125	0.09375	0.135416667	0.043010753	0.0625	0.105263158	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.114583333	0.563333333	0.563333333
SE	0.03375386	0.02974911	0.03492238	0.02103781	0.02470529	0.03148648	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.03250865	0.05031728	0.05031728

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

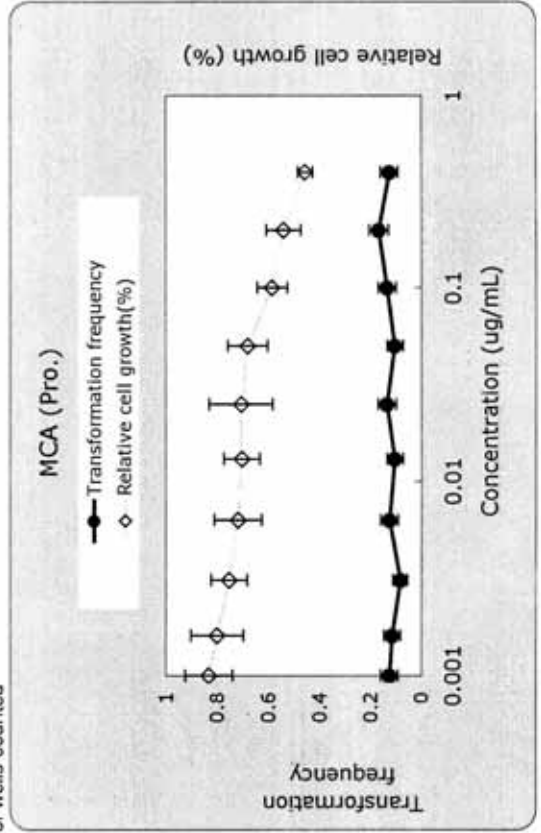
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	MCA
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control	Positive Control
			0.1% DMSO	0.0031	0.0063	0.013	0.025	0.05	0.1	0.2	0.4	0.1% DMSO		
1	0.035	0.456	0.413	0.462	0.391	0.410	0.392	0.371	0.320	0.328	0.280	0.456	0.567	
2	0.045	0.513	0.508	0.491	0.476	0.504	0.386	0.418	0.427	0.348	0.310	0.513	0.674	
3	0.040	0.536	0.532	0.516	0.557	0.440	0.495	0.445	0.396	0.385	0.292	0.536	0.698	
4	0.039	0.547	0.528	0.487	0.465	0.479	0.555	0.471	0.380	0.371	0.311	0.547	0.632	
5	0.041	0.587	0.530	0.495	0.420	0.464	0.450	0.472	0.378	0.382	0.320	0.587	0.646	
6	0.038	0.542	0.565	0.470	0.423	0.403	0.495	0.468	0.385	0.341	0.310	0.542	0.630	
7	0.044	0.513	0.481	0.438	0.452	0.436	0.423	0.418	0.363	0.324	0.301	0.513	0.614	
8	0.042	0.423	0.406	0.391	0.407	0.391	0.343	0.362	0.339	0.282	0.275	0.423	0.632	
Average	0.041	0.515	0.495	0.469	0.449	0.441	0.442	0.428	0.374	0.348	0.300	0.515	0.637	
SD	0.003	0.053	0.058	0.039	0.053	0.039	0.070	0.044	0.033	0.038	0.016	0.053	0.039	
Average - Blank	0.000	0.474	0.455	0.428	0.408	0.400	0.402	0.388	0.333	0.308	0.259	0.474	0.596	
Relative Cell Growth (%)		100.0	95.9	90.3	86.1	84.4	84.8	81.8	70.2	64.9	54.7	100.0	125.7	
SD of Relative Cell Growth		11.1	12.2	8.3	11.1	8.3	14.8	9.3	7.0	8.1	3.4	11.1	8.2	

Transformation Assay	Giemsa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	Transformation frequency*	SE	Concentrations (ug/ml)										Negative Control	Positive Control
							0.001	0.0016	0.0031	0.0063	0.013	0.025	0.05	0.1	0.2	0.4		
		0	96	0	0.000		0.001	0.0016	0.0031	0.0063	0.013	0.025	0.05	0.1	0.2	0.4	0.1% DMSO	TPA (50ng/mL)
		96	96	96	0.104166667		0	96	96	96	96	96	96	96	96	96	0	0
		12	11	8	0.083333333		12	12	12	10	13	13	10	13	16	12	12	12
		0.125	0.114583333	0.083333333	0.104166667		0.125	0.104166667	0.083333333	0.125	0.104166667	0.135416667	0.104166667	0.135416667	0.166666667	0.125	0.125	0.125
		0.03375386	0.03250865	0.02820847	0.03375386		0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code: MCA
Institution: Lab 3
Test Number

Cell Growth Assay Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
	Blank	0.0001	0.001	0.003	0.01	0.03	0.1	0.3	1	3		
1	0.110	1.064	0.989	1.005	1.006	0.845	0.816	0.689	0.622	0.668	1.196	1.852
2	0.104	1.034	1.039	0.980	0.898	0.865	0.869	0.667	0.668	0.723	1.329	1.772
3	0.110	1.058	1.046	0.917	0.850	0.879	0.736	0.675	0.677	0.683	1.341	1.709
4	0.128	1.083	0.932	1.069	0.907	0.853	0.839	0.722	0.707	0.698	1.219	1.925
5	0.123	1.072	1.052	0.993	0.882	0.911	0.791	0.638	0.711	0.724	1.234	1.779
6	0.124	1.012	1.048	1.006	0.997	0.960	0.862	0.728	0.713	0.745	1.239	1.809
7	0.146	1.102	1.074	1.016	0.942	0.910	0.936	0.692	0.778	0.742	1.240	1.807
8	0.141	1.105	1.023	0.997	0.945	0.961	0.739	0.638	0.735	0.705	1.358	1.798
Average	0.123	1.066	1.025	0.998	0.928	0.898	0.824	0.681	0.701	0.711	#DIV/0!	1.806
SD	0.015	0.032	0.045	0.042	0.055	0.045	0.068	0.034	0.047	0.027	#DIV/0!	0.063
Average - Blank	0.000	0.943	0.902	0.875	0.805	0.775	0.700	0.558	0.578	0.588	#DIV/0!	1.683
Relative Cell Growth (%)		100.0	95.7	92.7	85.4	82.2	74.3	59.2	61.3	62.3	#DIV/0!	100.0
SD of Relative Cell Growth		3.4	4.8	4.5	5.8	4.8	7.2	3.6	5.0	2.9	#DIV/0!	6.6

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	0.0001	0.001	0.003	0.01	0.03	0.1	0.3	1	3	0		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	8	7	5	3	1	2	0	0	0	0	6	59
Transformation frequency*	0.083333333	0.072916667	0.052083333	0.03125	0.010416667	0.020833333	0	0	0	0	0.0625	0.614583333
SE	0.02820847	0.02653610	0.02267769	0.01775805	0.01036227	0.01457713	0.00000000	0.00000000	0.00000000	0.00000000	0.02470529	0.04967296

* No. of wells having foci/ No. of wells counted

MCA (Pro.)

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Remarks

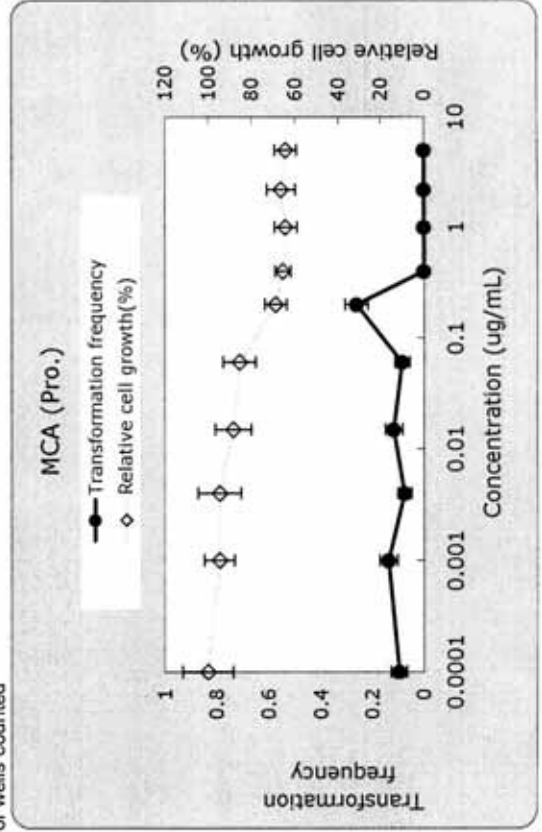
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	MCA
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)											
Crystal Violet Staining	Blank	0.5% DMSO	0.001	0.004	0.015	0.06	0.2	0.4	1	2.2	5	Negative Control	Positive Control
OD _{540nm}		0.0001										0.1% DMSO	TPA (50ng/mL)
1	0.062	0.262	0.251	0.244	0.219	0.228	0.204	0.194	0.185	0.184	0.182	0.281	0.423
2	0.065	0.262	0.267	0.242	0.257	0.248	0.214	0.215	0.202	0.208	0.209	0.300	0.469
3	0.065	0.260	0.271	0.294	0.262	0.247	0.210	0.210	0.204	0.228	0.209	0.318	0.520
4	0.063	0.272	0.262	0.265	0.267	0.243	0.219	0.214	0.208	0.228	0.208	0.324	0.498
5	0.066	0.325	0.281	0.293	0.265	0.260	0.223	0.205	0.223	0.208	0.211	0.318	0.477
6	0.071	0.305	0.287	0.288	0.272	0.272	0.216	0.221	0.210	0.221	0.211	0.333	0.483
7	0.063	0.301	0.288	0.292	0.280	0.278	0.242	0.212	0.216	0.207	0.221	0.311	0.474
8	0.061	0.312	0.297	0.285	0.271	0.265	0.216	0.214	0.215	0.216	0.212	0.310	0.466
Average	0.065	0.287	0.276	0.275	0.262	0.255	0.218	0.211	0.208	0.213	0.208	0.312	0.476
SD	0.003	0.026	0.015	0.022	0.019	0.017	0.011	0.008	0.011	0.014	0.011	0.016	0.028
Average - Blank	0.000	0.223	0.211	0.211	0.197	0.191	0.154	0.146	0.143	0.148	0.143	0.247	0.412
Relative Cell Growth (%)		100.0	94.7	94.6	88.4	85.5	68.9	65.6	64.3	66.4	64.3	100.0	166.4
SD of Relative Cell Growth		11.7	6.9	9.9	8.3	7.4	5.0	3.6	5.2	6.5	5.0	7.1	12.5

Transformation Assay		Concentrations (ug/ml)											
Glensa Staining	Blank	0.5% DMSO	0.001	0.004	0.015	0.06	0.2	0.4	1	2.2	5	Negative Control	Positive Control
No. of wells excluded		0.0001										0	0
No. of wells counted		95	96	96	96	96	96	0	0	96	96	96	96
No. of wells having foci		9	13	7	11	8	25					8	52
Transformation frequency*		0.094736842	0.135416667	0.072916667	0.114583333	0.083333333	0.260416667	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	0.083333333	0.541666667
SE		0.03004589	0.03492238	0.02853610	0.03250865	0.02820847	0.04479116	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	0.02820847	0.05085354

* No. of wells having foci/ No. of wells counted



Remarks	

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

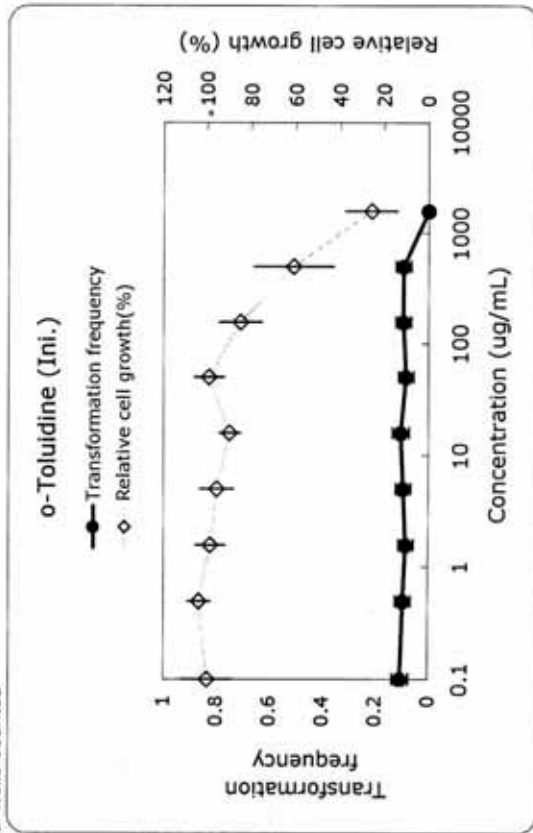
Chemical Code:	o-Toluidine
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/ml)	
	Blank	0.1	0.5	1.58	5	15.8	50	158	500	1580			
Crystal Violet Staining OD _{540nm}													
1	0.104	0.705	0.774	0.737	0.754	0.688	0.713	0.692	0.541	0.315	0.739	0.267	
2	0.115	0.872	0.785	0.792	0.718	0.674	0.745	0.660	0.499	0.255	0.701	0.297	
3	0.117	0.787	0.702	0.747	0.652	0.670	0.688	0.616	0.401	0.232	0.549	0.263	
4	0.114	0.709	0.738	0.685	0.679	0.632	0.659	0.677	0.401	0.272	0.628	0.284	
5	0.134	0.692	0.733	0.672	0.662	0.671	0.719	0.570	0.413	0.235	0.708	0.296	
6	0.141	0.652	0.718	0.690	0.716	0.618	0.780	0.654	0.394	0.235	0.606	0.295	
7	0.139	0.689	0.758	0.696	0.660	0.667	0.701	0.537	0.624	0.263	0.721	0.309	
8	0.144	0.706	0.785	0.720	0.770	0.707	0.765	0.685	0.661	0.443	0.781	0.323	
Average	0.126	0.727	0.749	0.717	0.701	0.666	0.721	0.636	0.492	0.281	0.679	0.292	
SD	0.015	0.070	0.031	0.040	0.045	0.029	0.040	0.057	0.107	0.071	0.077	0.020	
Average - Blank	0.000	0.601	0.623	0.591	0.575	0.540	0.595	0.510	0.366	0.155	0.553	0.166	
Relative Cell Growth (%)		100.0	103.8	98.5	95.8	89.9	99.1	85.0	60.9	25.9	100.0	30.0	
SD of Relative Cell Growth		11.6	5.2	6.6	7.5	4.8	6.7	9.5	17.9	11.8	12.9	3.3	

Transformation Assay

Transformation Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control
	Blank	0.1	0.5	1.58	5	15.8	50	158	500	1580		
Giemsa Staining												
No. of wells excluded	0	0	0	0	0	0	0	0	0	42	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	54	96	96
No. of wells having foci	10	9	8	8	9	10	8	9	9	TOX	8	44
Transformation frequency*	0.104166667	0.09375	0.083333333	0.083333333	0.09375	0.104166667	0.083333333	0.09375	0.09375	#VALUE!	0.083333333	0.458333333
SE	0.03117758	0.02974911	0.02820847	0.02974911	0.02974911	0.03117758	0.02820847	0.02974911	0.02974911	#VALUE!	0.02820847	0.05085354

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

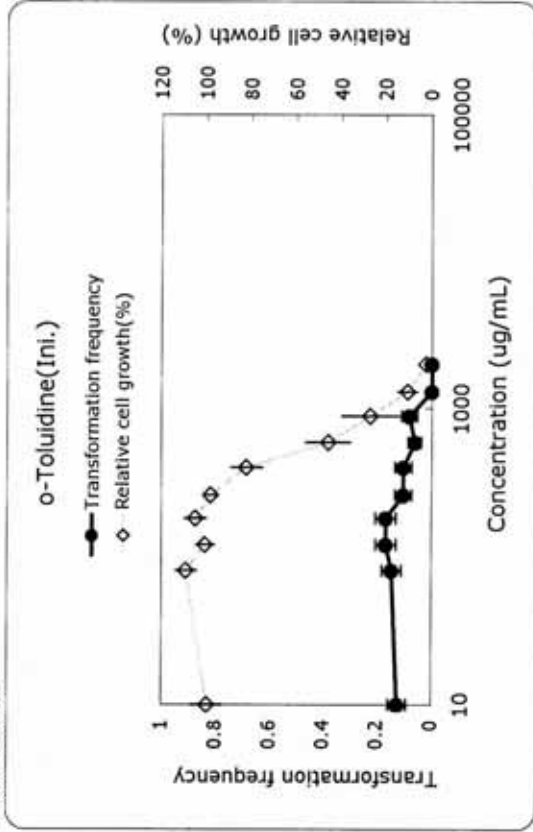
The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	o-Toluidine
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining		0.5% DMSO										0.1% DMSO		MCA (1ug/mL)	
OD _{540nm}		Blank	10	80	120	180	260	400	590	890	1300	2000			
1	0.054	0.510	0.539	0.530	0.572	0.511	0.521	0.380	0.217	0.142	0.074	0.543	0.328		
2	0.056	0.500	0.593	0.551	0.583	0.548	0.472	0.293	0.164	0.105	0.067	0.574	0.320		
3	0.053	0.515	0.592	0.572	0.568	0.546	0.448	0.272	0.144	0.091	0.063	0.596	0.372		
4	0.052	0.576	0.609	0.573	0.587	0.545	0.433	0.244	0.142	0.089	0.065	0.571	0.356		
5	0.052	0.552	0.603	0.561	0.591	0.541	0.432	0.237	0.142	0.088	0.061	0.597	0.396		
6	0.052	0.565	0.597	0.521	0.589	0.540	0.426	0.237	0.157	0.096	0.063	0.565	0.371		
7	0.053	0.596	0.608	0.549	0.541	0.542	0.437	0.253	0.196	0.104	0.061	0.575	0.291		
8	0.054	0.543	0.575	0.528	0.527	0.514	0.489	0.313	0.328	0.135	0.069	0.544	0.367		
Average	0.053	0.545	0.590	0.548	0.570	0.536	0.457	0.279	0.186	0.106	0.065	0.571	0.350		
SD	0.001	0.034	0.020	0.024	0.015	0.015	0.034	0.049	0.063	0.021	0.004	0.020	0.034		
Average - Blank	0.000	0.491	0.536	0.495	0.517	0.483	0.404	0.225	0.133	0.053	0.012	0.517	0.297		
Relative Cell Growth (%)		100.0	109.1	100.7	105.1	98.2	82.2	45.9	27.1	10.8	2.5	100.0	57.4		
SD of Relative Cell Growth		7.0	4.7	4.1	4.8	3.0	6.9	10.0	12.9	4.3	0.9	4.1	7.0		

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining		0.5% DMSO										0.1% DMSO		MCA (1ug/mL)	
No. of wells excluded		Blank	10	80	120	180	260	400	590	890	1300	2000			
No. of wells counted	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells having foci	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96
Transformation frequency*	12	14	16	16	16	10	10	6	8	0	0	0	10	10	68
SE	0.125	0.145833333	0.166666667	0.166666667	0.166666667	0.104166667	0.104166667	0.0625	0.083333333	0	0	0	0.104166667	0.104166667	0.708333333
* No. of wells having foci/ No. of wells counted	0.03375386	0.03602169	0.03803629	0.03803629	0.03803629	0.03117758	0.03117758	0.02470529	0.02820847	0.00000000	0.00000000	0.00000000	0.03117758	0.03117758	0.04639024



Remarks

Acceptance Criteria (1)

Negative Control < 15 and 40 < Positive Control

Pass

Acceptance Criteria (2)

Concurrent Cell Growth Assay

Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

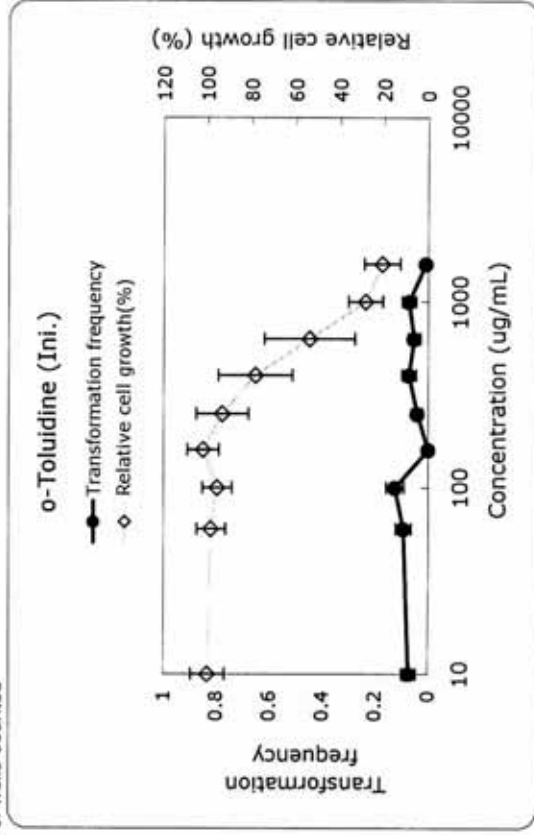
Chemical Code:	o-Toluidine
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining		OD _{590nm}										0.1% DMSO		MCA (1ug/mL)	
Blank		10	60	100	160	250	400	630	1000	1600					
1	0.106	1.205	1.256	1.174	1.198	1.328	1.217	1.005	0.478	0.383	0.918	0.379			
2	0.102	1.058	1.230	1.091	1.219	1.172	1.115	0.698	0.436	0.271	0.660	0.363			
3	0.107	1.139	1.139	1.128	1.131	1.088	0.780	0.662	0.320	0.287	0.774	0.294			
4	0.106	1.116	1.044	0.972	1.055	1.013	0.820	0.472	0.364	0.247	0.556	0.293			
5	0.105	1.153	1.108	1.179	1.190	0.911	0.758	0.406	0.233	0.329	0.536	0.329			
6	0.099	1.151	1.121	1.075	1.134	1.035	0.789	0.536	0.339	0.324	0.673	0.339			
7	0.108	1.299	1.144	1.112	1.203	1.039	0.903	0.628	0.394	0.371	0.884	0.398			
8	0.108	1.227	1.116	1.180	1.316	1.142	1.070	0.967	0.556	0.486	1.067	0.442			
Average	0.105	1.160	1.145	1.114	1.181	1.091	0.932	0.672	0.403	0.325	0.759	0.355			
SD	0.003	0.081	0.068	0.070	0.077	0.125	0.178	0.217	0.082	0.085	0.187	0.051			
Average - Blank	0.000	1.055	1.040	1.009	1.076	0.986	0.826	0.567	0.298	0.220	0.653	0.250			
Relative Cell Growth (%)		100.0	98.6	95.6	102.0	93.5	78.3	53.7	28.2	20.9	100.0	38.2			
SD of Relative Cell Growth		7.7	6.5	6.7	7.3	11.9	16.8	20.6	7.8	8.0	17.7	4.9			

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining		10	60	100	160	250	400	630	1000	1600					
No. of wells excluded		0	0	0	0	0	0	0	0	0	0	0	0	0	
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96	96	96	
No. of wells having foci		7	9	12	0	4	7	5	7	1	5	5	5	66	
Transformation frequency*		0.072916667	0.09375	0.125	0	0.041666667	0.072916667	0.052083333	0.072916667	0.010416667	0.052083333	0.052083333	0.052083333	0.6875	
SE		0.02653610	0.02974911	0.03375386	0.00000000	0.02039469	0.02653610	0.02267769	0.02653610	0.01036227	0.02267769	0.02267769	0.02267769	0.04730704	

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

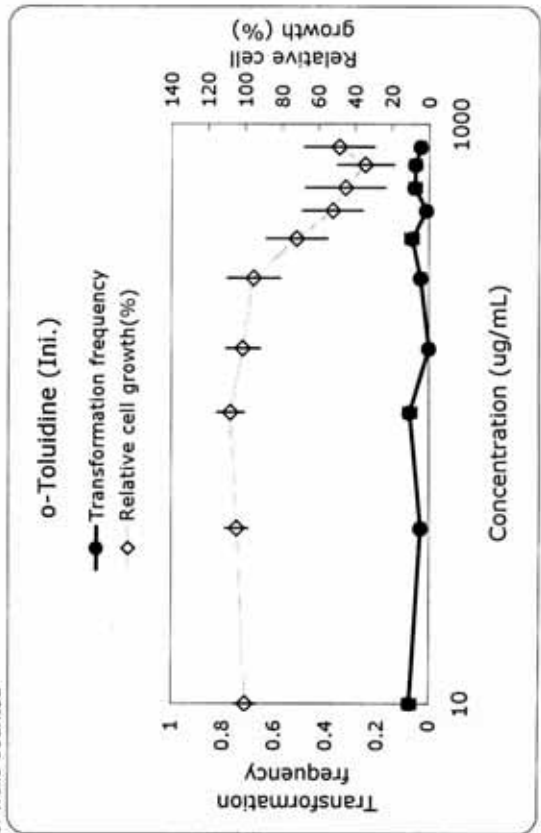
Chemical Code:	o-Toluidine
Institution:	Lab 4
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)	
	Blank	10	40	100	166.67	291.67	400	500	600	720			833.33
Crystal Violet Staining													
OD _{540nm}													
1	0.062	0.275	0.295	0.304	0.323	0.285	0.269	0.236	0.232	0.182	0.256	0.275	0.139
2	0.065	0.280	0.301	0.296	0.312	0.291	0.230	0.163	0.164	0.132	0.172	0.280	0.110
3	0.065	0.282	0.320	0.292	0.289	0.273	0.182	0.144	0.121	0.121	0.179	0.282	0.123
4	0.063	0.301	0.296	0.310	0.272	0.246	0.198	0.158	0.124	0.114	0.128	0.301	0.116
5	0.066	0.310	0.317	0.325	0.269	0.251	0.196	0.159	0.133	0.118	0.136	0.310	0.122
6	0.071	0.300	0.297	0.303	0.320	0.269	0.226	0.172	0.134	0.118	0.159	0.300	0.117
7	0.063	0.304	0.286	0.324	0.303	0.302	0.239	0.201	0.177	0.154	0.155	0.304	0.132
8	0.061	0.300	0.323	0.342	0.285	0.350	0.294	0.243	0.254	0.210	0.224	0.300	0.146
Average	0.065	0.294	0.304	0.312	0.297	0.283	0.229	0.185	0.169	0.144	0.176	0.294	0.126
SD	0.003	0.013	0.014	0.017	0.021	0.033	0.038	0.038	0.050	0.036	0.044	0.013	0.012
Average - Blank	0.000	0.230	0.240	0.248	0.232	0.219	0.165	0.120	0.104	0.079	0.112	0.230	0.061
Relative Cell Growth (%)		100.0	104.5	107.8	101.1	95.4	71.8	52.3	45.4	34.5	48.6	100.0	26.6
SD of Relative Cell Growth		5.7	6.0	7.4	9.1	14.4	16.6	16.4	21.6	15.5	19.0	5.7	5.4

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	10	40	100	166.67	291.67	400	500	600	720			833.33
Giemsa Staining													
No. of wells excluded	0	0	0	0	0	0	4	0	3	0	0	0	0
No. of wells counted	96	96	96	96	96	96	92	96	93	96	96	96	96
No. of wells having foci	7	3	7	7	3	3	6	1	5	5	3	7	50
Transformation frequency*	0.072916667	0.03125	0.072916667	0.072916667	0.03125	0.065217391	0.010416667	0.053763441	0.052083333	0.03125	0.03125	0.072916667	0.520833333
SE	0.02653610	0.01775805	0.02653610	0.02653610	0.01775805	0.02574205	0.01036227	0.02338848	0.02267769	0.01775805	0.01775805	0.02653610	0.05098672

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

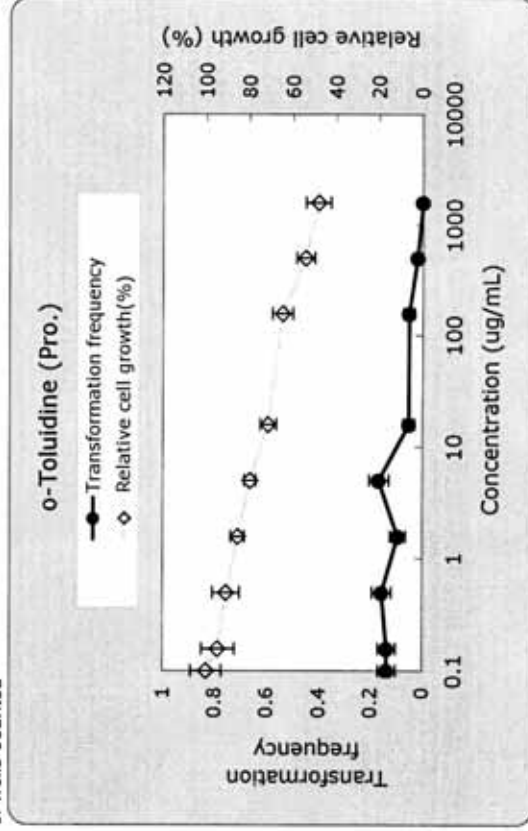
Chemical Code:	o-Toluidine
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Crystal Violet Staining												0.1% DMSO	TPA (50ng/mL)
OD _{590nm}	Blank	0.1	0.158	0.5	1.58	5	15.8	50	158	500	1580		
1	0.121	0.508	0.486	0.493	0.428	0.408	0.362	0.334	0.300	0.314	0.300	0.363	0.378
2	0.128	0.505	0.485	0.429	0.421	0.399	0.355	0.342	0.279	0.304	0.279	0.358	0.382
3	0.120	0.506	0.475	0.421	0.421	0.390	0.364	0.325	0.273	0.292	0.273	0.352	0.383
4	0.117	0.494	0.478	0.441	0.430	0.400	0.376	0.354	0.283	0.301	0.283	0.350	0.383
5	0.109	0.455	0.443	0.442	0.420	0.403	0.389	0.364	0.279	0.316	0.279	0.341	0.374
6	0.109	0.452	0.432	0.431	0.439	0.414	0.378	0.361	0.285	0.318	0.285	0.353	0.368
7	0.106	0.454	0.411	0.450	0.410	0.409	0.384	0.349	0.290	0.311	0.290	0.357	0.364
8	0.111	0.461	0.463	0.461	0.445	0.421	0.393	0.379	0.340	0.341	0.340	0.363	0.394
Average	0.115	0.479	0.459	0.446	0.427	0.406	0.375	0.351	0.291	0.312	0.291	0.355	0.378
SD	0.008	0.026	0.028	0.023	0.011	0.010	0.014	0.017	0.021	0.015	0.021	0.007	0.010
Average - Blank	0.000	0.364	0.344	0.331	0.312	0.290	0.260	0.236	0.176	0.197	0.176	0.240	0.263
Relative Cell Growth (%)	100.0	94.4	90.8	85.6	79.7	71.4	64.8	54.1	48.3	54.1	48.3	100.0	109.9
SD of Relative Cell Growth	7.1	7.6	6.3	3.1	2.6	3.7	4.8	4.0	5.9	4.0	5.9	2.0	2.6

Transformation Assay

Transformation Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining												0.1% DMSO	TPA (50ng/mL)
No. of wells excluded	Blank	0.1	0.158	0.5	1.58	5	15.8	50	158	500	1580		
No. of wells counted	0	0	0	0	0	0	0	0	0	0	53	0	0
No. of wells having foci	96	96	96	96	96	96	96	96	96	96	43	96	96
Transformation frequency*	0.135416667	0.135416667	0.15625	0.09375	0.166666667	0.052083333	0.052083333	0.020833333	0.020833333	0.020833333	0.027083333	0.09375	0.510416667
SE	0.03492238	0.03492238	0.03705794	0.02974911	0.03803629	0.02267769	0.02267769	0.01457713	0.01457713	0.01457713	0.02267769	0.02974911	0.05101996

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)

Negative Control < 20 and 40 < Positive Control

Pass

Acceptance Criteria (2)

Concurrent Cell Growth Assay

Pass

The Final Data

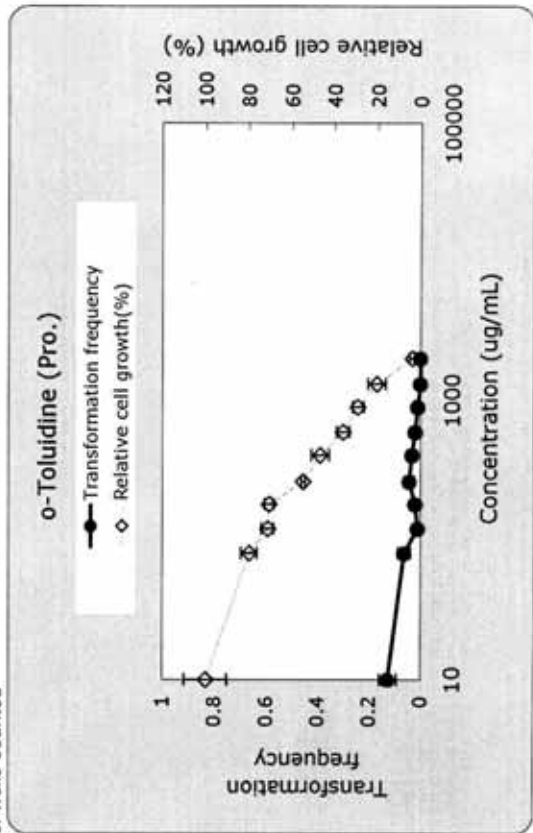
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	o-Toluidine
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining	Blank	10	80	120	180	260	400	590	890	1300	2000	0.1% DMSO	TPA (50ng/mL)		
OD _{540nm}															
1	0.054	0.404	0.367	0.330	0.354	0.279	0.263	0.220	0.187	0.162	0.080	0.450	0.609		
2	0.056	0.471	0.397	0.354	0.352	0.283	0.257	0.207	0.173	0.142	0.068	0.547	0.728		
3	0.053	0.455	0.405	0.357	0.352	0.278	0.265	0.193	0.164	0.125	0.063	0.556	0.829		
4	0.052	0.497	0.398	0.363	0.344	0.287	0.256	0.191	0.165	0.121	0.066	0.560	0.794		
5	0.052	0.532	0.390	0.356	0.350	0.277	0.250	0.187	0.160	0.118	0.064	0.654	0.765		
6	0.052	0.502	0.384	0.354	0.352	0.280	0.230	0.199	0.170	0.129	0.065	0.599	0.818		
7	0.053	0.459	0.375	0.339	0.338	0.276	0.222	0.200	0.179	0.136	0.064	0.530	0.703		
8	0.054	0.429	0.362	0.337	0.330	0.281	0.229	0.219	0.189	0.161	0.073	0.514	0.640		
Average	0.053	0.469	0.385	0.349	0.347	0.280	0.247	0.202	0.173	0.137	0.068	0.551	0.736		
SD	0.001	0.041	0.016	0.012	0.009	0.004	0.017	0.012	0.011	0.017	0.006	0.060	0.081		
Average - Blank	0.000	0.415	0.332	0.296	0.293	0.227	0.193	0.149	0.120	0.084	0.015	0.498	0.683		
Relative Cell Growth (%)		100.0	79.8	71.1	70.6	54.6	46.5	35.8	28.9	20.1	3.5	100.0	137.0		
SD of Relative Cell Growth		10.0	3.7	2.8	2.1	0.9	4.1	3.0	2.6	4.1	1.4	14.4	19.5		

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Transformation frequency*	SE	10	80	120	180	260	400	590	890	1300	2000	0.1% DMSO	TPA (50ng/mL)		
No. of wells excluded		0	0	0	0	0	0	0	0	0	0	0	0		
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96	96		
No. of wells having foci		12	6	1	2	4	3	2	1	0	0	8	36		
Transformation frequency*		0.125	0.0625	0.010416667	0.020833333	0.041666667	0.03125	0.020833333	0.010416667	0	0	0.083333333	0.375		
SE		0.03375386	0.02470529	0.01036227	0.01457713	0.02039469	0.01775805	0.01457713	0.01036227	0.00000000	0.00000000	0.02820947	0.04941059		

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

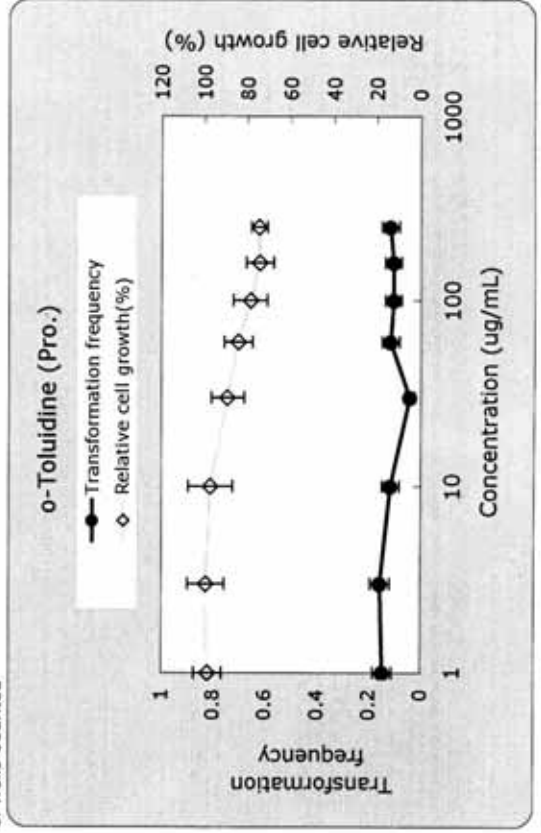
Chemical Code:	o-Toluidine
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Crystal Violet Staining		0.5% DMSO										0.1% DMSO	TPA (50ng/mL)
OD _{550nm}	Blank	0.1	1	3	10	30	60	100	160	250	250	TPA (50ng/mL)	
1	0.106	0.891	0.917	0.996	0.956	0.842	0.770	0.703	0.704	0.678	0.764	1.310	
2	0.102	0.906	0.896	0.897	0.836	0.828	0.772	0.692	0.671	0.696	0.780	1.221	
3	0.107	0.881	0.877	0.859	0.838	0.787	0.730	0.709	0.662	0.690	0.718	1.158	
4	0.106	0.886	0.807	0.812	0.728	0.692	0.677	0.643	0.643	0.631	0.668	1.053	
5	0.105	0.870	0.801	0.816	0.759	0.734	0.718	0.651	0.645	0.631	0.691	1.181	
6	0.099	0.830	0.782	0.789	0.826	0.744	0.700	0.678	0.623	0.687	0.736	1.163	
7	0.108	0.787	0.877	0.865	0.901	0.786	0.767	0.722	0.645	0.672	0.785	1.242	
8	0.108	0.877	0.873	0.849	0.924	0.856	0.833	0.834	0.776	0.704	0.809	1.232	
Average	0.105	0.866	0.854	0.860	0.846	0.784	0.746	0.704	0.671	0.674	0.744	1.195	
SD	0.003	0.039	0.050	0.065	0.079	0.057	0.050	0.059	0.049	0.028	0.049	0.076	
Average - Blank	0.000	0.761	0.749	0.755	0.741	0.679	0.641	0.599	0.566	0.569	0.639	1.090	
Relative Cell Growth (%)		100.0	98.4	99.3	97.4	89.2	84.2	78.7	74.4	74.7	100.0	170.6	
SD of Relative Cell Growth		5.1	6.5	8.5	10.3	7.5	6.5	7.8	6.4	3.7	6.5	10.0	

Transformation Assay

Transformation Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining		0.1										0.1% DMSO	TPA (50ng/mL)
No. of wells excluded	No. of wells counted	0.1	1	3	10	30	60	100	160	250	250	TPA (50ng/mL)	
0	96	0	0	0	0	0	0	0	0	0	0	0	
13	14	15	11	4	11	11	10	10	10	11	5	46	
Transformation frequency*	0.135416667	0.145833333	0.15625	0.114583333	0.041666667	0.114583333	0.104166667	0.104166667	0.104166667	0.114583333	0.052083333	0.479166667	
SE	0.03492238	0.03602169	0.03705794	0.03250865	0.02039469	0.03250865	0.03117758	0.03117758	0.03117758	0.03250865	0.02267769	0.05098672	

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

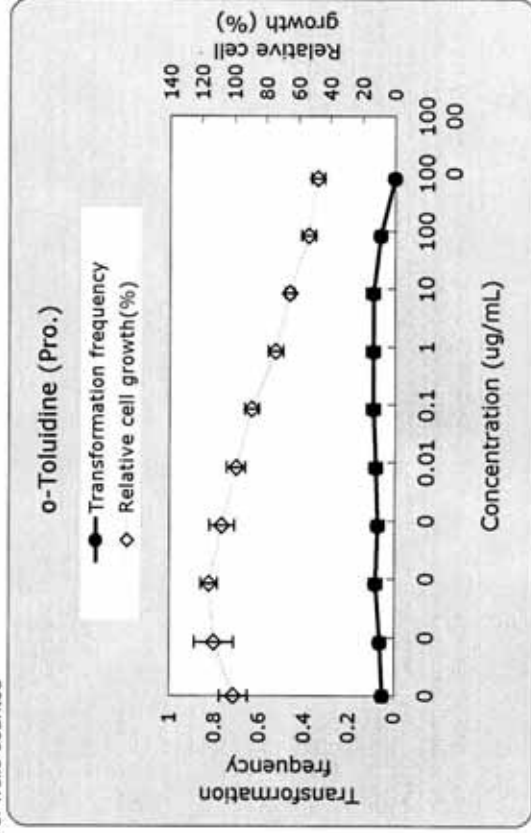
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	o-Toluidine
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining	Blank	0.1% DMSO	8.33E-06	0.0000833	0.000833	0.00833	0.0833	0.833	8.33	83.33	833.33	833.33	0.1% DMSO	TPA (50ng/mL)	
1	0.078	0.225	0.243	0.269	0.240	0.244	0.223	0.211	0.190	0.184	0.164	0.164	0.225	0.223	
2	0.074	0.242	0.244	0.276	0.250	0.261	0.229	0.199	0.183	0.161	0.160	0.160	0.242	0.244	
3	0.073	0.241	0.268	0.271	0.260	0.244	0.234	0.207	0.185	0.171	0.159	0.159	0.241	0.228	
4	0.076	0.237	0.270	0.275	0.269	0.254	0.235	0.198	0.194	0.167	0.151	0.151	0.237	0.221	
5	0.072	0.247	0.285	0.278	0.285	0.238	0.222	0.215	0.182	0.168	0.150	0.150	0.247	0.223	
6	0.074	0.263	0.307	0.294	0.264	0.254	0.216	0.203	0.186	0.158	0.148	0.148	0.263	0.226	
7	0.074	0.272	0.277	0.272	0.263	0.234	0.237	0.189	0.192	0.169	0.168	0.168	0.272	0.234	
8	0.079	0.257	0.260	0.263	0.257	0.235	0.234	0.203	0.193	0.165	0.163	0.163	0.257	0.233	
Average	0.075	0.248	0.269	0.275	0.261	0.245	0.229	0.203	0.188	0.168	0.158	0.158	0.248	0.229	
SD	0.002	0.015	0.021	0.009	0.013	0.010	0.008	0.008	0.005	0.008	0.007	0.007	0.015	0.008	
Average - Blank	0.000	0.173	0.194	0.200	0.186	0.170	0.154	0.128	0.113	0.093	0.083	0.083	0.173	0.154	
Relative Cell Growth (%)		100.0	112.3	115.4	107.3	98.5	88.9	74.0	65.3	53.6	47.8	47.8	100.0	89.0	
SD of Relative Cell Growth		8.7	12.3	5.2	7.7	5.7	4.4	4.7	2.8	4.4	4.3	4.3	8.7	4.4	

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining	Blank	0.1% DMSO	8.33E-06	0.0000833	0.000833	0.00833	0.0833	0.833	8.33	83.33	833.33	833.33	0.1% DMSO	TPA (50ng/mL)	
No. of wells excluded		0	0	0	0	0	0	0	0	0	0	0	0	0	
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96	96	96	
No. of wells having foci		5	6	8	7	8	9	9	9	6	0	0	5	50	
Transformation frequency*		0.052083333	0.0625	0.083333333	0.072916667	0.083333333	0.09375	0.09375	0.09375	0.0625	0	0	0.052083333	0.520833333	
SE		0.02267769	0.02470529	0.02820847	0.02853610	0.02820847	0.02974911	0.02974911	0.02974911	0.02470529	0.00000000	0.00000000	0.02267769	0.05098672	

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

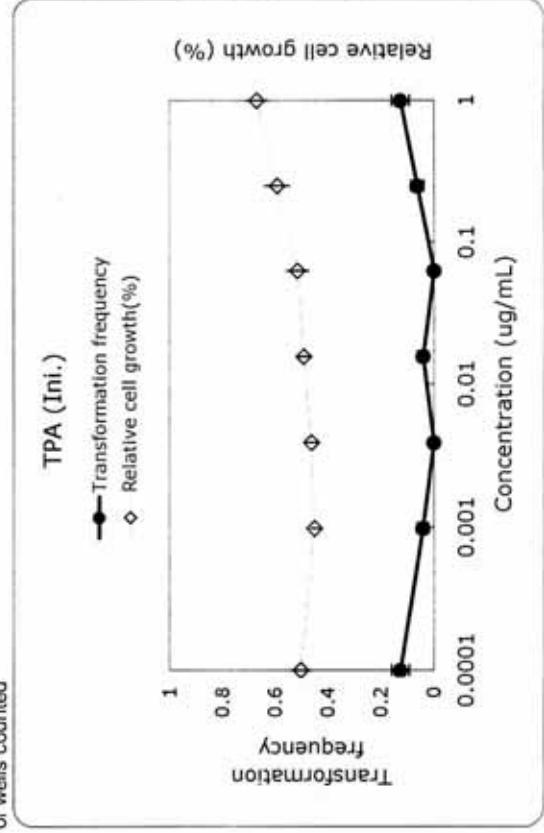
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	TPA
Institution:	Lab 1
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/ml)		
		0.0001	0.000977	0.00391	0.0156	0.0625	0.25	0.958	1.088	1.088	1.088				
Crystal Violet Staining															
OD _{590nm}															
1	0.095	0.765	0.719	0.709	0.720	0.789	0.958	1.088	0.765	0.763	0.763	0.409			
2	0.097	0.722	0.660	0.683	0.747	0.807	0.869	0.975	0.722	0.050	0.050	0.519			
3	0.095	0.838	0.705	0.654	0.783	0.774	0.976	1.025	0.838	#DIV/0!	#DIV/0!	0.451			
4	0.091	0.746	0.679	0.708	0.729	0.761	0.887	1.025	0.746	#DIV/0!	#DIV/0!	0.449			
5	0.083	0.773	0.684	0.683	0.752	0.762	0.892	1.036	0.773	#DIV/0!	#DIV/0!	0.343			
6	0.088	0.745	0.692	0.740	0.770	0.681	0.848	0.993	0.745	#DIV/0!	#DIV/0!	0.395			
7	0.087	0.827	0.724	0.713	0.734	0.870	0.775	0.950	0.827	#DIV/0!	#DIV/0!	0.347			
8	0.094	0.688	0.698	0.731	0.802	0.829	0.908	0.928	0.688	#DIV/0!	#DIV/0!	0.317			
Average	0.091	0.763	0.695	0.711	0.751	0.784	0.889	0.992	0.763	#DIV/0!	#DIV/0!	0.404			
SD	0.005	0.050	0.021	0.032	0.031	0.056	0.063	0.055	0.050	#DIV/0!	#DIV/0!	0.068			
Average - Blank	0.000	0.672	0.604	0.620	0.659	0.693	0.798	0.900	0.672	#DIV/0!	#DIV/0!	0.313			
Relative Cell Growth (%)															
SD of Relative Cell Growth															
		100.0	89.9	92.3	98.2	103.1	118.8	134.0	100.0	#DIV/0!	#DIV/0!	46.5			
		7.5	3.2	4.7	4.6	8.3	9.4	8.2	7.5	#DIV/0!	#DIV/0!	10.1			

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	0.0001	0.000977	0.00391	0.0156	0.0625	0.25	0.958	1.088	1.088	1.088			
Giemsa Staining													
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	12	4	0	4	0	0	6	12	12	12	12	54	54
Transformation frequency*	0.125	0.041666667	0	0.041666667	0	0.0625	0.125	0.125	0.125	0.125	0.125	0.5625	0.5625
SE	0.03375386	0.02039469	0.00000000	0.02039469	0.00000000	0.02470529	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.05063079	0.05063079

* No. of wells having foci/ No. of wells counted



TPA (Ini.)

● Transformation frequency
◇ Relative cell growth(%)

Acceptance Criteria (1)

Negative Control < 15 and 40 < Positive Control

Pass

Acceptance Criteria (2)

Concurrent Cell Growth Assay

Pass

The Final Data

Remarks

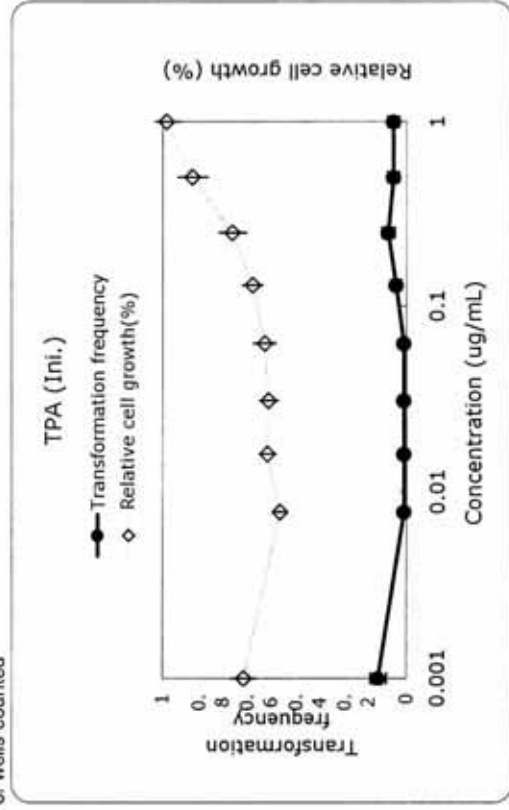
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	TPA
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining	Blank	0.001	0.016	0.031	0.063	0.13	0.25	0.5	1	0.1% DMSO	MCA (1ug/mL)	0.1% DMSO	MCA (1ug/mL)		
1	0.049	0.447	0.433	0.436	0.457	0.482	0.533	0.657	0.684	0.532	0.315	0.532	0.315		
2	0.052	0.430	0.441	0.469	0.450	0.488	0.531	0.648	0.717	0.581	0.303	0.581	0.303		
3	0.050	0.410	0.452	0.466	0.424	0.544	0.590	0.752	0.772	0.537	0.273	0.537	0.273		
4	0.059	0.408	0.481	0.444	0.518	0.547	0.518	0.643	0.774	0.498	0.301	0.498	0.301		
5	0.068	0.420	0.450	0.483	0.505	0.519	0.532	0.637	0.786	0.517	0.314	0.517	0.314		
6	0.080	0.466	0.486	0.479	0.486	0.490	0.619	0.689	0.780	0.552	0.261	0.552	0.261		
7	0.082	0.421	0.498	0.476	0.465	0.486	0.613	0.726	0.774	0.554	0.295	0.554	0.295		
8	0.080	0.404	0.450	0.405	0.453	0.476	0.545	0.656	0.714	0.466	0.325	0.466	0.325		
Average	0.065	0.426	0.461	0.457	0.470	0.504	0.561	0.676	0.750	0.530	0.298	0.530	0.298		
SD	0.014	0.021	0.024	0.027	0.031	0.029	0.040	0.042	0.039	0.036	0.022	0.036	0.022		
Average - Blank	0.000	0.361	0.396	0.392	0.405	0.439	0.496	0.611	0.685	0.465	0.233	0.465	0.233		
Relative Cell Growth (%)		77.6	85.3	84.4	87.1	94.5	106.8	131.5	147.5	100.0	50.2	100.0	50.2		
SD of Relative Cell Growth		4.6	5.1	5.8	6.7	6.2	8.5	9.1	8.4	7.7	4.7	7.7	4.7		

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining	Blank	0.001	0.016	0.031	0.063	0.13	0.25	0.5	1	0.1% DMSO	MCA (1ug/mL)	0.1% DMSO	MCA (1ug/mL)		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0		
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96		
No. of wells having foci	11	1	1	1	1	4	7	5	5	11	66	11	66		
Transformation frequency*	0.114583333	0.010416667	0.010416667	0.010416667	0.010416667	0.041666667	0.072916667	0.052083333	0.052083333	0.114583333	0.6875	0.032508865	0.04730704		
SE	0.032508865	0.01036227	0.01036227	0.01036227	0.01036227	0.02039469	0.02653610	0.02267769	0.02267769	0.032508865	0.04730704	0.032508865	0.04730704		

*No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

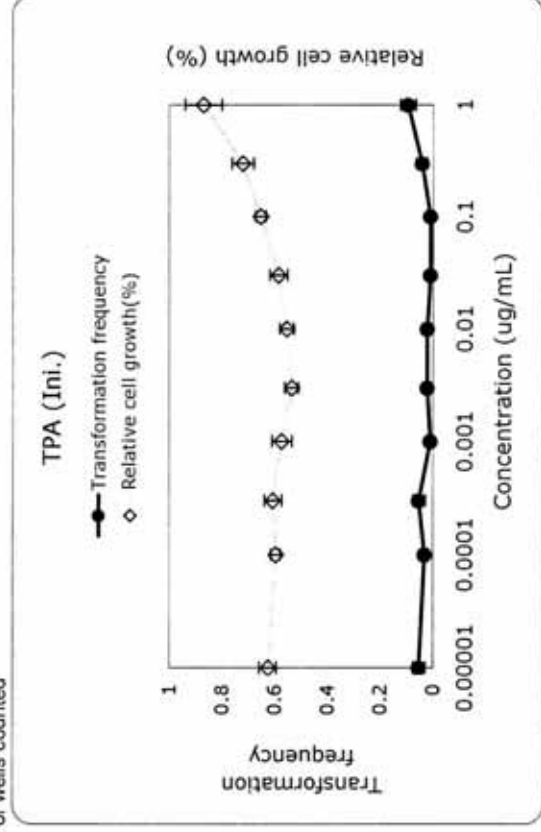
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	TPA
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control	Positive Control	
Crystal Violet Staining		0.00001	0.00003	0.0001	0.0003	0.001	0.003	0.01	0.03	0.1	0.3	1	0.1% DMSO	MCA (1ug/mL)
1	Blank	0.078	0.913	0.964	0.872	0.885	0.872	0.892	0.942	0.973	1.113	1.423	0.934	0.507
2		0.060	0.861	0.880	0.838	0.851	0.838	0.836	0.911	0.995	1.054	1.369	0.862	0.469
3		0.048	0.914	0.838	0.787	0.877	0.787	0.774	0.879	0.988	0.984	1.139	0.950	0.480
4		0.053	0.911	0.905	0.803	0.866	0.803	0.823	0.833	0.933	1.007	1.251	0.885	0.432
5		0.051	0.856	0.851	0.768	0.823	0.768	0.827	0.818	0.961	1.149	1.233	0.978	0.446
6		0.055	0.900	0.901	0.813	0.933	0.813	0.793	0.813	0.926	1.050	1.198	0.954	0.477
7		0.064	0.893	0.957	0.766	0.751	0.766	0.842	0.898	0.995	1.022	1.205	0.982	0.499
8		0.073	0.859	0.927	0.794	0.865	0.794	0.862	0.900	0.973	1.104	1.344	0.887	0.552
Average		0.060	0.888	0.903	0.805	0.856	0.805	0.831	0.873	0.968	1.060	1.270	0.929	0.483
SD		0.011	0.026	0.046	0.036	0.053	0.036	0.037	0.047	0.027	0.057	0.098	0.045	0.037
Average - Blank		0.000	0.828	0.843	0.745	0.796	0.745	0.771	0.813	0.908	1.000	1.210	0.869	0.423
Relative Cell Growth (%)			95.3	97.0	85.7	91.6	85.7	88.7	93.6	104.5	115.1	139.3	100.0	48.6
SD of Relative Cell Growth			2.9	5.3	4.1	6.1	4.3	4.3	5.4	3.1	6.6	11.3	5.2	4.3

Transformation Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining		0.00001	0.0001	0.0003	0.001	0.003	0.01	0.03	0.1	0.3	1	0.1% DMSO	MCA (1ug/mL)
No. of wells excluded		0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci		5	3	5	1	2	2	1	1	4	9	5	53
Transformation frequency*		0.052083333	0.03125	0.052083333	0.010416667	0.020833333	0.020833333	0.010416667	0.010416667	0.041666667	0.09375	0.052083333	0.552083333
SE		0.02267769	0.01775805	0.02267769	0.01036227	0.01457713	0.01457713	0.01036227	0.01036227	0.02039469	0.02974911	0.02267769	0.05075342

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

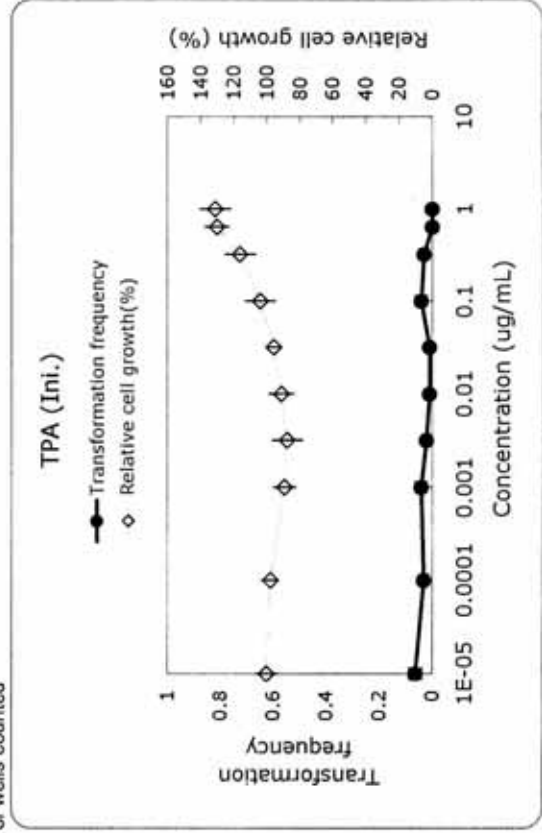
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	TPA
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)											
Crystal Violet Staining OD _{550nm}	Blank	0.1% DMSO											
		0.00001	0.0001	0.001	0.0032	0.01	0.032	0.1	0.32	0.64	1	Negative Control 0.1% DMSO	Positive Control MCA (1ug/ml)
1	0.078	0.259	0.276	0.250	0.277	0.279	0.266	0.291	0.354	0.348	0.345	0.259	0.137
2	0.074	0.290	0.270	0.281	0.268	0.262	0.275	0.276	0.326	0.367	0.358	0.290	0.175
3	0.073	0.300	0.290	0.268	0.254	0.270	0.280	0.311	0.309	0.345	0.329	0.300	0.166
4	0.076	0.298	0.284	0.282	0.246	0.250	0.289	0.283	0.318	0.374	0.356	0.298	0.177
5	0.072	0.290	0.294	0.279	0.277	0.295	0.285	0.293	0.314	0.340	0.392	0.290	0.163
6	0.074	0.285	0.294	0.246	0.249	0.277	0.268	0.278	0.292	0.366	0.348	0.285	0.164
7	0.074	0.298	0.293	0.260	0.290	0.250	0.285	0.325	0.343	0.330	0.375	0.298	0.177
8	0.079	0.292	0.272	0.260	0.234	0.277	0.292	0.318	0.327	0.356	0.341	0.292	0.144
Average	0.075	0.289	0.284	0.266	0.262	0.270	0.280	0.297	0.323	0.353	0.356	0.289	0.163
SD	0.002	0.013	0.010	0.014	0.019	0.016	0.009	0.019	0.019	0.015	0.020	0.013	0.015
Average - Blank	0.000	0.214	0.209	0.191	0.187	0.195	0.205	0.222	0.248	0.278	0.281	0.214	0.088
Relative Cell Growth (%)		100.0	97.7	89.1	87.4	91.1	95.7	103.8	115.9	130.0	131.1	100.0	41.0
SD of Relative Cell Growth		6.2	4.7	6.6	8.9	7.3	4.4	8.8	9.1	7.1	9.4	6.2	7.0

Transformation Assay		Concentrations (ug/ml)											
Giemsa Staining	No. of wells excluded	0.1% DMSO											
		0.00001	0.0001	0.001	0.0032	0.01	0.032	0.1	0.32	0.64	1	Negative Control 0.1% DMSO	Positive Control
No. of wells counted	0	0	0	0	0	0	0	0	0	0	96	0	0
No. of wells having foci	6	96	96	96	96	96	96	96	96	96	0	96	96
Transformation frequency* SE	0.0625 0.02470529	0.03125 0.01775805	0.041666667 0.02039469	0.020833333 0.01457713	0.010416667 0.01036227	0.010416667 0.01036227	0.041666667 0.01036227	0.041666667 0.02039469	0.03125 0.01775805	0.03125 0.01775805	#DIV/0! #DIV/0!	0.0625 0.02470529	0.447916667 0.05075342

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

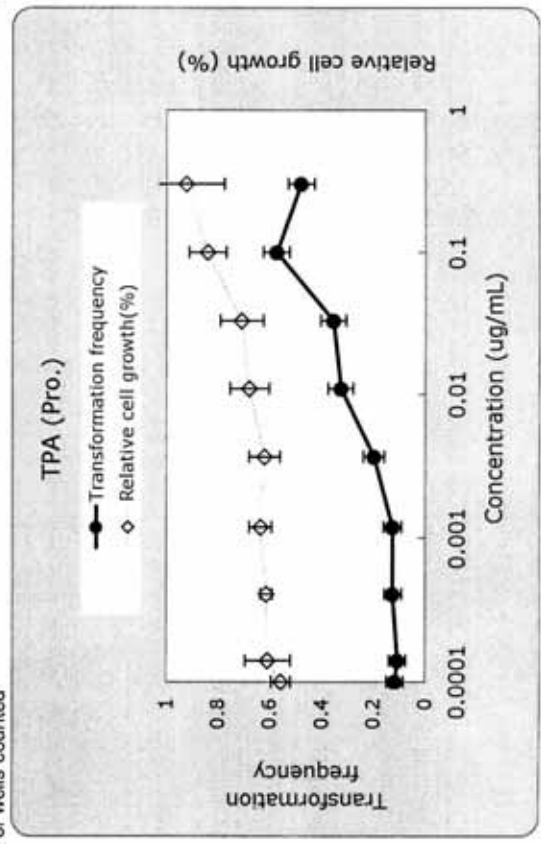
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	TPA
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)											
Crystal Violet Staining OD _{550nm}	Blank	0.1% DMSO										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
		0.0001	0.00014	0.00041	0.0012	0.0037	0.011	0.033	0.1	0.3	0.570		
1	0.029	0.325	0.321	0.337	0.365	0.346	0.376	0.356	0.514	0.570	0.325	0.370	
2	0.027	0.330	0.360	0.357	0.389	0.395	0.370	0.418	0.480	0.602	0.330	0.480	
3	0.030	0.317	0.299	0.360	0.352	0.328	0.418	0.436	0.493	0.585	0.317	0.456	
4	0.031	0.286	0.442	0.359	0.388	0.393	0.388	0.423	0.456	0.443	0.286	0.468	
5	0.030	0.300	0.316	0.352	0.360	0.353	0.453	0.456	0.466	0.468	0.300	0.339	
6	0.028	0.334	0.372	0.364	0.382	0.382	0.399	0.413	0.497	0.586	0.334	0.331	
7	0.034	0.338	0.363	0.354	0.322	0.331	0.362	0.385	0.468	0.443	0.338	0.347	
8	0.033	0.341	0.315	0.331	0.350	0.317	0.322	0.325	0.391	0.421	0.341	0.356	
Average	0.030	0.321	0.349	0.352	0.364	0.356	0.386	0.402	0.471	0.515	0.321	0.353	
SD	0.002	0.019	0.046	0.012	0.023	0.031	0.039	0.043	0.037	0.077	0.019	0.022	
Average - Blank	0.000	0.291	0.318	0.322	0.333	0.325	0.356	0.371	0.440	0.485	0.291	0.323	
Relative Cell Growth (%)		100.0	109.3	110.4	114.5	111.8	122.2	127.5	151.3	166.4	100.0	110.9	
SD of Relative Cell Growth		6.7	15.9	4.0	7.8	10.6	13.4	14.9	12.8	26.6	6.7	7.6	

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control					
Giemsa Staining	No. of wells excluded	0.0001										0.1		0					
		1	95	10	12	12	12	12	12	12	12	12	12	12	1	96	96	96	
Transformation frequency*		0.115789474	0.105263158	0.125	0.126315789	0.197916667	0.326315789	0.354166667	0.574468085	0.479166667	0	0	0	0	0.115789474	0.03282847	0.05099588	0.05099588	0.05099588
SE		0.03282847	0.03148648	0.03375386	0.03408351	0.04066449	0.04810447	0.04881221	0.05099588	0.05099588	0.00000000	0.00000000	0.00000000	0.00000000	0.03282847	0.03282847	0.03282847	0.03282847	0.05099588

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

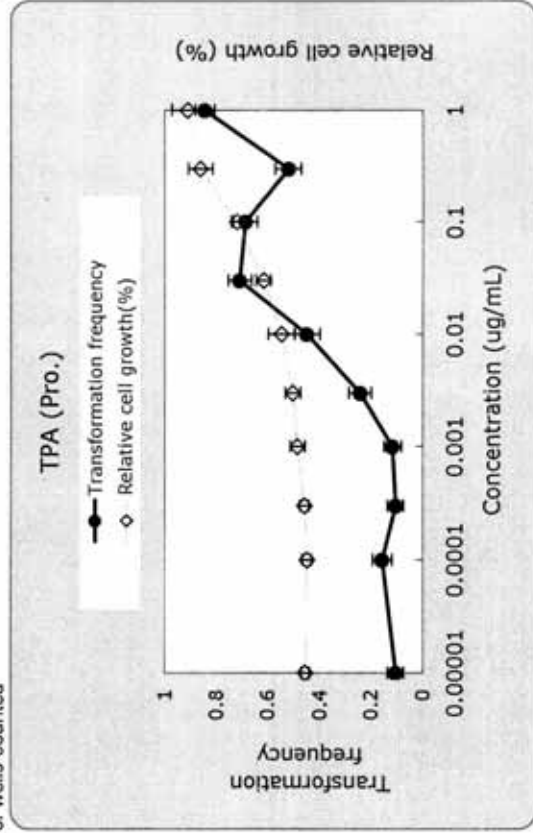
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	TPA
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)													
	Blank	0.00001	0.00003	0.0001	0.0003	0.001	0.003	0.01	0.03	0.1	0.3	1	Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
Crystal Violet Staining OD _{540nm}														
1	0.095	0.998	0.959	0.962	1.100	0.992	1.100	1.211	1.259	1.445	1.859	1.944	0.998	1.358
2	0.098	0.985	0.942	0.925	1.096	1.055	1.096	1.283	1.218	1.442	1.819	1.971	0.985	1.248
3	0.101	0.943	1.018	0.960	1.031	1.002	1.031	1.094	1.299	1.509	1.768	1.854	0.943	1.421
4	0.104	0.971	0.969	0.913	1.055	1.028	1.055	1.205	1.319	1.428	1.678	1.788	0.971	1.332
5	0.106	0.932	0.972	0.993	1.068	1.068	1.068	1.099	1.209	1.472	1.609	1.658	0.932	1.319
6	0.104	0.902	0.910	0.945	1.071	0.958	1.071	1.096	1.330	1.439	1.679	1.795	0.902	1.288
7	0.106	0.939	0.934	0.943	0.956	1.058	0.956	1.020	1.189	1.481	1.657	1.805	0.939	1.429
8	0.108	0.957	0.972	0.895	0.966	0.915	0.966	1.005	1.227	1.364	1.668	1.647	0.957	1.290
Average	0.103	0.953	0.960	0.942	1.043	1.010	1.043	1.127	1.256	1.448	1.717	1.808	0.953	1.336
SD	0.004	0.031	0.032	0.031	0.055	0.054	0.055	0.098	0.054	0.043	0.088	0.117	0.031	0.064
Average - Blank	0.000	0.851	0.857	0.839	0.940	0.907	0.940	1.024	1.154	1.345	1.614	1.705	0.851	1.233
Relative Cell Growth (%)	100.0	98.7	100.7	106.6	110.5	106.6	110.5	120.4	135.6	158.1	189.8	200.4	100.0	144.9
SD of Relative Cell Growth	3.6	3.6	3.8	3.6	6.5	6.3	6.5	11.5	6.3	5.1	10.3	13.8	3.6	7.5

Transformation Assay	Concentrations (ug/ml)											
Giemsa Staining	0.00001	0.0001	0.0003	0.001	0.003	0.01	0.03	0.1	0.3	1	Negative Control	Positive Control
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	10	15	10	11	23	43	68	66	50	81	10	75
Transformation frequency*	0.104166667	0.15625	0.104166667	0.114583333	0.239583333	0.447916667	0.708333333	0.6875	0.520833333	0.84375	0.104166667	0.78125
SE	0.03117758	0.03705794	0.03117758	0.03250865	0.04356307	0.05075342	0.04639024	0.04730704	0.05098672	0.03705794	0.03117758	0.04219232

*No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

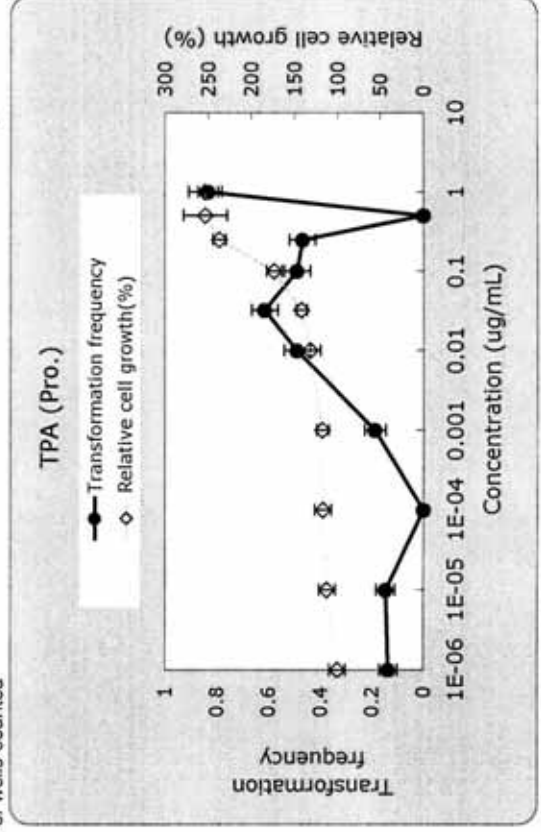
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	TPA
Institution:	Lab 4
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)										Negative Control	Positive Control			
		0.1% DMSO	0.00001	0.0001	0.001	0.01	0.032	0.1	0.25	0.5	1			0.1% DMSO	TPA (50ng/mL)	
Crystal Violet Staining																
OD _{540nm}																
1	0.077	0.275	0.344	0.333	0.312	0.362	0.382	0.456	0.603	0.595	0.645	0.275	0.359			
2	0.096	0.283	0.300	0.326	0.337	0.328	0.365	0.458	0.598	0.623	0.620	0.283	0.374			
3	0.088	0.299	0.309	0.338	0.334	0.367	0.389	0.449	0.573	0.572	0.559	0.299	0.404			
4	0.085	0.326	0.342	0.366	0.320	0.353	0.396	0.495	0.607	0.615	0.691	0.326	0.380			
5	0.086	0.308	0.319	0.305	0.350	0.398	0.373	0.449	0.587	0.556	0.604	0.308	0.412			
6	0.078	0.324	0.347	0.350	0.334	0.340	0.407	0.441	0.564	0.633	0.603	0.324	0.416			
7	0.082	0.288	0.326	0.345	0.359	0.374	0.389	0.460	0.586	0.700	0.661	0.288	0.420			
8	0.097	0.283	0.302	0.309	0.335	0.398	0.395	0.428	0.602	0.698	0.608	0.283	0.351			
Average	0.086	0.298	0.324	0.334	0.335	0.365	0.387	0.454	0.590	0.624	0.624	0.298	0.389			
SD	0.007	0.020	0.019	0.021	0.015	0.025	0.013	0.019	0.015	0.053	0.041	0.020	0.027			
Average - Blank	0.000	0.212	0.238	0.248	0.249	0.279	0.301	0.368	0.504	0.538	0.538	0.212	0.303			
Relative Cell Growth (%)		100.0	112.0	116.9	117.4	131.5	141.8	173.7	237.4	253.5	253.5	100.0	143.0			
SD of Relative Cell Growth		9.2	9.0	9.7	7.1	11.8	6.3	9.2	7.3	24.9	19.2	9.2	12.8			

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	0.000001	0.00001	0.0001	0.001	0.01	0.032	0.1	0.25	0.5	1			0.00001
Giemsa Staining													
No. of wells excluded	0	96	0	5	0	0	0	0	0	96	0	0	0
No. of wells counted	96	96	0	91	96	96	96	96	96	0	96	96	96
No. of wells having foci	13	14	17	17	47	59	47	45	45	80	13	54	54
Transformation frequency*	0.135416667	0.145833333	#DIV/0!	0.186813187	0.489583333	0.614583333	0.489583333	0.46875	0.46875	#DIV/0!	0.833333333	0.135416667	0.5625
SE	0.03492238	0.03602169	#DIV/0!	0.04085811	0.05101996	0.04967296	0.05101996	0.05093127	0.05093127	#DIV/0!	0.03803629	0.03492238	0.05063079

*No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

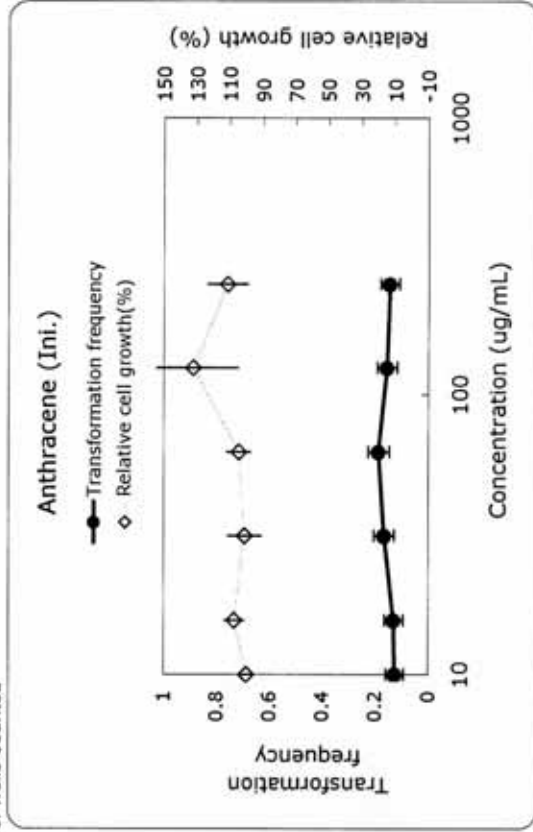
Chemical Code:	Anthracene
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)							
Crystal Violet Staining OD _{540nm}	Blank	0.5% DMSO							
		10	15.6	31.3	62.5	125	250	Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
1	0.099	0.686	0.698	0.652	0.660	0.624	0.700	0.631	0.213
2	0.105	0.639	0.688	0.716	0.774	1.065	0.803	0.684	0.285
3	0.096	0.665	0.728	0.669	0.671	0.982	0.720	0.742	0.311
4	0.100	0.710	0.783	0.618	0.681	0.906	0.847	0.653	0.309
5	0.102	0.674	0.704	0.754	0.746	0.910	0.714	0.609	0.346
6	0.098	0.717	0.762	0.768	0.757	0.867	0.788	0.718	0.353
7	0.099	0.690	0.750	0.729	0.710	0.983	0.639	0.687	0.285
8	0.100	0.702	0.711	0.627	0.703	0.656	0.812	0.616	0.173
Average	0.100	0.685	0.728	0.692	0.713	0.874	0.753	0.668	0.284
SD	0.003	0.026	0.034	0.058	0.042	0.157	0.070	0.048	0.062
Average - Blank	0.000	0.586	0.628	0.592	0.613	0.774	0.653	0.568	0.185
Relative Cell Growth (%)		100.0	107.3	101.1	104.7	132.2	111.5	100.0	32.5
SD of Relative Cell Growth		4.4	5.8	9.9	7.2	26.8	12.0	8.3	10.7

Transformation Assay		Concentrations (ug/ml)							
Giemsa Staining	No. of wells excluded	0.5% DMSO							
		10	15.6	31.3	62.5	125	250	Negative Control 0.1% DMSO	Positive Control
	0	3	0	0	0	0	0	0	0
No. of wells counted	96	93	96	96	96	96	96	96	96
No. of wells having foci	12	12	16	18	15	14	8	8	47
Transformation frequency*	0.125	0.129032258	0.166666667	0.1875	0.15625	0.145833333	0.083333333	0.083333333	0.489583333
SE	0.03375386	0.03476232	0.03803629	0.03983609	0.03705794	0.03602169	0.02820947	0.02820947	0.05101996

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

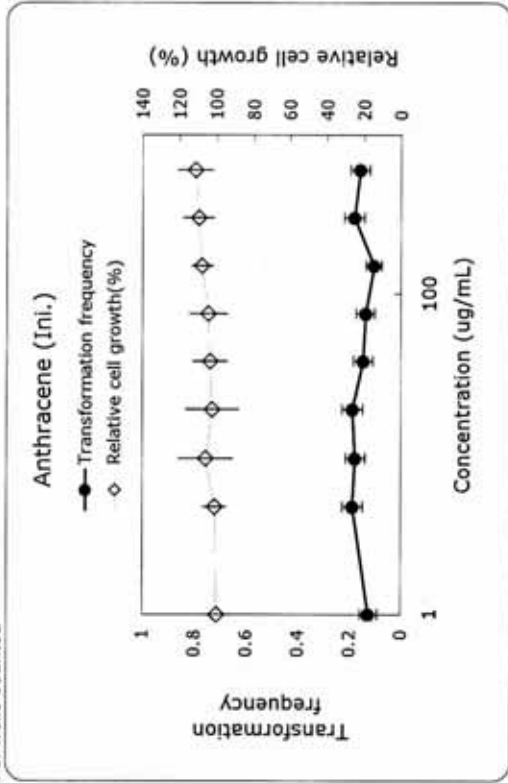
Chemical Code:	Anthracene
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control	Positive Control
			0.5% DMSO	4.7	9.4	19	38	75	150	300	600	0.1% DMSO		
OD _{550nm}			1	0.316	0.346	0.373	0.331	0.294	0.334	0.325	0.343	0.271	0.177	
1	0.028	0.028	0.316	0.346	0.373	0.331	0.294	0.334	0.325	0.343	0.271	0.177		
2	0.030	0.030	0.287	0.389	0.355	0.343	0.359	0.362	0.356	0.362	0.294	0.174		
3	0.026	0.026	0.307	0.326	0.284	0.302	0.311	0.299	0.299	0.345	0.272	0.180		
4	0.025	0.025	0.309	0.341	0.332	0.340	0.311	0.343	0.307	0.309	0.293	0.157		
5	0.032	0.032	0.305	0.280	0.288	0.282	0.284	0.309	0.363	0.304	0.302	0.174		
6	0.027	0.027	0.317	0.299	0.265	0.278	0.291	0.320	0.320	0.321	0.282	0.184		
7	0.040	0.040	0.330	0.304	0.280	0.309	0.351	0.314	0.337	0.318	0.307	0.212		
8	0.033	0.033	0.278	0.271	0.301	0.319	0.324	0.329	0.331	0.373	0.239	0.164		
Average	0.030	0.030	0.306	0.320	0.310	0.313	0.316	0.325	0.330	0.334	0.283	0.178		
SD	0.005	0.005	0.017	0.039	0.039	0.025	0.028	0.015	0.022	0.025	0.022	0.016		
Average - Blank	0.000	0.000	0.276	0.289	0.280	0.283	0.286	0.295	0.300	0.304	0.252	0.148		
Relative Cell Growth (%)			100.0	105.9	102.3	103.5	104.5	107.8	109.6	111.3	100.0	58.5		
SD of Relative Cell Growth			7.5	6.1	14.2	9.1	10.1	5.7	8.1	9.2	8.0	6.0		

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glensa Staining	1	4.7	9.4	19	38	75	150	300	600	0		
No. of wells excluded	0	0	0	0	0	1	0	2	1	0	0	0
No. of wells counted	96	96	96	96	96	95	96	94	95	96	96	96
No. of wells having foci	12	18	17	18	14	13	10	17	15	10	10	58
Transformation frequency*	0.125	0.1875	0.17708333	0.1875	0.14583333	0.13684211	0.10416667	0.18085106	0.15789474	0	0.104166667	0.604166667
SE	0.03375386	0.03983609	0.03896110	0.03983609	0.03602169	0.03526088	0.03117758	0.03969886	0.03741151	0.00000000	0.03117758	0.04991131

*No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass or Failure

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass or Failure

This Data is ...

Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Anthracene
Institution:	Lab 3
Test Number	

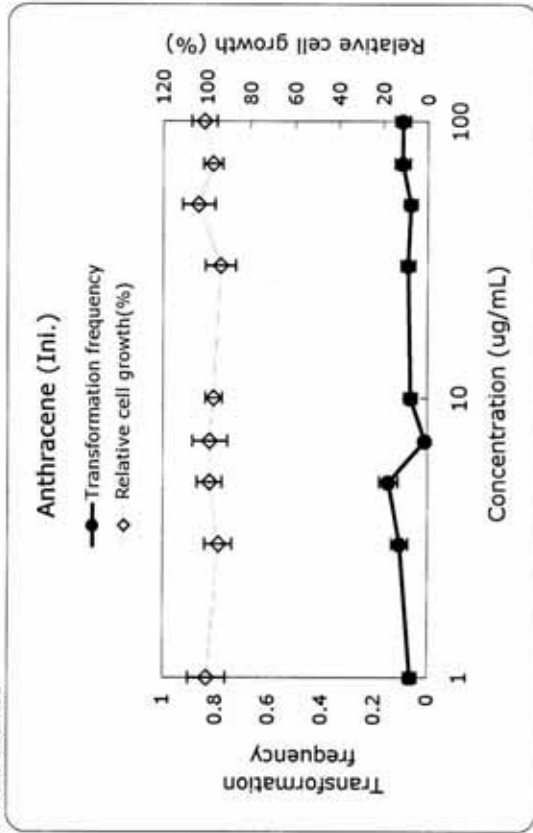
Cell Growth Assay		Concentrations (ug/ml)								Negative Control	Positive Control			
Crystal Violet Staining	Blank	0.5% DMSO								0.1% DMSO	MCA (1ug/mL)			
		1	3	5	7	10	30	50	70			100		
OD _{550nm}														
1	0.094	1.324	1.147	1.220	1.270	1.153	1.199	1.290	1.116	1.257	1.144	1.144	1.144	0.547
2	0.087	1.375	1.168	1.148	1.042	1.258	1.165	1.186	1.227	1.188	1.275	1.275	1.275	0.562
3	0.086	1.189	1.197	1.262	1.284	1.172	1.230	1.266	1.186	1.249	1.147	1.147	1.147	0.530
4	0.089	1.089	1.189	1.151	1.191	1.181	1.042	1.168	1.163	1.163	1.253	1.253	1.253	0.533
5	0.094	1.119	1.119	1.163	1.126	1.195	1.141	1.194	1.230	1.188	1.104	1.104	1.104	0.564
6	0.100	1.225	1.032	1.316	1.246	1.132	1.202	1.228	1.215	1.372	1.160	1.160	1.160	0.515
7	0.107	1.258	1.262	1.267	1.222	1.211	1.040	1.354	1.270	1.265	1.039	1.039	1.039	0.472
8	0.112	1.247	1.226	1.179	1.306	1.236	1.237	1.217	1.157	1.239	1.170	1.170	1.170	0.477
Average	0.096	1.228	1.168	1.213	1.211	1.192	1.157	1.270	1.196	1.240	1.162	1.162	1.162	0.525
SD	0.009	0.097	0.071	0.063	0.089	0.042	0.078	0.083	0.049	0.065	0.076	0.076	0.076	0.035
Average - Blank	0.000	1.132	1.071	1.117	1.115	1.096	1.061	1.173	1.100	1.144	1.065	1.065	1.065	0.429
Relative Cell Growth (%)		100.0	94.7	98.7	98.5	96.9	93.7	103.7	97.2	101.1	100.0	100.0	100.0	40.3
SD of Relative Cell Growth		8.6	6.2	5.6	7.9	3.7	6.9	7.3	4.3	5.8	6.7	6.7	6.7	3.1

Transformation Assay

Giemsa Staining	1	3	5	7	10	30	50	70	100	Negative Control	Positive Control
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	6	10	14	1	6	7	6	9	9	1	44
Transformation frequency*	0.0625	0.104166667	0.145833333	0.010416667	0.0625	0.072916667	0.0625	0.09375	0.09375	0.010416667	0.458333333
SE	0.02470529	0.03117758	0.03602169	0.01036227	0.02470529	0.02653610	0.02470529	0.02974911	0.02974911	0.01036227	0.05085354

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

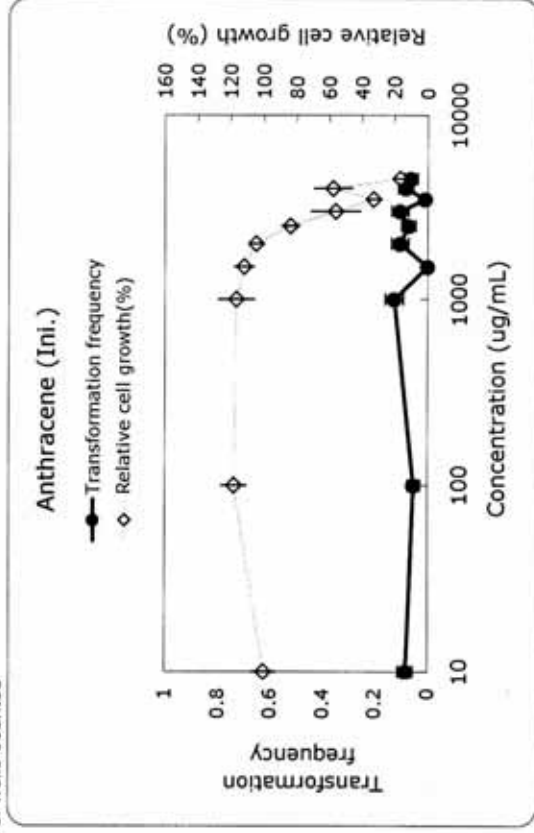
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Anthracene
Institution:	Lab 4
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)		
		10	100	1000	1500	2000	2500	3000	3500	4000	4500				
Crystal Violet Staining															
OD _{540nm}															
1	0.062	0.335	0.357	0.349	0.383	0.364	0.310	0.285	0.181	0.242	0.106	0.307	0.177		
2	0.065	0.337	0.423	0.381	0.388	0.355	0.292	0.199	0.155	0.225	0.125	0.329	0.192		
3	0.065	0.336	0.423	0.384	0.386	0.362	0.302	0.205	0.153	0.206	0.106	0.335	0.169		
4	0.063	0.373	0.400	0.390	0.383	0.371	0.279	0.183	0.145	0.225	0.107	0.338	0.183		
5	0.066	0.382	0.416	0.451	0.357	0.343	0.296	0.201	0.168	0.193	0.104	0.331	0.176		
6	0.071	0.327	0.399	0.390	0.368	0.360	0.306	0.199	0.150	0.221	0.117	0.346	0.170		
7	0.063	0.341	0.407	0.423	0.412	0.374	0.318	0.229	0.161	0.219	0.111	0.358	0.187		
8	0.061	0.369	0.393	0.410	0.397	0.380	0.327	0.297	0.154	0.303	0.124	0.347	0.180		
Average	0.065	0.350	0.402	0.397	0.384	0.364	0.304	0.225	0.158	0.229	0.113	0.336	0.179		
SD	0.003	0.021	0.031	0.017	0.012	0.012	0.015	0.043	0.011	0.033	0.008	0.015	0.008		
Average - Blank	0.000	0.286	0.338	0.333	0.320	0.299	0.239	0.160	0.094	0.165	0.048	0.272	0.115		
Relative Cell Growth (%)		100.0	118.3	116.5	112.0	104.8	83.8	56.1	32.9	57.7	16.8	100.0	42.2		
SD of Relative Cell Growth		7.4	7.5	10.7	5.9	4.1	5.3	15.0	4.0	11.6	3.0	5.3	2.8		

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	10	100	1000	1500	2000	2500	3000	3500	4000	4500		
No. of wells excluded	0	0	0	96	0	0	0	0	0	0	3	0
No. of wells counted	96	96	96	0	96	96	96	96	96	96	93	96
No. of wells having foci	8	5	12	0	10	7	10	1	8	6	5	52
Transformation frequency*	0.083333333	0.052083333	0.125	#DIV/0!	0.104166667	0.072916667	0.104166667	0.010416667	0.083333333	0.0625	0.053763441	0.541666667
SE	0.02820847	0.02267769	0.03375386	#DIV/0!	0.03117758	0.02653610	0.03117758	0.01036227	0.02820847	0.02470529	0.02338848	0.05085354

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

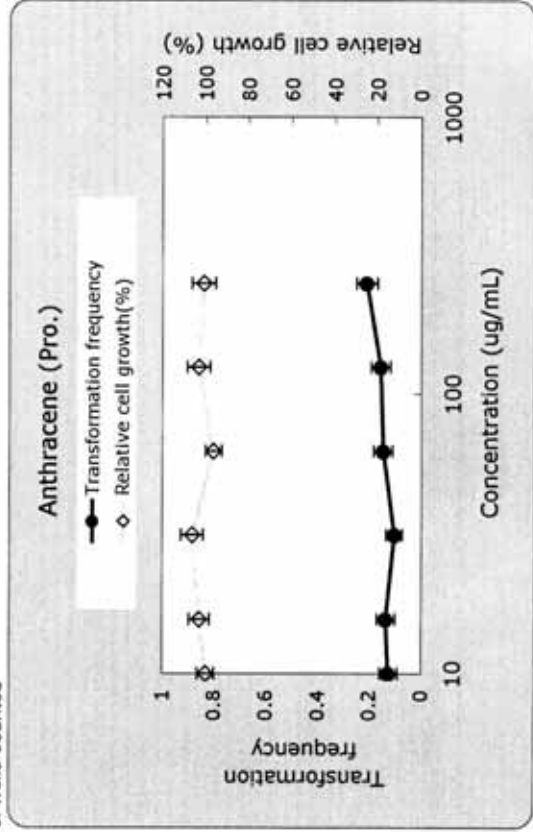
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Anthracene
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)						Negative Control	Positive Control
Crystal Violet Staining		OD _{550nm}						0.1% DMSO	TPA (50ng/mL)
Blank		10	15.6	31.3	62.5	125	250		
1	0.098	0.507	0.495	0.499	0.487	0.498	0.492	0.478	0.813
2	0.096	0.495	0.530	0.540	0.499	0.493	0.488	0.579	0.857
3	0.107	0.464	0.519	0.476	0.465	0.489	0.537	0.590	0.839
4	0.103	0.504	0.511	0.534	0.475	0.526	0.491	0.587	0.866
5	0.098	0.503	0.481	0.518	0.473	0.475	0.501	0.553	0.821
6	0.103	0.493	0.516	0.519	0.494	0.518	0.498	0.601	0.862
7	0.105	0.494	0.488	0.518	0.467	0.498	0.457	0.575	0.801
8	0.104	0.475	0.483	0.522	0.458	0.535	0.500	0.579	0.846
Average	0.102	0.492	0.503	0.516	0.477	0.504	0.496	0.568	0.838
SD	0.004	0.015	0.018	0.020	0.015	0.020	0.022	0.039	0.024
Average - Blank	0.000	0.390	0.401	0.414	0.376	0.402	0.394	0.466	0.736
Relative Cell Growth (%)		100.0	102.8	106.1	96.3	103.1	100.9	100.0	158.0
SD of Relative Cell Growth		3.8	4.7	5.2	3.7	5.2	5.6	10.0	6.2

Transformation Assay		Concentrations (ug/ml)						Negative Control	Positive Control
Giemsa Staining		No. of wells having foci							
Blank		10	15.6	31.3	62.5	125	250		
No. of wells excluded	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96
No. of wells having foci	12	13	10	14	15	20	20	10	66
Transformation frequency*	0.125	0.135416667	0.104166667	0.145833333	0.15625	0.208333333	0.208333333	0.104166667	0.6875
SE	0.03375386	0.03492238	0.03117758	0.03602169	0.03705794	0.04144908	0.04144908	0.03117758	0.04730704

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

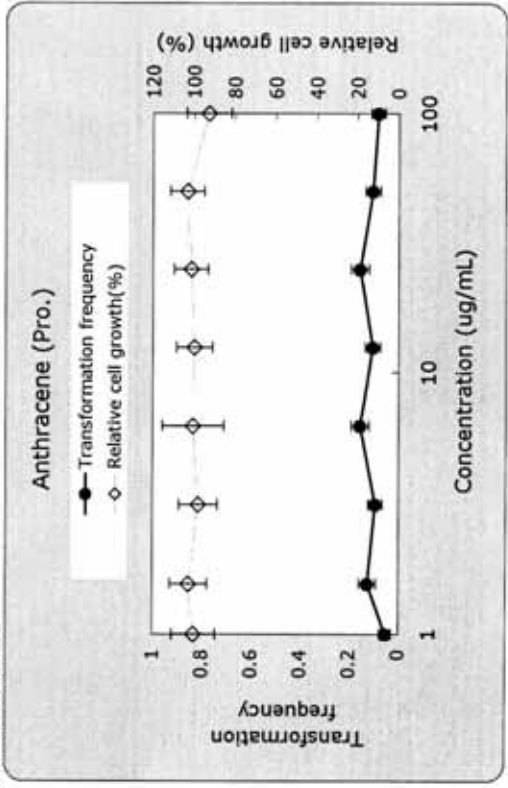
Chemical Code:	Anthracene
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Crystal Violet Staining												0.1% DMSO	TPA (50ng/mL)
OD _{560nm}	Blank	1.56	3.13	6.25	12.5	25	50	100					
1	0.054	0.361	0.327	0.301	0.306	0.324	0.328	0.301			0.414	0.442	
2	0.050	0.361	0.317	0.371	0.344	0.382	0.382	0.353			0.389	0.491	
3	0.047	0.361	0.383	0.436	0.375	0.357	0.387	0.301			0.442	0.531	
4	0.047	0.390	0.340	0.325	0.363	0.372	0.361	0.334			0.359	0.527	
5	0.045	0.351	0.325	0.345	0.347	0.356	0.369	0.393			0.420	0.550	
6	0.048	0.383	0.390	0.362	0.338	0.395	0.397	0.348			0.470	0.574	
7	0.051	0.379	0.361	0.388	0.346	0.344	0.341	0.325			0.455	0.567	
8	0.056	0.302	0.329	0.303	0.395	0.329	0.338	0.296			0.388	0.472	
Average	0.050	0.361	0.347	0.354	0.352	0.357	0.363	0.331	#DIV/0!	#DIV/0!	0.417	0.519	
SD	0.004	0.027	0.028	0.046	0.027	0.025	0.025	0.033	#DIV/0!	#DIV/0!	0.038	0.047	
Average - Blank	0.000	0.311	0.297	0.304	0.302	0.308	0.313	0.282	#DIV/0!	#DIV/0!	0.367	0.470	
Relative Cell Growth (%)		100.0	97.8	100.2	99.5	101.4	103.2	92.8	#DIV/0!	#DIV/0!	100.0	127.8	
SD of Relative Cell Growth		10.7	9.2	15.0	8.8	8.2	8.3	10.9	#DIV/0!	#DIV/0!	12.4	15.5	

Transformation Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining												0	0
No. of wells excluded	No. of wells counted	1.56	3.13	6.25	12.5	25	50	100					
0	96	0	0	0	0	0	0	0			0	0	
5	96	12	9	15	10	15	10	8			96	96	
No. of wells having foci											96	96	
Transformation frequency*		0.05208333	0.09375	0.15625	0.10416667	0.15625	0.10416667	0.08333333	0	0	0.104166667	0.427083333	
SE		0.02267769	0.02974911	0.03705794	0.03117758	0.03705794	0.03117758	0.02820847	0.00000000	0.00000000	0.03117758	0.05048547	

*No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

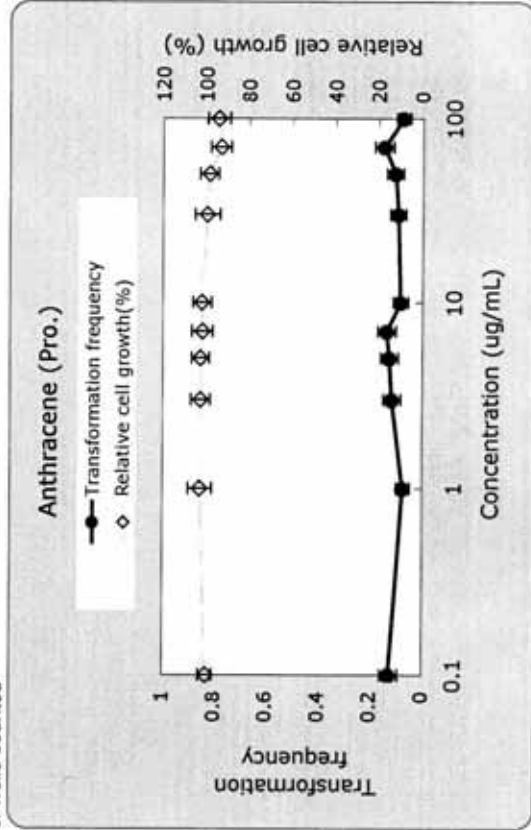
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Anthracene
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining		0.5% DMSO										0.1% DMSO		TPA (50ng/mL)	
OD _{550nm}		Blank	1	3	5	7	10	30	50	70	100	0.1% DMSO		TPA (50ng/mL)	
1	0.098		1.045	1.080	1.073	1.063	1.005	1.005	1.010	0.973	0.989	1.193	1.367		
2	0.103		1.040	1.110	1.160	1.062	1.085	1.074	1.037	0.962	0.957	1.163	1.379		
3	0.109		1.115	1.018	1.082	1.052	1.057	1.118	1.024	0.991	0.987	1.192	1.387		
4	0.111		1.029	1.142	1.054	1.042	1.071	1.092	1.119	1.068	1.095	1.241	1.540		
5	0.112		1.076	1.115	1.099	1.048	1.147	1.104	1.091	0.929	0.952	1.231	1.525		
6	0.117		1.180	1.096	1.100	1.049	1.062	1.080	1.007	1.043	0.975	1.275	1.407		
7	0.122		1.124	1.037	1.086	1.155	1.086	1.002	1.056	0.997	1.038	1.156	1.419		
8	0.125		1.084	1.081	1.025	1.152	1.112	0.970	1.032	0.997	1.048	1.220	1.436		
Average	0.112		1.087	1.085	1.085	1.078	1.078	1.056	1.047	0.995	1.005	1.209	1.433		
SD	0.009		0.051	0.041	0.039	0.047	0.042	0.055	0.040	0.044	0.050	0.040	0.066		
Average - Blank	0.000		0.975	0.973	0.973	0.966	0.966	0.944	0.935	0.883	0.893	1.097	1.320		
Relative Cell Growth (%)			102.7	102.6	102.6	101.8	101.8	99.5	98.6	93.1	94.1	100.0	120.4		
SD of Relative Cell Growth			5.4	4.3	4.1	5.0	4.4	5.8	4.2	4.6	5.3	4.3	6.9		

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining		0.1										0		0	
No. of wells excluded		0	1	3	5	7	10	30	50	70	100	0		0	
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96		96	
No. of wells having foci		12	7	11	12	13	8	9	10	14	7	11		45	
Transformation frequency*		0.125	0.072916667	0.114583333	0.125	0.135416667	0.083333333	0.09375	0.104166667	0.145833333	0.072916667	0.114583333		0.46875	
SE		0.03375386	0.026553610	0.03250865	0.03375386	0.03492238	0.02820847	0.02974911	0.03117758	0.03602169	0.02653610	0.03250865		0.05093127	

*No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

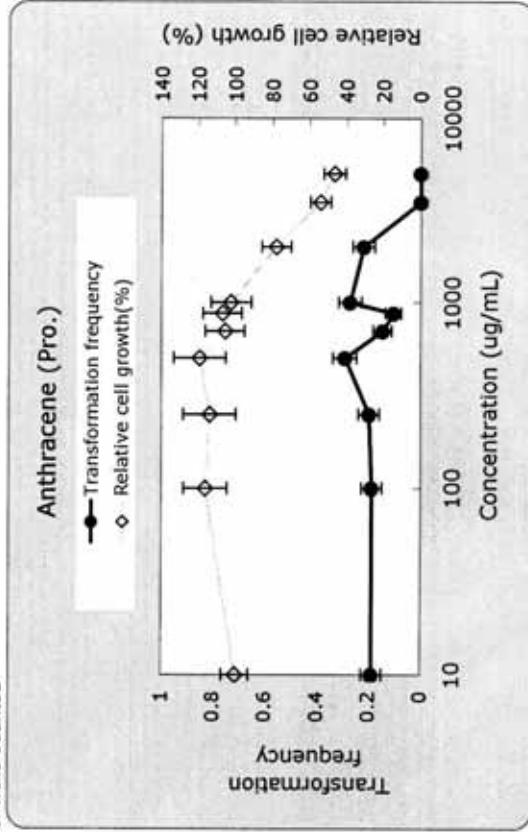
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Anthracene
Institution:	Lab 4
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control	Positive Control	
Crystal Violet Staining	Blank	10	0.5% DMSO	100	250	500	700	875	1000	2000	3500	5000	0.1% DMSO	TPA (50ng/mL)
OD _{540nm}														
1	0.077	0.258	0.272	0.285	0.272	0.285	0.259	0.275	0.245	0.216	0.174	0.175	0.374	0.399
2	0.096	0.281	0.330	0.321	0.309	0.309	0.304	0.303	0.309	0.228	0.190	0.163	0.398	0.403
3	0.088	0.297	0.314	0.298	0.321	0.321	0.282	0.338	0.306	0.256	0.188	0.173	0.355	0.462
4	0.085	0.286	0.358	0.321	0.358	0.371	0.313	0.274	0.308	0.229	0.188	0.168	0.354	0.422
5	0.086	0.299	0.322	0.331	0.322	0.347	0.306	0.298	0.282	0.253	0.208	0.191	0.366	0.422
6	0.078	0.280	0.326	0.322	0.329	0.329	0.302	0.288	0.280	0.257	0.203	0.186	0.368	0.418
7	0.082	0.302	0.302	0.360	0.302	0.332	0.324	0.313	0.301	0.251	0.202	0.179	0.362	0.435
8	0.097	0.277	0.277	0.301	0.277	0.294	0.281	0.302	0.286	0.239	0.197	0.196	0.344	0.423
Average	0.086	0.285	0.313	0.317	0.313	0.323	0.296	0.299	0.290	0.241	0.194	0.179	0.365	0.423
SD	0.007	0.015	0.028	0.023	0.028	0.028	0.021	0.021	0.022	0.016	0.011	0.011	0.016	0.020
Average - Blank	0.000	0.199	0.227	0.231	0.227	0.237	0.210	0.213	0.203	0.155	0.108	0.093	0.279	0.337
Relative Cell Growth (%)		100.0	113.9	116.2	113.9	119.3	105.7	107.0	102.3	77.9	54.1	46.6	100.0	120.7
SD of Relative Cell Growth		7.4	14.3	11.7	14.3	14.1	10.4	10.5	10.9	7.8	5.5	5.8	8.3	9.8

Transformation Assay		Concentrations (ug/ml)										Negative Control	Positive Control	
Giemsa Staining	Blank	10	0.5% DMSO	100	250	500	700	875	1000	2000	3500	5000	0.1% DMSO	TPA (50ng/mL)
No. of wells excluded		0		0	0	0	0	0	0	0	96	96	0	0
No. of wells counted		96		96	96	96	96	96	96	96	0	0	96	96
No. of wells having foci		18		19	19	28	14	10	26	21	0	0	12	56
Transformation frequency*		0.1875		0.197916667	0.197916667	0.291666667	0.145833333	0.104166667	0.270833333	0.21875	#DIV/0!	#DIV/0!	0.125	0.583333333
SE		0.03983609		0.03983609	0.04066449	0.04639024	0.03602169	0.03117758	0.04535538	0.04219232	#DIV/0!	#DIV/0!	0.03375386	0.05031728

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Phenanthrene
Institution:	Lab 1
Test Number	

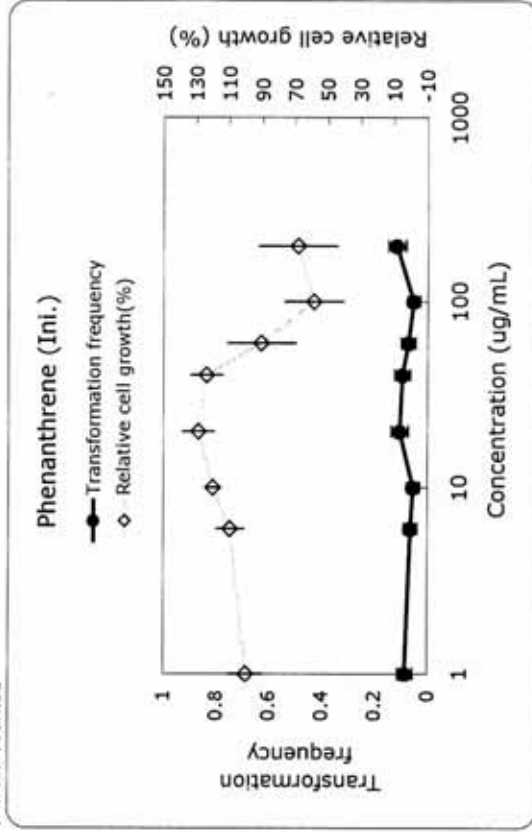
Cell Growth Assay		Concentrations (ug/ml)						Negative Control	Positive Control			
Crystal Violet Staining		Blank	0.1% DMSO	6	10	20	40	60	100	200	0.1% DMSO	MCA (1ug/mL)
OD _{590nm}												
1	0.093	0.434	0.464	0.563	0.547	0.610	0.561	0.450	0.529	0.434	0.268	
2	0.100	0.449	0.541	0.542	0.586	0.583	0.421	0.363	0.372	0.449	0.237	
3	0.096	0.464	0.488	0.551	0.615	0.559	0.331	0.285	0.335	0.464	0.255	
4	0.100	0.448	0.488	0.541	0.643	0.545	0.346	0.293	0.270	0.448	0.246	
5	0.101	0.476	0.494	0.548	0.566	0.509	0.429	0.261	0.272	0.476	0.293	
6	0.103	0.517	0.528	0.574	0.578	0.552	0.477	0.248	0.297	0.517	0.228	
7	0.104	0.464	0.524	0.533	0.530	0.540	0.445	0.309	0.343	0.464	0.243	
8	0.115	0.545	0.553	0.543	0.595	0.613	0.505	0.352	0.435	0.545	0.233	
Average	0.102	0.475	0.510	0.549	0.583	0.564	0.439	0.320	0.357	0.475	0.250	
SD	0.007	0.038	0.031	0.013	0.036	0.036	0.077	0.066	0.089	0.038	0.021	
Average - Blank	0.000	0.373	0.409	0.448	0.481	0.462	0.338	0.219	0.255	0.373	0.149	
Relative Cell Growth (%)		100.0	109.5	120.0	128.9	123.9	90.6	58.6	68.4	100.0	39.9	
SD of Relative Cell Growth		10.1	8.3	3.6	9.7	9.6	20.6	17.7	23.7	10.1	5.7	

Transformation Assay

Giemsa Staining		Concentrations (ug/ml)						Negative Control	Positive Control			
		Blank	0.1% DMSO	6	10	20	40	60	100	200	0.1% DMSO	MCA (1ug/mL)
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	8	6	5	10	9	7	5	11	8	8	42	
Transformation frequency*	0.0833333333	0.0625	0.0520833333	0.104166667	0.09375	0.072916667	0.0520833333	0.1145833333	0.0833333333	0.0833333333	0.4375	
SE	0.02820847	0.02470529	0.022267769	0.03117758	0.02974911	0.02653610	0.02267769	0.03250865	0.00000000	0.02820847	0.05063079	

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

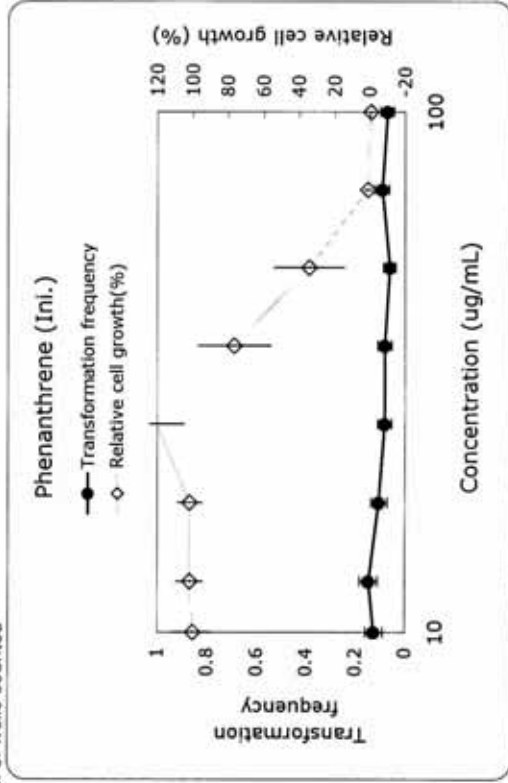
Chemical Code:	Phenanthrene
Institution:	Lab 2
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Crystal Violet Staining	Blank	10	12.5	17.7	25	35.4	50	70.7	100	100	0.1% DMSO	MCA (Iug/mL)	
OD _{540nm}													
1	0.074	0.412	0.443	0.442	0.453	0.493	0.377	0.084	0.078	0.078	0.475	0.313	
2	0.070	0.441	0.467	0.510	0.619	0.619	0.510	0.470	0.074	0.074	0.475	0.313	
3	0.075	0.462	0.509	0.476	0.519	0.286	0.476	0.067	0.072	0.072	0.551	0.279	
4	0.079	0.523	0.470	0.518	0.618	0.400	0.181	0.077	0.063	0.063	0.482	0.312	
5	0.072	0.530	0.508	0.464	0.596	0.268	0.195	0.073	0.059	0.059	0.473	0.325	
6	0.075	0.527	0.486	0.486	0.619	0.446	0.143	0.083	0.067	0.067	0.483	0.298	
7	0.070	0.493	0.494	0.523	0.577	0.332	0.142	0.074	0.075	0.075	0.526	0.336	
8	0.074	0.455	0.471	0.478	0.508	0.384	0.285	0.095	0.079	0.079	0.459	0.257	
Average	0.074	0.480	0.487	0.487	0.563	0.385	0.213	0.078	0.071	0.071	0.491	0.304	
SD	0.003	0.044	0.030	0.028	0.063	0.084	0.080	0.009	0.007	0.007	0.031	0.026	
Average - Blank	0.000	0.407	0.414	0.414	0.490	0.311	0.140	0.005	-0.003	-0.003	0.417	0.231	
Relative Cell Growth (%)		100.0	101.7	101.7	120.4	76.5	34.4	1.2	-0.7	-0.7	100.0	55.2	
SD of Relative Cell Growth		10.9	7.4	6.9	15.5	20.6	19.8	2.1	1.7	1.7	7.6	6.3	

Transformation Assay		Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	Blank	10	12.5	17.7	25	35.4	50	70.7	100	100	0.1% DMSO	MCA (Iug/mL)	
No. of wells excluded		0	0	0	0	0	0	0	0	0	0	0	
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96	
No. of wells having foci		12	14	10	8	8	6	9	7	7	11	69	
Transformation frequency*		0.125	0.14583333	0.10416667	0.08333333	0.08333333	0.0625	0.09375	0.07291667	0.07291667	0.114583333	0.71875	
SE		0.03375386	0.03602169	0.03117758	0.02820847	0.02820847	0.02470529	0.02974911	0.02653610	0.02653610	0.03250865	0.04588805	

*No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

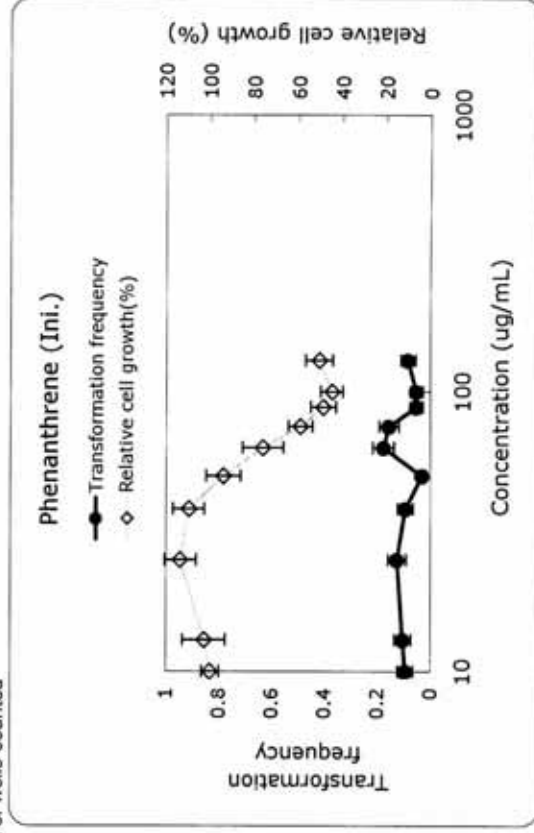
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Phenanthrene
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{560nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
		Blank	10	13	25	38	50	63	75	88	100		
1	0.102	1.079	1.068	1.101	1.102	0.981	0.974	0.646	0.644	0.543	0.555	1.079	0.408
2	0.109	1.005	1.018	1.159	1.183	0.961	0.778	0.618	0.488	0.505	0.608	1.005	0.412
3	0.113	0.970	1.221	1.206	1.199	1.052	0.779	0.628	0.508	0.467	0.499	0.970	0.452
4	0.116	1.010	1.012	1.151	1.096	0.946	0.766	0.650	0.544	0.495	0.509	1.010	0.398
5	0.118	1.028	1.047	1.224	1.029	0.833	0.689	0.600	0.591	0.533	0.597	1.028	0.413
6	0.124	1.034	0.993	1.135	1.098	0.932	0.766	0.715	0.521	0.463	0.576	1.034	0.424
7	0.127	0.972	1.057	1.021	1.020	0.934	0.789	0.592	0.537	0.531	0.542	0.972	0.422
8	0.119	1.029	0.912	1.111	1.099	1.052	0.859	0.722	0.595	0.602	0.668	1.029	0.447
Average	0.116	1.016	1.041	1.139	1.103	0.961	0.800	0.646	0.554	0.517	0.569	1.016	0.422
SD	0.008	0.035	0.088	0.064	0.063	0.071	0.084	0.049	0.052	0.045	0.056	0.035	0.019
Average - Blank	0.000	0.900	0.925	1.023	0.987	0.845	0.684	0.530	0.438	0.401	0.453	0.900	0.306
Relative Cell Growth (%)		100.0	102.8	113.6	109.7	93.9	76.0	58.9	48.6	44.6	50.4	100.0	34.0
SD of Relative Cell Growth		3.9	9.7	7.1	7.0	7.9	9.3	5.4	5.8	5.0	6.2	3.9	2.1

Transformation Assay	Giemsa Staining	Concentrations (ug/ml)										Negative Control	Positive Control
		10	13	25	38	50	63	75	88	100	130		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	9	10	12	9	3	17	15	5	5	5	8	9	42
Transformation frequency*	0.09375	0.104166667	0.125	0.09375	0.03125	0.177083333	0.15625	0.052083333	0.052083333	0.052083333	0.083333333	0.09375	0.4375
SE	0.02974911	0.03117758	0.03375386	0.02974911	0.01775805	0.03896110	0.03705794	0.02267769	0.02267769	0.02267769	0.02820847	0.02974911	0.05063079

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Phenanthrene
Institution:	Lab 4
Test Number	

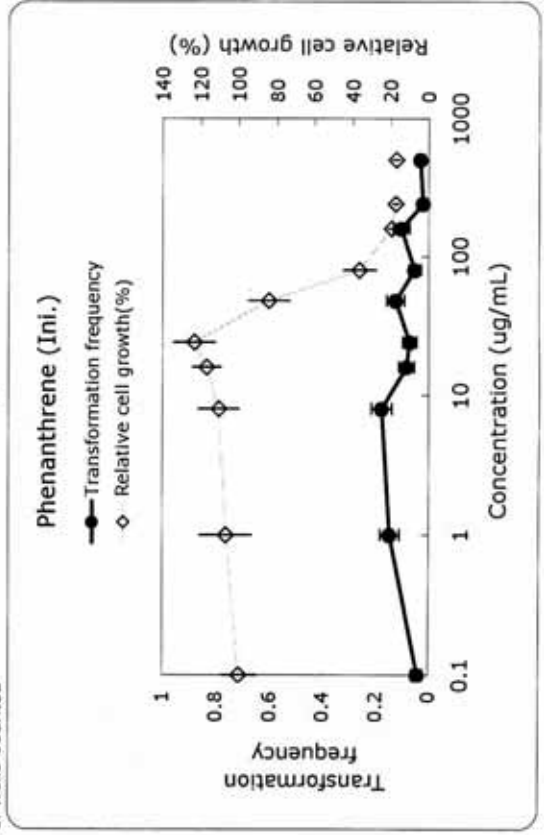
Cell Growth Assay		Concentrations (ug/ml)										Negative Control	Positive Control	
Crystal Violet Staining	Blank	0.1	0.5% DMSO	1	8	16	24	48	80	160	240	500	0.1% DMSO	MCA (1ug/mL)
OD _{590nm}														
1	0.072	0.292		0.286	0.323	0.338	0.364	0.288	0.199	0.124	0.115	0.116	0.309	0.156
2	0.071	0.295		0.318	0.350	0.342	0.398	0.290	0.148	0.122	0.111	0.115	0.293	0.143
3	0.074	0.320		0.364	0.333	0.361	0.367	0.296	0.145	0.115	0.120	0.115	0.334	0.132
4	0.075	0.296		0.307	0.365	0.356	0.344	0.276	0.151	0.114	0.114	0.116	0.316	0.142
5	0.073	0.328		0.332	0.349	0.328	0.385	0.251	0.141	0.123	0.117	0.113	0.321	0.156
6	0.074	0.329		0.371	0.313	0.367	0.329	0.224	0.149	0.122	0.117	0.117	0.286	0.143
7	0.075	0.319		0.302	0.325	0.352	0.369	0.255	0.157	0.116	0.112	0.114	0.291	0.139
8	0.076	0.269		0.295	0.286	0.318	0.326	0.277	0.179	0.117	0.107	0.104	0.263	0.140
Average	0.074	0.306		0.322	0.331	0.345	0.360	0.269	0.159	0.119	0.114	0.114	0.302	0.144
SD	0.001	0.021		0.031	0.025	0.017	0.025	0.025	0.020	0.004	0.004	0.004	0.023	0.008
Average - Blank	0.000	0.232		0.248	0.257	0.272	0.286	0.196	0.085	0.045	0.040	0.040	0.228	0.070
Relative Cell Growth (%)		100.0		106.8	110.6	116.9	123.3	84.3	36.5	19.5	17.4	17.2	100.0	30.8
SD of Relative Cell Growth		9.2		13.6	10.6	7.3	10.9	10.6	8.6	1.8	1.7	1.9	9.8	3.6

Transformation Assay

Giemsa Staining	0.1	1	8	16	24	48	80	160	240	500	Negative Control	Positive Control
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	2	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	94	96
No. of wells having foci	4	14	17	8	7	12	5	10	2	3	9	41
Transformation frequency*	0.041666667	0.145833333	0.177083333	0.083333333	0.072916667	0.125	0.052083333	0.104166667	0.020833333	0.03125	0.095744681	0.427083333
SE	0.02039469	0.03602169	0.03896110	0.02820847	0.02653610	0.03375396	0.02267769	0.03117758	0.01457713	0.01775805	0.03034862	0.05048547

* No. of wells having foci/ No. of wells counted

Phenanthrene (Ini.)
Transformation frequency
Relative cell growth (%)



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Remarks

Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Phenanthrene
Institution:	Lab 4
Test Number	

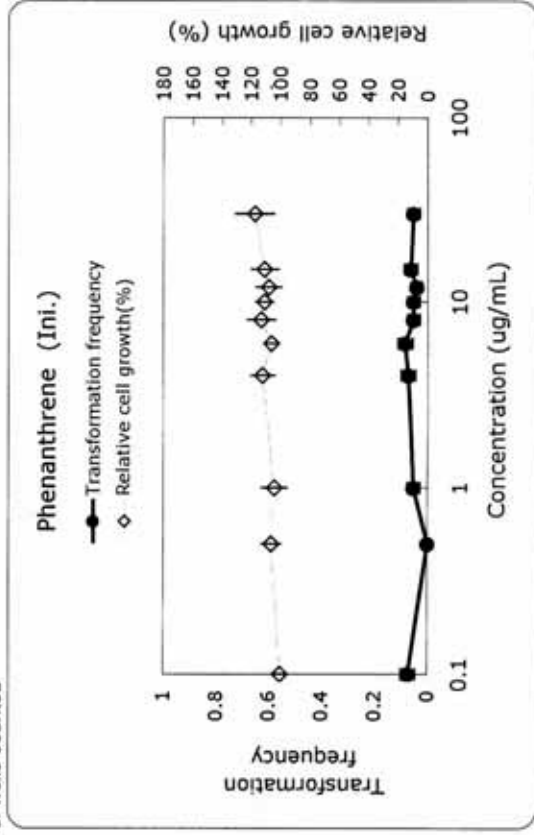
Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)	
	Blank	0.1	0.5	1	4	6	8	10	12	15			30
Crystal Violet Staining													
OD _{590nm}													
1	0.062	0.303	0.318	0.298	0.325	0.320	0.306	0.316	0.303	0.336	0.296	0.254	0.174
2	0.065	0.305	0.324	0.291	0.330	0.312	0.353	0.346	0.348	0.337	0.342	0.288	0.164
3	0.065	0.321	0.316	0.331	0.346	0.326	0.348	0.351	0.365	0.337	0.367	0.291	0.178
4	0.063	0.332	0.334	0.335	0.350	0.349	0.345	0.363	0.335	0.319	0.370	0.311	0.172
5	0.066	0.319	0.350	0.328	0.338	0.331	0.337	0.329	0.339	0.334	0.380	0.286	0.174
6	0.071	0.312	0.316	0.341	0.368	0.341	0.378	0.356	0.360	0.362	0.388	0.303	0.185
7	0.063	0.322	0.335	0.356	0.384	0.345	0.384	0.352	0.310	0.392	0.400	0.312	0.170
8	0.061	0.315	0.360	0.330	0.333	0.335	0.344	0.341	0.330	0.330	0.341	0.271	0.160
Average	0.065	0.316	0.332	0.326	0.347	0.332	0.349	0.344	0.336	0.343	0.361	0.290	0.172
SD	0.003	0.010	0.016	0.022	0.020	0.013	0.024	0.015	0.022	0.023	0.033	0.020	0.008
Average - Blank	0.000	0.252	0.267	0.262	0.282	0.268	0.285	0.280	0.272	0.279	0.296	0.225	0.108
Relative Cell Growth (%)		100.0	106.2	104.0	112.2	106.5	113.2	111.2	108.0	110.8	117.6	100.0	47.8
SD of Relative Cell Growth		3.8	6.5	8.6	8.0	5.0	9.6	6.1	8.7	9.1	13.2	7.9	3.1

Transformation Assay

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	0.1	0.5	1	4	6	8	10	12	15	30			
Giemsa Staining													
No. of wells excluded	0	96	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	0	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	7		5	7	8	5	5	5	4	6	5	1	45
Transformation frequency*	0.072916667	0.052083333	0.072916667	0.072916667	0.083333333	0.052083333	0.052083333	0.052083333	0.041666667	0.0625	0.052083333	0.010416667	0.46875
SE	0.02653610	0.02267769	0.02653610	0.02653610	0.02820847	0.02267769	0.02267769	0.02267769	0.02039469	0.02470529	0.02267769	0.01036227	0.05093127

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Phenanthrene
Institution:	Lab 1
Test Number	

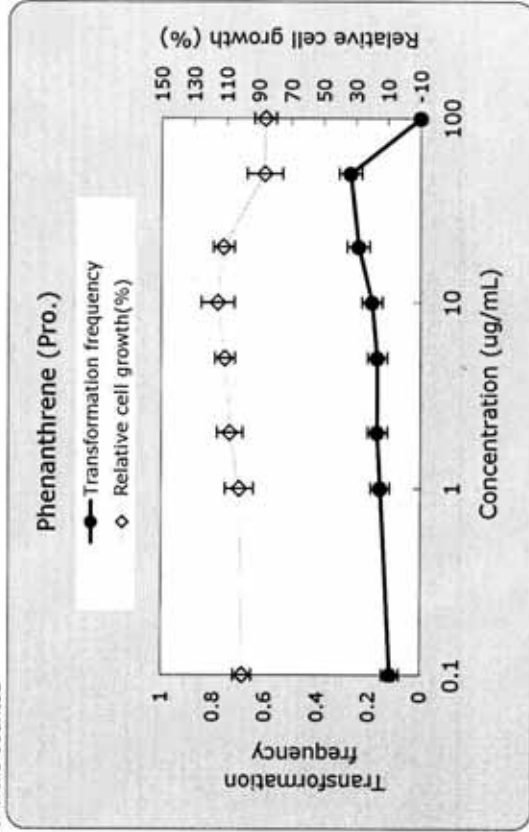
Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/ml)	
	Blank	0.1	1	2	5	10	20	50	100	100			
Crystal Violet Staining OD _{540nm}													
1	0.099	0.477	0.491	0.474	0.510	0.504	0.549	0.486	0.484	0.484	0.477	0.839	
2	0.108	0.474	0.477	0.488	0.515	0.547	0.540	0.449	0.441	0.441	0.474	0.809	
3	0.111	0.498	0.438	0.511	0.510	0.529	0.522	0.386	0.416	0.416	0.498	0.817	
4	0.104	0.481	0.510	0.527	0.546	0.526	0.499	0.407	0.444	0.444	0.481	0.850	
5	0.106	0.509	0.492	0.531	0.537	0.519	0.561	0.397	0.416	0.416	0.509	0.798	
6	0.111	0.501	0.524	0.525	0.573	0.594	0.555	0.467	0.423	0.423	0.501	0.822	
7	0.109	0.468	0.515	0.564	0.526	0.614	0.504	0.431	0.420	0.420	0.468	0.812	
8	0.112	0.530	0.549	0.554	0.558	0.577	0.565	0.497	0.471	0.471	0.530	0.803	
Average	0.108	0.492	0.500	0.522	0.534	0.551	0.537	0.440	0.439	0.439	0.492	0.819	
SD	0.004	0.021	0.033	0.030	0.023	0.039	0.026	0.042	0.026	0.026	0.021	0.018	
Average - Blank	0.000	0.385	0.392	0.414	0.427	0.444	0.429	0.333	0.332	0.332	0.385	0.711	
Relative Cell Growth (%)			101.9	107.7	110.9	115.3	111.6	86.4	86.3	86.3	100.0	184.9	
SD of Relative Cell Growth			8.7	7.9	6.1	10.2	6.7	10.8	6.8	6.8	5.5	4.6	

Transformation Assay

Giemsa Staining	Concentrations (ug/ml)										Negative Control	Positive Control
	0.1	1	2	5	10	20	50	100	100	100		
No. of wells excluded	0	0	0	0	0	0	0	0	53	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	43	96	96	96
No. of wells having foci	11	15	16	16	18	23	26	TOX	TOX	96	11	53
Transformation frequency*	0.114583333	0.15625	0.166666667	0.166666667	0.1875	0.239583333	0.270833333	#VALUE!	0	0	0.114583333	0.552083333
SE	0.03250865	0.03705794	0.03803629	0.03803629	0.03983609	0.04356307	0.04535538	#VALUE!	0.00000000	0.00000000	0.03250865	0.05075342

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

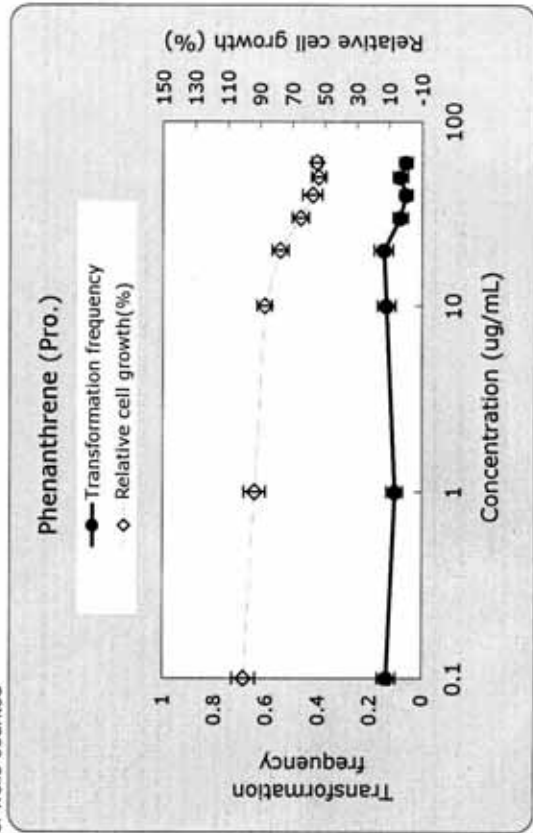
Chemical Code:	Phenanthrene
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)											Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)	
	Blank	0.1	1	10	20	30	40	50	60					
Crystal Violet Staining														
OD _{590nm}														
1	0.117	0.779	0.794	0.729	0.666	0.647	0.536	0.564	0.538	0.538	0.538	0.779	1.166	
2	0.118	0.801	0.855	0.805	0.718	0.565	0.539	0.464	0.484	0.484	0.484	0.801	1.047	
3	0.108	0.824	0.806	0.713	0.622	0.546	0.522	0.503	0.479	0.479	0.479	0.824	1.099	
4	0.108	0.851	0.803	0.699	0.733	0.554	0.478	0.512	0.514	0.514	0.514	0.851	1.046	
5	0.125	0.868	0.778	0.758	0.672	0.568	0.514	0.463	0.497	0.497	0.497	0.868	1.039	
6	0.128	0.939	0.687	0.753	0.651	0.609	0.581	0.521	0.540	0.540	0.540	0.939	1.026	
7	0.104	0.810	0.812	0.743	0.653	0.553	0.467	0.489	0.515	0.515	0.515	0.810	1.084	
8	0.111	0.807	0.756	0.730	0.661	0.614	0.593	0.502	0.518	0.518	0.518	0.807	1.112	
Average	0.115	0.835	0.786	0.741	0.672	0.582	0.529	0.502	0.511	0.511	0.511	0.835	1.077	
SD	0.009	0.051	0.049	0.032	0.036	0.037	0.044	0.033	0.023	0.023	0.023	0.051	0.047	
Average - Blank	0.000	0.720	0.672	0.626	0.557	0.467	0.414	0.387	0.396	0.396	0.396	0.720	0.963	
Relative Cell Growth (%)														
SD of Relative Cell Growth		100.0	93.3	87.0	77.4	64.9	57.5	53.8	55.0	55.0	55.0	100.0	133.7	
		7.0	6.8	4.5	5.1	5.1	6.1	4.5	3.1	3.1	3.1	7.0	6.6	

Transformation Assay	Concentrations (ug/ml)											Negative Control	Positive Control
Giemsa Staining	0.1	1	10	20	30	40	50	60					
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	13	10	13	14	8	6	8	6	6	6	6	13	63
Transformation frequency*	0.135416667	0.104166667	0.135416667	0.145833333	0.083333333	0.0625	0.083333333	0.0625	0.0625	0.0625	0.0625	0.135416667	0.65625
SE	0.03492238	0.03117758	0.03492238	0.03602169	0.02820847	0.02470529	0.02820847	0.02470529	0.02470529	0.02470529	0.02470529	0.03492238	0.04847529

* No. of wells having foci/ No. of wells counted

Remarks	



Acceptance Criteria (1)	
Negative Control < 20 and 40 < Positive Control	Pass

Acceptance Criteria (2)	
Concurrent Cell Growth Assay	Pass

The Final Data	
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Data Sheet for Cell Transformation Assay on Promotion Assay

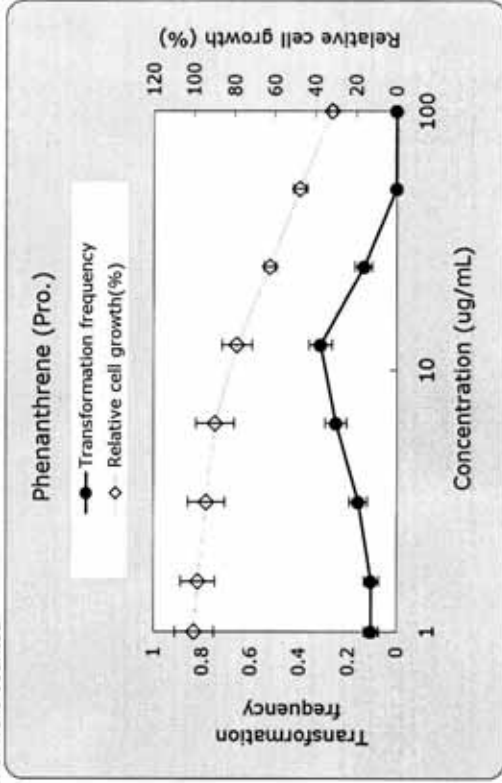
Chemical Code:	Phenanthrene
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
		Blank	0.5% DMSO	1	1.56	3.13	6.25	12.5	25	50	100		
1	0.068	0.359	0.345	0.357	0.320	0.271	0.281	0.220	0.173	0.472	0.452		
2	0.065	0.392	0.350	0.372	0.360	0.331	0.274	0.208	0.183	0.457	0.520		
3	0.063	0.441	0.434	0.420	0.357	0.321	0.277	0.216	0.167	0.441	0.538		
4	0.067	0.404	0.403	0.372	0.372	0.333	0.269	0.228	0.176	0.450	0.519		
5	0.062	0.407	0.404	0.365	0.411	0.352	0.281	0.218	0.163	0.423	0.500		
6	0.064	0.427	0.411	0.404	0.391	0.341	0.261	0.222	0.173	0.441	0.512		
7	0.059	0.410	0.403	0.416	0.378	0.342	0.256	0.228	0.163	0.455	0.439		
8	0.065	0.347	0.368	0.345	0.321	0.330	0.284	0.244	0.166	0.399	0.416		
Average	0.064	0.398	0.393	0.379	0.364	0.328	0.273	0.223	0.171	0.442	0.487		
SD	0.003	0.032	0.029	0.030	0.032	0.025	0.010	0.011	0.007	0.023	0.045		
Average - Blank	0.000	0.334	0.328	0.315	0.300	0.264	0.209	0.159	0.106	0.378	0.423		
Relative Cell Growth (%)	100.0	98.2	94.1	89.6	89.6	78.8	62.5	47.5	31.8	100.0	111.8		
SD of Relative Cell Growth	9.5	8.5	9.0	9.5	9.5	7.4	3.0	3.2	2.1	6.8	13.4		

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	1	1.56	3.13	6.25	12.5	25	50	100	0	0		
No. of wells excluded	0	0	1	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	95	96	96	96	96	96	96	96	96	96
No. of wells having foci	10	10	15	24	30	13	0	0	0	13	69	69
Transformation frequency*	0.10416667	0.10416667	0.15789474	0.25	0.3125	0.13541667	0.00000000	0.00000000	0.00000000	0.00000000	0.71875	0.71875
SE	0.03117758	0.03117758	0.03741151	0.04419417	0.04730704	0.03492238	0.00000000	0.00000000	0.00000000	0.03492238	0.04588805	0.04588805

* No. of wells having foci/ No. of wells counted

Remarks	



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

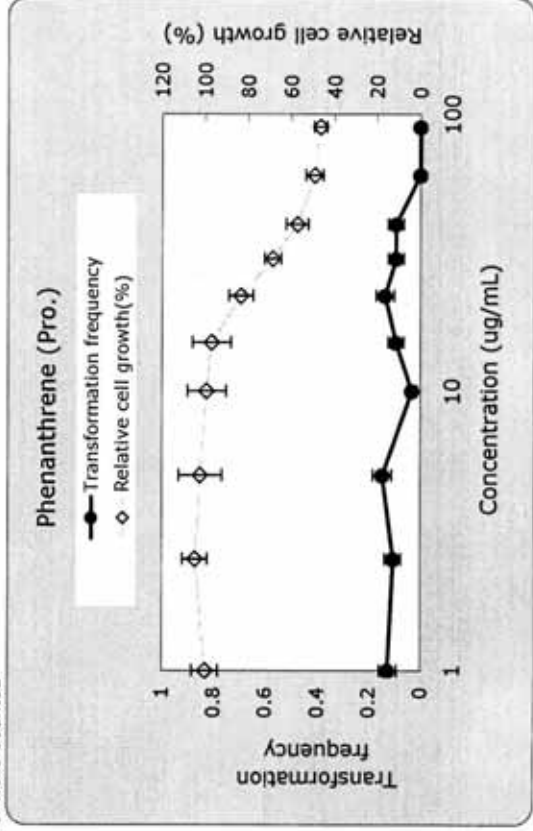
Chemical Code:	Phenanthrene
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
		0.1% DMSO										0.1% DMSO		TPA (50ng/mL)	
Crystal Violet Staining	Blank	1	2.5	5	10	15	22	30	40	60	90	0.1% DMSO	TPA (50ng/mL)		
1	0.386	0.975	1.029	0.882	0.913	0.921	0.870	0.777	0.729	0.677	0.673	0.975	1.285		
2	0.358	1.090	1.050	0.990	1.088	1.005	0.915	0.818	0.727	0.682	0.629	1.090	1.325		
3	0.340	1.003	0.975	1.101	1.012	0.999	0.873	0.792	0.689	0.699	0.656	1.003	1.312		
4	0.317	0.985	1.025	1.020	1.061	0.895	0.912	0.737	0.683	0.652	0.660	0.985	1.318		
5	0.330	0.984	1.014	1.073	0.989	0.996	0.937	0.813	0.715	0.636	0.625	0.984	1.316		
6	0.291	0.963	1.087	1.046	0.977	1.006	0.919	0.794	0.698	0.673	0.615	0.963	1.209		
7	0.329	0.978	1.074	1.028	0.993	1.078	0.851	0.798	0.789	0.620	0.649	0.978	1.257		
8	0.344	1.020	0.994	0.978	0.923	0.936	0.833	0.798	0.696	0.664	0.647	1.020	1.290		
Average	0.337	1.000	1.031	1.015	0.995	0.980	0.889	0.791	0.716	0.663	0.644	1.000	1.289		
SD	0.028	0.040	0.038	0.067	0.060	0.059	0.037	0.025	0.034	0.026	0.020	0.040	0.039		
Average - Blank	0.000	0.663	0.694	0.678	0.658	0.643	0.552	0.454	0.379	0.326	0.307	0.663	0.952		
Relative Cell Growth (%)		100.0	104.7	102.3	99.2	96.9	83.3	68.5	57.2	49.2	46.4	100.0	143.6		
SD of Relative Cell Growth		6.1	5.8	10.1	9.1	8.9	5.6	3.8	5.2	3.9	3.0	6.1	5.9		

Transformation Assay

Giemsa Staining		Concentrations (ug/ml)										Negative Control		Positive Control	
		0.1% DMSO										0.1% DMSO		TPA (50ng/mL)	
No. of wells excluded	No. of wells counted	1	2.5	5	10	15	22	30	40	60	90	0.1% DMSO	TPA (50ng/mL)		
0	96	0	0	0	0	0	0	0	0	0	0	0	0		
12	10	14	3	9	9	13	9	9	9	0	0	12	70		
Transformation frequency*		0.104166667	0.145833333	0.03125	0.09375	0.135416667	0.09375	0.09375	0.09375	0	0	0.125	0.729166667		
SE		0.03117758	0.03602169	0.01775805	0.02974911	0.03492238	0.02974911	0.02974911	0.02974911	0.00000000	0.00000000	0.03375386	0.045355538		

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

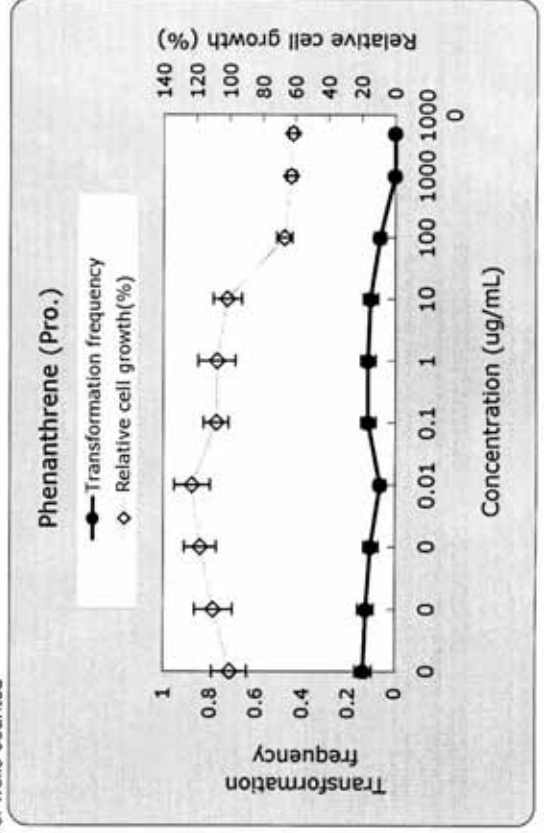
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Phenanthrene
Institution:	Lab 4
Test Number	

Cell Growth Assay	Crystal Violet Staining	Concentrations (ug/ml)										Negative Control	Positive Control
		0.5% DMSO	0.0001	0.001	0.01	0.1	1	10	100	1000	5000		
	Blank	0.077	0.284	0.304	0.336	0.303	0.288	0.280	0.217	0.206	0.219	0.317	0.385
1		0.096	0.299	0.331	0.335	0.297	0.318	0.308	0.219	0.226	0.212	0.326	0.387
2		0.088	0.287	0.370	0.364	0.313	0.305	0.293	0.241	0.218	0.224	0.350	0.396
3		0.085	0.306	0.342	0.361	0.329	0.313	0.331	0.221	0.224	0.225	0.332	0.364
4		0.086	0.308	0.346	0.365	0.347	0.333	0.329	0.231	0.223	0.221	0.390	0.383
5		0.078	0.306	0.352	0.359	0.322	0.348	0.289	0.233	0.225	0.211	0.383	0.397
6		0.082	0.344	0.340	0.370	0.323	0.343	0.305	0.244	0.224	0.217	0.348	0.340
7		0.097	0.268	0.318	0.301	0.309	0.284	0.290	0.226	0.219	0.219	0.311	0.392
8		0.086	0.300	0.338	0.349	0.318	0.317	0.303	0.229	0.220	0.219	0.345	0.381
Average		0.007	0.023	0.021	0.024	0.016	0.024	0.019	0.010	0.007	0.005	0.029	0.019
SD		0.000	0.214	0.252	0.263	0.232	0.230	0.217	0.143	0.134	0.132	0.258	0.294
Relative Cell Growth (%)		100.0	109.8	117.5	122.7	108.1	107.6	101.2	66.7	62.7	61.8	100.0	113.9
SD of Relative Cell Growth		10.5	11.4	9.6	11.0	7.5	11.1	8.8	4.7	3.1	2.5	13.7	9.0

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	0.00001	0.0001	0.001	0.01	0.1	1	10	100	1000	5000		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	13	12	10	6	11	11	10	6	0	0	15	67
Transformation frequency*	0.135416667	0.125	0.104166667	0.0625	0.114583333	0.114583333	0.104166667	0.0625	#DIV/0!	#DIV/0!	0.15625	0.697916667
SE	0.03492238	0.03375386	0.03117758	0.02470529	0.03250865	0.03250865	0.03117758	0.02470529	#DIV/0!	#DIV/0!	0.03705794	0.04686294

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Annex 11

**Results Submitted from Laboratories
in
the Phase II
of
Validation Study
of
96-Well Method**

Data Sheet for Cell Transformation Assay on Initiation Assay

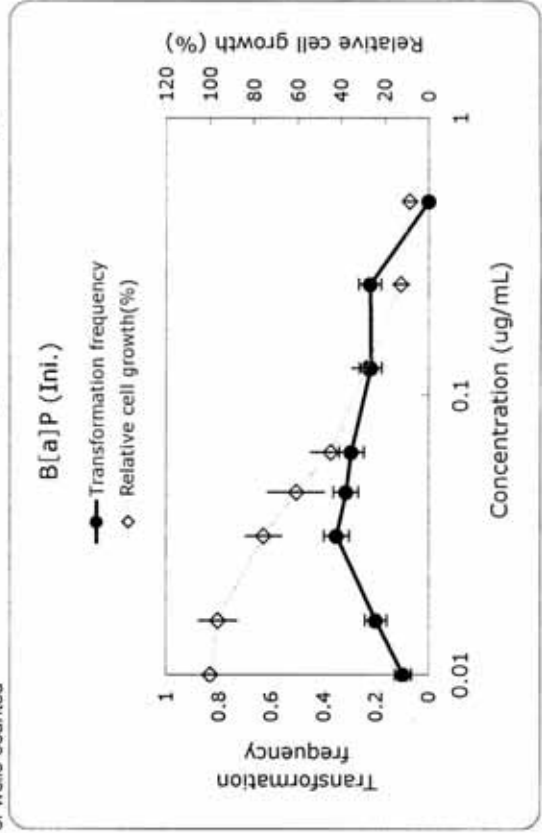
Chemical Code:	B[a]P
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)											Negative Control	Positive Control
Crystal Violet Staining	Blank	0.01	0.0156	0.0313	0.045	0.0625	0.125	0.25	0.5	0.1% DMSO			0.1% DMSO	MCA (1ug/mL)
1	0.072	0.665	0.691	0.462	0.298	0.248	0.186	0.102	0.094	0.665			0.665	0.254
2	0.073	0.680	0.702	0.550	0.562	0.367	0.295	0.172	0.153	0.680			0.680	0.445
3	0.072	0.716	0.675	0.568	0.465	0.395	0.261	0.153	0.130	0.716			0.716	0.377
4	0.075	0.688	0.635	0.492	0.433	0.416	0.205	0.149	0.143	0.688			0.688	0.300
5	0.079	0.739	0.619	0.570	0.420	0.387	0.243	0.177	0.123	0.739			0.739	0.358
6	0.080	0.694	0.610	0.528	0.502	0.379	0.296	0.156	0.142	0.694			0.694	0.329
7	0.082	0.696	0.773	0.568	0.430	0.354	0.286	0.174	0.140	0.696			0.696	0.353
8	0.078	0.703	0.698	0.626	0.521	0.290	0.257	0.164	0.115	0.703			0.703	0.410
Average	0.076	0.698	0.675	0.546	0.454	0.355	0.254	0.156	0.130	0.698			0.698	0.353
SD	0.004	0.023	0.054	0.051	0.080	0.057	0.041	0.024	0.019	0.023			0.023	0.060
Average - Blank	0.000	0.621	0.599	0.469	0.378	0.278	0.177	0.080	0.054	0.621			0.621	0.277
Relative Cell Growth (%)		100.0	96.4	75.5	60.8	44.8	28.5	12.8	8.6	100.0			100.0	44.6
SD of Relative Cell Growth		3.6	8.6	8.2	12.9	9.2	6.6	3.9	3.0	3.6			3.6	9.7

Transformation Assay		Concentrations (ug/ml)											Negative Control	Positive Control
Giemsa Staining		0.01	0.0156	0.0313	0.045	0.0625	0.125	0.25	0.5	0.1% DMSO			0.1% DMSO	MCA (1ug/mL)
No. of wells excluded		3	1	2	1	1	1	2	8	3			3	0
No. of wells counted		93	95	94	95	95	95	94	88	93			93	96
No. of wells having foci		9	19	33	30	28	21	21	TOX	9			9	50
Transformation frequency*		0.096774194	0.2	0.35106383	0.315789474	0.294736842	0.221052632	0.223404255	#VALUE!	0.096774194			0.096774194	0.520833333
SE		0.03065748	0.04103913	0.04923003	0.04769050	0.04677685	0.04257357	0.04296149	#VALUE!	0.03065748			0.03065748	0.05098672

* No. of wells having foci/ No. of wells counted

Remarks	



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

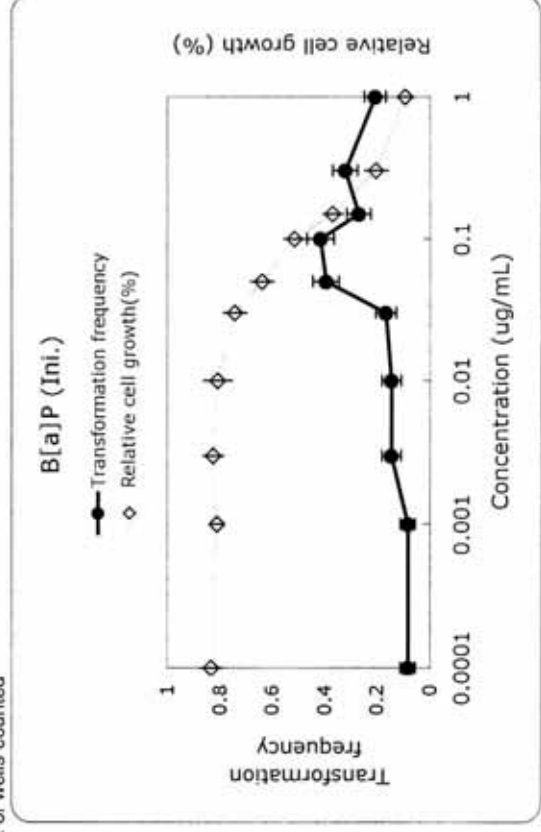
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	B[a]P
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{590nm}	Concentrations (ug/ml)											Negative Control		Positive Control	
		Blank	0.0001	0.001	0.003	0.01	0.03	0.05	0.1	0.15	0.3	1	0.1% DMSO	MCA (Iug/mL)		
1	0.128	1.319	1.393	1.368	1.246	1.160	1.168	1.022	0.622	0.337	0.224	1.319	0.817			
2	0.135	1.389	1.345	1.333	1.410	1.324	1.065	0.939	0.674	0.471	0.299	1.389	0.844			
3	0.135	1.385	1.384	1.456	1.499	1.288	1.025	0.898	0.679	0.445	0.250	1.385	0.826			
4	0.137	1.456	1.388	1.343	1.315	1.277	1.190	0.983	0.672	0.453	0.327	1.456	0.827			
5	0.135	1.399	1.398	1.447	1.475	1.229	1.201	0.846	0.679	0.575	0.252	1.399	0.922			
6	0.125	1.469	1.476	1.420	1.346	1.284	1.157	0.887	0.797	0.465	0.290	1.469	0.880			
7	0.117	1.527	1.388	1.423	1.421	1.326	1.065	0.870	0.720	0.386	0.281	1.527	0.813			
8	0.118	1.442	1.336	1.504	1.361	1.386	1.101	0.992	0.776	0.448	0.279	1.442	0.754			
Average	0.129	1.423	1.389	1.412	1.384	1.284	1.122	0.930	0.702	0.448	0.275	1.423	0.835			
SD	0.008	0.064	0.042	0.059	0.084	0.068	0.066	0.064	0.059	0.069	0.032	0.064	0.050			
Average - Blank	0.000	1.295	1.260	1.283	1.255	1.156	0.993	0.801	0.574	0.319	0.147	1.295	0.707			
Relative Cell Growth (%)		100.0	97.3	99.1	97.0	89.3	76.7	61.9	44.3	24.6	11.3	100.0	54.6			
SD of Relative Cell Growth		4.9	3.3	4.6	6.5	5.2	5.1	4.9	4.5	5.3	2.5	4.9	3.8			

Transformation Assay	Concentrations (ug/ml)											Negative Control		Positive Control	
Glensa Staining	0.0001	0.001	0.003	0.01	0.03	0.05	0.1	0.15	0.3	1	0.1% DMSO	MCA (Iug/mL)			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0			
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96			
No. of wells having foci	8	14	14	14	16	38	40	26	31	20	8	54			
Transformation frequency* SE	0.083333333 0.02820847	0.145833333 0.03602169	0.145833333 0.03602169	0.145833333 0.03602169	0.166666667 0.03803629	0.395833333 0.04991131	0.416666667 0.05031728	0.270833333 0.04535538	0.322916667 0.04723332	0.208333333 0.04144908	0.083333333 0.02820847	0.5625 0.05063079			

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

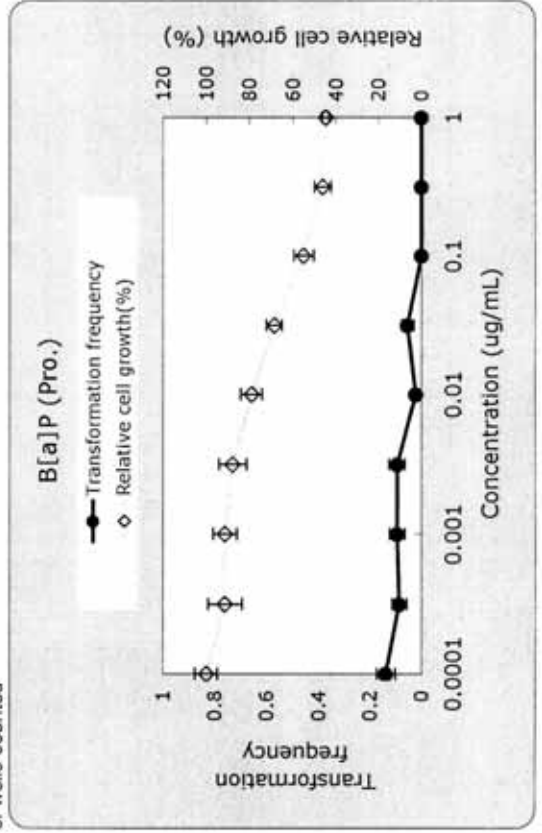
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	B[a]P
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining	Blank	0.0001	0.000316	0.001	0.00316	0.01	0.0316	0.1	0.316	1	0.1% DMSO	TPA (50ng/mL)			
1	0.074	0.672	0.658	0.579	0.597	0.543	0.473	0.374	0.323	0.314	0.672	0.860			
2	0.072	0.716	0.656	0.640	0.625	0.522	0.498	0.424	0.333	0.344	0.716	0.764			
3	0.075	0.634	0.645	0.621	0.597	0.531	0.444	0.441	0.358	0.338	0.634	0.883			
4	0.078	0.674	0.599	0.615	0.534	0.541	0.500	0.397	0.362	0.340	0.674	0.890			
5	0.079	0.633	0.621	0.553	0.614	0.601	0.465	0.400	0.320	0.342	0.633	0.932			
6	0.081	0.657	0.528	0.644	0.527	0.525	0.478	0.397	0.381	0.344	0.657	0.885			
7	0.078	0.622	0.594	0.591	0.608	0.507	0.461	0.381	0.358	0.341	0.622	0.917			
8	0.080	0.670	0.567	0.627	0.602	0.528	0.498	0.360	0.332	0.338	0.670	0.864			
Average	0.077	0.660	0.609	0.609	0.588	0.537	0.477	0.397	0.346	0.338	#DIV/0!	0.874			
SD	0.003	0.030	0.046	0.032	0.037	0.028	0.020	0.026	0.022	0.010	#DIV/0!	0.030			
Average - Blank	0.000	0.583	0.531	0.532	0.511	0.460	0.400	0.320	0.269	0.261	#DIV/0!	0.797			
Relative Cell Growth (%)		100.0	91.2	91.2	87.7	79.0	68.7	54.9	46.1	44.7	#DIV/0!	136.8			
SD of Relative Cell Growth		5.2	7.9	5.4	6.3	4.8	3.5	4.5	3.7	1.7	#DIV/0!	8.7			

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining	Blank	0.0001	0.000316	0.001	0.00316	0.01	0.0316	0.1	0.316	1	0.1% DMSO	TPA (50ng/mL)			
No. of wells excluded		1	2	0	1	1	3	8	3	96	1	0			
No. of wells counted		95	94	96	95	95	93	88	93	0	95	96			
No. of wells having foci		13	8	9	9	2	5	TOX	TOX	TOX	13	56			
Transformation frequency*		0.136842105	0.085106383	0.09375	0.094736842	0.021052632	0.053763441	#VALUE!	#VALUE!	#VALUE!	0.136842105	0.583333333			
SE		0.03526088	0.02878077	0.02974911	0.03004589	0.01472893	0.02338848	#VALUE!	#VALUE!	#VALUE!	0.03526088	0.05031728			

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

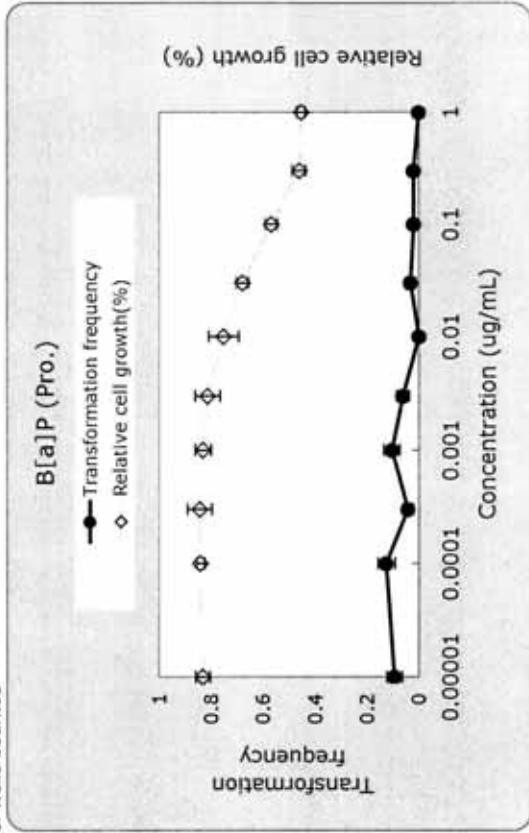
Chemical Code:	B[a]P
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining	Blank	0.00001	0.00003	0.0001	0.0003	0.01	0.03	0.1	0.3	1	1	0.1% DMSO	TPA (50ng/mL)		
1	0.128	1.250	1.212	1.286	1.267	1.081	1.026	0.863	0.714	0.736	1.250	1.552			
2	0.139	1.360	1.291	1.283	1.154	1.104	1.095	0.925	0.758	0.742	1.360	1.596			
3	0.138	1.270	1.337	1.336	1.255	1.339	1.105	0.957	0.757	0.789	1.270	1.633			
4	0.147	1.272	1.407	1.322	1.289	1.173	1.075	0.932	0.786	0.787	1.272	1.731			
5	0.128	1.296	1.380	1.342	1.238	1.205	1.070	0.906	0.805	0.741	1.296	1.708			
6	0.131	1.336	1.260	1.280	1.293	1.143	1.040	0.952	0.811	0.775	1.336	1.659			
7	0.123	1.266	1.280	1.293	1.394	1.213	1.112	0.943	0.819	0.774	1.266	1.764			
8	0.120	1.285	1.293	1.220	1.236	1.177	1.091	0.907	0.746	0.766	1.285	1.645			
Average	0.132	1.292	1.308	1.290	1.266	1.179	1.077	0.923	0.775	0.764	1.292	1.661			
SD	0.009	0.038	0.064	0.043	0.068	0.079	0.031	0.031	0.037	0.021	0.038	0.071			
Average - Blank	0.000	1.172	1.176	1.158	1.134	1.048	0.945	0.791	0.643	0.632	1.160	1.529			
Relative Cell Growth (%)	100.0	101.3	97.7	99.8	97.7	90.3	81.5	68.2	55.4	54.5	100.0	131.8			
SD of Relative Cell Growth	3.2	5.5	5.8	3.7	5.8	6.8	2.6	2.7	3.2	1.8	3.2	6.1			

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Glenssa Staining	Blank	0.00001	0.00003	0.0001	0.0003	0.01	0.03	0.1	0.3	1	0.1% DMSO	TPA (50ng/mL)			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0			
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96			
No. of wells having foci	9	12	4	10	6	0	3	2	2	Tox.	9	72			
Transformation frequency*	0.09375	0.125	0.041666667	0.104166667	0.0625	0	0.03125	0.020833333	0.020833333	#VALUE!	0.09375	0.75			
SE	0.02974911	0.03375386	0.02039469	0.03117758	0.02470529	0.00000000	0.01775805	0.01457713	0.01457713	#VALUE!	0.02974911	0.04419417			

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

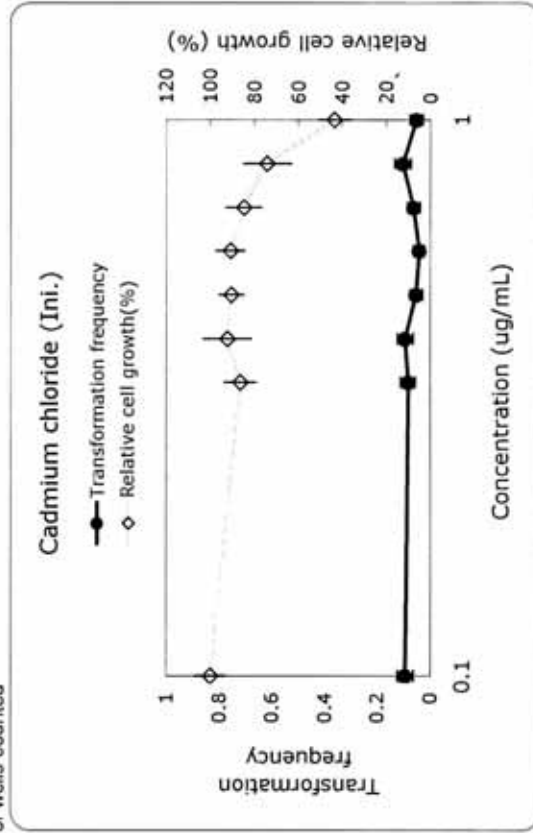
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Cadmium chloride
Institution:	Lab 1
Test Number	

Cell Growth Assay		Concentrations (ug/ml)									
Crystal Violet Staining OD _{540nm}	Blank	5% Water									
		0.1	0.335	0.402	0.482	0.579	0.694	0.833	1	Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
1	0.094	0.809	0.632	0.765	0.701	0.704	0.638	0.480	0.329	0.817	0.320
2	0.108	0.768	0.739	0.740	0.696	0.750	0.689	0.599	0.340	0.806	0.322
3	0.142	0.713	0.635	0.763	0.783	0.691	0.615	0.727	0.446	0.760	0.330
4	0.120	0.742	0.684	0.661	0.675	0.659	0.648	0.581	0.436	0.705	0.320
5	0.107	0.723	0.675	0.638	0.678	0.647	0.658	0.572	0.433	0.837	0.318
6	0.124	0.791	0.646	0.820	0.733	0.738	0.688	0.648	0.448	0.801	0.362
7	0.110	0.778	0.687	0.733	0.686	0.756	0.785	0.636	0.371	0.754	0.358
8	0.106	0.853	0.760	0.628	0.715	0.745	0.646	0.563	0.402	0.851	0.286
Average	0.114	0.772	0.682	0.721	0.708	0.711	0.671	0.601	0.401	#DIV/0!	0.327
SD	0.015	0.046	0.047	0.071	0.036	0.043	0.052	0.072	0.048	#DIV/0!	0.024
Average - Blank	0.000	0.658	0.568	0.607	0.595	0.597	0.557	0.487	0.287	#DIV/0!	0.213
Relative Cell Growth (%)		100.0	86.3	92.2	90.3	90.8	84.6	74.0	43.6	100.0	31.5
SD of Relative Cell Growth		7.1	7.1	10.8	5.4	6.5	7.9	11.0	7.3	7.4	3.7

Transformation Assay		Concentrations (ug/ml)											
Glensa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	5% Water									
				0.1	0.335	0.402	0.482	0.579	0.694	0.833	1	0	Negative Control
Transformation frequency*		0.09375	0.083333333	0.09375	0.052083333	0.041666667	0.0625	0.105263158	0.053191489	0	0	0.145833333	0.489583333
SE		0.02974911	0.02820847	0.02974911	0.02267769	0.02039469	0.02470529	0.03148648	0.02314665	0.00000000	0.00000000	0.03602169	0.05101996

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

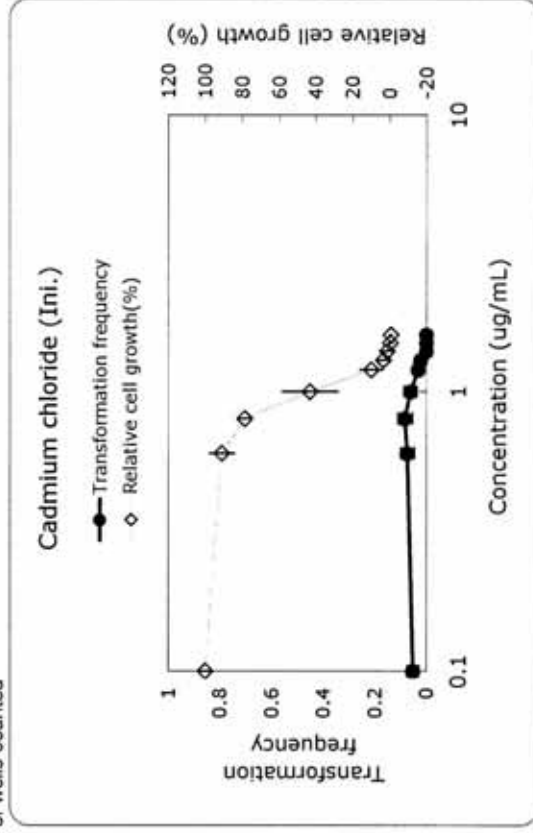
Chemical Code:	Cadmium chloride
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control
	Blank	5% Water										
Crystal Violet Staining		0.1	0.6	0.8	1	1.2	1.3	1.4	1.5	1.6		
OD _{540nm}												
1	0.107	1.314	1.090	1.075	0.570	0.169	0.116	0.097	0.092	0.094	1.154	0.517
2	0.108	1.286	1.325	1.030	0.612	0.328	0.118	0.101	0.102	0.101	1.203	0.615
3	0.109	1.246	1.216	1.062	0.933	0.226	0.131	0.128	0.099	0.100	1.309	0.608
4	0.114	1.339	1.137	1.060	0.762	0.318	0.143	0.114	0.100	0.100	1.219	0.736
5	0.119	1.326	1.192	1.079	0.746	0.147	0.148	0.132	0.107	0.107	1.347	0.665
6	0.115	1.355	1.252	0.993	0.626	0.175	0.260	0.210	0.138	0.118	1.293	0.530
7	0.113	1.275	1.146	1.152	0.500	0.316	0.145	0.132	0.117	0.120	1.435	0.596
8	0.115	1.382	1.284	0.998	0.328	0.191	0.179	0.118	0.114	0.113	1.377	0.580
Average	0.113	1.315	1.205	1.056	0.635	0.234	0.155	0.129	0.109	0.107	#DIV/0!	0.606
SD	0.004	0.045	0.080	0.051	0.183	0.075	0.047	0.035	0.014	0.009	#DIV/0!	0.095
Average - Blank	0.000	1.203	1.093	0.944	0.522	0.121	0.043	0.017	-0.004	-0.006	#DIV/0!	0.493
Relative Cell Growth (%)		100.0	90.8	78.4	43.4	10.1	3.5	1.4	-0.3	-0.5	#DIV/0!	100.0
SD of Relative Cell Growth		3.7	6.6	4.2	15.2	6.3	3.9	2.9	1.2	0.8	#DIV/0!	7.9

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glensa Staining	0.1	0.6	0.8	1	1.2	1.3	1.4	1.5	1.6	0		
No. of wells excluded	0	0	0	0	0	2	13	83	96	96	0	0
No. of wells counted	96	96	96	96	96	94	83	96	96	96	96	96
No. of wells having foci	5	7	8	6	3	2	Tox.	Tox.	Tox.	Tox.	7	57
Transformation frequency*	0.052083333	0.072916667	0.083333333	0.0625	0.03125	0.021276596	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.072916667	0.59375
SE	0.02267769	0.02653610	0.02820847	0.02470529	0.01775805	0.01488391	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.02653610	0.05012598

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

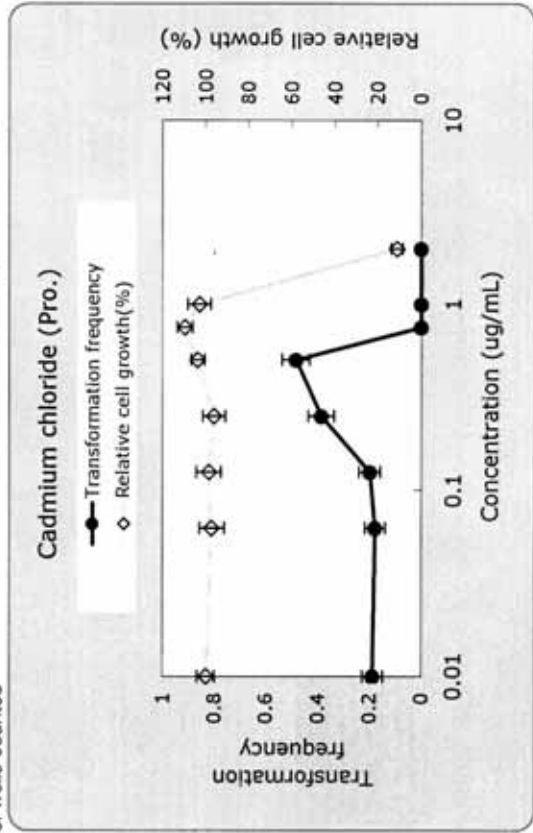
Chemical Code:	Cadmium chloride
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control	Positive Control
			5% Water	0.01	0.0625	0.125	0.25	0.5	0.75	1		
		0.108	0.733	0.766	0.700	0.778	0.829	0.859	0.781	0.181	0.783	1.135
		0.098	0.778	0.767	0.782	0.734	0.800	0.836	0.730	0.186	0.818	1.288
		0.112	0.819	0.769	0.734	0.788	0.800	0.870	0.803	0.201	0.819	1.107
		0.100	0.790	0.787	0.794	0.716	0.854	0.816	0.834	0.218	0.721	1.360
		0.120	0.792	0.715	0.812	0.709	0.769	0.814	0.819	0.187	0.694	1.265
		0.115	0.781	0.713	0.800	0.802	0.807	0.884	0.814	0.177	0.745	1.356
		0.129	0.779	0.784	0.780	0.772	0.828	0.828	0.831	0.159	0.734	1.312
		0.126	0.816	0.834	0.802	0.777	0.834	0.865	0.831	0.205	0.801	1.306
	Average	0.114	0.786	0.767	0.776	0.760	0.810	0.851	0.805	0.189	0.764	1.266
	SD	0.011	0.027	0.039	0.039	0.035	0.021	0.023	0.035	0.018	0.047	0.095
	Average - Blank	0.000	0.673	0.653	0.662	0.646	0.697	0.738	0.692	0.076	0.651	1.153
	Relative Cell Growth (%)	100.0	97.2	98.4	96.1	96.1	103.6	109.7	102.9	11.3	100.0	177.1
	SD of Relative Cell Growth	4.0	5.8	5.7	5.2	5.2	3.2	3.5	5.2	2.7	7.0	14.2

Transformation Assay	Concentrations (ug/ml)											
Giemsa Staining	0.01	0.0625	0.125	0.25	0.5	0.75	1	2	0	0	0	0
No. of wells excluded	0	0	0	0	5	25	63	96	0	0	0	0
No. of wells counted	96	96	96	96	91	71	33	0	96	96	96	96
No. of wells having foci	18	17	19	37	44	TOX	TOX	TOX	0	0	0	0
Transformation frequency*	0.1875	0.177083333	0.197916667	0.385416667	0.483516484	#VALUE!	#VALUE!	#VALUE!	0	0	0.177083333	0.802083333
SE	0.03983609	0.03896110	0.04066449	0.04967296	0.05238575	#VALUE!	#VALUE!	#VALUE!	0.00000000	0.00000000	0.03896110	0.04066449

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

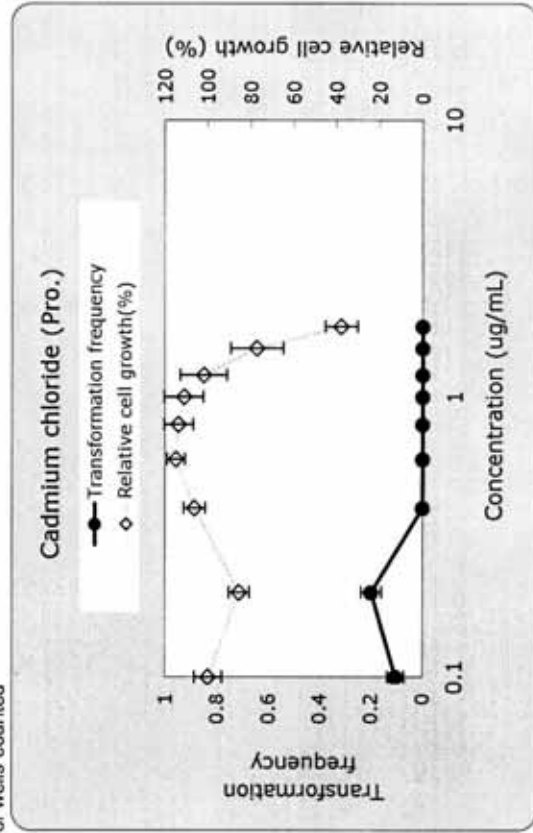
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Cadmium chloride
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)								Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
		Blank	5% Water 0.1	0.2	0.4	0.6	0.8	1	1.2		
1	0.108	1.216	1.058	1.415	1.409	1.410	1.370	1.264	0.996	1.284	1.780
2	0.118	1.237	1.110	1.306	1.437	1.338	1.224	1.110	0.950	1.144	2.119
3	0.118	1.310	1.097	1.323	1.483	1.374	1.504	1.254	1.049	1.281	2.028
4	0.115	1.241	1.081	1.302	1.493	1.442	1.365	1.292	0.726	1.253	1.994
5	0.118	1.296	1.111	1.425	1.429	1.506	1.438	1.359	1.161	1.298	1.813
6	0.114	1.298	1.238	1.426	1.558	1.507	1.558	1.505	1.189	1.315	2.132
7	0.118	1.461	1.159	1.332	1.464	1.501	1.518	1.477	1.062	1.345	2.016
8	0.122	1.308	1.151	1.436	1.508	1.574	1.437	1.297	1.084	1.320	2.000
Average	0.116	1.296	1.126	1.371	1.473	1.457	1.427	1.320	1.027	1.280	1.985
SD	0.004	0.076	0.056	0.060	0.048	0.079	0.107	0.127	0.145	0.062	0.128
Average - Blank	0.000	1.180	1.009	1.254	1.356	1.340	1.310	1.203	0.911	1.164	1.869
Relative Cell Growth (%)		100.0	85.6	106.3	115.0	113.6	111.1	102.0	77.2	100.0	160.6
SD of Relative Cell Growth		6.4	4.8	5.1	4.1	6.7	9.1	10.8	12.3	7.6	10.8

Transformation Assay	Concentrations (ug/ml)								Negative Control	Positive Control
Giemsa Staining	0.1	0.2	0.4	0.6	0.8	1	1.2	1.5		
No. of wells excluded	0	0	8							0
No. of wells counted	96	96	88							96
No. of wells having foci	10	19	25 (Tox.)							13
Transformation frequency*	0.104166667	0.197916667	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.135416667
SE	0.03117758	0.04066449	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.03492238

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Failure

The Progressing Data

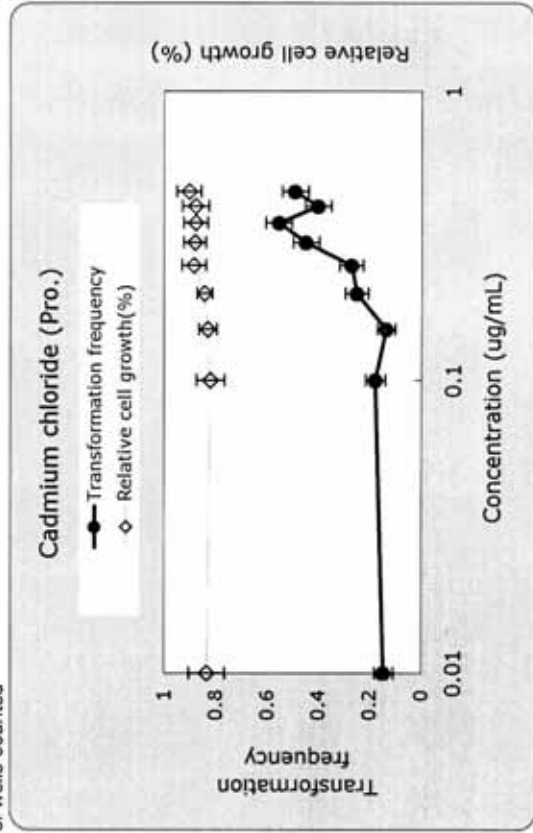
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Cadmium chloride
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{490nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
		Blank	5% Water 0.01	0.1	0.15	0.2	0.25	0.3	0.35	0.4	0.45		
1	0.081	1.189	1.127	1.127	1.127	1.168	1.239	1.175	1.250	1.336	1.107	1.440	
2	0.084	1.312	1.106	1.148	1.142	1.250	1.195	1.176	1.220	1.261	1.155	1.526	
3	0.090	1.227	1.102	1.255	1.189	1.141	1.214	1.302	1.311	1.461	1.352	1.461	
4	0.086	1.113	1.209	1.209	1.214	1.293	1.251	1.234	1.122	1.347	1.073	1.518	
5	0.085	1.194	1.162	1.165	1.185	1.329	1.233	1.247	1.211	1.255	1.144	1.551	
6	0.081	1.090	1.283	1.161	1.221	1.267	1.218	1.245	1.238	1.225	1.046	1.522	
7	0.086	1.238	1.210	1.169	1.174	1.236	1.358	1.317	1.328	1.266	1.103	1.632	
8	0.090	1.034	1.102	1.112	1.240	1.236	1.171	1.151	1.164	1.148	1.114	1.609	
Average	0.085	1.175	1.157	1.168	1.187	1.240	1.235	1.231	1.231	1.264	1.137	1.532	
SD	0.003	0.090	0.072	0.046	0.039	0.062	0.056	0.060	0.069	0.062	0.094	0.066	
Average - Blank	0.000	1.089	1.071	1.083	1.101	1.155	1.150	1.146	1.145	1.179	1.051	1.447	
Relative Cell Growth (%)		100.0	98.4	99.4	101.1	106.0	105.5	105.2	105.1	108.2	100.0	137.6	
SD of Relative Cell Growth		8.3	6.6	4.2	3.6	5.7	5.1	5.5	6.3	5.7	8.6	6.0	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glensa Staining	0.01	0.1	0.15	0.2	0.25	0.3	0.35	0.4	0.45	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	1	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	95	96	96	96	96
No. of wells having foci	14	17	13	24	26	43	53	38	47	7	7	60
Transformation frequency*	0.145833333	0.177083333	0.135416667	0.25	0.270833333	0.447916667	0.552083333	0.4	0.489583333	0	0.072916667	0.625
SE	0.03602169	0.03896110	0.03492238	0.04419417	0.04535538	0.05075342	0.05075342	0.05026247	0.05101996	0.00000000	0.02653610	0.04941059

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

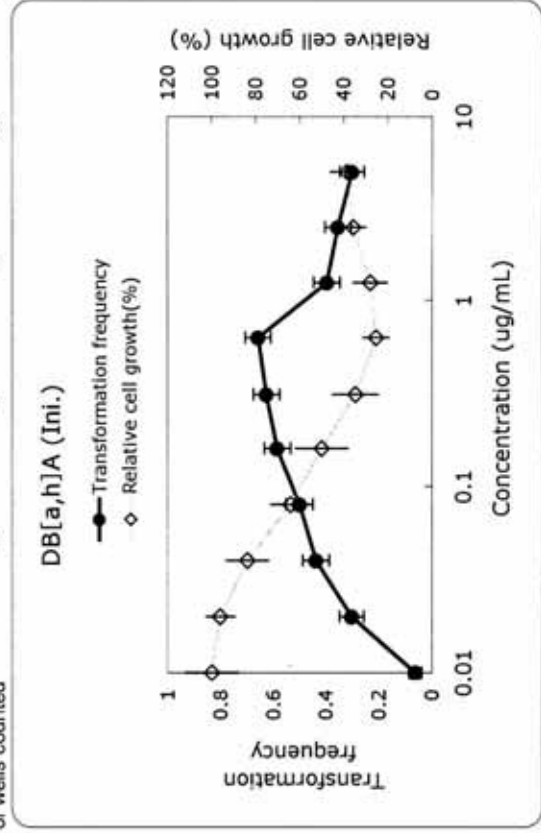
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	DB[a,h]A
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining	OD _{540nm}	Concentrations (ug/ml)										Negative Control	Positive Control
			Blank	0.1% DMSO	0.02	0.04	0.08	0.16	0.31	0.63	1.25	2.5		
1	0.105	0.215	0.211	0.229	0.193	0.188	0.152	0.149	0.156	0.159	0.159	0.215	0.174	
2	0.103	0.241	0.238	0.226	0.199	0.191	0.167	0.137	0.147	0.154	0.167	0.241	0.164	
3	0.097	0.240	0.235	0.206	0.190	0.173	0.137	0.137	0.129	0.149	0.152	0.240	0.171	
4	0.102	0.232	0.228	0.217	0.170	0.162	0.166	0.135	0.139	0.147	0.157	0.232	0.181	
5	0.102	0.249	0.220	0.202	0.174	0.161	0.136	0.128	0.131	0.141	0.145	0.249	0.159	
6	0.113	0.232	0.227	0.194	0.198	0.155	0.144	0.129	0.132	0.137	0.139	0.232	0.159	
7	0.099	0.244	0.231	0.202	0.173	0.150	0.145	0.126	0.130	0.148	0.141	0.244	0.154	
8	0.100	0.202	0.225	0.211	0.187	0.157	0.132	0.140	0.144	0.156	0.159	0.202	0.170	
Average	0.103	0.232	0.227	0.211	0.186	0.167	0.147	0.135	0.139	0.149	0.152	0.232	0.167	
SD	0.005	0.016	0.009	0.012	0.012	0.015	0.013	0.008	0.010	0.007	0.010	0.016	0.009	
Average - Blank	0.000	0.129	0.124	0.108	0.083	0.065	0.045	0.033	0.036	0.046	0.050	0.129	0.064	
Relative Cell Growth (%)		100.0	96.1	83.8	64.1	49.9	34.6	25.1	27.8	35.8	38.5	100.0	49.4	
SD of Relative Cell Growth		12.3	6.6	9.5	9.0	11.9	10.3	5.8	7.6	5.8	7.7	12.3	7.0	

Transformation Assay	Giemsa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	Transformation frequency*	SE	Concentrations (ug/ml)										Negative Control	Positive Control
							0.01	0.02	0.04	0.08	0.16	0.31	0.63	1.25	2.5	5		
		0	96	6	0.0625	0.02470529	0.01	0.02	0.04	0.08	0.16	0.31	0.63	1.25	2.5	5	0.0625	0.78125
		96	96	29	0.04686294	0.05063079	0.05103104	0.05031728	0.04941059	0.04881221	0.04686294	0.04941059	0.04881221	0.04686294	0.04686294	0.04686294	0.02470529	0.04219232

* No. of wells having foci/ No. of wells counted



DB[a,h]A (Ini.)

Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

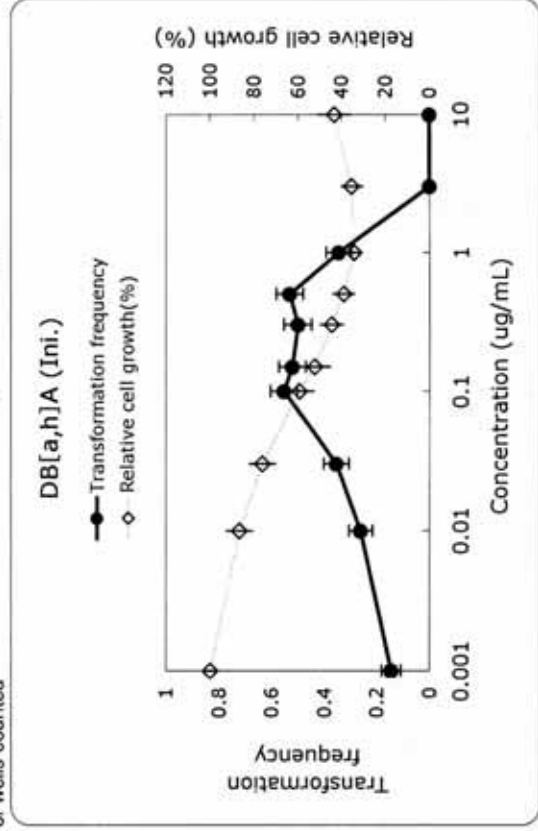
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	DB[a,h]A
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
			0.001	0.01	0.03	0.1	0.15	0.3	0.5	1	3	10		
1	0.119	0.001	1.215	1.083	0.825	0.887	0.722	0.573	0.511	0.604	0.622	1.390	0.785	
2	0.126	1.390	1.199	1.033	0.864	0.749	0.726	0.722	0.572	0.588	0.836	1.472	0.863	
3	0.127	1.472	1.282	1.263	0.916	0.873	0.611	0.582	0.572	0.520	0.687	1.359	1.040	
4	0.122	1.359	1.396	1.137	0.944	0.805	0.646	0.594	0.576	0.634	0.845	1.474	0.952	
5	0.123	1.474	1.269	1.143	0.887	0.651	0.779	0.626	0.541	0.575	0.643	1.462	0.870	
6	0.149	1.462	1.304	1.073	0.878	0.779	0.807	0.617	0.616	0.516	0.648	1.397	0.968	
7	0.128	1.397	1.251	1.181	1.079	0.801	0.723	0.651	0.617	0.571	0.609	1.474	0.924	
8	0.135	1.474	1.140	1.053	0.817	0.942	0.664	0.736	0.604	0.710	0.643	1.429	0.863	
Average	0.129	1.432	1.257	1.121	0.901	0.811	0.710	0.638	0.576	0.590	0.692	1.432	0.908	
SD	0.010	0.045	0.077	0.076	0.083	0.091	0.066	0.062	0.037	0.063	0.095	0.045	0.079	
Average - Blank	0.000	1.304	1.128	0.992	0.773	0.682	0.581	0.509	0.448	0.461	0.563	1.304	0.780	
Relative Cell Growth (%)		100.0	86.6	76.1	59.3	52.3	44.6	39.0	34.3	35.4	43.2	100.0	59.8	
SD of Relative Cell Growth		3.5	5.9	5.8	6.4	6.9	5.1	4.7	2.8	4.8	7.3	3.5	6.1	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Giemsa Staining	0.001	0.01	0.03	0.1	0.15	0.3	0.5	1	3	10			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	14	25	34	53	50	48	51	33	Tox.	Tox.	14	14	51
Transformation frequency*	0.145833333	0.260416667	0.354166667	0.552083333	0.520833333	0.5	0.53125	0.34375	#VALUE!	#VALUE!	0.145833333	0.53125	0.05093127
SE	0.03602169	0.04479116	0.04881221	0.05075342	0.05098672	0.05103104	0.05093127	0.04847529	#VALUE!	#VALUE!	0.03602169	0.05093127	0.05093127

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

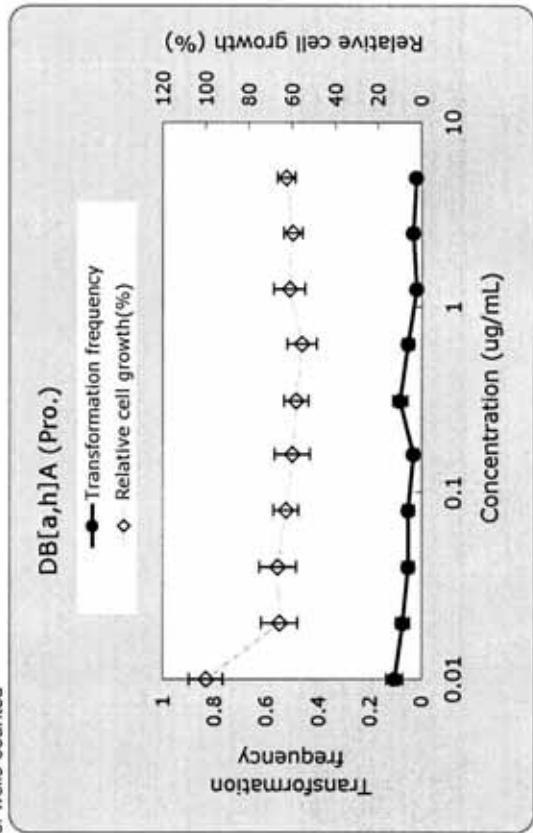
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	DB[a,h]A
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
	Blank	0.01	0.02	0.04	0.08	0.16	0.31	0.63	1.25	2.5	5	0.1% DMSO	TPA (50ng/mL)	
Crystal Violet Staining														
OD _{550nm}														
1	0.105	0.286	0.218	0.241	0.214	0.210	0.198	0.190	0.204	0.205	0.212	0.286	0.332	
2	0.103	0.292	0.234	0.253	0.234	0.235	0.223	0.221	0.216	0.224	0.210	0.292	0.370	
3	0.097	0.313	0.250	0.233	0.227	0.215	0.209	0.209	0.218	0.215	0.217	0.313	0.377	
4	0.102	0.286	0.240	0.231	0.237	0.216	0.215	0.216	0.219	0.221	0.223	0.286	0.383	
5	0.102	0.287	0.231	0.214	0.213	0.203	0.203	0.208	0.227	0.214	0.229	0.287	0.347	
6	0.113	0.303	0.220	0.229	0.215	0.240	0.207	0.219	0.243	0.222	0.232	0.303	0.348	
7	0.099	0.283	0.210	0.210	0.217	0.199	0.213	0.187	0.206	0.206	0.219	0.283	0.350	
8	0.100	0.264	0.204	0.207	0.206	0.201	0.204	0.200	0.203	0.204	0.216	0.264	0.333	
Average	0.103	0.289	0.226	0.227	0.220	0.215	0.211	0.206	0.217	0.214	0.220	0.289	0.355	
SD	0.005	0.014	0.016	0.016	0.011	0.015	0.010	0.013	0.013	0.008	0.008	0.014	0.019	
Average - Blank	0.000	0.187	0.123	0.125	0.118	0.112	0.109	0.104	0.114	0.111	0.117	0.187	0.252	
Relative Cell Growth (%)		100.0	66.0	66.8	63.1	60.1	58.3	55.5	61.3	59.6	62.8	100.0	135.2	
SD of Relative Cell Growth		7.7	8.4	8.6	5.9	8.2	5.5	6.9	7.2	4.3	4.2	7.7	10.4	

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
	Blank	0.01	0.02	0.04	0.08	0.16	0.31	0.63	1.25	2.5	5	0.1% DMSO	TPA (50ng/mL)	
Giemsa Staining														
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0	
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	
No. of wells having foci	10	7	5	5	5	3	8	5	2	3	2	10	50	
Transformation frequency*	0.104166667	0.072916667	0.052083333	0.052083333	0.052083333	0.03125	0.083333333	0.052083333	0.020833333	0.03125	0.020833333	0.104166667	0.520833333	
SE	0.03117758	0.02653610	0.02267769	0.02267769	0.02267769	0.01775805	0.02820847	0.02267769	0.01457713	0.01775805	0.01457713	0.03117758	0.05098672	

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

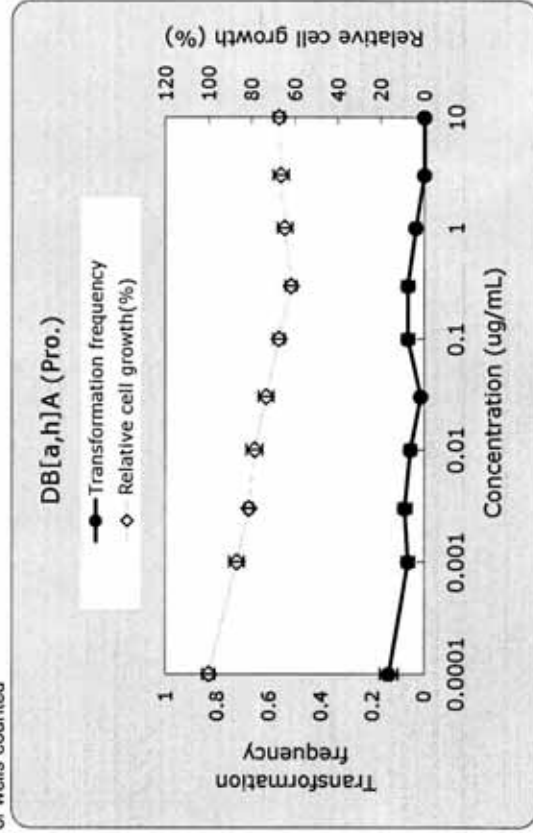
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	DB[a,h]A
Institution:	Lab 3
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)				
		0.0001	0.001	0.003	0.01	0.03	0.1	0.3	1	3	10						
Crystal Violet Staining																	
OD _{540nm}																	
1	0.111	1.311	1.162	1.110	1.056	0.993	0.911	0.868	0.917	0.906	0.939	1.311	1.735				
2	0.124	1.390	1.206	1.121	1.069	1.023	1.003	0.903	0.968	0.956	0.980	1.390	1.774				
3	0.121	1.360	1.186	1.110	1.162	1.101	0.955	0.921	0.932	0.957	0.951	1.360	1.816				
4	0.121	1.363	1.234	1.182	1.122	1.015	0.960	0.913	0.967	0.985	0.973	1.363	1.746				
5	0.109	1.346	1.258	1.124	1.107	1.066	0.973	0.834	0.933	0.990	1.015	1.346	1.615				
6	0.113	1.399	1.201	1.144	1.095	0.991	0.974	0.880	0.930	0.905	0.962	1.399	1.658				
7	0.108	1.417	1.237	1.123	1.148	1.075	0.965	0.921	0.894	0.980	0.962	1.417	1.607				
8	0.101	1.328	1.132	1.141	1.030	0.990	0.903	0.846	0.838	0.872	0.908	1.328	1.683				
Average	0.114	1.202	1.132	1.099	1.032	0.956	0.866	0.886	0.922	0.944	0.958	1.364	1.704				
SD	0.008	0.036	0.042	0.024	0.043	0.033	0.034	0.032	0.042	0.044	0.032	0.036	0.076				
Average - Blank	0.000	1.089	1.018	0.985	0.918	0.842	0.842	0.772	0.809	0.830	0.845	1.251	1.591				
Relative Cell Growth (%)		87.0	81.4	78.8	73.4	67.3	61.7	64.7	64.7	66.4	67.5	100.0	127.2				
SD of Relative Cell Growth		2.9	3.3	1.9	3.6	2.7	2.7	3.4	3.4	3.5	2.6	2.9	6.0				

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	0.0001	0.001	0.003	0.01	0.03	0.1	0.3	1	3	10			
Giemsa Staining													
No. of wells excluded	0	0	0	0	9	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	87	96	96	96	96	96	96	96	96
No. of wells having foci	13	6	7	5	1	6	6	3	0	0	0	13	73
Transformation frequency*	0.135416667	0.0625	0.072916667	0.052083333	0.011494253	0.0625	0.0625	0.03125	0	0	0	0.135416667	0.760416667
SE	0.03492238	0.02470529	0.02653610	0.02267769	0.01142800	0.02470529	0.02470529	0.01775805	0.00000000	0.00000000	0.00000000	0.03492238	0.04356307

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

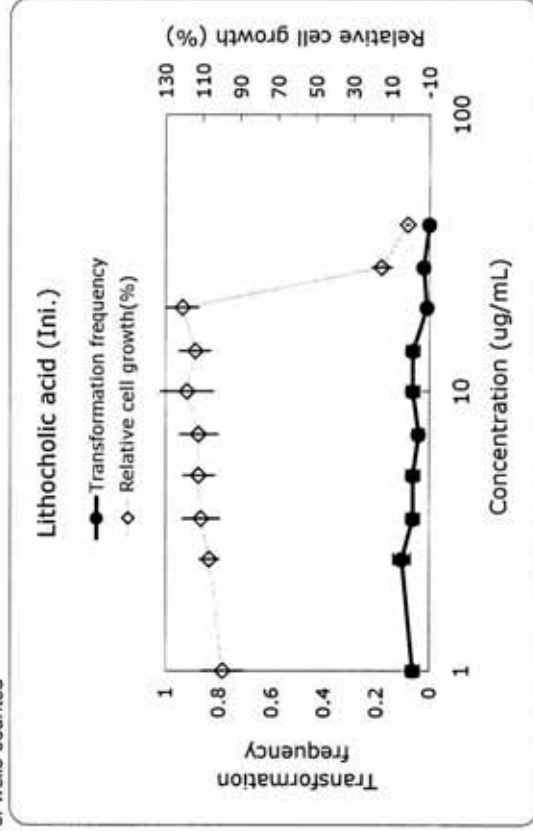
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Lithocholic acid
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/ml)			
	Blank	1	2.5	3.5	5	7	10	14	20	28			40		
Crystal Violet Staining OD _{540nm}															
1	0.052	0.236	0.269	0.261	0.292	0.300	0.305	0.284	0.316	0.086	0.060	0.236	0.190		
2	0.057	0.276	0.294	0.321	0.287	0.306	0.291	0.324	0.324	0.106	0.060	0.276	0.188		
3	0.060	0.275	0.279	0.296	0.320	0.302	0.332	0.295	0.325	0.085	0.060	0.275	0.168		
4	0.055	0.281	0.292	0.305	0.307	0.315	0.328	0.304	0.319	0.098	0.057	0.281	0.184		
5	0.055	0.280	0.285	0.283	0.295	0.289	0.318	0.307	0.312	0.083	0.055	0.280	0.184		
6	0.054	0.290	0.276	0.277	0.284	0.284	0.332	0.291	0.306	0.080	0.057	0.290	0.178		
7	0.052	0.247	0.272	0.297	0.279	0.285	0.268	0.287	0.294	0.086	0.060	0.247	0.208		
8	0.053	0.230	0.264	0.264	0.263	0.246	0.258	0.266	0.273	0.069	0.056	0.230	0.198		
Average	0.055	0.264	0.279	0.288	0.291	0.291	0.304	0.295	0.309	0.087	0.058	0.264	0.187		
SD	0.003	0.023	0.011	0.021	0.017	0.021	0.029	0.017	0.018	0.011	0.002	0.023	0.012		
Average - Blank	0.000	0.210	0.224	0.233	0.236	0.236	0.249	0.240	0.254	0.032	0.003	0.210	0.133		
Relative Cell Growth (%)		100.0	106.9	111.3	112.6	112.6	118.9	114.5	121.1	15.2	1.6	100.0	63.2		
SD of Relative Cell Growth		11.0	5.1	9.8	8.3	10.1	13.9	8.3	8.4	5.3	1.0	11.0	5.8		

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Giemsa Staining	1	2.5	3.5	5	7	10	14	20	28	40			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	6	10	6	6	4	6	6	6	2	2	6	6	52
Transformation frequency*	0.0625	0.104166667	0.0625	0.0625	0.041666667	0.0625	0.0625	0.0625	0.010416667	0.020833333	0.0625	0.0625	0.541666667
SE	0.02470529	0.03117758	0.02470529	0.02470529	0.02039469	0.02470529	0.02470529	0.02470529	0.01036227	0.01457713	0.02470529	0.02470529	0.05085354

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

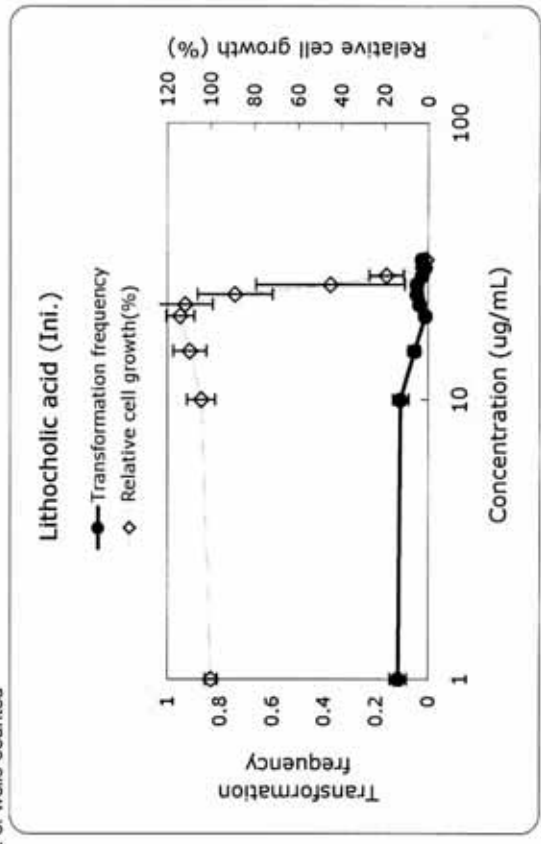
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Lithocholic acid
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control	Positive Control
			0.1% DMSO	10	15	20	22	24	26	28	30	32		
1	0.106	1.342	1.339	1.375	1.417	1.572	1.215	1.121	0.466	0.184	0.139	1.342	0.718	
2	0.113	1.364	1.390	1.354	1.575	1.611	1.437	1.165	0.358	0.164	0.128	1.364	0.838	
3	0.117	1.320	1.263	1.562	1.543	1.513	1.498	0.975	0.474	0.173	0.132	1.320	0.801	
4	0.116	1.297	1.361	1.593	1.609	1.522	1.332	0.885	0.374	0.132	0.126	1.297	0.734	
5	0.109	1.326	1.466	1.406	1.592	1.142	1.142	0.417	0.327	0.124	0.118	1.326	0.832	
6	0.112	1.383	1.484	1.358	1.576	1.167	0.914	0.357	0.300	0.133	0.115	1.383	0.775	
7	0.115	1.284	1.470	1.478	1.425	1.327	1.073	0.150	0.280	0.117	0.114	1.284	0.821	
8	0.119	1.352	1.327	1.502	1.481	1.501	0.971	0.211	0.186	0.115	0.116	1.352	0.713	
Average	0.113	1.334	1.388	1.453	1.507	1.476	1.198	0.660	0.346	0.143	0.124	1.334	0.779	
SD	0.004	0.033	0.080	0.094	0.078	0.152	0.213	0.419	0.096	0.027	0.009	0.033	0.052	
Average - Blank	0.000	1.220	1.274	1.340	1.394	1.362	1.084	0.547	0.232	0.029	0.010	1.220	0.666	
Relative Cell Growth (%)		100.0	104.4	109.8	114.2	111.6	88.9	44.8	19.0	2.4	0.8	100.0	54.6	
SD of Relative Cell Growth		2.7	6.5	7.7	6.4	12.5	17.4	34.4	7.8	2.2	0.8	2.7	4.2	

Transformation Assay	Giemsa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	Transformation frequency*	SE	Concentrations (ug/ml)										Negative Control	Positive Control
							1	10	15	20	22	24	26	28	30	32		
1	0	0	0	0	0.000	0	0	0	0	0	0	0	0	0	0	0		
96	96	96	96	96	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667	0.04166667		
11	10	5	1	3	0.03125	0.03125	0.03125	0.03125	0.03125	0.03125	0.03125	0.03125	0.03125	0.03125	0.03125	0.03125		
0.1145833333	0.1041666667	0.0520833333	0.0104166667	0.03125	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667	0.0416666667		
0.03250865	0.03117758	0.02267769	0.01036227	0.01775905	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469		

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

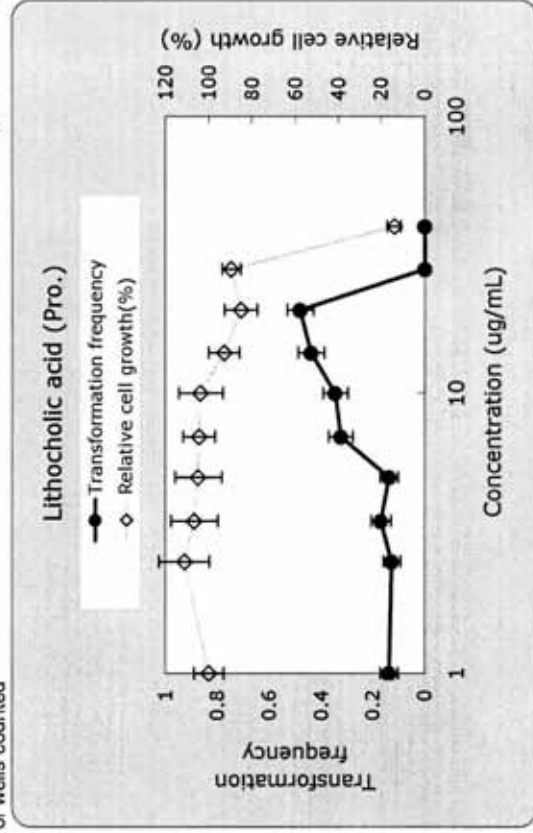
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Lithocholic acid
Institution:	Lab 2
Test Number	

Cell Growth Assay	Blank	Concentrations (ug/ml)										Positive Control				
		0.1% DMSO	2.5	3.5	5	7	10	14	20	28	40	0.1% DMSO	TPA (50ng/mL)			
Crystal Violet Staining	Blank															
OD _{540nm}																
1	0.052	0.306	0.280	0.267	0.268	0.295	0.293	0.268	0.249	0.293	0.081	0.306	0.392			
2	0.057	0.292	0.319	0.322	0.299	0.321	0.309	0.312	0.274	0.265	0.085	0.292	0.440			
3	0.060	0.304	0.346	0.329	0.335	0.323	0.344	0.293	0.249	0.294	0.089	0.304	0.417			
4	0.055	0.341	0.380	0.339	0.349	0.317	0.328	0.283	0.280	0.283	0.100	0.341	0.481			
5	0.055	0.317	0.333	0.355	0.338	0.334	0.345	0.299	0.303	0.266	0.095	0.317	0.482			
6	0.054	0.300	0.335	0.335	0.333	0.338	0.309	0.306	0.277	0.282	0.100	0.300	0.447			
7	0.052	0.306	0.321	0.333	0.325	0.324	0.326	0.283	0.265	0.276	0.091	0.306	0.457			
8	0.053	0.284	0.341	0.303	0.300	0.284	0.270	0.260	0.252	0.280	0.079	0.284	0.434			
Average	0.055	0.334	0.323	0.317	0.318	0.317	0.316	0.288	0.269	0.280	0.090	0.306	0.444			
SD	0.003	0.029	0.027	0.027	0.027	0.019	0.026	0.018	0.019	0.011	0.008	0.017	0.031			
Average - Blank	0.000	0.280	0.268	0.264	0.262	0.262	0.261	0.233	0.214	0.225	0.035	0.252	0.389			
Relative Cell Growth (%)		100.0	111.2	106.6	104.8	104.3	103.7	92.7	85.0	89.5	14.0	100.0	154.7			
SD of Relative Cell Growth		6.8	11.7	10.7	10.7	7.4	10.2	7.2	7.5	4.3	3.2	6.8	12.2			

Transformation Assay	Concentrations (ug/ml)										Positive Control	
Giemsa Staining	1	2.5	3.5	5	7	10	14	20	28	40	Negative Control	Positive Control
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	13	12	16	13	31	33	42	46	0	TOX	13	48
Transformation frequency*	0.135416667	0.125	0.166666667	0.135416667	0.322916667	0.34375	0.4375	0.479166667	0	#VALUE!	0.135416667	0.5
SE	0.03492238	0.03375386	0.03803629	0.03492238	0.04772332	0.04847529	0.05063079	0.05098672	0.00000000	#VALUE!	0.03492238	0.05103104

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

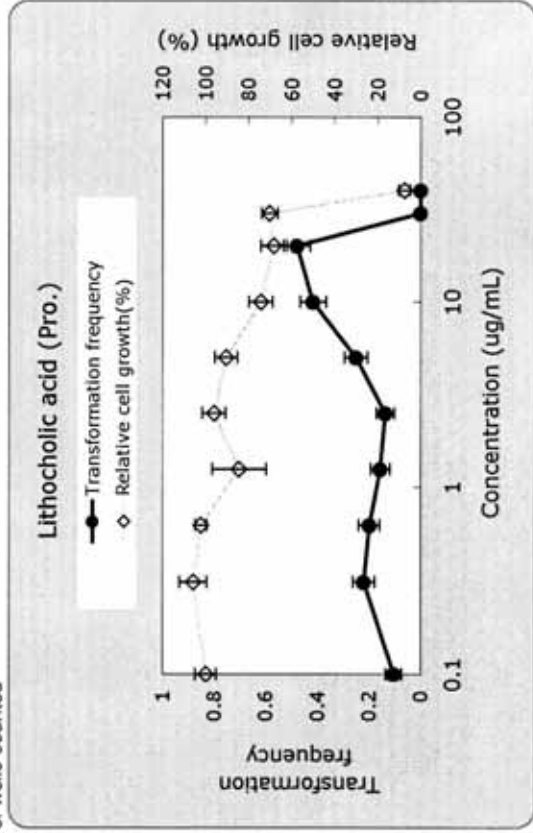
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Lithocholic acid
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	0.1	0.3125	0.625	1.25	2.5	5	10	20	30			40
Crystal Violet Staining													
OD _{540nm}													
1	0.118	1.286	1.321	1.344	1.308	1.299	1.198	1.010	1.044	1.028	0.178	1.286	1.718
2	0.125	1.337	1.533	1.400	0.972	1.335	1.211	1.186	1.065	0.981	0.199	1.337	1.589
3	0.128	1.337	1.353	1.373	1.026	1.362	1.210	1.082	1.023	1.023	0.261	1.337	1.749
4	0.125	1.309	1.399	1.403	1.335	1.345	1.361	0.992	1.017	1.021	0.179	1.309	1.680
5	0.121	1.403	1.493	1.395	0.997	1.256	1.303	1.014	0.927	1.016	0.267	1.403	1.579
6	0.119	1.423	1.418	1.369	1.083	1.385	1.229	0.991	0.916	0.914	0.205	1.423	1.707
7	0.117	1.439	1.515	1.438	1.314	1.198	1.223	1.021	0.855	0.980	0.204	1.439	1.665
8	0.106	1.295	1.377	1.338	1.247	1.243	1.147	0.983	0.920	0.919	0.200	1.295	1.582
Average	0.120	1.354	1.426	1.383	1.160	1.305	1.235	1.035	0.961	0.985	0.212	1.354	1.659
SD	0.007	0.060	0.079	0.033	0.156	0.068	0.067	0.068	0.073	0.046	0.034	0.060	0.067
Average - Blank	0.000	1.234	1.306	1.263	1.040	1.186	1.115	0.915	0.841	0.865	0.092	1.234	1.539
Relative Cell Growth (%)		100.0	105.9	102.3	84.3	96.1	90.4	74.2	68.2	70.1	7.4	100.0	124.7
SD of Relative Cell Growth		4.9	6.4	2.7	12.6	5.5	5.4	5.5	5.9	3.8	2.8	4.9	5.4

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Glenssa Staining	0.1	0.3125	0.625	1.25	2.5	5	10	20	30	40			0.1% DMSO
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	10	21	19	15	13	24	40	46	0	0	10	10	57
Transformation frequency*	0.104166667	0.21875	0.197916667	0.15625	0.135416667	0.25	0.416666667	0.479166667	0	0	0.104166667	0.59375	
SE	0.03117758	0.04219232	0.04066449	0.03705794	0.03492238	0.04419417	0.05031728	0.05098672	0.00000000	0.00000000	0.03117758	0.05012598	

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

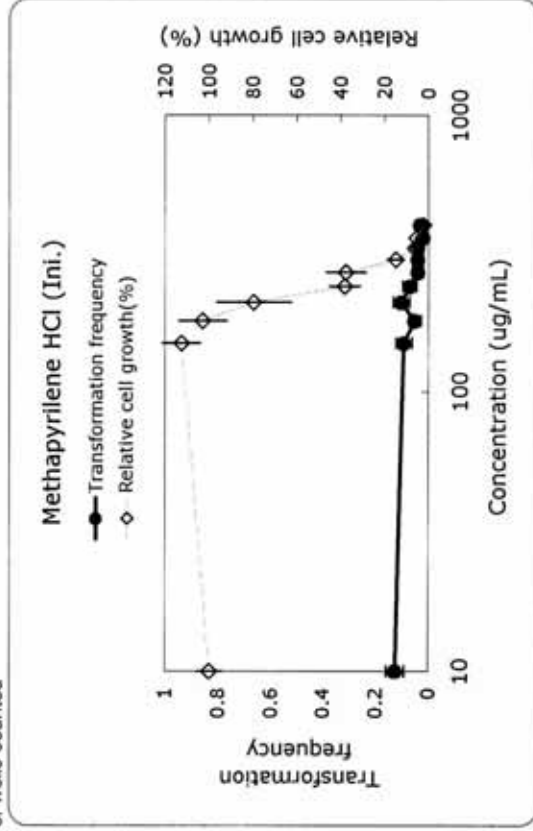
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Methapyrilene HCl
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/ml)			
	Blank	5% Water 10	150	180	210	240	270	300	330	360			400		
Crystal Violet Staining															
OD _{540nm}															
1	0.065	0.407	0.412	0.436	0.286	0.179	0.160	0.098	0.077	0.079	0.070	0.400	0.235		
2	0.062	0.409	0.480	0.398	0.340	0.197	0.190	0.116	0.089	0.080	0.071	0.346	0.286		
3	0.060	0.398	0.433	0.369	0.279	0.201	0.167	0.107	0.081	0.079	0.070	0.347	0.273		
4	0.059	0.432	0.471	0.402	0.469	0.181	0.173	0.097	0.080	0.079	0.069	0.400	0.259		
5	0.060	0.415	0.498	0.478	0.305	0.191	0.211	0.132	0.086	0.076	0.070	0.391	0.256		
6	0.060	0.409	0.458	0.480	0.355	0.203	0.196	0.127	0.089	0.081	0.070	0.396	0.258		
7	0.060	0.463	0.501	0.442	0.368	0.178	0.210	0.117	0.087	0.085	0.067	0.350	0.269		
8	0.065	0.421	0.459	0.428	0.367	0.254	0.261	0.132	0.084	0.087	0.067	0.347	0.249		
Average	0.061	0.419	0.464	0.429	0.346	0.198	0.196	0.116	0.084	0.081	0.069	0.372	0.261		
SD	0.002	0.020	0.031	0.039	0.061	0.025	0.032	0.014	0.004	0.004	0.001	0.026	0.016		
Average - Blank	0.000	0.358	0.403	0.368	0.285	0.137	0.135	0.054	0.023	0.019	0.008	0.311	0.199		
Relative Cell Growth (%)		100.0	112.5	102.8	79.6	38.2	37.6	15.2	6.4	5.4	2.2	100.0	64.1		
SD of Relative Cell Growth		5.7	8.6	10.8	17.0	6.9	9.1	4.0	1.2	1.0	0.4	7.4	4.3		

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Giemsa Staining	10	150	180	210	240	270	300	330	360	400			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	12	9	5	10	7	4	4	4	2	3	3	11	88
Transformation frequency*	0.125	0.09375	0.052083333	0.104166667	0.072916667	0.041666667	0.041666667	0.041666667	0.020833333	0.03125	0.03125	0.114583333	0.708333333
SE	0.03375386	0.02974911	0.02267769	0.03117758	0.02853610	0.02039469	0.02039469	0.02039469	0.01457713	0.01775805	0.01775805	0.03250865	0.04639024

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

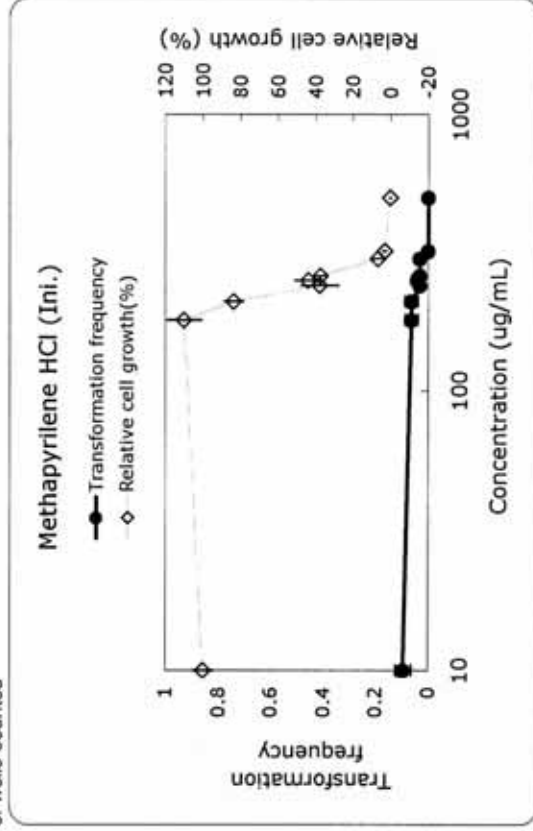
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Methapyrilene HCl
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (Tug/mL)
		Blank	5% Water	180	210	240	250	260	300	320	500		
1	0.122	1.247	1.553	1.054	0.528	0.600	0.523	0.223	0.152	0.126	1.276	0.705	
2	0.124	1.214	1.345	1.024	0.349	0.641	0.533	0.239	0.162	0.131	1.301	0.696	
3	0.123	1.231	1.253	1.087	0.454	0.487	0.586	0.197	0.158	0.128	1.308	0.739	
4	0.127	1.326	1.393	1.087	0.701	0.638	0.561	0.210	0.185	0.125	1.248	0.725	
5	0.123	1.389	1.464	1.139	0.568	0.667	0.520	0.227	0.164	0.123	1.320	0.833	
6	0.125	1.319	1.496	1.104	0.668	0.772	0.603	0.181	0.158	0.121	1.305	0.753	
7	0.118	1.276	1.264	1.026	0.600	0.598	0.549	0.170	0.138	0.117	1.274	0.803	
8	0.118	1.222	1.386	1.195	0.620	0.656	0.571	0.165	0.150	0.125	1.301	0.706	
Average	0.123	1.278	1.394	1.090	0.561	0.632	0.556	0.202	0.158	0.125	1.292	0.745	
SD	0.003	0.062	0.107	0.058	0.116	0.080	0.030	0.028	0.014	0.004	0.024	0.049	
Average - Blank	0.000	1.156	1.272	0.967	0.439	0.510	0.433	0.079	0.036	0.002	1.169	0.623	
Relative Cell Growth (%)		100.0	110.1	83.7	37.9	44.1	37.5	6.8	3.1	0.2	100.0	53.2	
SD of Relative Cell Growth		5.3	9.2	5.0	10.0	6.9	2.6	2.4	1.2	0.4	2.0	4.3	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	10	180	210	240	250	260	300	320	500	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	9	6	6	3	4	3	3	0	Tox.	0	12	46
Transformation frequency*	0.09375	0.0625	0.0625	0.03125	0.041666667	0.03125	0.03125	0.03125	0	#VALUE!	0.125	0.479166667
SE	0.02974911	0.02470529	0.02470529	0.01775805	0.02039469	0.01775805	0.01775805	0.01775805	0.00000000	#VALUE!	0.03375386	0.050998672

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Methapyrilene HCl
Institution:	Lab 2
Test Number	

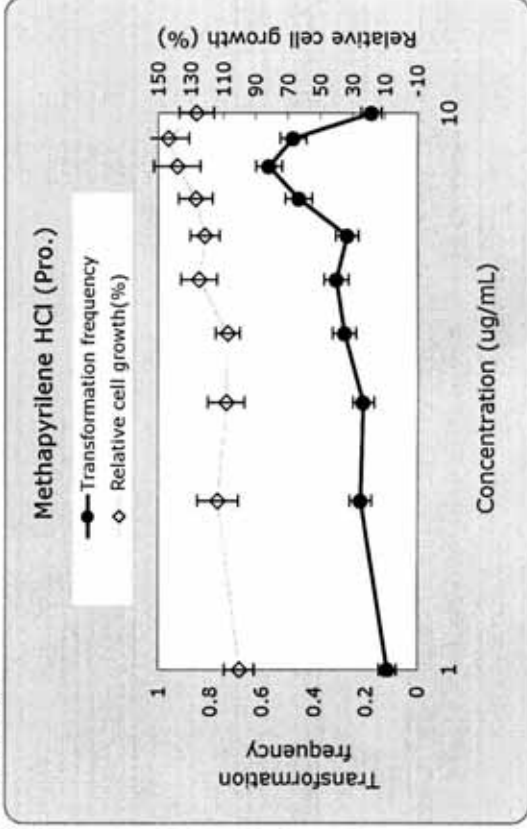
Cell Growth Assay		Concentrations (ug/ml)																				
Crystal Violet Staining OD _{510nm}	Blank	5% Water		8.8		13		20		30		44		67		100		150		Negative Control		Positive Control
		µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	TPA (50ng/mL)	
1	0.104	0.342	0.356	0.365	0.365	0.424	0.405	0.395	0.503	0.467	0.481	0.439	0.469	0.484	0.437	0.341	0.434	0.319	0.420	0.319	0.420	0.420
2	0.107	0.386	0.420	0.383	0.365	0.448	0.418	0.440	0.490	0.494	0.426	0.428	0.038	0.034	0.028	0.016	0.430	0.344	0.430	0.344	0.430	0.430
3	0.104	0.395	0.401	0.416	0.388	0.404	0.442	0.441	0.514	0.513	0.437	0.437	0.367	0.382	0.240	0.448	0.448	0.361	0.448	0.361	0.448	0.448
4	0.107	0.384	0.457	0.425	0.403	0.467	0.472	0.456	0.487	0.475	0.465	0.465	0.403	0.464	0.447	0.436	0.436	0.361	0.436	0.361	0.436	0.436
5	0.097	0.389	0.432	0.416	0.406	0.460	0.402	0.481	0.403	0.464	0.446	0.460	0.469	0.524	0.440	0.453	0.453	0.350	0.447	0.350	0.447	0.447
6	0.100	0.365	0.399	0.362	0.414	0.458	0.422	0.460	0.469	0.524	0.440	0.460	0.458	0.510	0.406	0.453	0.453	0.339	0.453	0.339	0.453	0.453
7	0.095	0.331	0.392	0.397	0.381	0.411	0.428	0.414	0.458	0.510	0.406	0.414	0.458	0.510	0.406	0.436	0.436	0.333	0.436	0.333	0.436	0.436
8	0.098	0.346	0.363	0.343	0.366	0.390	0.400	0.421	0.427	0.421	0.396	0.421	0.427	0.421	0.400	0.400	0.400	0.322	0.400	0.322	0.400	0.400
Average	0.102	0.367	0.403	0.388	0.386	0.433	0.424	0.439	0.469	0.484	0.437	0.439	0.469	0.484	0.437	0.341	0.434	0.319	0.420	0.319	0.420	0.420
SD	0.005	0.025	0.034	0.030	0.020	0.029	0.024	0.028	0.038	0.034	0.028	0.028	0.038	0.034	0.028	0.016	0.430	0.344	0.430	0.344	0.430	0.430
Average - Blank	0.000	0.266	0.301	0.287	0.285	0.331	0.322	0.337	0.367	0.382	0.336	0.337	0.367	0.382	0.240	0.332	0.332	0.240	0.332	0.240	0.332	0.332
Relative Cell Growth (%)		100.0	113.3	107.9	107.1	124.6	121.2	126.8	138.2	143.7	126.3	126.8	138.2	143.7	100.0	138.7	138.7	100.0	138.7	100.0	138.7	138.7
SD of Relative Cell Growth		9.3	12.7	11.3	7.5	11.0	9.1	10.4	14.4	12.7	10.6	10.4	14.4	12.7	6.0	6.5	6.5	6.0	6.5	6.0	6.5	6.5

Transformation Assay		Concentrations (ug/ml)																				
Giemsa Staining	µg/mL	5.9		8.8		13		20		30		44		67		100		150		Negative Control		Positive Control
		µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	µg/mL	TPA (50ng/mL)	
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	11	21	20	27	30	30	26	44	55	46	17	44	55	46	17	49	49	12	49	12	49	49
Transformation frequency*	0.114583333	0.21875	0.208333333	0.28125	0.3125	0.270833333	0.458333333	0.458333333	0.572916667	0.479166667	0.177083333	0.458333333	0.572916667	0.479166667	0.177083333	0.510416667	0.510416667	0.03375386	0.510416667	0.03375386	0.510416667	0.510416667
SE	0.03250865	0.04219232	0.04144908	0.04588805	0.04730704	0.04535538	0.05085354	0.05085354	0.05098672	0.05098672	0.03896110	0.05085354	0.05098672	0.05098672	0.03896110	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386	0.03375386

* No. of wells having foci/ No. of wells counted

Remarks

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Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

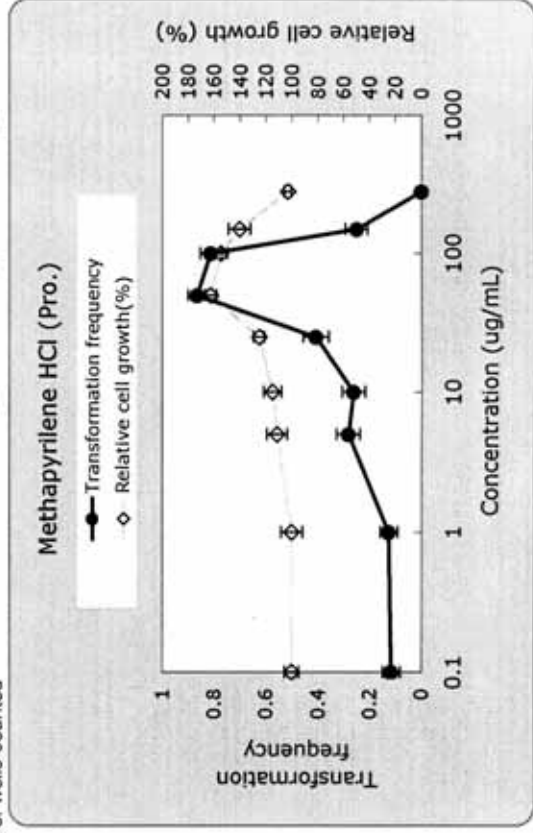
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Methapyrilene HCl
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	5% Water	1	5	10	25	50	100	150	280			0.1% DMSO
Crystal Violet Staining													
OD _{540nm}													
1	0.124	1.236	1.222	1.250	1.448	1.600	2.051	1.914	1.691	1.329	1.216	1.701	
2	0.134	1.266	1.301	1.467	1.372	1.540	2.023	1.958	1.757	1.349	1.331	1.521	
3	0.131	1.306	1.180	1.435	1.473	1.674	2.147	1.919	1.647	1.351	1.487	1.696	
4	0.129	1.303	1.299	1.499	1.475	1.651	2.053	1.881	1.807	1.344	1.419	1.801	
5	0.122	1.399	1.331	1.427	1.642	1.663	2.071	1.952	1.957	1.392	1.360	1.624	
6	0.122	1.288	1.304	1.568	1.432	1.545	2.022	2.020	1.845	1.286	1.334	1.678	
7	0.127	1.426	1.511	1.423	1.498	1.622	2.088	2.071	1.764	1.364	1.251	1.653	
8	0.113	1.278	1.359	1.498	1.564	1.606	1.968	1.990	1.873	1.390	1.396	1.691	
Average	0.125	1.313	1.313	1.446	1.488	1.613	2.053	1.963	1.793	1.351	1.349	1.671	
SD	0.007	0.066	0.099	0.093	0.083	0.051	0.053	0.062	0.100	0.034	0.088	0.079	
Average - Blank	0.000	1.188	1.321	1.321	1.363	1.487	1.928	1.838	1.667	1.225	1.224	1.545	
Relative Cell Growth (%)		100.0	100.1	111.2	114.8	125.3	162.3	154.8	140.4	103.2	100.0	126.3	
SD of Relative Cell Growth		5.5	8.3	7.8	7.0	4.3	4.4	5.2	8.4	2.9	7.4	6.7	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	0.1	1	5	10	25	50	100	150	280	0		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	11	12	27	25	39	83	78	24	Tox.	14	86	86
Transformation frequency*	0.114583333	0.125	0.28125	0.260416667	0.40625	0.864583333	0.8125	0.25	#VALUE!	0	0.145833333	0.895833333
SE	0.03250865	0.03375386	0.04588805	0.04479116	0.05012598	0.03492238	0.03983609	0.04419417	#VALUE!	0.00000000	0.03602169	0.03117758

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

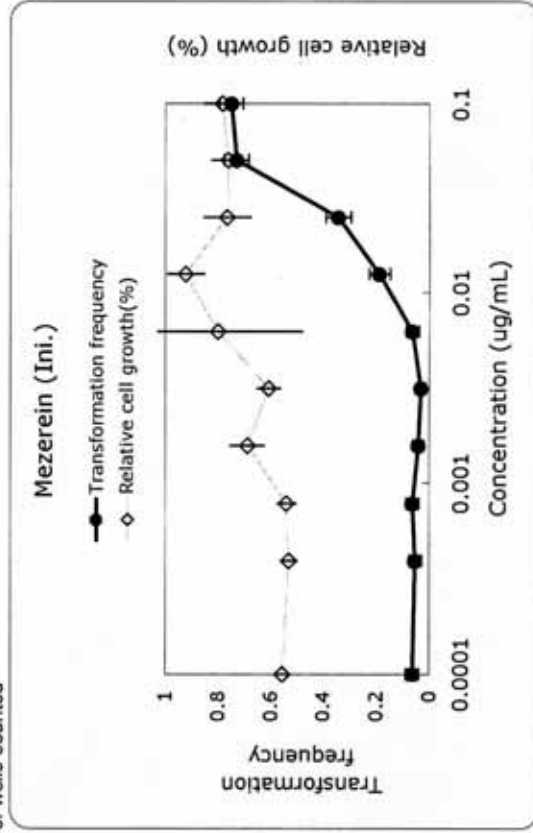
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Mezerein
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)												Negative Control		Positive Control
			0.000391	0.000781	0.001563	0.003125	0.00625	0.0125	0.025	0.05	0.1	0.1% DMSO	0.1	0.1	0.1	0.1	MCA (1ug/mL)
1	0.105	0.290	0.285	0.370	0.432	0.400	0.321	0.432	0.392	0.394	0.421	0.290	0.421	0.290	0.174		
2	0.103	0.290	0.280	0.381	0.435	0.459	0.323	0.435	0.403	0.388	0.361	0.302	0.361	0.302	0.180		
3	0.097	0.309	0.305	0.353	0.411	0.419	0.323	0.411	0.357	0.363	0.395	0.306	0.395	0.306	0.176		
4	0.102	0.303	0.294	0.356	0.423	0.434	0.352	0.423	0.414	0.363	0.370	0.302	0.370	0.302	0.173		
5	0.102	0.277	0.279	0.326	0.478	0.310	0.310	0.478	0.316	0.331	0.407	0.302	0.407	0.302	0.170		
6	0.113	0.295	0.314	0.339	0.312	0.429	0.312	0.395	0.376	0.395	0.363	0.303	0.363	0.303	0.178		
7	0.099	0.281	0.296	0.312	0.416	0.400	0.303	0.416	0.380	0.357	0.375	0.281	0.375	0.281	0.173		
8	0.100	0.284	0.300	0.339	0.448	0.437	0.306	0.448	0.354	0.386	0.351	0.311	0.351	0.311	0.181		
Average	0.103	0.291	0.294	0.347	0.430	0.386	0.319	0.430	0.374	0.372	0.380	0.300	0.380	0.300	0.176		
SD	0.005	0.011	0.012	0.023	0.015	0.113	0.015	0.025	0.031	0.022	0.025	0.010	0.025	0.010	0.004		
Average - Blank	0.000	0.189	0.192	0.244	0.327	0.284	0.216	0.327	0.271	0.270	0.278	0.197	0.278	0.197	0.073		
Relative Cell Growth (%)		95.7	97.2	124.0	109.7	143.9	166.1	137.8	15.9	11.4	141.0	100.0	141.0	100.0	37.1		
SD of Relative Cell Growth		4.8	6.3	11.5	7.9	57.3	12.9	15.9	12.5	11.4	12.5	4.8	12.5	4.8	1.9		

Transformation Assay	Giemsa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	Transformation frequency* SE	Concentrations (ug/ml)												Negative Control		Positive Control
						0.0001	0.000391	0.000781	0.001563	0.003125	0.00625	0.0125	0.025	0.05	0.1	0.1% DMSO	0.1	0.1	0.1	0.1
		0	0	0	0.000391	0.000781	0.001563	0.003125	0.00625	0.0125	0.025	0.05	0.1	0.1	0.1	0.1	0.1	0.1		
		96	96	96	0.0625	0.0625	0.041666667	0.03125	0.0625	0.1875	0.34375	0.729166667	0.75	0.75	0.75	0.75	0.75	0.75		
		6	5	6	0.02267769	0.02470529	0.02039469	0.01775805	0.02470529	0.03983609	0.04847529	0.04535538	0.04419417	0.04419417	0.04419417	0.04419417	0.04419417	0.04419417		
		100.0	95.7	97.2	124.0	109.7	143.9	166.1	137.8	15.9	11.4	141.0	100.0	141.0	100.0	141.0	100.0	37.1		
		4.8	6.3	11.5	7.9	57.3	12.9	15.9	12.5	11.4	12.5	4.8	12.5	4.8	1.9	1.9	1.9	1.9		

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

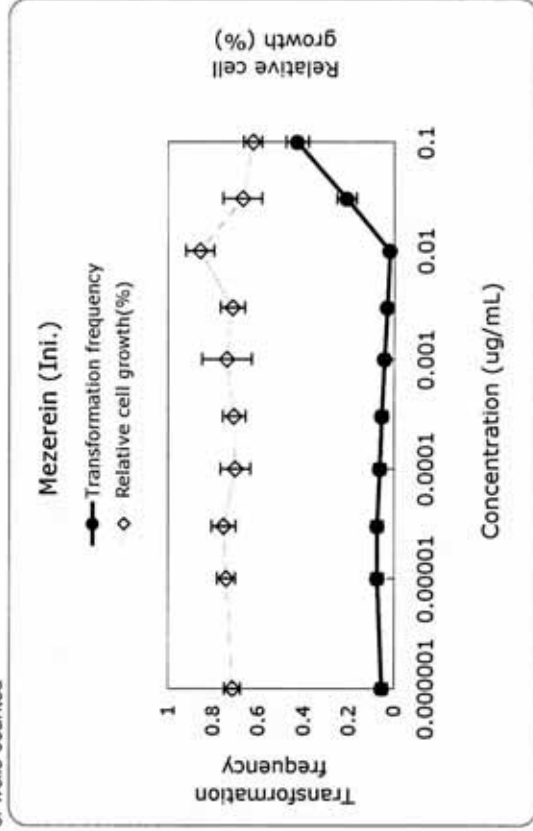
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Mezerein
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Crystal Violet Staining	Blank	0.000001	0.000003	0.00001	0.00003	0.0001	0.0003	0.001	0.003	0.01	0.03	0.1	0.1	0.1% DMSO	MCA (1ug/ml)
1	0.116	1.248	1.229	1.342	1.283	1.325	1.118	1.308	1.298	1.242	1.219	1.242	1.248	1.248	0.768
2	0.123	1.352	1.405	1.364	1.462	1.290	1.292	1.338	1.618	1.342	1.342	1.161	1.352	1.352	0.785
3	0.130	1.397	1.422	1.335	1.195	1.338	1.470	1.271	1.499	1.424	1.424	1.184	1.397	1.397	0.680
4	0.125	1.253	1.261	1.368	1.409	1.414	1.283	1.371	1.573	1.093	1.093	1.124	1.253	1.253	0.780
5	0.133	1.332	1.460	1.382	1.317	1.242	1.357	1.394	1.561	1.302	1.302	1.173	1.332	1.332	0.901
6	0.123	1.250	1.411	1.248	1.235	1.332	1.312	1.269	1.586	1.319	1.319	1.197	1.250	1.250	0.814
7	0.126	1.259	1.333	1.369	1.185	1.208	1.713	1.117	1.597	1.083	1.083	1.137	1.259	1.259	0.674
8	0.115	1.330	1.419	1.270	1.167	1.170	1.213	1.340	1.585	1.045	1.045	1.019	1.330	1.330	0.649
Average	0.124	1.347	1.368	1.282	1.282	1.290	1.345	1.301	1.540	1.228	1.228	1.155	1.303	1.303	0.756
SD	0.006	0.066	0.088	0.108	0.079	0.079	0.180	0.086	0.104	0.140	0.140	0.066	0.057	0.057	0.084
Average - Blank	0.000	1.223	1.244	1.158	1.166	1.166	1.221	1.177	1.416	1.031	1.031	0.874	1.179	1.179	0.633
Relative Cell Growth (%)		103.8	105.5	98.2	98.9	98.9	103.6	99.9	120.1	87.4	87.4	87.4	100.0	100.0	53.7
SD of Relative Cell Growth		4.9	7.4	9.2	6.7	6.7	15.3	7.3	8.8	11.9	11.9	5.6	4.9	4.9	7.2

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control	
Glenssa Staining	Blank	0.000001	0.000003	0.00001	0.00003	0.0001	0.0003	0.001	0.003	0.01	0.03	0.1	0.1	0.1% DMSO	MCA (1ug/ml)
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	5	7	7	6	5	5	4	3	2	2	20	41	5	5	66
Transformation frequency*	0.052083333	0.072916667	0.072916667	0.0625	0.052083333	0.041666667	0.03125	0.020833333	0.020833333	0.020833333	0.208333333	0.427083333	0.052083333	0.052083333	0.6875
SE	0.02267769	0.02653610	0.02653610	0.02470529	0.02267769	0.02039469	0.01775805	0.01457713	0.0144908	0.05048547	0.02267769	0.04730704	0.02267769	0.02267769	0.04730704

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

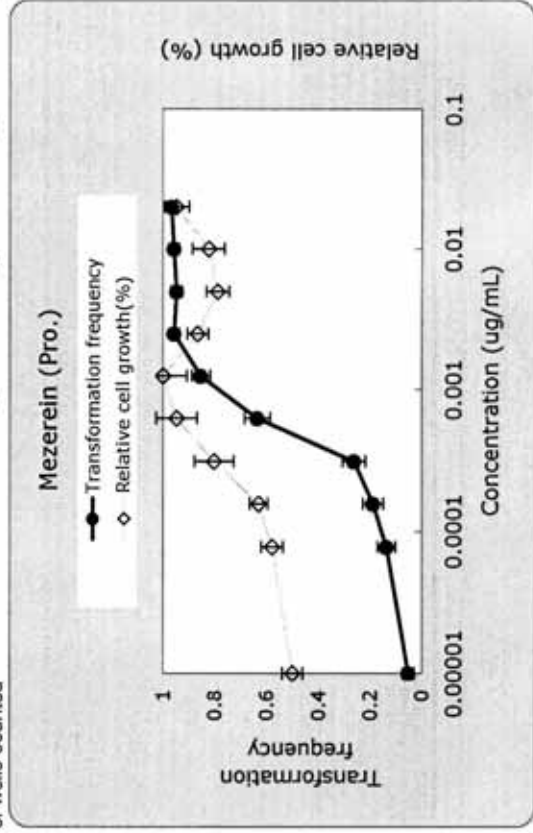
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Mezerein
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control		Positive Control
	Blank	0.00001	0.000078	0.000156	0.000313	0.000625	0.00125	0.0025	0.005	0.01	0.02	0.1% DMSO	TPA (50ng/mL)
Crystal Violet Staining													
OD _{540nm}													
1	0.052	0.285	0.311	0.330	0.368	0.461	0.500	0.492	0.462	0.460	0.511	0.285	0.444
2	0.057	0.299	0.351	0.346	0.469	0.505	0.501	0.425	0.411	0.445	0.481	0.299	0.478
3	0.060	0.303	0.308	0.367	0.456	0.490	0.512	0.468	0.418	0.475	0.481	0.303	0.486
4	0.055	0.308	0.328	0.342	0.470	0.487	0.620	0.460	0.413	0.419	0.481	0.308	0.482
5	0.055	0.279	0.332	0.370	0.452	0.527	0.551	0.472	0.434	0.438	0.541	0.279	0.469
6	0.054	0.310	0.352	0.358	0.397	0.567	0.503	0.452	0.403	0.439	0.498	0.310	0.508
7	0.052	0.277	0.332	0.364	0.425	0.516	0.514	0.457	0.414	0.387	0.527	0.277	0.441
8	0.053	0.258	0.299	0.331	0.422	0.452	0.497	0.470	0.449	0.473	0.486	0.258	0.416
Average	0.055	0.290	0.327	0.351	0.432	0.501	0.525	0.462	0.426	0.442	0.501	0.290	0.466
SD	0.003	0.018	0.019	0.016	0.036	0.037	0.042	0.019	0.021	0.029	0.023	0.018	0.030
Average - Blank	0.000	0.296	0.272	0.296	0.378	0.446	0.470	0.407	0.371	0.387	0.446	0.235	0.411
Relative Cell Growth (%)		100.0	115.6	126.0	160.6	189.6	199.9	173.2	157.7	164.7	189.7	100.0	174.7
SD of Relative Cell Growth		7.7	8.3	6.8	15.5	15.7	17.9	8.2	8.8	12.4	9.9	7.7	12.6

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control
	Blank	0.00001	0.000078	0.000156	0.000313	0.000625	0.00125	0.0025	0.005	0.01	0.02	0.1% DMSO	TPA (50ng/mL)
Giemsa Staining													
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	5	13	18	25	61	82	92	91	92	92	93	5	46
Transformation frequency*	0.052083333	0.135416667	0.1875	0.260416667	0.635416667	0.854166667	0.958333333	0.947916667	0.958333333	0.958333333	0.96875	0.052083333	0.479166667
SE	0.02267769	0.03492238	0.03983609	0.04479116	0.04912382	0.03602169	0.02039469	0.02267769	0.02039469	0.02039469	0.01775805	0.02267769	0.05098672

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

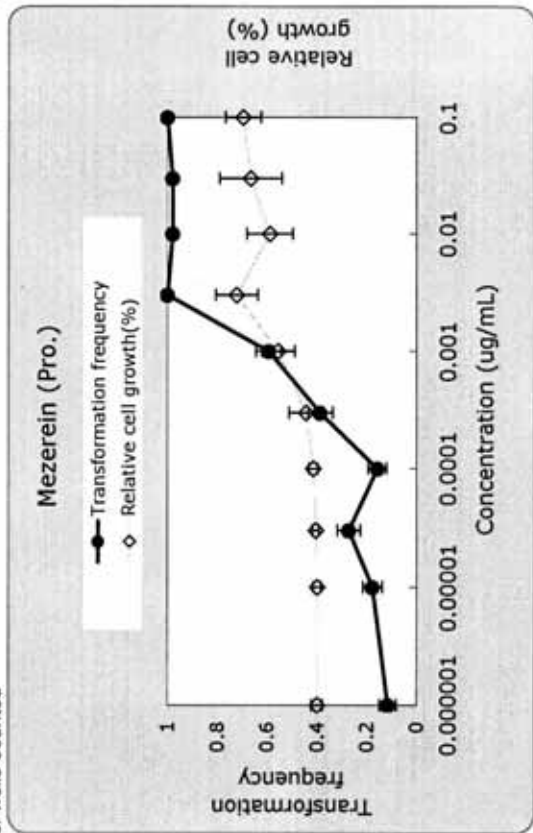
Chemical Code:	Mezerein
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)												
		Blank	0.000001	0.000003	0.00001	0.00003	0.0001	0.0003	0.001	0.01	0.03	0.1	Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
Crystal Violet Staining	OD _{540nm}													
1	0.123	1.326	1.285	1.360	1.244	1.580	2.101	1.650	2.089	2.524	1.712	1.326	1.825	
2	0.134	1.332	1.281	1.272	1.311	1.526	1.783	2.274	2.028	2.379	2.426	1.332	1.837	
3	0.138	1.310	1.298	1.316	1.350	1.478	1.882	2.327	1.952	2.231	2.150	1.310	1.734	
4	0.134	1.305	1.239	1.303	1.321	1.470	1.470	2.180	1.824	2.076	2.290	1.305	1.901	
5	0.126	1.302	1.267	1.341	1.411	1.541	1.781	2.365	1.907	2.085	2.192	1.302	1.627	
6	0.128	1.354	1.396	1.343	1.346	1.505	1.656	2.313	1.912	1.943	2.206	1.354	1.578	
7	0.127	1.312	1.314	1.339	1.405	1.486	1.751	2.452	1.854	2.073	2.092	1.312	1.618	
8	0.113	1.143	1.297	1.230	1.353	1.304	1.586	2.339	1.211	1.293	2.226	1.143	1.539	
Average	0.128	1.298	1.297	1.313	1.343	1.424	1.751	2.238	1.847	2.076	2.162	1.298	1.707	
SD	0.008	0.065	0.046	0.044	0.053	0.200	0.192	0.250	0.271	0.368	0.207	0.065	0.135	
Average - Blank	0.000	1.169	1.185	1.185	1.215	1.296	1.623	2.110	1.719	1.948	2.034	1.170	1.580	
Relative Cell Growth (%)		100.0	99.9	101.3	103.8	110.8	138.7	180.3	146.9	166.4	173.8	100.0	135.0	
SD of Relative Cell Growth		5.5	3.9	3.7	4.6	17.1	16.4	21.3	23.2	31.4	17.7	5.5	11.6	

Transformation Assay		Concentrations (ug/ml)											
Glenssa Staining	0.000001	0.000003	0.00001	0.00003	0.0001	0.0003	0.001	0.003	0.01	0.03	0.1	Negative Control 0.1% DMSO	Positive Control
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	11	17	26	15	57	94	94	94	94	94	96	11	48
Transformation frequency*	0.114583333	0.177083333	0.270833333	0.15625	0.385416667	0.59375	1	0.979166667	0.979166667	0.979166667	1	0.114583333	0.5
SE	0.03250865	0.03896110	0.04535538	0.03705794	0.04967296	0.05012598	0.00000000	0.01457713	0.01457713	0.01457713	0.00000000	0.03250865	0.05103104

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

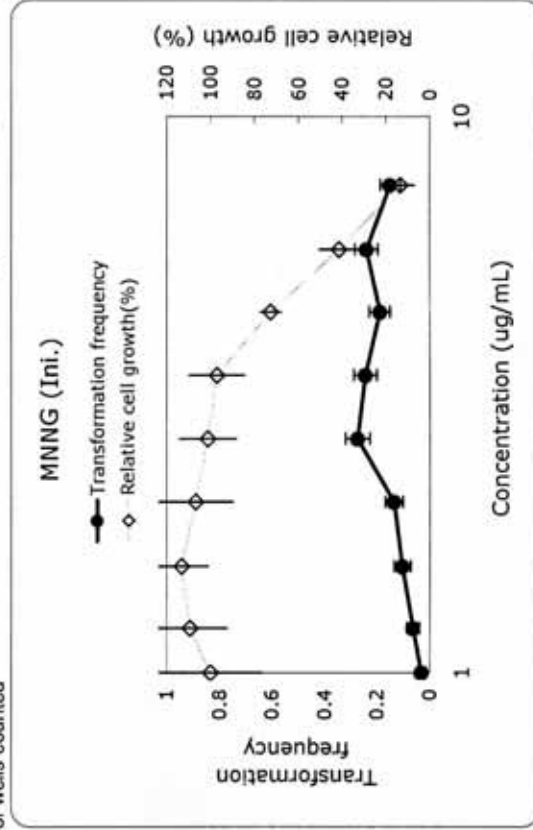
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	MNNG
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/ml)	
	Blank	5% Water	1.2	1.55	2.02	2.62	3.41	4.43	5.75	7.5			
Crystal Violet Staining OD _{540nm}													
1	0.090	0.733	0.758	0.659	0.777	0.728	0.769	0.515	0.321	0.144	0.681	0.323	
2	0.096	0.638	0.814	0.731	0.795	0.756	0.685	0.509	0.332	0.154	0.740	0.341	
3	0.088	0.480	0.757	0.796	0.710	0.686	0.684	0.551	0.438	0.181	0.700	0.364	
4	0.090	0.607	0.578	0.776	0.627	0.695	0.609	0.463	0.263	0.172	0.789	0.340	
5	0.087	0.554	0.576	0.692	0.573	0.640	0.633	0.518	0.315	0.127	0.769	0.300	
6	0.089	0.610	0.663	0.695	0.594	0.517	0.597	0.486	0.331	0.150	0.772	0.350	
7	0.086	0.809	0.797	0.694	0.699	0.641	0.528	0.487	0.278	0.248	0.684	0.343	
8	0.089	0.877	0.790	0.867	0.832	0.696	0.663	0.519	0.326	0.159	0.776	0.298	
Average	0.089	0.664	0.717	0.739	0.701	0.670	0.646	0.506	0.326	0.167	0.739	0.332	
SD	0.003	0.133	0.097	0.069	0.096	0.073	0.072	0.027	0.052	0.037	0.044	0.024	
Average - Blank	0.000	0.574	0.627	0.649	0.612	0.581	0.557	0.417	0.236	0.078	0.650	0.243	
Relative Cell Growth (%)		100.0	109.3	113.1	106.5	101.1	97.0	72.6	41.1	13.5	100.0	37.4	
SD of Relative Cell Growth		23.2	17.0	12.1	16.8	12.7	12.5	4.7	9.1	6.4	7.7	4.1	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Giemsa Staining	1	1.2	1.55	2.02	2.62	3.41	4.43	5.75	7.5	0		
No. of wells excluded	0	0	1	0	1	2	1	0	2	0	0	2
No. of wells counted	96	96	95	96	95	94	95	96	94	96	96	94
No. of wells having foci	3	6	10	13	26	23	18	23	14	14	9	44
Transformation frequency*	0.03125	0.0625	0.105263158	0.135416667	0.273684211	0.244680851	0.189473684	0.239583333	0.14893617	0	0.09375	0.468085106
SE	0.01775805	0.02470529	0.03148648	0.03492238	0.04574312	0.04434057	0.04020649	0.04356307	0.03672123	0.00000000	0.02974911	0.05146590

* No. of wells having foci/ No. of wells counted



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass
The Final Data

Remarks

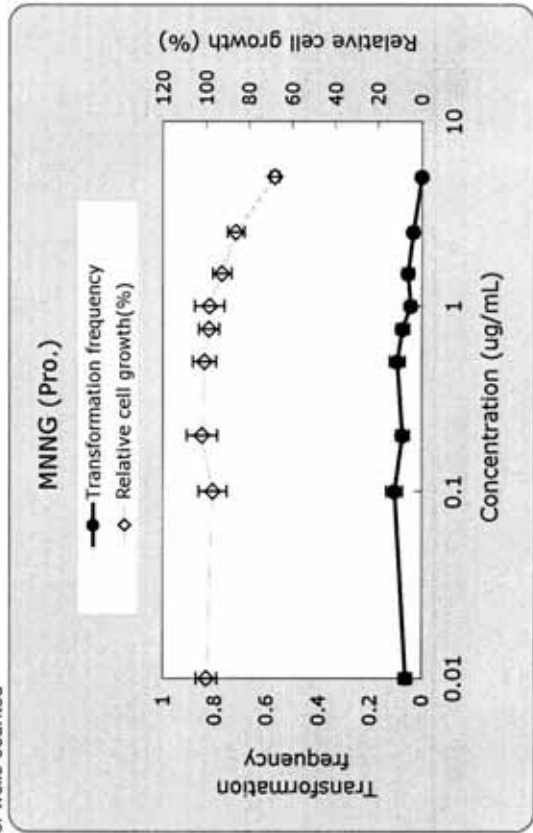
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	MNNG
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
		Blank	5% Water 0.01	0.1	0.2	0.5	0.75	1	1.5	2.5	5		
1	0.116		1.173	1.131	1.206	1.229	1.223	1.227	1.162	1.116	0.924	1.271	1.595
2	0.122		1.271	1.288	1.314	1.290	1.261	1.099	1.117	1.046	0.927	1.260	1.530
3	0.123		1.255	1.222	1.277	1.258	1.285	1.304	1.220	1.164	0.866	1.216	1.738
4	0.128		1.354	1.293	1.249	1.196	1.181	1.215	1.275	1.086	0.893	1.287	1.752
5	0.123		1.310	1.300	1.402	1.343	1.247	1.318	1.174	1.104	0.923	1.269	1.727
6	0.124		1.267	1.328	1.213	1.281	1.265	1.263	1.170	1.164	0.916	1.365	1.870
7	0.121		1.269	1.254	1.429	1.321	1.367	1.325	1.235	1.111	0.945	1.198	1.780
8	0.117		1.329	1.136	1.320	1.381	1.286	1.331	1.205	1.178	0.898	1.507	1.801
Average	0.122		1.279	1.244	1.301	1.287	1.264	1.260	1.195	1.121	0.912	1.297	1.724
SD	0.004		0.055	0.075	0.082	0.061	0.054	0.079	0.049	0.045	0.025	0.099	0.111
Average - Blank	0.000		1.157	1.122	1.180	1.166	1.143	1.139	1.073	0.999	0.790	1.175	1.602
Relative Cell Growth (%)			100.0	97.0	102.0	100.8	98.8	98.4	92.8	86.4	68.3	100.0	136.4
SD of Relative Cell Growth			4.8	6.5	7.1	5.2	4.7	6.8	4.3	3.9	2.1	8.5	9.6

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Giemsa Staining	0.01	0.1	0.2	0.5	0.75	1	1.5	2.5	5	0			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	6	10	7	9	7	4	5	3	0	0	0	7	62
Transformation frequency*	0.0625	0.104166667	0.072916667	0.09375	0.072916667	0.041666667	0.052083333	0.03125	0	0	0	0.072916667	0.645833333
SE	0.02470529	0.03117758	0.02853610	0.02974911	0.02853610	0.02039469	0.02267769	0.01775905	0.00000000	0.00000000	0.00000000	0.02853610	0.04881221

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

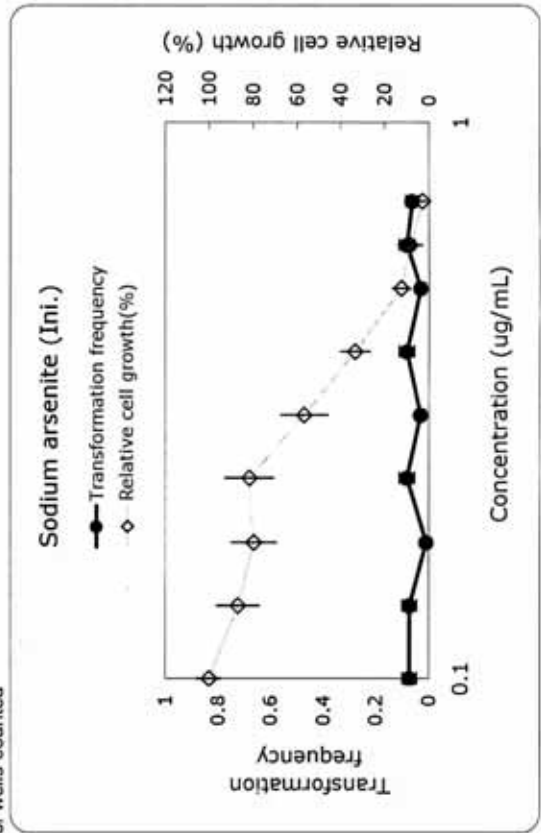
Chemical Code:	Sodium arsenite
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control		Positive Control		
			5% Water	0.1	0.135	0.175	0.228	0.296	0.385	0.5	0.6	0.72	0.808	0.912	0.300
1	0.109	0.873	0.794	0.777	0.809	0.687	0.439	0.259	0.261	0.149					
2	0.119	0.882	0.774	0.752	0.844	0.566	0.421	0.222	0.198	0.144					
3	0.112	0.820	0.775	0.753	0.685	0.590	0.303	0.196	0.132	0.137					
4	0.109	0.889	0.851	0.723	0.630	0.498	0.375	0.228	0.183	0.154					
5	0.112	0.861	0.662	0.576	0.605	0.503	0.361	0.186	0.156	0.125					
6	0.110	0.844	0.805	0.713	0.721	0.455	0.321	0.177	0.160	0.136					
7	0.123	0.765	0.732	0.727	0.718	0.488	0.314	0.214	0.147	0.112					
8	0.123	0.882	0.650	0.593	0.721	0.465	0.356	0.171	0.185	0.133					
Average	0.115	0.852	0.755	0.702	0.717	0.532	0.361	0.207	0.178	0.136	#DIV/0!				
SD	0.006	0.042	0.070	0.075	0.081	0.078	0.049	0.030	0.040	0.013	#DIV/0!				
Average - Blank	0.000	0.737	0.641	0.587	0.602	0.417	0.247	0.092	0.063	0.022	#DIV/0!				
Relative Cell Growth (%)		100.0	86.9	79.6	81.6	56.5	33.4	12.5	8.6	2.9	#DIV/0!				
SD of Relative Cell Growth		5.7	9.5	10.2	11.0	10.6	6.7	4.0	5.4	1.8	#DIV/0!				

Transformation Assay	Glenssa Staining	0.1	0.135	0.175	0.228	0.296	0.385	0.5	0.6	0.72	0	Negative Control	Positive Control
No. of wells excluded		0	0	0	0	0	0	0	0	1	0	0	0
No. of wells counted		96	96	96	96	96	96	96	96	95	96	96	96
No. of wells having foci		7	7	1	8	3	8	3	8	6	0	9	41
Transformation frequency*		0.072916667	0.072916667	0.010416667	0.083333333	0.03125	0.083333333	0.03125	0.083333333	0.063157895	0	0.09375	0.427083333
SE		0.02653610	0.02653610	0.01036227	0.02820847	0.01775905	0.02820847	0.01775905	0.02820847	0.02495659	0.00000000	0.02974911	0.05048547

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

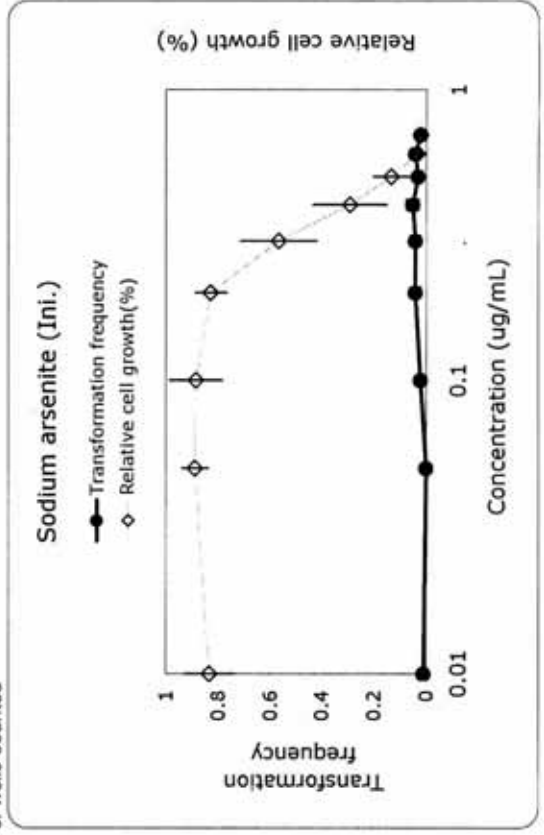
Chemical Code:	Sodium arsenite
Institution:	Lab 3
Test Number	

Cell Growth Assay		Concentrations (ug/ml)										Negative Control		Positive Control			
		Blank	5% Water	0.01	0.05	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0.1% DMSO	MCA (1ug/mL)			
Crystal Violet Staining	OD _{540nm}																
1		0.073	0.976	1.063	1.216	1.226	0.848	0.743	0.285	0.148	0.136	1.149	0.679				
2		0.074	1.093	1.135	1.192	1.069	0.545	0.677	0.217	0.105	0.084	1.206	0.719				
3		0.075	1.073	1.199	1.053	1.035	1.000	0.415	0.271	0.101	0.092	1.189	0.696				
4		0.077	1.355	1.087	1.087	1.022	0.641	0.358	0.188	0.113	0.096	1.193	0.669				
5		0.077	1.143	1.175	1.215	1.026	0.890	0.360	0.420	0.113	0.127	1.157	0.588				
6		0.076	1.031	1.158	1.375	1.143	0.989	0.353	0.179	0.125	0.086	1.136	0.538				
7		0.079	1.026	1.131	1.187	1.072	0.636	0.261	0.171	0.105	0.113	1.113	0.723				
8		0.079	1.096	1.266	0.978	1.151	0.632	0.323	0.197	0.129	0.083	1.194	0.522				
Average		0.076	1.099	1.168	1.163	1.093	0.773	0.436	0.241	0.117	0.102	1.167	0.642				
SD		0.002	0.115	0.062	0.122	0.073	0.180	0.175	0.084	0.016	0.021	0.033	0.081				
Average - Blank		0.000	1.023	1.092	1.087	1.017	0.696	0.360	0.165	0.041	0.026	1.091	0.566				
Relative Cell Growth (%)			100.0	106.7	106.2	99.4	68.1	35.2	16.1	4.0	2.5	100.0	51.8				
SD of Relative Cell Growth			11.3	6.0	11.9	7.2	17.5	17.1	8.2	1.5	2.0	3.2	7.9				

Transformation Assay		Concentrations (ug/ml)										Negative Control		Positive Control		
		0.01	0.05	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0	96	96	96	0	0
Giemsa Staining		0.01	0.05	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0	0	0	0	0	0
No. of wells excluded		0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci		1	0	2	4	4	5	3	4	2	0	3	0	3	0	52
Transformation frequency*		0.010416667	0	0.020833333	0.041666667	0.041666667	0.052083333	0.03125	0.041666667	0.020833333	0	0.03125	0.541666667	0.03125	0.541666667	0.05085354
SE		0.01036227	0.0000000	0.01457713	0.02039469	0.02039469	0.02267769	0.01775805	0.02039469	0.01457713	0.00000000	0.01775805	0.05085354	0.01775805	0.05085354	

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

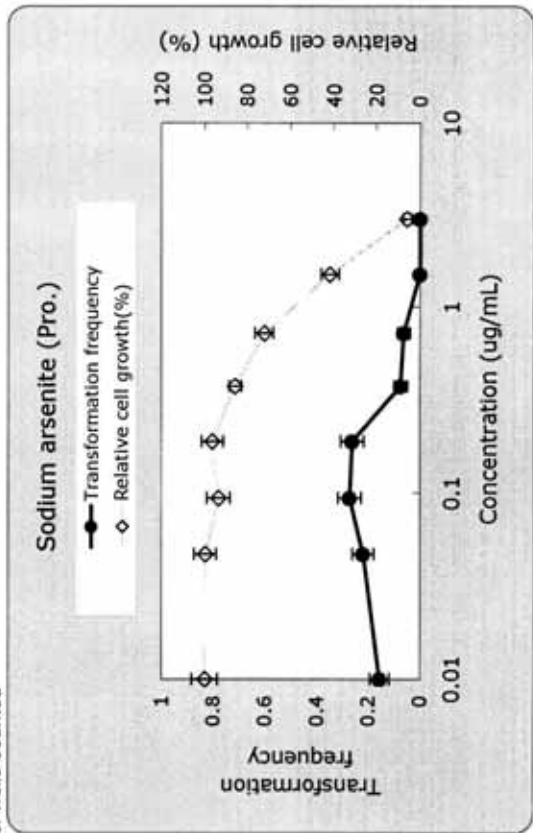
Chemical Code:	Sodium arsenite
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control	Positive Control
			5% Water	0.01	0.0469	0.0938	0.188	0.375	0.725	1.5		
1	0.107	0.707	0.754	0.697	0.730	0.617	0.564	0.399	0.146	0.699	1.039	
2	0.114	0.736	0.702	0.638	0.630	0.618	0.542	0.327	0.148	0.804	1.183	
3	0.105	0.666	0.698	0.676	0.658	0.629	0.526	0.343	0.130	0.750	1.026	
4	0.110	0.738	0.692	0.626	0.685	0.614	0.520	0.336	0.135	0.688	1.272	
5	0.112	0.665	0.646	0.688	0.701	0.628	0.525	0.364	0.143	0.719	1.101	
6	0.119	0.713	0.706	0.636	0.671	0.595	0.498	0.364	0.166	0.745	1.127	
7	0.116	0.652	0.681	0.638	0.682	0.592	0.550	0.380	0.148	0.697	1.203	
8	0.112	0.736	0.720	0.706	0.686	0.641	0.577	0.343	0.151	0.702	1.163	
Average	0.112	0.702	0.700	0.663	0.680	0.617	0.538	0.357	0.146	#DIV/0!	1.139	
SD	0.005	0.036	0.031	0.032	0.029	0.017	0.026	0.024	0.011	#DIV/0!	0.083	
Average - Blank	0.000	0.590	0.588	0.551	0.569	0.505	0.426	0.245	0.034	#DIV/0!	1.027	
Relative Cell Growth (%)		100.0	99.7	93.5	96.4	85.6	72.2	41.6	5.8	#DIV/0!	167.4	
SD of Relative Cell Growth		6.1	5.2	5.4	5.0	2.8	4.3	4.1	1.8	#DIV/0!	14.1	

Transformation Assay	0.01	0.0469	0.0938	0.188	0.375	0.725	1.5	3	0	0	0	Negative Control	Positive Control
Giemsa Staining	0.01	0.0469	0.0938	0.188	0.375	0.725	1.5	3	0	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	43	96	96	96	96	96	96
No. of wells counted	96	96	96	96	96	96	53	0	0	0	0	14	72
No. of wells having foci	15	21	26	25	7	6	TOX	TOX	TOX	TOX	TOX	14	72
Transformation frequency*	0.15625	0.21875	0.270833333	0.260416667	0.072916667	0.0625	#VALUE!	#VALUE!	0	0	0	0.145833333	0.75
SE	0.03705794	0.04219232	0.04535538	0.04479116	0.02653610	0.02470529	#VALUE!	#VALUE!	0.00000000	0.00000000	0.00000000	0.03602169	0.04419417

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Sodium arsenite
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
	Blank	5% Water	0.01	0.025	0.05	0.1	0.25	0.5	1	1.5	2	0.1% DMSO	TPA (50ng/mL)	
Crystal Violet Staining														
OD _{560nm}														
1	0.078	1.147	1.142	0.942	1.220	1.099	1.032	0.774	0.611	0.142	0.954	1.723		
2	0.078	1.034	1.194	1.166	1.107	1.132	0.971	0.775	0.610	0.151	0.979	1.599		
3	0.081	1.166	1.228	0.998	1.157	1.140	1.027	0.780	0.685	0.279	1.024	1.595		
4	0.083	1.275	1.125	1.165	1.209	1.176	0.937	0.762	0.613	0.288	0.994	1.538		
5	0.082	1.228	1.224	1.111	1.213	1.220	0.915	0.770	0.659	0.233	1.061	1.703		
6	0.081	1.231	1.175	1.174	1.119	1.206	0.931	0.778	0.664	0.215	1.039	1.642		
7	0.084	1.181	1.236	1.114	1.171	1.245	1.005	0.778	0.678	0.277	1.050	1.672		
8	0.081	1.117	1.182	1.229	1.126	1.122	0.913	0.726	0.624	0.204	0.968	1.612		
Average	0.081	1.172	1.188	1.112	1.165	1.168	0.966	0.768	0.643	0.224	1.009	1.636		
SD	0.002	0.076	0.041	0.096	0.045	0.052	0.049	0.018	0.032	0.057	0.040	0.062		
Average - Blank	0.000	1.091	1.107	1.031	1.084	1.087	0.885	0.687	0.562	0.143	0.928	1.555		
Relative Cell Growth (%)														
SD of Relative Cell Growth														
		6.9	3.7	8.8	4.2	4.8	4.5	1.6	2.9	5.2	3.7	5.7		
		100.0	101.5	94.5	99.3	99.6	81.1	62.9	51.5	13.1	100.0	167.6		
		0.01	0.025	0.05	0.1	0.25	0.5	1	1.5	2	0	0		
		0	0	0	0	0	0	96	96	96	96	96		
		12	10	11	8	5	0	Tox	Tox	Tox	7	68		
		0.125	0.104166667	0.114583333	0.083333333	0.052083333	0	#VALUE!	#VALUE!	#VALUE!	0.072916667	0.708333333		
		0.03375386	0.03117758	0.03250965	0.02820847	0.02267769	0.00000000	#VALUE!	#VALUE!	#VALUE!	0.02653610	0.04639024		

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
Giemsa Staining	0.01	0.025	0.05	0.1	0.25	0.5	1	1.5	2	2	0	0	0	
No. of wells excluded	0	0	0	0	0	0	0	96	96	96	96	96	96	
No. of wells counted	96	96	96	96	96	96	96	Tox	Tox	Tox	7	68	68	
No. of wells having foci	12	10	11	8	5	0	0	Tox	Tox	Tox	7	68	68	
Transformation frequency*	0.125	0.104166667	0.114583333	0.083333333	0.052083333	0	0	#VALUE!	#VALUE!	#VALUE!	0.072916667	0.708333333		
SE	0.03375386	0.03117758	0.03250965	0.02820847	0.02267769	0.00000000	0	#VALUE!	#VALUE!	#VALUE!	0.02653610	0.04639024		

* No. of wells having foci/ No. of wells counted

Sodium arsenite (Pro.)

Transformation frequency
 Relative cell growth(%)

Acceptance Criteria (1)

Negative Control < 20 and 40 < Positive Control

Pass

Acceptance Criteria (2)

Concurrent Cell Growth Assay

Pass

The Final Data

Remarks	

Data Sheet for Cell Transformation Assay on Initiation Assay

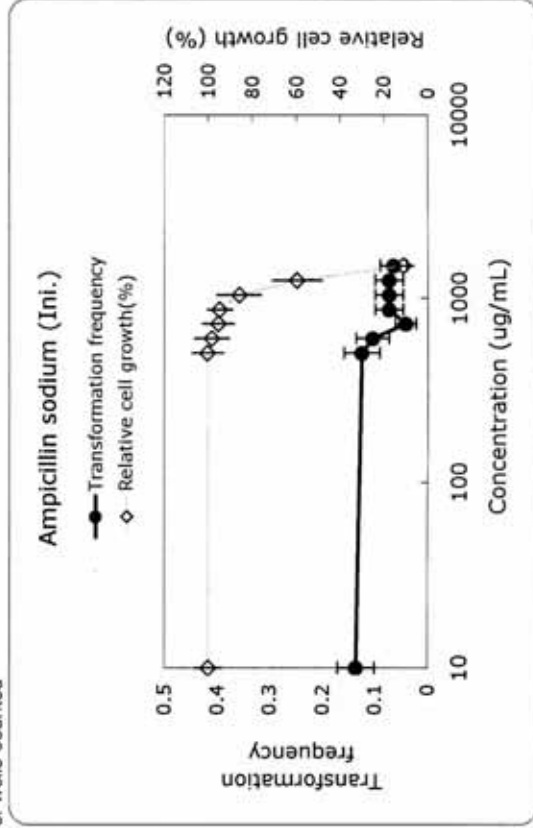
Chemical Code:	Ampicillin sodium
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control
	Blank	5% Water	10	502	603	724	868	1040	1250	1500		
Crystal Violet Staining	Blank	10	502	603	724	868	1040	1250	1500			
OD _{540nm}												
1	0.137	0.692	0.736	0.701	0.690	0.696	0.641	0.554	0.234		0.677	0.392
2	0.133	0.694	0.708	0.624	0.632	0.622	0.618	0.506	0.167		0.660	0.374
3	0.129	0.697	0.609	0.663	0.646	0.665	0.522	0.367	0.178		0.683	0.359
4	0.132	0.663	0.699	0.656	0.660	0.677	0.661	0.466	0.231		0.674	0.379
5	0.137	0.647	0.678	0.642	0.654	0.614	0.552	0.451	0.192		0.716	0.333
6	0.140	0.700	0.697	0.698	0.597	0.644	0.573	0.423	0.223		0.686	0.345
7	0.151	0.659	0.717	0.685	0.710	0.692	0.665	0.432	0.178		0.717	0.325
8	0.159	0.756	0.669	0.761	0.714	0.660	0.655	0.529	0.203		0.711	0.341
Average	0.140	0.689	0.689	0.679	0.663	0.659	0.611	0.466	0.201	#DIV/0!	0.691	0.356
SD	0.010	0.034	0.039	0.043	0.040	0.030	0.055	0.061	0.026	#DIV/0!	0.021	0.024
Average - Blank	0.000	0.549	0.549	0.539	0.523	0.519	0.471	0.326	0.061	#DIV/0!	0.551	0.216
Relative Cell Growth (%)		100.0	100.1	98.2	95.3	94.6	85.9	59.5	11.1	#DIV/0!	100.0	39.3
SD of Relative Cell Growth		6.2	7.0	7.8	7.3	5.5	10.0	11.2	4.8	#DIV/0!	3.9	4.4

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
	10	502	603	724	868	1040	1250	1500	0	0		
Glensa Staining	10	502	603	724	868	1040	1250	1500	0	0		
No. of wells excluded	0	0	0	0	0	0	0	2	96	96		
No. of wells counted	96	96	96	96	96	96	96	96	96	96		
No. of wells having foci	13	12	10	4	7	7	7	6	6	6		
Transformation frequency*	0.135416667	0.125	0.104166667	0.041666667	0.072916667	0.072916667	0.072916667	0.063829787	0	0		
SE	0.03492238	0.03375386	0.03117758	0.02039469	0.02653610	0.02653610	0.02653610	0.02521304	0.00000000	0.00000000		

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

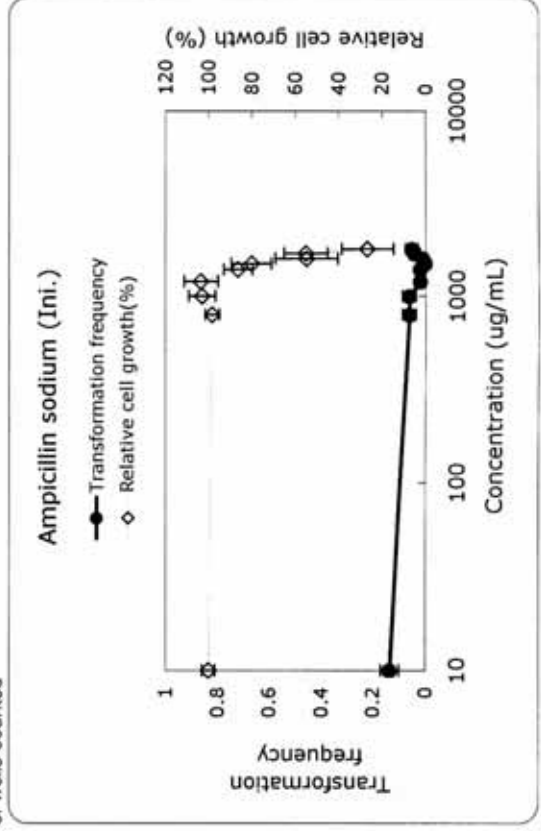
Chemical Code:	Ampicillin sodium
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control		Positive Control	
			5% Water	800	1000	1200	1400	1500	1600	1700	1800	0.1% DMSO	MCA (1ug/mL)	
1	0.115	1.126	1.163	1.218	1.389	1.112	1.096	0.510	0.799	0.258	1.175	0.578		
2	0.119	1.148	1.103	1.236	1.141	0.965	0.912	0.814	0.622	0.307	1.180	0.687		
3	0.123	1.215	1.146	1.276	1.150	0.975	0.970	0.548	0.716	0.425	1.253	0.659		
4	0.123	1.180	1.130	1.206	1.206	1.012	0.795	0.565	0.797	0.324	1.067	0.626		
5	0.111	1.178	1.156	1.157	1.220	0.941	1.030	0.824	0.656	0.346	1.146	0.742		
6	0.116	1.209	1.207	1.139	1.148	1.106	1.050	0.843	0.722	0.427	1.244	0.673		
7	0.108	1.195	1.167	1.202	1.275	1.042	0.990	0.866	0.784	0.426	1.164	0.676		
8	0.106	1.170	1.199	1.310	1.182	1.110	0.881	0.600	0.492	0.672	1.100	0.682		
Average	0.115	1.178	1.159	1.209	1.214	1.033	0.966	0.696	0.699	0.398	#DIV/0!	0.665		
SD	0.006	0.030	0.034	0.065	0.084	0.070	0.099	0.153	0.106	0.127	#DIV/0!	0.048		
Average - Blank	0.000	1.063	1.044	1.093	1.099	0.918	0.850	0.581	0.583	0.283	#DIV/0!	0.550		
Relative Cell Growth (%)		100.0	98.2	102.9	103.4	86.4	80.0	54.9	54.9	26.6	#DIV/0!	100.0		
SD of Relative Cell Growth		2.8	3.2	6.1	7.9	6.6	9.3	14.4	10.0	12.0	#DIV/0!	6.0		

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
Glenssa Staining	10	800	1000	1200	1400	1500	1600	1700	1800	1800	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	13	6	6	2	2	0	1	4	5	5	13	54	54	54
Transformation frequency*	0.135416667	0.0625	0.0625	0.020833333	0.020833333	0.000000000	0.010416667	0.041666667	0.052083333	0.052083333	0	0.135416667	0.5625	0.5625
SE	0.03492238	0.02470529	0.02470529	0.01457713	0.01457713	0.000000000	0.01036227	0.02039469	0.02267769	0.000000000	0.03492238	0.05063079	0.05063079	0.05063079

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

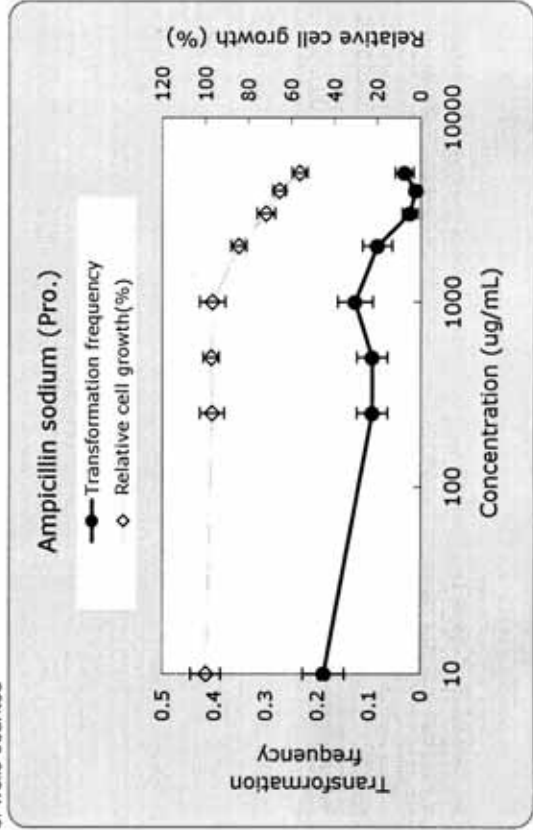
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Ampicillin sodium
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control		Positive Control	
			5% Water	250	500	1000	2000	3000	4000	5000	0.1% DMSO	TPA (50ng/mL)		
1	0.153	0.692	0.727	0.700	0.671	0.638	0.582	0.555	0.498	0.760	0.838			
2	0.152	0.648	0.712	0.729	0.744	0.634	0.552	0.516	0.488	0.680	0.879			
3	0.154	0.712	0.676	0.703	0.725	0.627	0.558	0.522	0.466	0.707	0.849			
4	0.170	0.750	0.690	0.695	0.672	0.635	0.563	0.518	0.448	0.677	0.935			
5	0.151	0.743	0.668	0.675	0.674	0.680	0.539	0.525	0.480	0.674	0.894			
6	0.163	0.772	0.719	0.737	0.699	0.617	0.574	0.533	0.449	0.684	0.964			
7	0.154	0.750	0.703	0.729	0.706	0.616	0.544	0.513	0.486	0.727	0.963			
8	0.169	0.731	0.768	0.708	0.756	0.644	0.611	0.558	0.499	0.684	0.927			
Average	0.158	0.725	0.708	0.710	0.706	0.636	0.565	0.530	0.477	0.699	0.906			
SD	0.008	0.040	0.032	0.021	0.033	0.020	0.023	0.017	0.020	0.030	0.049			
Average - Blank	0.000	0.567	0.550	0.551	0.548	0.478	0.407	0.372	0.319	0.541	0.748			
Relative Cell Growth (%)	100.0	97.0	97.3	96.7	96.7	84.4	71.9	65.6	56.2	100.0	138.3			
SD of Relative Cell Growth	7.0	5.6	3.7	5.9	3.6	4.1	3.1	3.6	3.6	5.4	8.6			

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
Glenssa Staining	10	250	500	1000	2000	3000	4000	5000	0	0	0	0	0	0
No. of wells excluded	0	0	0	1	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	95	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	18	9	9	12	8	2	1	3	0	0	0	0	0	0
Transformation frequency*	0.1875	0.09375	0.09375	0.126315789	0.083333333	0.020833333	0.010416667	0.03125	0	0	0	0	0	0
SE	0.03983609	0.02974911	0.02974911	0.03408351	0.02820847	0.01457713	0.01036227	0.01775805	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

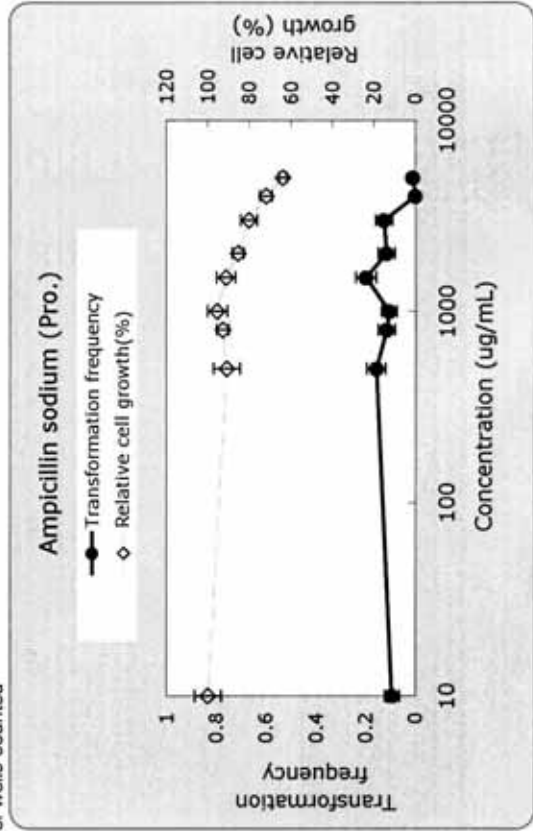
Chemical Code:	Ampicillin sodium
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)								Negative Control		Positive Control	
		Blank	5% Water	10	500	800	1000	1500	2000	3000	4000	5000	0.1% DMSO
1	0.121		1.307	1.194	1.264	1.261	1.215	1.180	1.032	0.990	0.936	1.013	1.623
2	0.130		1.318	1.315	1.289	1.346	1.237	1.222	1.186	1.017	0.921	1.277	1.770
3	0.128		1.478	1.265	1.338	1.354	1.298	1.182	1.148	1.079	0.971	1.221	1.712
4	0.127		1.394	1.321	1.363	1.377	1.398	1.266	1.203	1.034	0.895	1.171	1.703
5	0.118		1.345	1.249	1.285	1.436	1.295	1.238	1.137	0.992	0.935	1.258	1.746
6	0.119		1.403	1.449	1.344	1.307	1.235	1.167	1.136	1.055	0.929	1.282	1.565
7	0.117		1.368	1.241	1.269	1.346	1.263	1.233	1.148	1.035	0.956	1.310	1.776
8	0.112		1.546	1.199	1.276	1.253	1.313	1.188	1.149	1.091	0.928	1.197	1.695
Average	0.122		1.395	1.279	1.304	1.335	1.282	1.210	1.143	1.037	0.934	#DIV/0!	1.699
SD	0.006		0.082	0.083	0.039	0.060	0.058	0.035	0.051	0.037	0.023	#DIV/0!	0.073
Average - Blank	0.000		1.273	1.158	1.182	1.214	1.160	1.088	1.021	0.915	0.812	#DIV/0!	1.577
Relative Cell Growth (%)			100.0	90.9	92.8	95.3	91.1	85.4	80.2	71.9	63.8	#DIV/0!	144.1
SD of Relative Cell Growth			6.4	6.5	3.0	4.7	4.6	2.8	4.0	2.9	1.8	#DIV/0!	5.7

Transformation Assay	Concentrations (ug/ml)												Negative Control		Positive Control	
Glmsa Staining	10	500	800	1000	1500	2000	3000	4000	5000	5000	5000	0	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	9	15	11	10	19	11	12	0	1	0	1	0	0	0	0	0
Transformation frequency*	0.09375	0.15625	0.114583333	0.104166667	0.197916667	0.114583333	0.125	0	0.010416667	0	0.010416667	0	0	0.145833333	0.8125	0.03983609
SE	0.02974911	0.03705794	0.03250865	0.03117758	0.04066449	0.03250865	0.03375396	0.00000000	0.01036227	0.00000000	0.01036227	0.00000000	0	0.145833333	0.03602169	0.03983609

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

This Data is ...

Data Sheet for Cell Transformation Assay on Initiation Assay

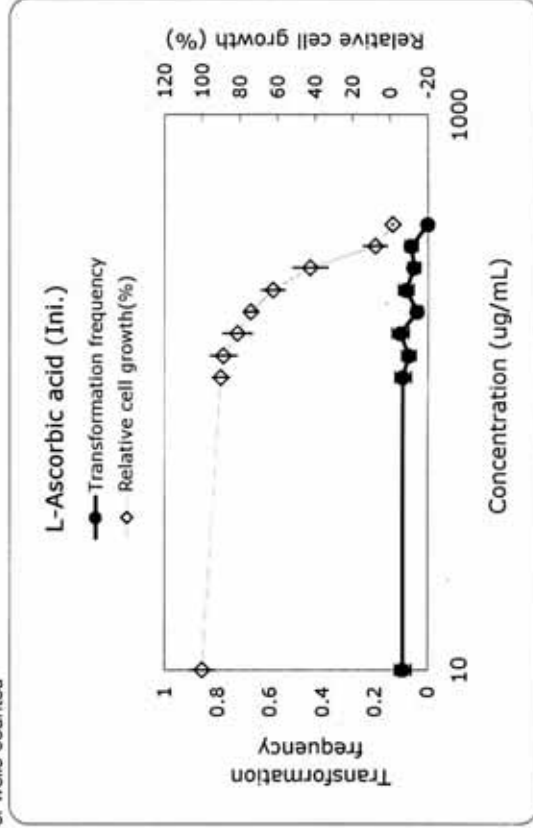
Chemical Code:	L-Ascorbic acid
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	5% Water	112	134	161	193	231	278	333	400			0.1% DMSO
Crystal Violet Staining													
OD _{540nm}													
1	0.093	0.855	0.817	0.759	0.735	0.694	0.677	0.433	0.137	0.113	0.851	0.315	
2	0.108	0.872	0.772	0.707	0.707	0.634	0.614	0.377	0.128	0.103	0.818	0.446	
3	0.112	0.814	0.812	0.813	0.666	0.666	0.625	0.395	0.186	0.111	0.786	0.450	
4	0.127	0.846	0.823	0.779	0.838	0.710	0.528	0.452	0.260	0.113	0.933	0.414	
5	0.126	0.908	0.795	0.821	0.812	0.689	0.576	0.429	0.200	0.110	0.823	0.388	
6	0.135	0.953	0.877	0.855	0.761	0.695	0.604	0.564	0.170	0.118	0.786	0.466	
7	0.137	0.916	0.827	0.875	0.681	0.706	0.550	0.387	0.141	0.110	0.798	0.398	
8	0.131	0.942	0.777	0.801	0.756	0.734	0.616	0.544	0.233	0.117	0.888	0.385	
Average	0.121	0.888	0.813	0.801	0.745	0.691	0.599	0.448	0.182	0.112	0.835	0.408	
SD	0.015	0.049	0.033	0.053	0.060	0.030	0.047	0.071	0.047	0.005	0.052	0.048	
Average - Blank	0.000	0.767	0.691	0.680	0.623	0.570	0.478	0.327	0.061	-0.009	0.714	0.287	
Relative Cell Growth (%)		100.0	90.1	88.7	81.3	74.3	62.3	42.6	7.9	-1.2	100.0	40.1	
SD of Relative Cell Growth		6.4	4.3	7.0	7.9	3.9	6.1	9.2	6.2	0.6	6.8	6.3	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glmsa Staining	10	112	134	161	193	231	278	333	400	0		
No. of wells excluded	0	0	0	0	0	0	0	0	20	20	0	0
No. of wells counted	96	96	96	96	96	96	96	96	76	96	96	96
No. of wells having foci	9	9	7	10	4	8	5	6	TOX	6	10	41
Transformation frequency*	0.09375	0.09375	0.072916667	0.104166667	0.041666667	0.083333333	0.052083333	0.0625	#VALUE!	0	0.104166667	0.427083333
SE	0.02974911	0.02974911	0.02653610	0.03117758	0.02039469	0.02820847	0.02267769	0.02470529	#VALUE!	0.00000000	0.03117758	0.05048547

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

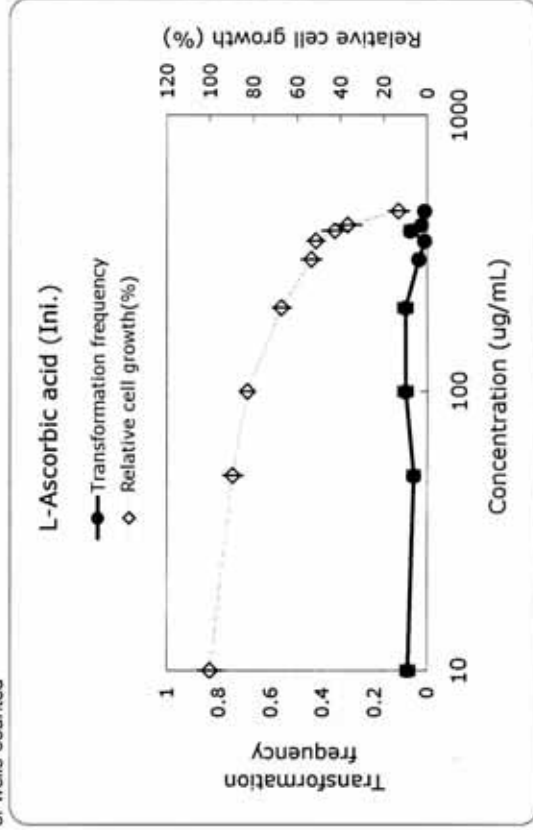
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	L-Ascorbic acid
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Blank	Concentrations (ug/ml)								Negative Control 0.1% DMSO	Positive Control MCA (1ug/ml)
			5% Water	10	50	100	200	300	350	400		
1	0.128	1.448	1.292	1.250	1.047	0.904	0.867	0.807	0.605	0.366	1.424	0.802
2	0.133	1.484	1.389	1.197	1.092	0.816	0.752	0.570	0.640	0.332	1.381	0.947
3	0.131	1.297	1.267	1.245	1.033	0.855	0.784	0.669	0.546	0.333	1.481	0.868
4	0.130	1.417	1.290	1.191	0.932	0.725	0.840	0.681	0.569	0.279	1.407	0.794
5	0.130	1.432	1.245	1.185	1.012	0.780	0.751	0.627	0.716	0.313	1.363	0.768
6	0.127	1.471	1.223	1.222	0.938	0.812	0.739	0.698	0.579	0.226	1.379	0.942
7	0.132	1.544	1.375	1.232	1.004	0.813	0.872	0.639	0.473	0.194	1.353	0.774
8	0.128	1.369	1.272	1.143	0.974	0.907	0.787	0.768	0.719	0.361	1.297	0.817
Average	0.130	1.433	1.294	1.208	1.004	0.827	0.799	0.682	0.606	0.301	#DIV/0!	0.839
SD	0.002	0.075	0.059	0.036	0.055	0.061	0.054	0.076	0.084	0.063	#DIV/0!	0.072
Average - Blank	0.000	1.303	1.164	1.078	0.874	0.697	0.669	0.553	0.476	0.171	#DIV/0!	0.709
Relative Cell Growth (%)		100.0	89.4	82.8	67.1	53.5	51.4	42.4	36.5	13.1	#DIV/0!	56.5
SD of Relative Cell Growth		5.8	4.5	2.8	4.2	4.7	4.1	5.9	6.4	4.8	#DIV/0!	5.5

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glensa Staining	10	50	100	200	300	350	400	450	0	450	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	7	5	8	8	3	1	2	1	7	1	7	80
Transformation frequency*	0.072916667	0.052083333	0.083333333	0.083333333	0.03125	0.010416667	0.020833333	0.010416667	0	0.020833333	0.010416667	0.625
SE	0.02653610	0.02267769	0.02820847	0.02820847	0.01775905	0.01036227	0.02470529	0.01457713	0.01036227	0.00000000	0.02653610	0.04941059

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

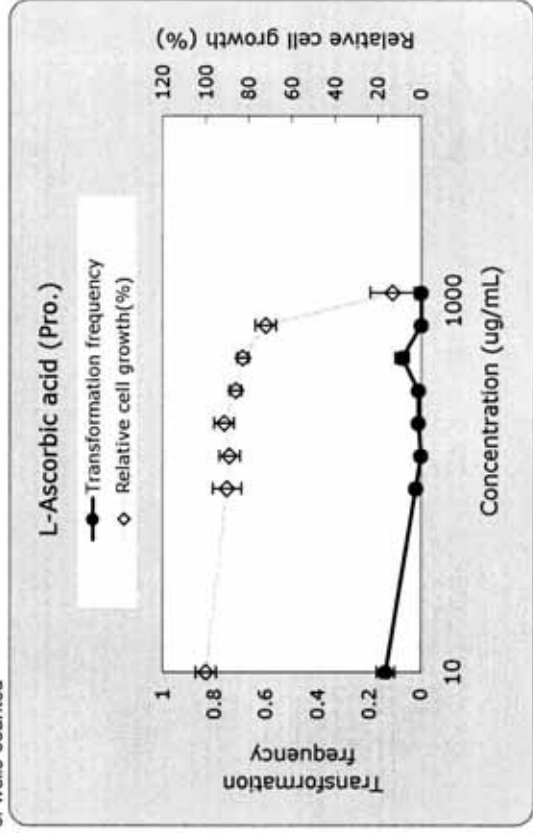
Chemical Code:	L-Ascorbic acid
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)								Negative Control	Positive Control		
	Blank	5% Water	96.6	145	217	326	489	733			1100	
Crystal Violet Staining												
OD _{540nm}												
1	0.091	0.718	0.647	0.668	0.662	0.696	0.644	0.573	0.232	0.741	0.978	
2	0.099	0.776	0.748	0.703	0.747	0.685	0.669	0.610	0.292	0.716	0.979	
3	0.111	0.794	0.725	0.708	0.713	0.703	0.664	0.610	0.118	0.718	0.960	
4	0.116	0.796	0.668	0.733	0.717	0.691	0.637	0.561	0.187	0.737	0.963	
5	0.112	0.779	0.716	0.720	0.726	0.702	0.686	0.589	0.164	0.753	0.975	
6	0.128	0.827	0.788	0.760	0.742	0.674	0.690	0.582	0.156	0.795	1.037	
7	0.120	0.763	0.719	0.659	0.712	0.704	0.661	0.572	0.305	0.757	0.977	
8	0.118	0.791	0.699	0.699	0.767	0.647	0.669	0.658	0.156	0.745	0.974	
Average	0.112	0.781	0.714	0.706	0.723	0.688	0.665	0.594	0.201	0.745	0.980	
SD	0.012	0.031	0.044	0.033	0.031	0.019	0.018	0.031	0.068	0.025	0.024	
Average - Blank	0.000	0.669	0.602	0.594	0.611	0.576	0.553	0.483	0.089	0.633	0.869	
Relative Cell Growth (%)		100.0	90.0	88.9	91.4	86.1	82.7	72.2	13.4	100.0	137.1	
SD of Relative Cell Growth		4.7	6.6	4.9	4.7	2.9	2.7	4.7	10.2	3.7	3.6	

Transformation Assay	Concentrations (ug/ml)								Negative Control	Positive Control	
Glenssa Staining	10	96.6	145	217	326	489	733	1100			
No. of wells excluded	0	0	0	0	0	1	1	94	96	0	0
No. of wells counted	96	96	96	96	96	95	95	2	0	96	96
No. of wells having foci	13	2	0	1	1	7	7	TOX	TOX	15	56
Transformation frequency*	0.135416667	0.020833333	0	0.010416667	0.010416667	0.073684211	0.073684211	#VALUE!	#VALUE!	0	0
SE	0.03492238	0.01457173	0.00000000	0.01036227	0.01036227	0.02680433	0.02680433	#VALUE!	#VALUE!	0.00000000	0.00000000

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Progressing Data

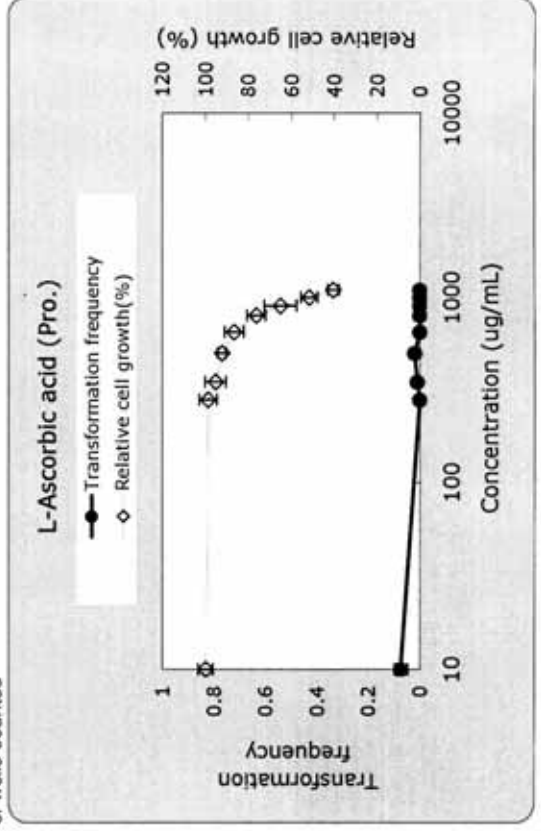
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	L-Ascorbic acid
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control		Positive Control	
			5% Water	280	350	500	650	800	900	1000	1100	0.1% DMSO	TPA (50ng/mL)	
1	0.127	1.336	1.380	1.324	1.273	1.168	1.118	1.004	0.778	0.628	1.305	1.728		
2	0.133	1.405	1.377	1.357	1.272	1.231	1.124	1.004	0.705	0.634	1.371	1.740		
3	0.134	1.410	1.348	1.360	1.312	1.158	1.112	1.026	0.761	0.571	1.391	1.656		
4	0.128	1.425	1.331	1.195	1.311	1.177	1.060	0.778	0.768	0.594	1.428	1.806		
5	0.126	1.390	1.391	1.384	1.254	1.158	0.972	0.871	0.731	0.633	1.442	1.765		
6	0.127	1.382	1.447	1.300	1.288	1.251	1.049	0.857	0.868	0.590	1.357	1.718		
7	0.121	1.364	1.300	1.330	1.312	1.058	1.058	1.035	0.754	0.682	1.284	1.690		
8	0.118	1.290	1.294	1.270	1.203	1.133	1.078	0.927	0.794	0.615	1.341	1.596		
Average	0.127	1.375	1.359	1.315	1.279	1.207	1.078	0.938	0.770	0.631	#DIV/0!	1.712		
SD	0.005	0.044	0.051	0.060	0.023	0.054	0.054	0.095	0.048	0.033	#DIV/0!	0.065		
Average - Blank	0.000	1.249	1.232	1.188	1.152	1.081	0.952	0.811	0.643	0.504	#DIV/0!	1.586		
Relative Cell Growth (%)		100.0	98.7	95.2	92.3	86.5	76.2	65.0	51.5	40.4	#DIV/0!	128.1		
SD of Relative Cell Growth		3.6	4.1	4.8	1.8	4.4	4.4	7.6	3.9	2.6	#DIV/0!	5.2		

Transformation Assay	Concentrations (ug/ml)								Negative Control		Positive Control	
Giemsa Staining	10	280	350	500	650	800	900	1000	1100	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	7	0	1	2	Tox.	Tox.	Tox.	Tox.	Tox.	Tox.	21	85
Transformation frequency*	0.072916667	0	0.010416667	0.020833333	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.21875	0.885416667
SE	0.02653610	0.00000000	0.01036227	0.01457713	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	#VALUE!	0.04219232	0.03250865

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Failure
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Failure

The Progressing Data

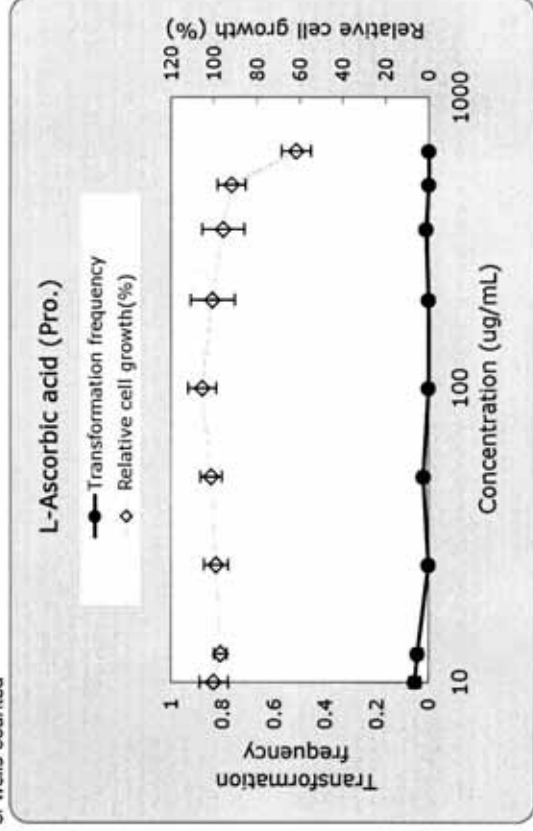
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	L-Ascorbic acid
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)								Negative Control		Positive Control	
		Blank	5% Water	10	12.5	25	50	100	200	350	500	650	0.1% DMSO
1	0.076	0.502	0.470	0.519	0.474	0.503	0.431	0.399	0.430	0.350	0.594	0.501	0.594
2	0.079	0.485	0.494	0.494	0.526	0.511	0.519	0.497	0.483	0.388	0.669	0.507	0.669
3	0.089	0.497	0.473	0.496	0.498	0.467	0.437	0.465	0.463	0.334	0.623	0.474	0.623
4	0.079	0.509	0.500	0.514	0.485	0.547	0.514	0.488	0.476	0.319	0.707	0.580	0.707
5	0.077	0.485	0.497	0.499	0.537	0.514	0.550	0.503	0.506	0.334	0.570	0.523	0.570
6	0.078	0.470	0.487	0.443	0.512	0.533	0.514	0.492	0.449	0.298	0.619	0.584	0.619
7	0.084	0.557	0.468	0.483	0.489	0.549	0.525	0.536	0.472	0.362	0.660	0.576	0.660
8	0.083	0.474	0.484	0.497	0.503	0.537	0.508	0.455	0.425	0.321	0.532	0.513	0.532
Average	0.081	0.497	0.484	0.493	0.503	0.520	0.500	0.479	0.463	0.338	0.622	0.532	0.622
SD	0.004	0.028	0.013	0.023	0.021	0.027	0.043	0.041	0.027	0.028	0.057	0.042	0.057
Average - Blank	0.000	0.417	0.404	0.413	0.422	0.440	0.419	0.399	0.382	0.258	0.541	0.452	0.541
Relative Cell Growth (%)		100.0	96.8	99.0	101.3	105.5	100.6	95.7	91.8	61.8	119.8	100.0	119.8
SD of Relative Cell Growth		6.6	3.0	5.6	5.1	6.6	10.2	9.8	6.6	6.7	13.6	10.1	13.6

Transformation Assay	Glenssa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	Transformation frequency*	SE	Concentrations (ug/ml)								Negative Control		Positive Control	
							10	12.5	25	50	100	200	350	500	650	0	0	0
		0	96	5	0.0520833333	0.02267769	0.02039469	0.00000000	0.01457713	0.00000000	0.00000000	0.010416667	0.0833333333	0.0833333333	0	0	0	0
		4	96	2	0.0208333333	0.01457713	0.00000000	0.01457713	0.00000000	0.00000000	0.01036227	0.0833333333	0.0833333333	0	0	96	96	45
		0	96	0	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0	0	96	45
		0	96	0	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0.00000000	0	0	96	45

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

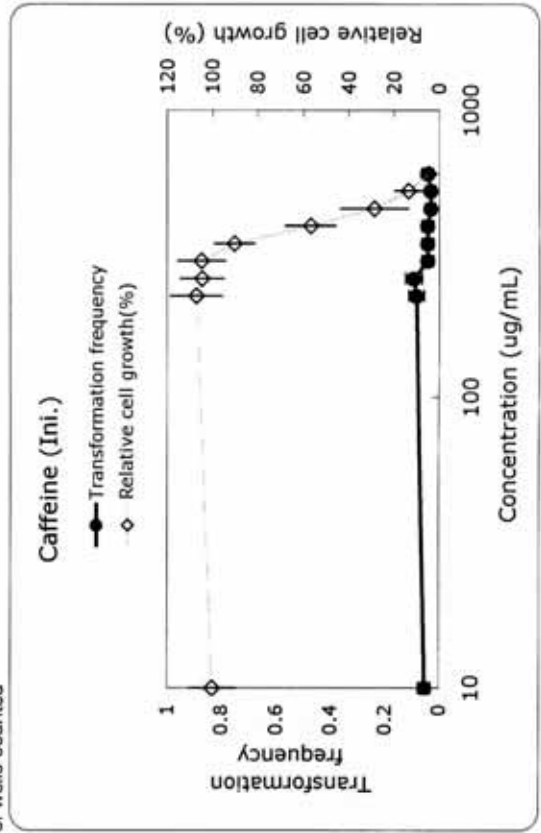
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Caffeine
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
		Blank	5% Water	226	259	298	343	395	454	522	600		
1	0.065	0.342	0.331	0.363	0.295	0.214	0.102	0.082	0.069	0.334	0.232		
2	0.062	0.386	0.368	0.362	0.344	0.241	0.127	0.097	0.077	0.325	0.209		
3	0.060	0.401	0.358	0.423	0.316	0.218	0.157	0.115	0.075	0.336	0.219		
4	0.059	0.377	0.370	0.414	0.351	0.269	0.134	0.100	0.088	0.315	0.216		
5	0.060	0.393	0.424	0.408	0.314	0.263	0.133	0.136	0.077	0.355	0.202		
6	0.060	0.347	0.368	0.350	0.321	0.229	0.191	0.117	0.072	0.334	0.201		
7	0.060	0.356	0.382	0.345	0.365	0.246	0.235	0.086	0.073	0.311	0.198		
8	0.065	0.309	0.377	0.362	0.369	0.170	0.105	0.083	0.072	0.299	0.197		
Average	0.061	0.364	0.378	0.378	0.334	0.233	0.148	0.102	0.075	0.326	0.209		
SD	0.002	0.031	0.029	0.031	0.027	0.034	0.045	0.019	0.006	0.018	0.012		
Average - Blank	0.000	0.303	0.316	0.317	0.273	0.172	0.087	0.041	0.014	0.265	0.148		
Relative Cell Growth (%)	100.0	107.2	104.5	104.8	90.2	56.8	28.6	13.4	4.6	100.0	55.9		
SD of Relative Cell Growth	10.2	11.4	9.7	10.3	8.8	11.1	15.0	6.4	1.9	5.8	4.1		

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glenssa Staining	10	226	259	298	343	395	454	522	600	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	5	8	9	4	4	4	3	3	4	4	4	55
Transformation frequency*	0.052083333	0.083333333	0.09375	0.041666667	0.041666667	0.041666667	0.03125	0.03125	0.041666667	0	0.041666667	0.572916667
SE	0.02267769	0.02820847	0.02974911	0.02039469	0.02039469	0.02039469	0.01775805	0.01775805	0.02039469	0.00000000	0.02039469	0.05048547

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

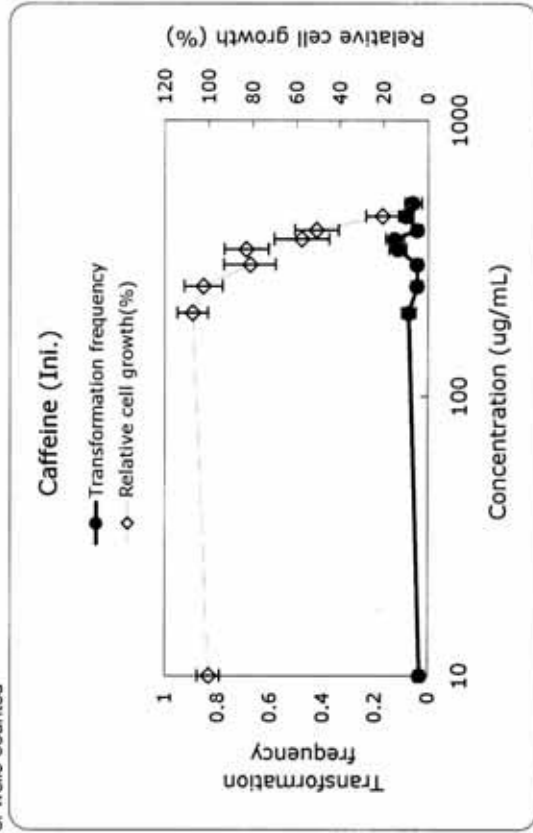
Chemical Code:	Caffeine
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control		
	Blank	5% Water	10	200	250	300	340	370	400	450			500	0.1% DMSO
Crystal Violet Staining	Blank	10												
OD _{540nm}														
1	0.085	1.226	1.252	1.188	0.916	1.061	0.629	0.718	0.208	0.100	1.107	0.572		
2	0.091	1.177	1.214	1.277	1.186	1.023	0.578	0.595	0.231	0.186	1.221	0.600		
3	0.097	1.193	1.386	1.108	1.100	1.050	0.803	0.748	0.290	0.143	1.284	0.613		
4	0.098	1.252	1.264	1.332	0.815	1.055	0.774	0.760	0.402	0.176	1.145	0.573		
5	0.094	1.153	1.408	1.166	0.989	1.110	0.692	0.769	0.408	0.245	1.250	0.672		
6	0.091	1.130	1.284	1.147	0.994	0.888	1.014	0.479	0.268	0.148	1.154	0.599		
7	0.098	1.300	1.251	1.378	1.089	0.791	0.711	0.537	0.437	0.180	1.199	0.533		
8	0.092	1.170	1.192	1.227	0.848	1.097	0.639	0.636	0.316	0.157	1.219	0.471		
Average	0.093	1.200	1.281	1.224	0.992	1.009	0.730	0.655	0.320	0.167	1.197	0.579		
SD	0.004	0.056	0.077	0.097	0.129	0.111	0.137	0.111	0.086	0.042	0.059	0.059		
Average - Blank	0.000	1.107	1.188	1.131	0.899	0.916	0.637	0.562	0.227	0.074	1.104	0.486		
Relative Cell Growth (%)		100.0	107.3	102.2	81.2	82.8	57.5	50.8	20.5	6.7	100.0	44.0		
SD of Relative Cell Growth		5.1	7.0	8.7	11.7	10.1	12.4	10.0	7.8	3.8	5.3	5.3		

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Glensa Staining	10	200	250	300	340	370	400	450	500			0
No. of wells excluded		0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted		96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci		3	7	4	4	11	12	4	8	6	2	41	
Transformation frequency*		0.03125	0.072916667	0.041666667	0.041666667	0.114583333	0.125	0.041666667	0.083333333	0.0625	0.020833333	0.427083333	
SE		0.01775805	0.02653610	0.02039469	0.02039469	0.03250865	0.03375386	0.02039469	0.02820847	0.02470529	0.01457713	0.05048547	

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Caffeine
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	5% Water	18	26	40	59	89	130	200	300			0.1% DMSO
Crystal Violet Staining													
OD _{490nm}													
1	0.067	0.285	0.284	0.297	0.260	0.243	0.222	0.212	0.183	0.161	0.257	0.367	
2	0.067	0.318	0.308	0.314	0.288	0.285	0.269	0.273	0.221	0.187	0.287	0.388	
3	0.065	0.325	0.331	0.346	0.333	0.313	0.280	0.249	0.227	0.182	0.304	0.391	
4	0.065	0.362	0.370	0.332	0.338	0.294	0.258	0.251	0.197	0.175	0.294	0.418	
5	0.066	0.339	0.340	0.312	0.326	0.314	0.261	0.254	0.258	0.177	0.303	0.403	
6	0.065	0.326	0.341	0.334	0.290	0.273	0.281	0.273	0.191	0.170	0.284	0.404	
7	0.063	0.308	0.297	0.281	0.316	0.292	0.257	0.234	0.199	0.166	0.279	0.376	
8	0.063	0.281	0.261	0.270	0.263	0.252	0.233	0.214	0.191	0.166	0.243	0.358	
Average	0.065	0.318	0.317	0.311	0.302	0.284	0.258	0.245	0.208	0.173	#DIV/0!	0.388	
SD	0.002	0.027	0.035	0.027	0.031	0.026	0.021	0.024	0.025	0.009	#DIV/0!	0.020	
Average - Blank	0.000	0.253	0.251	0.246	0.237	0.219	0.193	0.180	0.143	0.108	#DIV/0!	0.323	
Relative Cell Growth (%)		100.0	99.4	97.1	93.6	86.6	76.1	71.1	56.6	42.7	100.0	149.4	
SD of Relative Cell Growth		10.6	14.0	10.5	12.2	10.2	8.3	9.3	10.0	3.5	8.5	8.0	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
	Blank	5% Water	18	26	40	59	89	130	200	300		
Glensa Staining												
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	6	5	4	2	2	5	3	4	3	3	7	68
Transformation frequency*	0.0625	0.052083333	0.041666667	0.020833333	0.020833333	0.0625	0.03125	0.041666667	0.03125	0.03125	0.072916667	0.708333333
SE	0.02470529	0.02267769	0.02039469	0.01457713	0.01457713	0.02470529	0.01775805	0.02039469	0.01775805	0.01775805	0.02653610	0.04639024

* No. of wells having foci/ No. of wells counted

Caffeine (Pro.)

● Transformation frequency
 ◇ Relative cell growth(%)

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

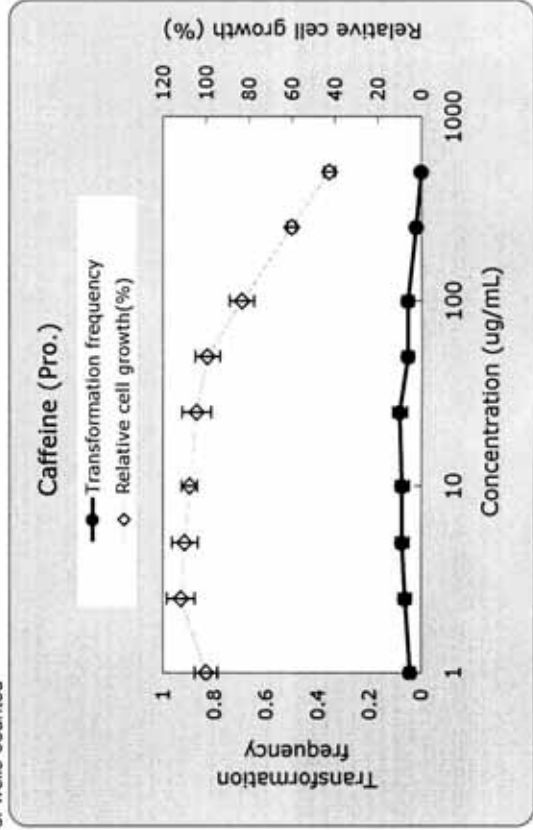
Chemical Code:	Caffeine
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)	
	Blank	5% Water	2.5	5	10	25	50	100	250	500			
Crystal Violet Staining													
OD _{540nm}													
1	0.084	0.995	1.194	1.271	1.136	1.000	0.959	0.966	0.696	0.477	1.170	1.614	
2	0.087	1.041	1.165	1.183	1.194	1.234	1.085	0.943	0.661	0.522	1.105	1.498	
3	0.087	1.127	1.181	1.141	1.148	1.133	1.154	0.904	0.696	0.507	1.095	1.594	
4	0.089	1.158	1.221	1.157	1.196	1.134	1.101	0.915	0.683	0.544	1.190	1.522	
5	0.088	1.105	1.176	1.207	1.193	1.119	1.085	0.977	0.724	0.556	1.154	1.541	
6	0.084	1.081	1.368	1.137	1.224	1.193	1.137	0.903	0.700	0.507	1.152	1.490	
7	0.091	1.117	1.187	1.298	1.167	1.138	1.080	0.970	0.716	0.528	1.110	1.600	
8	0.087	1.130	1.197	1.158	1.116	1.153	1.091	0.813	0.674	0.482	1.144	1.479	
Average	0.087	1.094	1.211	1.194	1.172	1.138	1.087	0.926	0.694	0.515	1.140	1.542	
SD	0.002	0.053	0.066	0.061	0.036	0.068	0.058	0.056	0.021	0.028	0.034	0.054	
Average - Blank	0.000	1.007	1.124	1.107	1.085	1.051	0.999	0.839	0.607	0.428	1.053	1.455	
Relative Cell Growth (%)		100.0	111.6	109.9	107.7	104.3	99.2	83.3	60.2	42.5	100.0	138.2	
SD of Relative Cell Growth		5.3	6.5	6.0	3.6	6.7	5.8	5.6	2.1	2.8	3.3	5.3	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glensa Staining	1	2.5	5	10	25	50	100	250	500			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	4	6	7	7	8	5	5	2	0	0	11	54
Transformation frequency*	0.041666667	0.0625	0.072916667	0.072916667	0.083333333	0.052083333	0.052083333	0.020833333	0	0	0.114583333	0.5625
SE	0.02039469	0.02470529	0.02653610	0.02653610	0.02820847	0.02267769	0.02267769	0.01457713	0.00000000	0.00000000	0.03250865	0.05063079

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

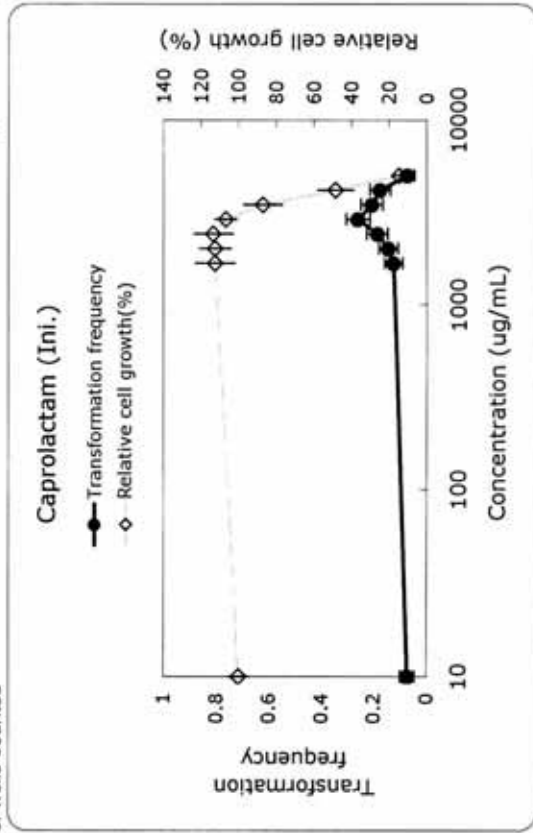
Chemical Code:	Caprolactam
Institution:	Lab 1
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
	Blank	5% Water	1670	2010	2410	2890	3470	4170	5000	5000		
Crystal Violet Staining OD _{540nm}	Blank	10	1670	2010	2410	2890	3470	4170	5000	5000	0.765	0.340
1	0.150	0.748	0.913	0.895	0.890	0.864	0.666	0.458	0.230	0.230	0.704	0.374
2	0.150	0.788	0.794	0.855	0.870	0.794	0.735	0.485	0.256	0.256	0.697	0.337
3	0.158	0.745	0.874	0.876	0.784	0.828	0.707	0.473	0.288	0.288	0.689	0.370
4	0.158	0.782	0.796	0.826	0.945	0.755	0.807	0.515	0.259	0.259	0.680	0.378
5	0.167	0.843	0.817	0.776	0.783	0.846	0.607	0.498	0.293	0.293	0.671	0.371
6	0.173	0.819	0.806	0.871	0.845	0.832	0.754	0.509	0.265	0.265	0.702	0.380
7	0.172	0.776	0.940	0.830	0.853	0.849	0.692	0.457	0.237	0.237	0.749	0.331
8	0.204	0.760	0.941	0.948	0.951	0.827	0.657	0.331	0.227	0.227		
Average	0.167	0.783	0.860	0.860	0.865	0.824	0.703	0.466	0.257	0.257	#DIV/0!	0.360
SD	0.018	0.034	0.065	0.051	0.064	0.035	0.062	0.059	0.025	0.025	#DIV/0!	0.020
Average - Blank	0.000	0.616	0.694	0.693	0.699	0.658	0.537	0.299	0.090	0.090	#DIV/0!	0.194
Relative Cell Growth (%)		100.0	112.6	112.5	113.4	106.8	87.1	48.6	14.7	14.7	#DIV/0!	35.8
SD of Relative Cell Growth		5.5	10.5	8.3	10.3	5.6	10.1	9.5	4.0	4.0	#DIV/0!	3.3

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
	Blank	5% Water	1670	2010	2410	2890	3470	4170	5000	5000		
Giemsa Staining	Blank	10	1670	2010	2410	2890	3470	4170	5000	5000	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	7	12	14	18	25	20	17	7	7	7	9	43
Transformation frequency*	0.072916667	0.125	0.145833333	0.1875	0.260416667	0.208333333	0.177083333	0.072916667	0.072916667	0.072916667	0.09375	0.447916667
SE	0.02653610	0.03375386	0.03602169	0.03983609	0.04479116	0.04144908	0.03896110	0.02653610	0.00000000	0.00000000	0.02974911	0.05075342

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

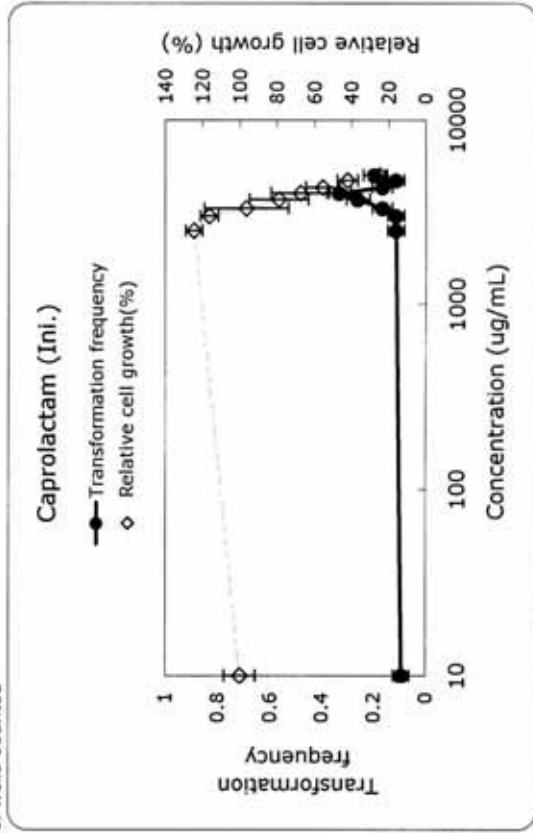
Chemical Code:	Caprolactam
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (µg/mL)	
	Blank	5% Water	2500	3000	3300	3700	4000	4300	4700	5000			
Crystal Violet Staining													
OD _{540nm}													
1	0.086	1.188	1.542	1.434	0.584	0.822	0.708	0.551	0.593	0.398	1.292	0.545	
2	0.097	1.365	1.515	1.345	1.386	0.634	0.537	0.762	0.677	0.321	1.318	0.559	
3	0.089	1.359	1.488	1.420	1.297	1.025	0.871	0.823	0.595	0.387	1.243	0.564	
4	0.085	1.211	1.471	1.337	1.306	1.170	1.102	0.628	0.496	0.363	1.144	0.539	
5	0.090	1.214	1.442	1.394	1.228	1.078	0.989	0.721	0.538	0.387	1.142	0.517	
6	0.090	1.152	1.434	1.507	1.316	1.050	0.809	0.788	0.507	0.292	1.201	0.520	
7	0.095	1.169	1.529	1.390	1.235	1.133	0.954	0.634	0.603	0.443	1.180	0.551	
8	0.088	1.104	1.577	1.429	1.088	0.966	0.861	0.839	0.514	0.396	1.180	0.500	
Average	0.090	1.220	1.500	1.407	1.180	0.985	0.854	0.718	0.565	0.373	#DIV/0!	0.537	
SD	0.004	0.094	0.050	0.054	0.256	0.178	0.175	0.104	0.062	0.048	#DIV/0!	0.022	
Average - Blank	0.000	1.130	1.410	1.317	1.090	0.895	0.764	0.628	0.475	0.283	#DIV/0!	0.447	
Relative Cell Growth (%)		100.0	124.7	116.5	96.4	79.2	67.6	55.6	42.1	25.1	#DIV/0!	39.8	
SD of Relative Cell Growth		8.3	4.4	4.8	22.7	15.7	15.5	9.2	5.5	4.2	#DIV/0!	2.0	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
	Glenssa Staining	2500	3000	3300	3700	4000	4300	4700	5000			
No. of wells excluded	10	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	9	11	11	16	25	32	16	11	11	19	8	65
Transformation frequency*	0.09375	0.114583333	0.114583333	0.166666667	0.260416667	0.333333333	0.166666667	0.114583333	0.197916667	0.197916667	0.083333333	0.677083333
SE	0.02974911	0.03250865	0.03250865	0.03803629	0.04479116	0.04811252	0.03803629	0.03250865	0.04066449	0.04066449	0.02820847	0.04772332

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

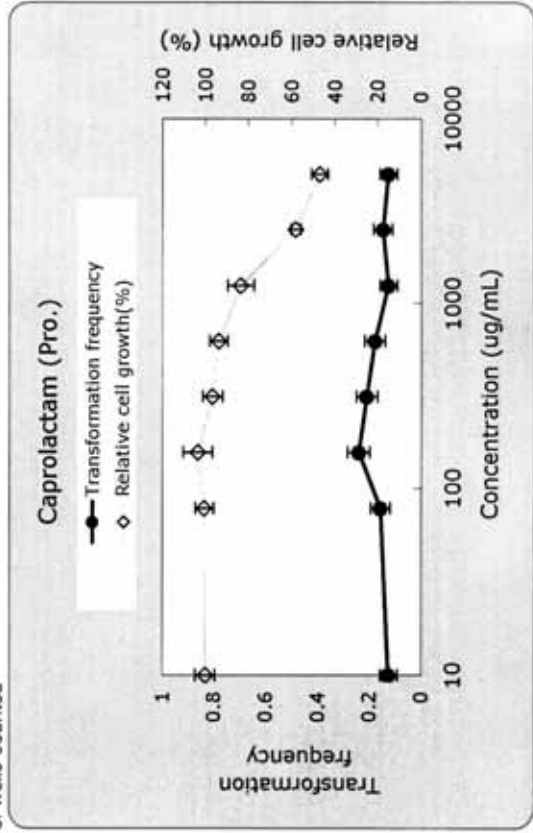
Chemical Code:	Caprolactam
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)								Negative Control	Positive Control	
			5% Water	78.1	156	313	625	1250	2500	5000			
OD _{540nm}	10												
1	0.121	0.706	0.708	0.682	0.648	0.631	0.540	0.457	0.412	0.622	0.799		
2	0.121	0.661	0.681	0.665	0.685	0.612	0.567	0.441	0.395	0.654	0.867		
3	0.119	0.662	0.653	0.750	0.633	0.657	0.599	0.427	0.358	0.661	0.895		
4	0.122	0.688	0.696	0.637	0.673	0.643	0.568	0.442	0.368	0.676	0.922		
5	0.120	0.636	0.702	0.704	0.655	0.671	0.607	0.454	0.370	0.643	0.806		
6	0.125	0.677	0.648	0.742	0.670	0.653	0.565	0.435	0.369	0.682	0.870		
7	0.125	0.660	0.657	0.706	0.617	0.611	0.597	0.446	0.386	0.694	0.831		
8	0.130	0.716	0.697	0.688	0.692	0.665	0.650	0.462	0.413	0.674	0.874		
Average	0.123	0.678	0.680	0.697	0.659	0.643	0.587	0.446	0.384	0.663	0.858		
SD	0.004	0.026	0.024	0.038	0.026	0.023	0.034	0.012	0.021	0.023	0.043		
Average - Blank	0.000	0.555	0.557	0.574	0.536	0.520	0.464	0.323	0.261	0.540	0.735		
Relative Cell Growth (%)	100.0	103.3	100.4	103.3	96.6	93.6	83.5	58.1	47.0	100.0	136.0		
SD of Relative Cell Growth	4.7	4.4	4.4	6.8	4.6	4.1	6.1	2.1	3.8	4.2	7.7		

Transformation Assay	Concentrations (ug/ml)								Negative Control	Positive Control	
Glenssa Staining	10	78.1	156	313	625	1250	2500	5000			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	12	15	23	20	17	12	14	12	12	14	53
Transformation frequency*	0.125	0.15625	0.239583333	0.208333333	0.177083333	0.125	0.145833333	0.125	0	0.145833333	0.65625
SE	0.03375386	0.03705794	0.04356307	0.04144908	0.03896110	0.03375386	0.03602169	0.03375386	0.00000000	0.03602169	0.04847529

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

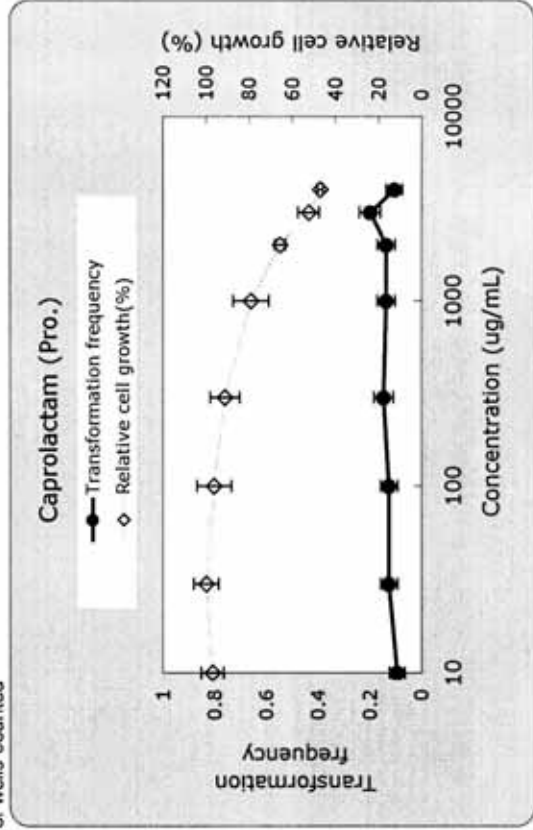
Chemical Code:	Caprolactam
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Blank	Concentrations (ug/ml)						Negative Control		Positive Control		
			5% Water	10	30	100	300	1000	2000	3000	4000	0.1% DMSO	TPA (50ng/mL)
1		0.069	1.092	1.090	1.082	1.138	1.053	0.902	0.785	0.633	0.574	1.109	1.452
2		0.076	1.226	1.237	1.232	1.125	1.111	0.940	0.797	0.623	0.570	1.131	1.589
3		0.080	1.154	1.147	1.150	1.156	1.115	0.955	0.780	0.681	0.586	1.347	1.591
4		0.084	1.224	1.142	1.085	1.200	1.043	0.970	0.844	0.653	0.607	1.178	1.607
5		0.087	1.199	1.052	1.205	1.145	1.170	1.016	0.792	0.710	0.598	1.201	1.493
6		0.087	1.119	1.115	1.187	1.146	1.082	0.959	0.804	0.672	0.594	1.136	1.627
7		0.086	1.157	1.129	1.146	1.197	1.105	1.051	0.799	0.686	0.611	1.117	1.515
8		0.081	1.186	1.180	1.255	0.924	0.920	0.752	0.756	0.538	0.602	1.102	1.624
Average		0.081	1.170	1.137	1.169	1.129	1.075	0.943	0.795	0.650	0.592	1.165	1.562
SD		0.006	0.048	0.056	0.062	0.087	0.074	0.090	0.025	0.053	0.014	0.081	0.066
Average - Blank		0.000	1.088	1.055	1.088	1.048	0.994	0.862	0.713	0.568	0.511	1.084	1.481
Relative Cell Growth (%)		100.0	97.0	99.9	96.3	91.3	79.2	82.2	65.5	52.2	46.9	100.0	136.6
SD of Relative Cell Growth		4.4	5.1	5.7	8.0	6.8	8.2	8.2	2.3	4.9	1.3	7.4	6.1

Transformation Assay	Concentrations (ug/ml)						Negative Control		Positive Control				
Giemsa Staining	1	10	30	100	300	1000	2000	3000	4000	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	10	9	12	12	14	13	13	19	10	10	11	54	54
Transformation frequency*	0.104166667	0.09375	0.125	0.125	0.145833333	0.135416667	0.135416667	0.197916667	0.104166667	0	0.114583333	0.5625	0.5625
SE	0.03117758	0.02974911	0.03375386	0.03375386	0.03602169	0.03492238	0.03492238	0.04066449	0.03117758	0.00000000	0.03250865	0.05063079	0.05063079

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

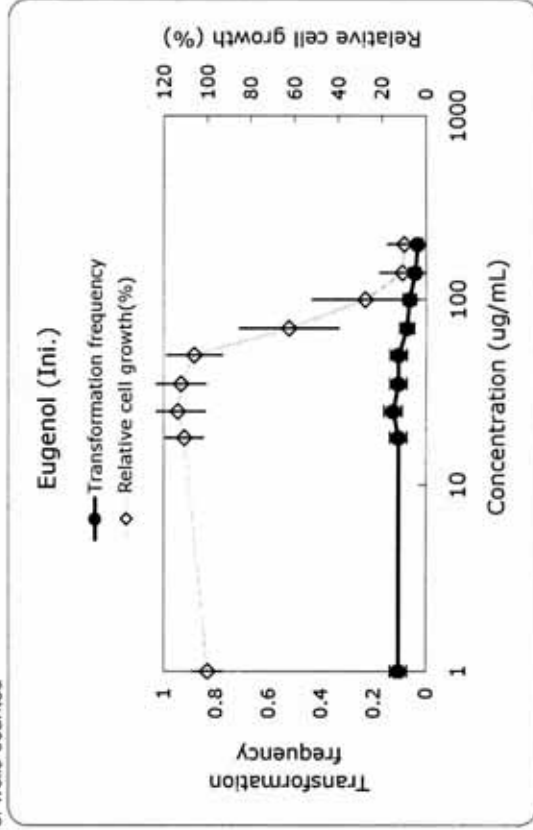
Chemical Code:	Eugenol
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	0.1% DMSO	18	25	35	50	70	100	140	200			0.1% DMSO
Crystal Violet Staining													
OD _{490nm}													
1	0.065	0.383	0.450	0.414	0.429	0.478	0.462	0.365	0.182	0.160	0.383	0.197	
2	0.062	0.424	0.486	0.461	0.425	0.473	0.306	0.163	0.103	0.094	0.424	0.156	
3	0.060	0.432	0.414	0.408	0.465	0.381	0.231	0.118	0.097	0.085	0.432	0.105	
4	0.059	0.436	0.407	0.537	0.533	0.390	0.274	0.112	0.087	0.087	0.436	0.120	
5	0.060	0.423	0.486	0.487	0.445	0.446	0.255	0.113	0.079	0.081	0.423	0.125	
6	0.060	0.401	0.469	0.420	0.432	0.384	0.233	0.125	0.081	0.086	0.401	0.109	
7	0.060	0.422	0.448	0.471	0.493	0.421	0.208	0.124	0.088	0.082	0.422	0.133	
8	0.065	0.371	0.429	0.471	0.414	0.487	0.278	0.150	0.068	0.094	0.371	0.146	
Average	0.061	0.412	0.449	0.459	0.455	0.433	0.281	0.159	0.099	0.096	0.412	0.136	
SD	0.002	0.024	0.031	0.044	0.041	0.044	0.080	0.085	0.035	0.026	0.024	0.030	
Average - Blank	0.000	0.350	0.387	0.397	0.393	0.371	0.220	0.097	0.038	0.035	0.350	0.075	
Relative Cell Growth (%)			110.6	113.5	112.3	106.0	62.7	27.8	10.7	9.9	100.0	21.4	
SD of Relative Cell Growth			8.7	12.5	11.6	12.7	22.7	24.4	10.1	7.5	6.8	8.6	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
	Blank	0.1% DMSO	18	25	35	50	70	100	140	200		
Glensa Staining												
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	10	10	12	10	10	7	4	6	4	3	10	64
Transformation frequency*	0.104166667	0.104166667	0.125	0.104166667	0.104166667	0.072916667	0.041666667	0.0625	0.041666667	0.03125	0.104166667	0.666666667
SE	0.03117758	0.03117758	0.03375386	0.03117758	0.03117758	0.02653610	0.02470529	0.02470529	0.02039469	0.01775805	0.03117758	0.04811252

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

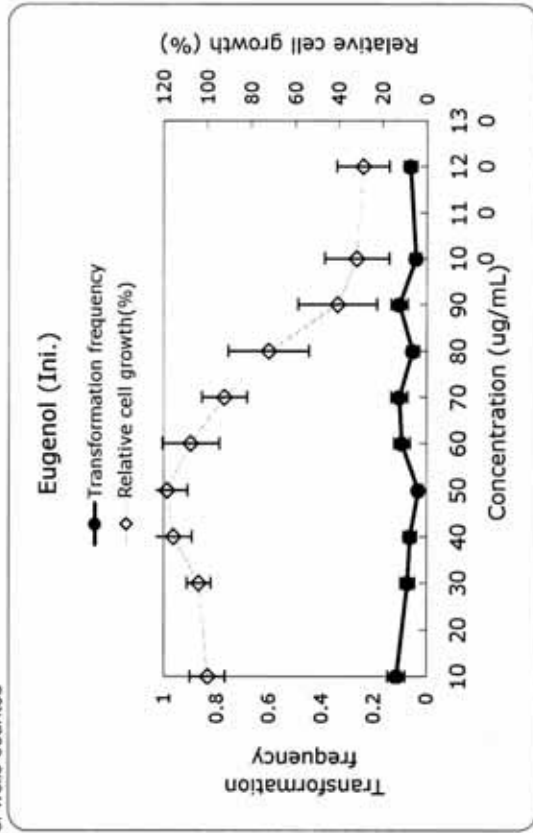
Chemical Code:	Eugenol
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)											Negative Control	Positive Control			
	Blank	10	30	40	50	60	70	80	90	100	120			0.1% DMSO	MCA (1ug/mL)	
Crystal Violet Staining																
OD _{540nm}																
1	0.087	0.781	0.757	0.817	0.828	0.899	0.777	0.777	0.503	0.472	0.366	0.781	0.239			
2	0.088	0.747	0.791	0.874	0.757	0.747	0.722	0.551	0.383	0.312	0.377	0.747	0.251			
3	0.089	0.717	0.686	0.810	0.890	0.798	0.659	0.475	0.295	0.283	0.311	0.717	0.228			
4	0.090	0.734	0.706	0.785	0.847	0.756	0.640	0.509	0.257	0.273	0.228	0.734	0.167			
5	0.086	0.677	0.741	0.807	0.790	0.769	0.625	0.415	0.220	0.190	0.221	0.677	0.178			
6	0.081	0.677	0.717	0.853	0.813	0.640	0.631	0.483	0.213	0.182	0.204	0.677	0.169			
7	0.084	0.648	0.710	0.742	0.883	0.665	0.586	0.447	0.345	0.252	0.217	0.648	0.177			
8	0.082	0.643	0.710	0.714	0.728	0.729	0.603	0.590	0.477	0.295	0.201	0.643	0.184			
Average	0.086	0.703	0.727	0.800	0.817	0.750	0.655	0.531	0.337	0.282	0.266	0.703	0.199			
SD	0.003	0.050	0.034	0.053	0.057	0.080	0.064	0.114	0.111	0.090	0.074	0.050	0.034			
Average - Blank	0.000	0.617	0.641	0.714	0.731	0.665	0.570	0.445	0.251	0.197	0.180	0.617	0.113			
Relative Cell Growth (%)		100.0	103.9	115.8	118.5	107.7	92.3	72.1	40.6	31.8	29.1	100.0	18.4			
SD of Relative Cell Growth		8.0	5.5	8.6	9.3	12.9	10.3	18.5	18.0	14.6	12.0	8.0	5.6			

Transformation Assay	Concentrations (ug/ml)											Negative Control	Positive Control
	Blank	10	30	40	50	60	70	80	90	100	120		
Glensa Staining													
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	11	7	6	6	3	9	10	5	10	4	6	11	59
Transformation frequency*	0.114583333	0.072916667	0.0625	0.0625	0.03125	0.09375	0.104166667	0.052083333	0.104166667	0.041666667	0.0625	0.114583333	0.614583333
SE	0.03250865	0.02653610	0.02470529	0.02470529	0.01775805	0.02974911	0.03117758	0.02267769	0.03117758	0.02039469	0.02470529	0.03250865	0.04967296

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

Data Sheet for Cell Transformation Assay on Promotion Assay

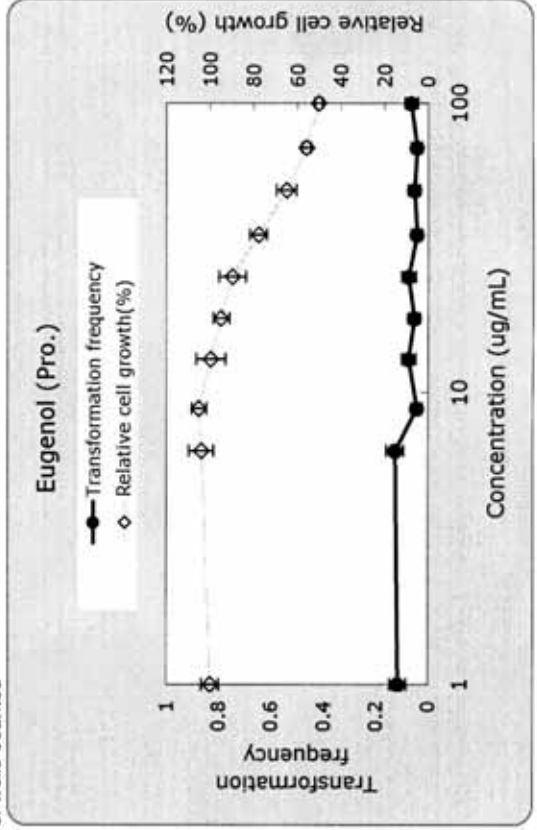
Chemical Code:	Eugenol
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	0.1% DMSO	6.3	8.8	13	18	25	35	50	70			100
Crystal Violet Staining													
OD _{540nm}													
1	0.060	0.304	0.301	0.317	0.280	0.292	0.276	0.239	0.211	0.194	0.190	0.304	0.280
2	0.063	0.303	0.330	0.316	0.331	0.297	0.312	0.264	0.227	0.207	0.187	0.303	0.283
3	0.063	0.301	0.336	0.316	0.301	0.309	0.286	0.254	0.243	0.204	0.183	0.301	0.274
4	0.065	0.331	0.322	0.338	0.304	0.301	0.294	0.254	0.218	0.197	0.184	0.331	0.264
5	0.065	0.307	0.330	0.312	0.292	0.294	0.276	0.256	0.218	0.206	0.186	0.307	0.281
6	0.066	0.310	0.305	0.322	0.318	0.295	0.282	0.262	0.225	0.195	0.182	0.310	0.258
7	0.065	0.301	0.313	0.322	0.306	0.294	0.266	0.259	0.227	0.196	0.194	0.301	0.278
8	0.064	0.304	0.302	0.315	0.319	0.277	0.267	0.240	0.208	0.199	0.182	0.304	0.273
Average	0.064	0.308	0.317	0.320	0.306	0.295	0.282	0.254	0.222	0.200	0.186	0.308	0.274
SD	0.002	0.010	0.014	0.008	0.016	0.009	0.015	0.009	0.011	0.005	0.004	0.010	0.009
Average - Blank	0.000	0.244	0.254	0.256	0.243	0.231	0.219	0.190	0.158	0.136	0.122	0.244	0.210
Relative Cell Growth (%)			104.0	105.0	99.5	94.8	89.6	77.8	64.9	55.7	50.1	100.0	86.2
SD of Relative Cell Growth		4.1	5.7	3.3	6.6	3.7	6.2	3.8	4.5	2.1	1.7	4.1	3.6

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	0.1% DMSO	6.3	8.8	13	18	25	35	50	70			100
Glensa Staining													
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	11	12	4	7	7	5	7	4	5	4	6	11	48
Transformation frequency*	0.114583333	0.125	0.041666667	0.072916667	0.052083333	0.072916667	0.041666667	0.041666667	0.052083333	0.041666667	0.0625	0.114583333	0.5
SE	0.03250865	0.03375386	0.02039469	0.02653610	0.02267769	0.02653610	0.02267769	0.02039469	0.02267769	0.02039469	0.02470529	0.03250865	0.05103104

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

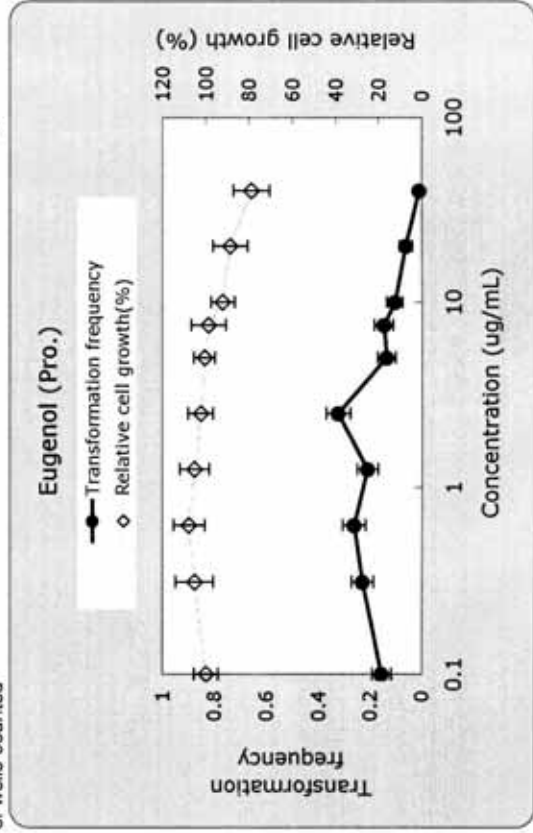
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Eugenol
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining	Blank	Concentrations (ug/ml)										Negative Control	Positive Control	
			0.1% DMSO	0.1	0.3125	0.625	1.25	2.5	5	7.5	10	20			40
1	0.114	0.114	1.373	1.373	1.373	1.571	1.351	1.238	1.319	1.218	1.174	1.131	0.982	1.373	1.409
2	0.127	0.127	1.397	1.310	1.396	1.396	1.374	1.433	1.400	1.311	1.262	1.127	1.045	1.397	1.423
3	0.134	0.134	1.399	1.332	1.420	1.420	1.416	1.415	1.414	1.317	1.239	1.223	1.001	1.399	1.624
4	0.126	0.126	1.319	1.497	1.517	1.396	1.467	1.378	1.378	1.343	1.311	1.218	1.087	1.319	1.605
5	0.124	0.124	1.448	1.568	1.375	1.375	1.545	1.452	1.318	1.340	1.405	1.305	1.212	1.448	1.507
6	0.119	0.119	1.518	1.623	1.499	1.499	1.595	1.435	1.468	1.477	1.290	1.303	1.200	1.518	1.733
7	0.115	0.115	1.297	1.481	1.640	1.479	1.445	1.445	1.375	1.418	1.347	1.428	1.240	1.297	1.640
8	0.109	0.109	1.345	1.453	1.463	1.463	1.448	1.448	1.487	1.532	1.276	1.213	1.194	1.345	1.726
Average	0.121	0.121	1.387	1.455	1.485	1.485	1.452	1.417	1.395	1.370	1.288	1.244	1.120	1.387	1.583
SD	0.008	0.008	0.072	0.111	0.091	0.091	0.085	0.074	0.062	0.101	0.070	0.100	0.103	0.072	0.125
Average - Blank	0.000	0.000	1.266	1.334	1.364	1.364	1.331	1.296	1.274	1.249	1.167	1.123	0.999	1.266	1.462
Relative Cell Growth (%)			100.0	105.3	107.8	105.2	102.3	102.3	100.6	98.6	92.2	88.7	78.9	100.0	115.5
SD of Relative Cell Growth			5.6	8.8	7.2	6.7	5.8	5.8	4.9	8.0	5.5	7.9	8.2	5.6	9.9

Transformation Assay	Giemsa Staining	No. of wells excluded	No. of wells counted	No. of wells having foci	Transformation frequency*	SE	Concentrations (ug/ml)										Negative Control	Positive Control					
							0.1	0.3125	0.625	1.25	2.5	5	7.5	10	20	40			0.1% DMSO	TPA (50ng/mL)			
							0.1	0.3125	0.625	1.25	2.5	5	7.5	10	20	40	0	0	0	0	0	0	
		96	96	96	0.260416667	0.022916667	0.229166667	0.260416667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	96	96	96	96	96	96
		15	15	15	0.04289635	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	0.04479116	15	15	15	15	15	15	
					0.15625	0.03705794	0.15625	0.229166667	0.260416667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.322916667	0.15625	0.15625	0.15625	0.15625	0.15625	0.15625
					0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794	0.03705794

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
 Negative Control < 20 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Progressing Data

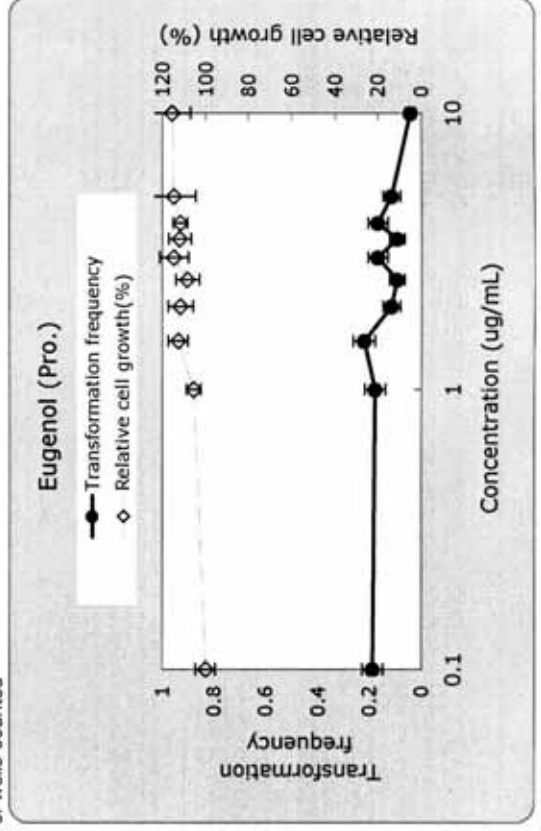
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Eugenol
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)	
		Blank	0.1	1	1.5	2	2.5	3	3.5	4	5			10
1	0.064	0.064	0.612	0.578	0.617	0.610	0.594	0.601	0.612	0.640	0.624	0.653	0.612	0.798
2	0.064	0.064	0.542	0.628	0.665	0.581	0.590	0.677	0.625	0.625	0.674	0.620	0.542	0.765
3	0.064	0.064	0.568	0.606	0.654	0.633	0.615	0.688	0.640	0.653	0.621	0.670	0.568	0.767
4	0.068	0.068	0.591	0.605	0.632	0.651	0.645	0.676	0.671	0.637	0.617	0.660	0.591	0.764
5	0.067	0.067	0.584	0.607	0.676	0.665	0.601	0.660	0.606	0.666	0.649	0.671	0.584	0.753
6	0.068	0.068	0.572	0.618	0.655	0.656	0.649	0.620	0.639	0.634	0.771	0.750	0.572	0.837
7	0.069	0.069	0.601	0.588	0.608	0.662	0.665	0.681	0.639	0.611	0.638	0.623	0.601	0.821
8	0.069	0.069	0.556	0.619	0.641	0.643	0.613	0.621	0.682	0.642	0.630	0.613	0.556	0.769
Average	0.067	0.067	0.578	0.606	0.644	0.638	0.622	0.653	0.639	0.639	0.653	0.658	0.578	0.784
SD	0.002	0.002	0.023	0.017	0.023	0.029	0.028	0.034	0.026	0.017	0.051	0.044	0.023	0.031
Average - Blank	0.000	0.000	0.540	0.577	0.571	0.571	0.555	0.586	0.573	0.572	0.586	0.591	0.512	0.718
Relative Cell Growth (%)		100.0	105.4	112.8	111.6	108.5	114.6	114.6	111.9	111.8	114.6	115.5	100.0	140.3
SD of Relative Cell Growth		4.6	3.2	4.6	5.7	5.5	6.6	6.6	5.2	3.3	10.0	8.6	4.6	6.0

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Glensa Staining	0.1	1	1.5	2	2.5	3	3.5	4	5	10			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	18	17	21	11	9	16	9	16	11	4	18	83	83
Transformation frequency*	0.1875	0.177083333	0.21875	0.114583333	0.09375	0.166666667	0.09375	0.166666667	0.114583333	0.041666667	0.1875	0.864583333	0.864583333
SE	0.03983609	0.03896110	0.04219232	0.03250865	0.02974911	0.03803629	0.02974911	0.03803629	0.03250865	0.02039469	0.03983609	0.03492238	0.03492238

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

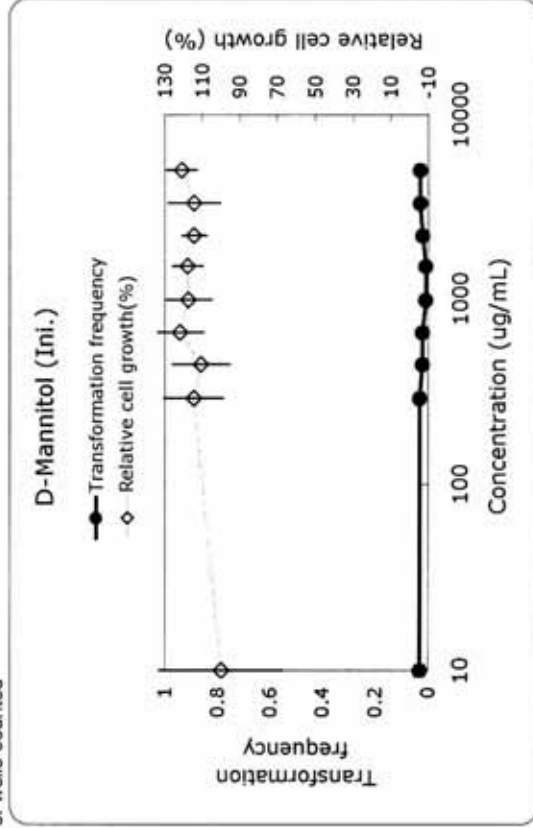
Chemical Code:	D-Mannitol
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)	
	Blank	5% Water	290	440	660	990	1500	2200	3300	5000			
Crystal Violet Staining	Blank	10											
OD _{540nm}													
1	0.104	0.299	0.308	0.275	0.307	0.276	0.297	0.309	0.305	0.301		0.257	0.174
2	0.107	0.299	0.339	0.311	0.288	0.313	0.333	0.304	0.308	0.335		0.322	0.211
3	0.104	0.340	0.301	0.311	0.318	0.292	0.306	0.325	0.368	0.318		0.329	0.212
4	0.107	0.290	0.316	0.304	0.335	0.343	0.312	0.318	0.295	0.333		0.318	0.188
5	0.097	0.321	0.345	0.297	0.359	0.335	0.326	0.304	0.290	0.314		0.318	0.215
6	0.100	0.328	0.321	0.351	0.343	0.333	0.338	0.316	0.320	0.345		0.290	0.199
7	0.095	0.234	0.299	0.317	0.308	0.318	0.304	0.316	0.293	0.318		0.240	0.216
8	0.098	0.158	0.252	0.259	0.330	0.312	0.311	0.286	0.300	0.311		0.148	0.183
Average	0.102	0.284	0.310	0.303	0.324	0.315	0.316	0.310	0.310	0.322	#DIV/0!	0.278	0.200
SD	0.005	0.060	0.029	0.028	0.023	0.023	0.015	0.012	0.025	0.015	#DIV/0!	0.062	0.016
Average - Blank	0.000	0.182	0.209	0.202	0.222	0.214	0.214	0.208	0.208	0.220	#DIV/0!	0.176	0.098
Relative Cell Growth (%)		100.0	114.6	110.7	121.9	117.4	117.7	114.3	114.4	121.0	#DIV/0!	100.0	55.7
SD of Relative Cell Growth		33.0	15.8	15.2	12.5	12.4	8.1	6.6	13.9	8.0	#DIV/0!	33.9	8.9

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Glensa Staining	10	290	440	660	990	1500	2200	3300	5000			
No. of wells excluded		0	0	2	0	0	0	1	0	0	0	0	0
No. of wells counted		96	96	94	96	96	96	95	96	96	96	96	96
No. of wells having foci		3	3	2	2	1	1	2	3	3	4	4	57
Transformation frequency*		0.03125	0.03125	0.021276596	0.020833333	0.010416667	0.010416667	0.021052632	0.03125	0.03125	0.03125	0.041666667	0.59375
SE		0.01775805	0.01775805	0.01488391	0.01457713	0.01036227	0.01036227	0.01472893	0.01775805	0.01775805	0.02039469	0.05012598	

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

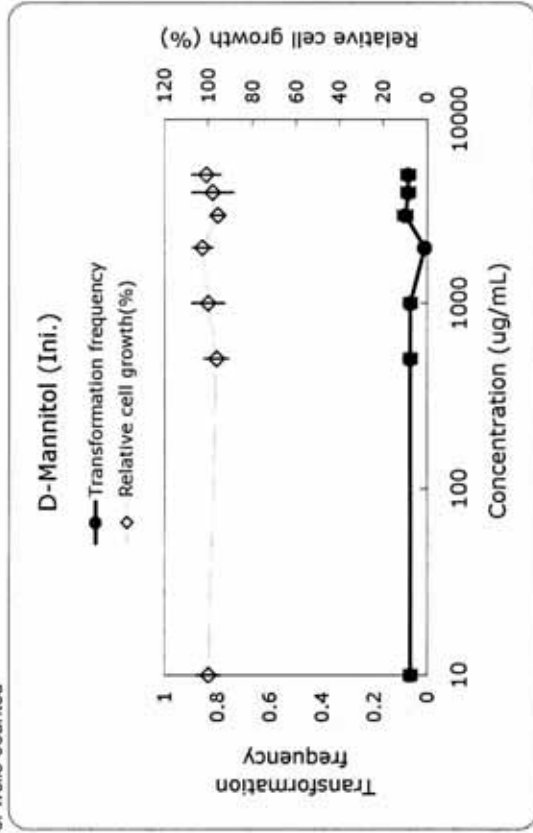
Chemical Code:	D-Mannitol
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)								Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)
		Blank	5% Water	10	500	1000	2000	3000	4000		
1	0.118	1.304	1.168	1.191	1.428	1.295	1.451	1.373	1.231	0.537	
2	0.120	1.270	1.282	1.178	1.274	1.263	1.306	1.235	1.336	0.643	
3	0.119	1.290	1.217	1.373	1.307	1.207	1.196	1.271	1.270	0.571	
4	0.122	1.245	1.312	1.317	1.295	1.231	1.177	1.478	1.337	0.534	
5	0.121	1.370	1.351	1.292	1.332	1.234	1.133	1.296	1.231	0.528	
6	0.123	1.285	1.208	1.359	1.335	1.313	1.335	1.263	1.278	0.666	
7	0.125	1.446	1.288	1.405	1.379	1.280	1.314	1.362	1.331	0.571	
8	0.123	1.303	1.328	1.404	1.410	1.274	1.407	1.327	1.369	0.526	
Average	0.121	1.314	1.269	1.315	1.345	1.262	1.290	1.326	#DIV/0!	0.572	
SD	0.002	0.064	0.065	0.089	0.055	0.036	0.113	0.078	#DIV/0!	0.052	
Average - Blank	0.000	1.193	1.148	1.194	1.224	1.141	1.169	1.204	#DIV/0!	0.451	
Relative Cell Growth (%)		100.0	96.2	100.1	102.6	95.6	98.0	101.0	#DIV/0!	38.3	
SD of Relative Cell Growth		5.4	5.4	7.5	4.6	3.0	9.4	6.6	#DIV/0!	4.5	

Transformation Assay	Concentrations (ug/ml)										
Giemsa Staining	10	500	1000	2000	3000	4000	5000	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	6	6	6	1	8	7	7	7	10	10	46
Transformation frequency*	0.0625	0.0625	0.0625	0.010416667	0.083333333	0.072916667	0.072916667	0.072916667	0.104166667	0.104166667	0.479166667
SE	0.02470529	0.02470529	0.02470529	0.01036227	0.02820847	0.02653610	0.02653610	0.02653610	0.03117758	0.03117758	0.05098672

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

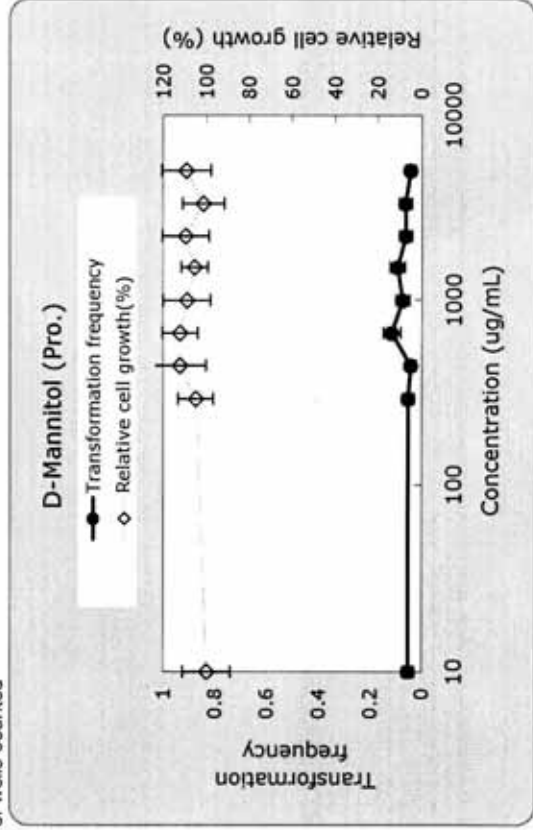
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	D-Mannitol
Institution:	Lab 2
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)								Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
		Blank	5% Water	290	440	660	990	1500	2200		
1	0.104	0.330	0.350	0.355	0.366	0.388	0.348	0.399	0.359	0.355	0.500
2	0.107	0.339	0.397	0.395	0.412	0.385	0.380	0.404	0.381	0.393	0.482
3	0.104	0.357	0.363	0.421	0.376	0.399	0.381	0.399	0.335	0.380	0.458
4	0.107	0.412	0.361	0.428	0.401	0.381	0.385	0.359	0.396	0.345	0.467
5	0.097	0.347	0.371	0.392	0.398	0.407	0.363	0.401	0.330	0.339	0.466
6	0.100	0.373	0.399	0.356	0.385	0.341	0.372	0.335	0.376	0.330	0.485
7	0.095	0.355	0.352	0.391	0.395	0.389	0.377	0.384	0.352	0.342	0.440
8	0.098	0.323	0.345	0.346	0.350	0.328	0.342	0.349	0.335	0.352	0.448
Average	0.102	0.355	0.367	0.386	0.385	0.377	0.369	0.379	0.358	0.378	0.468
SD	0.005	0.028	0.021	0.031	0.020	0.028	0.016	0.027	0.024	0.029	0.020
Average - Blank	0.000	0.253	0.266	0.284	0.284	0.276	0.267	0.277	0.257	0.277	0.367
Relative Cell Growth (%)		100.0	105.0	112.3	112.2	109.0	105.5	109.6	101.4	109.3	156.2
SD of Relative Cell Growth		11.1	8.2	12.1	8.1	11.0	6.3	10.7	9.7	11.4	7.9

Transformation Assay	Concentrations (ug/ml)								Negative Control	Positive Control
Giemsa Staining	10	290	440	660	990	1500	2200	3300		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	5	5	4	11	7	9	6	6	4	8
Transformation frequency*	0.052083333	0.052083333	0.041666667	0.114583333	0.072916667	0.09375	0.0625	0.0625	0.041666667	0
SE	0.02267769	0.02267769	0.02039469	0.03250865	0.02653610	0.02974911	0.02470529	0.02470529	0.02039469	0.00000000

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

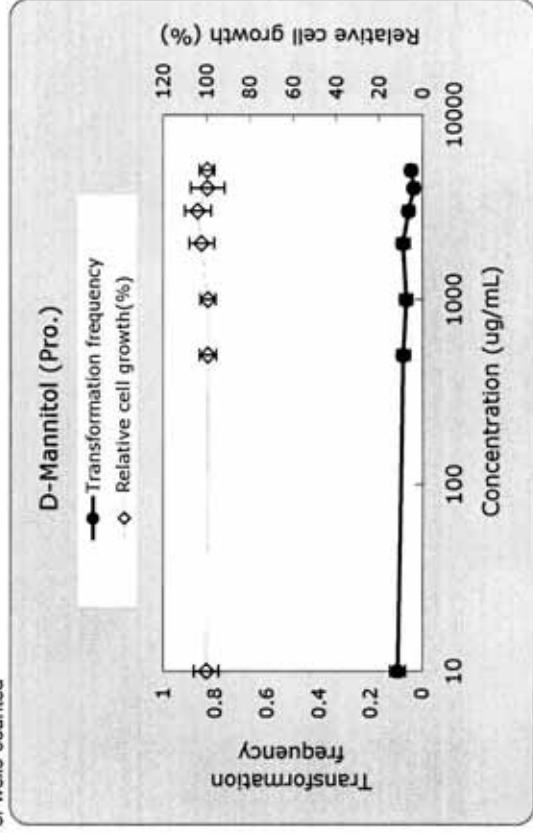
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	D-Mannitol
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)								Negative Control		Positive Control	
		Blank	5% Water	10	500	1000	2000	3000	4000	5000	0.1% DMSO	TPA (50ug/mL)	
1	0.104		1.086	1.126	1.100	1.104	1.199	1.149	1.166	1.146	1.729		
2	0.109		1.113	1.163	1.116	1.186	1.085	1.215	1.166	1.088	1.648		
3	0.114		1.129	1.174	1.200	1.241	1.169	1.120	1.162	1.211	1.786		
4	0.114		1.159	1.175	1.179	1.220	1.276	1.227	1.109	1.181	1.682		
5	0.112		1.169	1.221	1.167	1.142	1.276	1.231	1.194	1.197	1.783		
6	0.111		1.248	1.126	1.198	1.184	1.216	1.217	1.157	1.163	1.840		
7	0.115		1.215	1.113	1.162	1.299	1.228	0.987	1.149	1.141	1.774		
8	0.117		1.248	1.210	1.183	1.171	1.251	1.179	1.236	1.311	1.750		
Average	0.112		1.171	1.164	1.163	1.193	1.213	1.166	1.167	#DIV/0!	1.180	1.749	
SD	0.004		0.061	0.040	0.037	0.060	0.063	0.082	0.036	#DIV/0!	0.065	0.062	
Average - Blank	0.000		1.059	1.052	1.051	1.081	1.101	1.054	1.055	#DIV/0!	1.068	1.637	
Relative Cell Growth (%)			100.0	99.3	99.3	102.1	103.9	99.5	99.7	#DIV/0!	100.0	153.3	
SD of Relative Cell Growth			5.8	3.8	3.5	5.7	6.0	7.8	3.4	#DIV/0!	6.2	5.8	

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
Giensa Staining	10	500	1000	2000	3000	4000	5000	5000	0	0	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	9	7	6	7	5	3	4	4	4	4	4	4	4	4
Transformation frequency*	0.09375	0.072916667	0.0625	0.072916667	0.052083333	0.03125	0.041666667	0.041666667	0.041666667	0.041666667	0.041666667	0.041666667	0.041666667	0.041666667
SE	0.02974911	0.02653610	0.02470529	0.02653610	0.02267769	0.01775805	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469	0.02039469

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass
Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

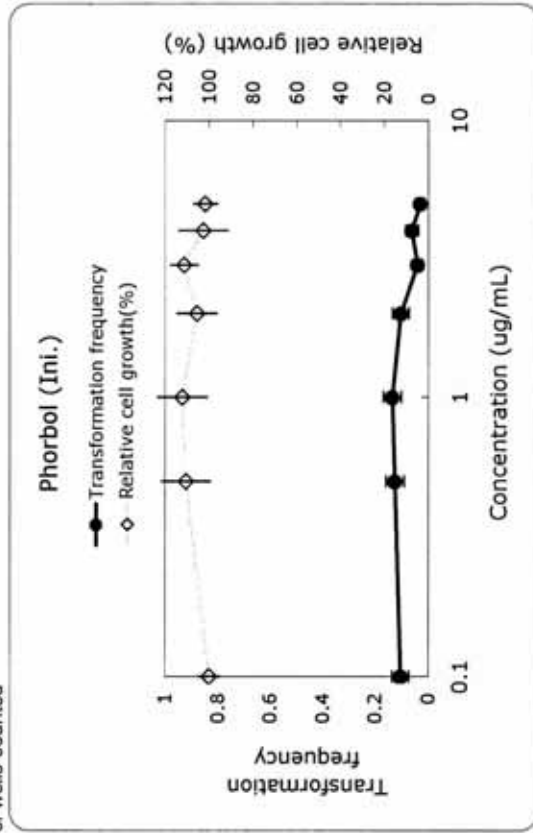
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Phorbol
Institution:	Lab 1
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{490nm}	Concentrations (ug/ml)					Negative Control 0.1% DMSO	Positive Control MCA (1ug/mL)				
		Blank	5% Water 0.1	0.5	1	2			3	4	5	
1	0.107		0.829	0.776	0.898	0.808	0.888	0.816	0.815		0.866	0.369
2	0.125		0.869	0.900	0.868	0.990	0.920	0.785	0.773		0.837	0.484
3	0.114		0.867	0.911	0.810	0.920	0.890	0.757	0.864		0.761	0.512
4	0.133		0.786	0.826	0.956	0.869	0.900	0.945	0.878		0.897	0.419
5	0.121		0.802	0.940	1.075	0.854	0.860	0.821	0.845		0.839	0.462
6	0.150		0.818	1.014	0.966	0.851	1.003	0.860	0.863		0.832	0.454
7	0.142		0.859	0.967	0.947	0.922	0.931	0.980	0.885		1.009	0.471
8	0.164		0.880	0.965	0.878	0.803	0.949	0.897	0.876		0.854	0.395
Average	0.132		0.839	0.912	0.925	0.877	0.918	0.858	0.850	#DIV/0!	#DIV/0!	0.446
SD	0.019		0.035	0.078	0.080	0.063	0.044	0.078	0.038	#DIV/0!	#DIV/0!	0.048
Average - Blank	0.000		0.707	0.780	0.793	0.745	0.786	0.726	0.718	#DIV/0!	#DIV/0!	0.314
Relative Cell Growth (%)			100.0	110.4	112.2	105.4	111.2	102.7	101.6	#DIV/0!	#DIV/0!	43.0
SD of Relative Cell Growth			4.9	11.1	11.3	9.0	6.3	11.0	5.4	#DIV/0!	#DIV/0!	6.8

Transformation Assay	Concentrations (ug/ml)					Negative Control	Positive Control
Glensa Staining	0.1	0.5	1	2	3		
No. of wells excluded	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96
No. of wells having foci	10	12	13	10	4	6	3
Transformation frequency*	0.104166667	0.125	0.135416667	0.104166667	0.041666667	0.0625	0.03125
SE	0.03117758	0.03375386	0.03492238	0.03117758	0.02039469	0.02470529	0.01775805
* No. of wells having foci/ No. of wells counted							

Remarks



Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

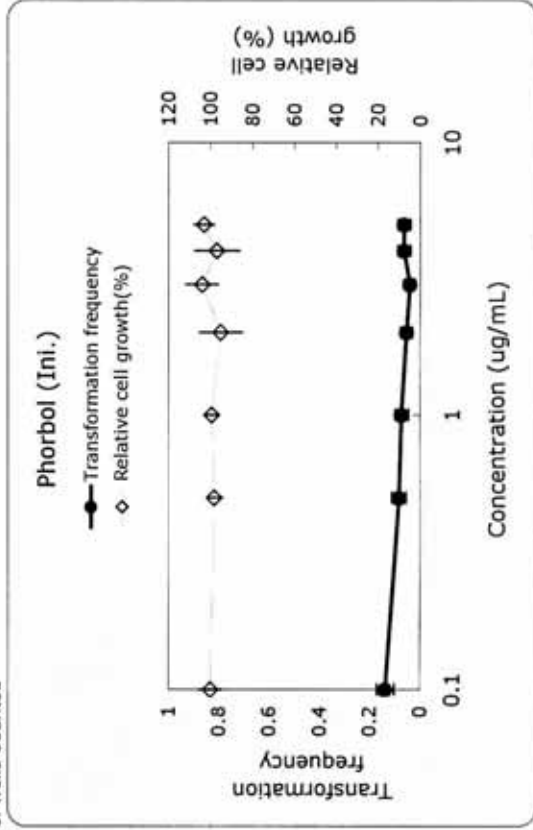
Data Sheet for Cell Transformation Assay on Initiation Assay

Chemical Code:	Phorbol
Institution:	Lab 3
Test Number	

Cell Growth Assay	5% Water		Concentrations (ug/ml)								Negative Control		Positive Control	
	Blank	0.1	0.5	1	2	3	4	5	5	0.1% DMSO	MCA (1ug/mL)			
Crystal Violet Staining														
OD _{540nm}														
1	0.125	1.211	1.190	1.273	0.976	1.243	0.969	1.301	1.301	1.261	0.687			
2	0.127	1.381	1.237	1.293	1.217	1.281	1.333	1.308	1.308	1.209	0.603			
3	0.126	1.197	1.299	1.284	1.243	1.327	1.206	1.345	1.345	1.279	0.606			
4	0.124	1.325	1.223	1.302	1.276	1.193	1.295	1.427	1.427	1.180	0.677			
5	0.119	1.338	1.274	1.201	1.314	1.401	1.239	1.280	1.280	1.186	0.722			
6	0.119	1.272	1.329	1.289	1.267	1.314	1.305	1.358	1.358	1.190	0.735			
7	0.116	1.248	1.284	1.321	1.362	1.429	1.348	1.257	1.257	1.308	0.671			
8	0.108	1.291	1.251	1.255	1.145	1.444	1.288	1.264	1.264	1.256	0.678			
Average	0.121	1.283	1.261	1.277	1.225	1.329	1.247	1.318	1.318	1.234	0.672			
SD	0.006	0.064	0.045	0.036	0.119	0.090	0.125	0.057	0.057	0.049	0.048			
Average - Blank	0.000	1.162	1.140	1.157	1.105	1.209	1.126	1.197	1.197	1.113	0.552			
Relative Cell Growth (%)		100.0	98.1	99.5	95.0	104.0	96.9	103.0	103.0	100.0	49.6			
SD of Relative Cell Growth		5.5	3.8	3.1	10.3	7.8	10.8	4.9	4.9	4.2	4.1			

Transformation Assay	Concentrations (ug/mL)										Negative Control		Positive Control	
	0.1	0.5	1	2	3	4	5	5	0	0	0	0	0	0
Giemsa Staining	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells excluded	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells counted	13	8	7	5	4	6	6	6	6	6	6	6	6	6
No. of wells having foci														
Transformation frequency*	0.135416667	0.083333333	0.072916667	0.052083333	0.041666667	0.0625	0.0625	0.0625	0.0625	0.0625	0.0625	0.0625	0.0625	0.0625
SE	0.03492238	0.02820847	0.02653610	0.02267769	0.02039469	0.02470529	0.02470529	0.02470529	0.02470529	0.02470529	0.02470529	0.02470529	0.02470529	0.02470529

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 15 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

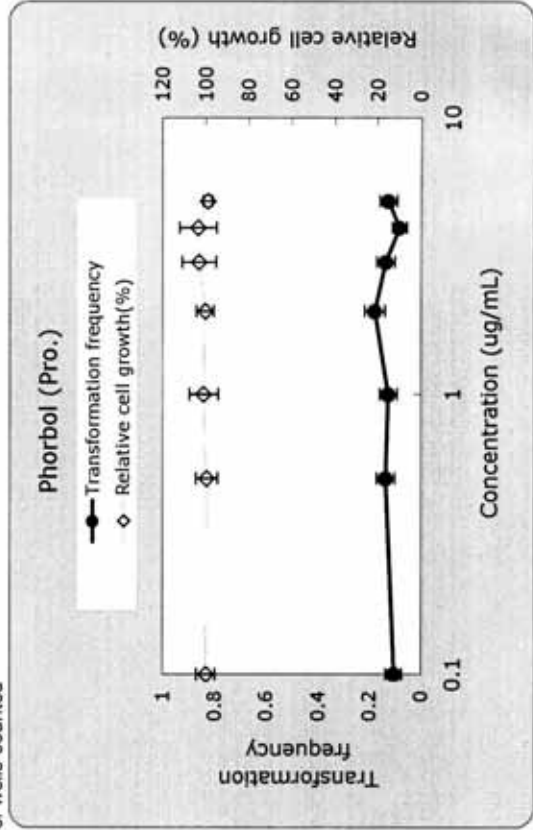
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Phorbol
Institution:	Lab 3
Test Number	

Cell Growth Assay	5% Water		Concentrations (ug/ml)								Negative Control		Positive Control	
	Blank	0.1	0.5	1	2	3	4	5	5	0.1% DMSO	TPA (50ng/mL)			
Crystal Violet Staining														
OD _{540nm}														
1	0.133	1.312	1.333	1.341	1.320	1.393	1.343	1.348	1.348	1.280	1.637			
2	0.139	1.429	1.466	1.491	1.337	1.433	1.323	1.399	1.399	1.499	1.749			
3	0.134	1.425	1.415	1.450	1.419	1.362	1.504	1.347	1.347	1.344	1.727			
4	0.134	1.438	1.375	1.318	1.361	1.357	1.605	1.364	1.364	1.333	1.804			
5	0.131	1.344	1.271	1.334	1.455	1.635	1.348	1.384	1.384	1.310	1.683			
6	0.134	1.358	1.418	1.458	1.397	1.369	1.425	1.376	1.376	1.280	1.751			
7	0.133	1.378	1.318	1.440	1.404	1.428	1.477	1.319	1.319	1.283	1.514			
8	0.126	1.307	1.348	1.262	1.320	1.319	1.307	1.349	1.349	1.225	1.642			
Average	0.133	1.374	1.368	1.387	1.377	1.412	1.417	1.361	1.361	#DIV/0!	#DIV/0!	1.688		
SD	0.004	0.052	0.063	0.083	0.050	0.098	0.105	0.025	0.025	#DIV/0!	#DIV/0!	0.091		
Average - Blank	0.000	1.241	1.235	1.254	1.244	1.279	1.284	1.228	1.228	#DIV/0!	#DIV/0!	1.555		
Relative Cell Growth (%)		100.0	99.5	101.0	100.2	103.1	103.4	98.9	98.9	100.0	131.1			
SD of Relative Cell Growth		4.2	5.1	6.7	4.0	7.9	8.5	2.0	2.0	6.6	7.3			

Transformation Assay	Concentrations (ug/ml)										Negative Control		Positive Control	
Glenssa Staining	0.1	0.5	1	2	3	4	5	5	0	0	0	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	10	13	12	17	13	8	12	12	12	11	11	11	81	
Transformation frequency*	0.104166667	0.135416667	0.125	0.177083333	0.135416667	0.083333333	0.125	0.125	0	0	0	0	0.84375	
SE	0.03117758	0.03492238	0.03375386	0.03896110	0.03492238	0.02820847	0.03375386	0.03375386	0.00000000	0.00000000	0.00000000	0.00000000	0.03705794	

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Data Sheet for Cell Transformation Assay on Initiation Assay

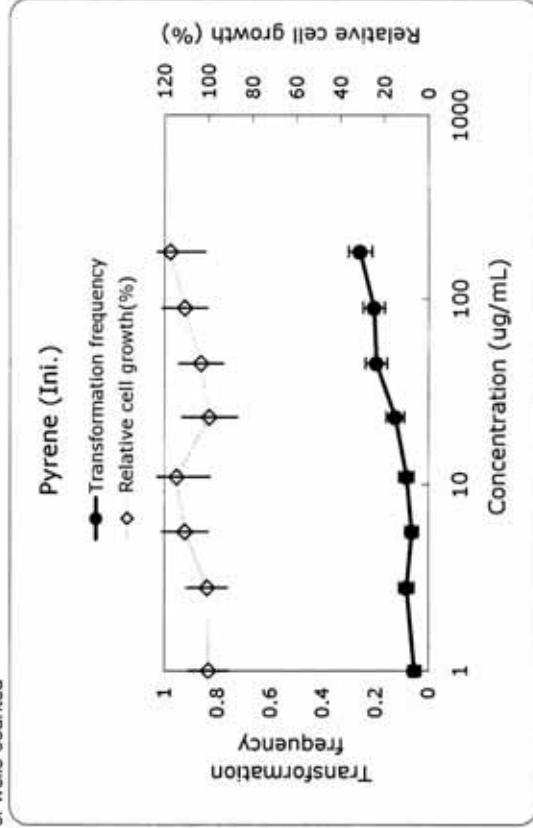
Chemical Code:	Pyrene
Institution:	Lab 2
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										
	Blank	0.5% DMSO	2.8	5.6	11	23	45	90	180	Negative Control	Positive Control
Crystal Violet Staining										0.1% DMSO	MCA (1ug/mL)
OD _{540nm}											
1	0.065	0.321	0.325	0.347	0.387	0.350	0.362	0.375	0.385	0.320	0.208
2	0.062	0.372	0.392	0.405	0.406	0.333	0.320	0.364	0.482	0.319	0.196
3	0.060	0.364	0.341	0.356	0.377	0.377	0.344	0.354	0.357	0.356	0.226
4	0.059	0.361	0.356	0.408	0.364	0.326	0.368	0.377	0.426	0.310	0.232
5	0.060	0.413	0.357	0.399	0.425	0.349	0.383	0.438	0.372	0.376	0.221
6	0.060	0.370	0.404	0.420	0.398	0.441	0.372	0.394	0.429	0.347	0.257
7	0.060	0.347	0.380	0.435	0.503	0.370	0.415	0.429	0.475	0.338	0.212
8	0.065	0.336	0.343	0.373	0.368	0.328	0.401	0.411	0.375	0.285	0.218
Average	0.061	0.361	0.362	0.393	0.404	0.359	0.371	0.393	0.413	#DIV/0!	0.221
SD	0.002	0.028	0.027	0.031	0.045	0.038	0.030	0.031	0.048	#DIV/0!	0.018
Average - Blank	0.000	0.299	0.301	0.332	0.342	0.298	0.309	0.331	0.351	#DIV/0!	0.160
Relative Cell Growth (%)		100.0	100.6	110.8	114.4	99.6	103.4	110.8	117.4	100.0	59.2
SD of Relative Cell Growth		9.2	9.1	10.4	15.1	12.7	10.1	10.3	16.0	9.6	6.1

Transformation Assay

Transformation Assay	Concentrations (ug/ml)										
Glenssa Staining	1	2.8	5.6	11	23	45	90	180	0	Negative Control	Positive Control
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	5	8	6	8	12	19	20	25	96	7	53
Transformation frequency*	0.052083333	0.083333333	0.0625	0.083333333	0.125	0.197916667	0.208333333	0.260416667	0	0.072916667	0.552083333
SE	0.02267769	0.02820847	0.02470529	0.02820847	0.03375396	0.04066449	0.04144908	0.04479116	0.00000000	0.02653610	0.05075342

* No. of wells having foci/ No. of wells counted



Remarks

--

Acceptance Criteria (1)

Negative Control < 15 and 40 < Positive Control

Pass

Acceptance Criteria (2)

Concurrent Cell Growth Assay

Pass

The Final Data

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Data Sheet for Cell Transformation Assay on Initiation Assay

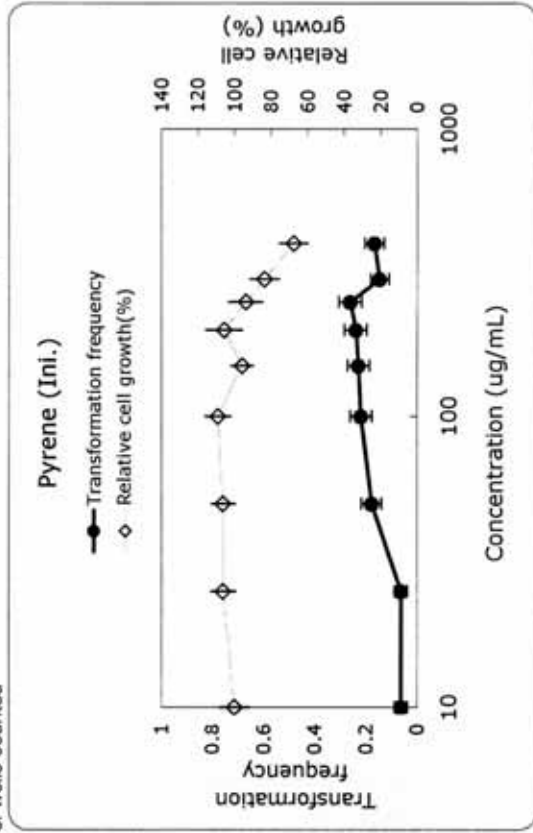
Chemical Code:	Pyrene
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
	Blank	0.5% DMSO	25	50	100	150	200	250	300	400			0.1% DMSO
Crystal Violet Staining													
OD _{490nm}													
1	0.077	0.442	0.445	0.485	0.503	0.464	0.507	0.442	0.414	0.370	0.554	0.377	
2	0.098	0.484	0.494	0.502	0.504	0.420	0.503	0.480	0.411	0.368	0.518	0.301	
3	0.079	0.489	0.500	0.447	0.478	0.444	0.476	0.435	0.370	0.343	0.529	0.324	
4	0.090	0.442	0.480	0.482	0.520	0.426	0.424	0.383	0.384	0.308	0.458	0.314	
5	0.085	0.470	0.444	0.471	0.455	0.407	0.438	0.411	0.364	0.296	0.525	0.275	
6	0.087	0.401	0.485	0.437	0.498	0.449	0.466	0.387	0.368	0.307	0.504	0.306	
7	0.085	0.458	0.447	0.468	0.463	0.464	0.442	0.439	0.446	0.348	0.497	0.330	
8	0.099	0.429	0.497	0.500	0.454	0.418	0.524	0.456	0.380	0.334	0.432	0.261	
Average	0.088	0.452	0.474	0.474	0.484	0.437	0.473	0.429	0.392	0.334	0.502	0.311	
SD	0.008	0.029	0.025	0.023	0.025	0.022	0.036	0.033	0.029	0.028	0.040	0.035	
Average - Blank	0.000	0.364	0.387	0.387	0.397	0.349	0.385	0.342	0.305	0.247	0.415	0.224	
Relative Cell Growth (%)		100.0	106.1	106.1	108.9	95.8	105.7	93.8	83.6	67.7	100.0	53.9	
SD of Relative Cell Growth		8.1	6.7	6.4	6.9	6.0	10.0	9.2	7.9	7.7	10.9	9.7	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Glensa Staining	10	25	50	100	150	200	250	300	400	0		
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	6	6	17	21	22	23	25	14	16	16	13	65
Transformation frequency*	0.0625	0.0625	0.177083333	0.21875	0.229166667	0.239583333	0.260416667	0.145833333	0.166666667	0	0.135416667	0.677083333
SE	0.02470529	0.02470529	0.03896110	0.04219232	0.04289635	0.04356307	0.04479116	0.03602169	0.03803629	0.00000000	0.03492238	0.04772332

* No. of wells having foci/ No. of wells counted

Remarks



Acceptance Criteria (1)
 Negative Control < 15 and 40 < Positive Control
 Pass

Acceptance Criteria (2)
 Concurrent Cell Growth Assay
 Pass

The Final Data

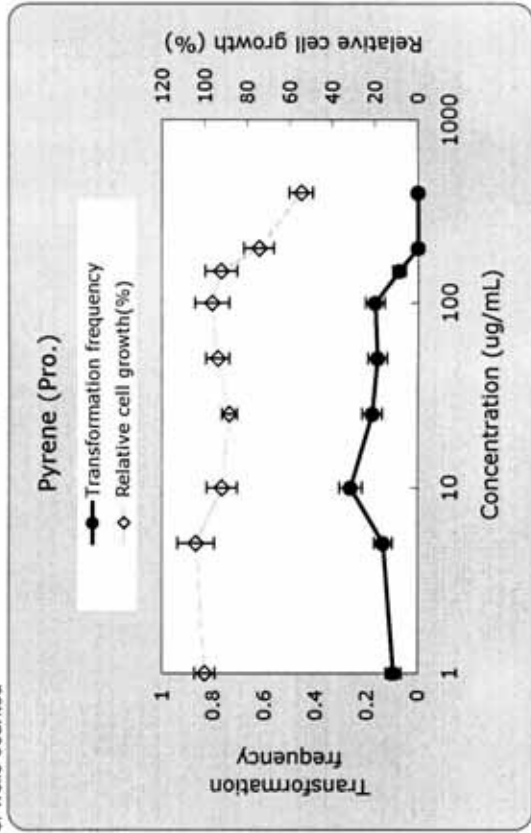
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Pyrene
Institution:	Lab 3
Test Number	

Cell Growth Assay	Crystal Violet Staining OD _{540nm}	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)
		Blank	1	5	10	25	50	100	150	200	400		
1	0.118	1.040	1.084	0.928	0.942	0.954	0.945	0.914	0.741	0.628	1.111	1.510	
2	0.125	1.138	1.222	1.052	1.009	1.070	1.037	0.979	0.780	0.622	1.255	1.646	
3	0.124	1.131	1.176	1.154	1.023	1.016	1.166	1.029	0.956	0.579	1.196	1.672	
4	0.125	1.109	1.216	1.034	1.004	1.097	1.072	1.042	0.886	0.688	1.357	1.516	
5	0.123	1.175	1.189	1.060	1.000	1.072	1.094	1.127	0.887	0.751	1.241	1.747	
6	0.122	1.145	1.277	1.066	1.033	1.118	1.171	1.112	0.854	0.679	1.340	1.880	
7	0.116	1.152	1.087	1.056	1.042	1.101	1.160	1.109	0.885	0.714	1.349	1.691	
8	0.110	1.054	1.023	0.953	0.961	1.025	1.016	1.013	0.908	0.663	1.246	1.581	
Average	0.120	1.118	1.159	1.038	1.002	1.057	1.083	1.041	0.862	0.666	#DIV/0!	1.655	
SD	0.005	0.048	0.086	0.070	0.035	0.055	0.081	0.074	0.070	0.055	#DIV/0!	0.123	
Average - Blank	0.000	0.998	1.039	0.918	0.881	0.936	0.962	0.920	0.742	0.545	#DIV/0!	1.535	
Relative Cell Growth (%)		100.0	104.1	92.0	88.3	93.8	96.5	92.2	74.4	54.6	#DIV/0!	134.5	
SD of Relative Cell Growth		4.8	8.6	7.0	3.5	5.5	8.2	7.4	7.0	5.5	#DIV/0!	12.4	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control
Transformation frequency*	0.09375	0.135416667	0.260416667	0.177083333	0.15625	0.166666667	0.072916667	#VALUE!	#VALUE!	#VALUE!	0	0.604166667
SE	0.02974911	0.03492238	0.04479116	0.03896110	0.03705794	0.03803629	0.02653610	#VALUE!	#VALUE!	#VALUE!	0.125	0.04991131
Giemsa Staining	1	5	10	25	50	100	150	200	400	0	0	0
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	9	13	25	17	15	16	7	Tox	Tox	Tox	12	58

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Progressing Data

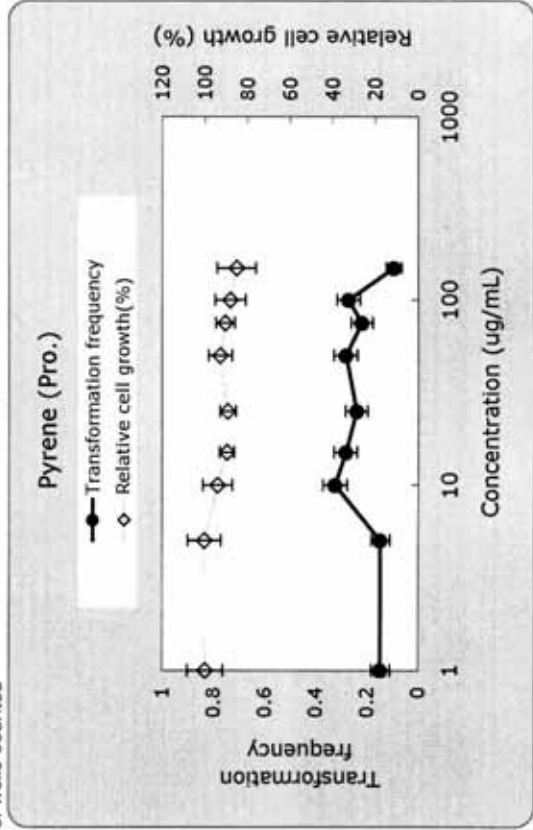
Data Sheet for Cell Transformation Assay on Promotion Assay

Chemical Code:	Pyrene
Institution:	Lab 3
Test Number	

Cell Growth Assay	Concentrations (ug/ml)										Negative Control 0.1% DMSO	Positive Control TPA (50ng/mL)		
	Blank	0.5% DMSO	1	5	10	15	25	50	75	100			150	
Crystal Violet Staining														
OD _{540nm}														
1	0.093	0.908	0.959	0.900	0.920	0.884	0.939	0.974	0.768	0.861		0.804	1.455	
2	0.091	0.992	0.979	0.966	0.961	0.946	1.060	0.886	0.969	0.936		0.991	1.643	
3	0.091	0.942	1.106	0.933	0.939	0.939	0.975	0.954	0.931	0.692		0.945	1.550	
4	0.098	1.091	1.060	1.085	0.908	0.953	0.896	0.938	0.900	0.913		0.958	1.493	
5	0.094	1.093	1.052	0.985	0.881	0.949	0.933	0.911	0.947	0.973		1.023	1.531	
6	0.092	1.112	1.075	0.975	0.964	0.888	0.927	0.949	0.954	0.903		1.001	1.575	
7	0.096	1.017	1.026	0.963	0.901	0.885	0.942	0.954	0.917	0.864		0.913	1.531	
8	0.088	0.963	0.884	0.877	0.882	0.876	0.912	0.847	0.854	0.880		0.962	1.529	
Average	0.093	1.015	1.018	0.961	0.920	0.915	0.948	0.927	0.905	0.878	#DIV/0!	0.950	1.538	
SD	0.003	0.077	0.073	0.063	0.033	0.034	0.051	0.042	0.066	0.084	#DIV/0!	0.068	0.056	
Average - Blank	0.000	0.922	0.925	0.868	0.827	0.822	0.855	0.834	0.812	0.785	#DIV/0!	0.857	1.446	
Relative Cell Growth (%)		100.0	100.3	94.1	89.7	89.2	92.8	90.4	88.1	85.1	#DIV/0!	100.0	168.7	
SD of Relative Cell Growth		8.3	7.9	6.8	3.5	3.7	5.5	4.6	7.2	9.1	#DIV/0!	7.4	6.0	

Transformation Assay	Concentrations (ug/ml)										Negative Control	Positive Control	
Glenssa Staining	1	5	10	15	25	50	75	100	150	0			
No. of wells excluded	0	0	0	0	0	0	0	0	0	0	0	0	0
No. of wells counted	96	96	96	96	96	96	96	96	96	96	96	96	96
No. of wells having foci	14	14	31	27	23	27	21	26	9	9	18	18	71
Transformation frequency*	0.145833333	0.145833333	0.322916667	0.28125	0.239583333	0.28125	0.21875	0.270833333	0.09375	0	0.1875	0.739583333	
SE	0.03602169	0.03602169	0.04772332	0.04588805	0.04588805	0.04588805	0.04219232	0.04535538	0.02974911	0.00000000	0.03983609	0.04479116	

* No. of wells having foci/ No. of wells counted



Remarks

Acceptance Criteria (1)
Negative Control < 20 and 40 < Positive Control
Pass

Acceptance Criteria (2)
Concurrent Cell Growth Assay
Pass

The Final Data

Annex 12

Recommended Protocol for the Bhas 42 Cell Transformation Assay (2012.7.27)

1. Introduction

Bhas 42 cells were established from BALB/c 3T3 A31-1-1 cells (BALB/c mouse embryo cells) by transfection of *v-Ha-ras* oncogene [1]. The cells are used as a sensitive and stable cell line for the cell transformation assay (CTA), the endpoint of which is focus formation of morphologically transformed cells [2]. The Bhas 42 CTA consists of an initiation assay and a promotion assay [3]. These assays can detect genotoxic and non-genotoxic carcinogens, respectively [4-8]. In the initiation assay, the cells are treated with chemicals in the beginning of growth phase and in the promotion assay the treatment is started at subconfluence of cell growth. Cell growth assay is carried out prior to the transformation assay for the determination of appropriate test concentrations, and concurrently with the transformation assay to confirm the toxic effects of chemical treatment (Fig. 1).

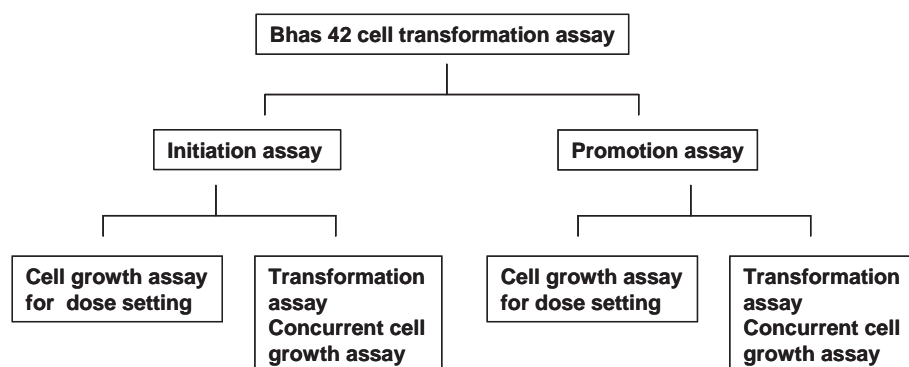


Fig. 1. Schematic of the Bhas 42 cell transformation assay.

There are two methods, a 6-well method and a 96-well method followed in the Bhas 42 CTA. The schematics of both methods are the same since their procedures are basically the same except for some differences associated with the use of 6-well micro-plates vs. 96-well micro-plates. Furthermore, similar results are obtained with both methods.

2. Materials and preparation

The assay itself and all the preliminary steps (i.e. cell stock preparation, reagent and treatment solution preparations) should be performed under sterile conditions.

2.1 Equipment

49 Micro-plates with 6 wells and 96 wells are used for the 6-well method and 96-well method, respectively.
50 Multichannel micropipettes are used for the 96-well method. In addition, general cell culture equipment is
51 necessary.

52
53

54 **2.2 Culture media**

55

56 Eagle's minimal essential medium (with 2.2 g/L NaHCO₃ and 0.292 g/L L-glutamine) supplemented with
57 10% fetal bovine serum (FBS) and antibiotics (100 units/mL penicillin G sodium and 100 µg/mL
58 streptomycin sulfate) (M10F) is used for the proliferation of Bhas 42 cells, cell cryo-preservation and the
59 first culture after cell thawing. Dulbecco's modified Eagle's medium/F12 (with 1.2 g/L NaHCO₃)
60 supplemented with 5% FBS and antibiotics (100 units/mL penicillin G sodium and 100 µg/mL
61 streptomycin sulfate) (DF5F) is used for second culture after cell thawing, cell growth assays and
62 transformation assays. (Other antibiotics could be used after examination for applicability.)

63
64

65 **2.3 Culture**

66

67 The cultures are maintained in a humidified incubator with a 5% CO₂ atmosphere at 37 °C.

68
69

70 **2.4 Reagents and solutions**

71

72 The following reagents and solutions are used:

73

- 74 ▪ 0.25% trypsin: Used for passage
- 75 ▪ 0.02% EDTA-PBS(-): Used for passage
- 76 ▪ Dimethylsulfoxide (DMSO): Used for cell cryo-preservation and as solvent for test chemicals
- 77 ▪ Formalin (37% formaldehyde): Used for fixing cells.
- 78 ▪ Methanol: Used for fixing cells.
- 79 ▪ Crystal violet (CV) solution: Used for staining cells in cell growth assays. In 50 mL of ethanol 1
80 g of crystal violet is dissolved, and the total volume is adjusted to 1 L with distilled water.
- 81 ▪ Dye extraction solution (0.02 mol/L HCl and 50% ethanol): Used for extracting CV in cell
82 growth assays. 480 mL distilled water + 500 mL ethanol + 20 mL 1 M HCl.
- 83 ▪ 5% Giemsa solution: Used for staining cells in transformation assays

84

85

86 **2.5 Chemicals**

87

88 *2.5.1 Test chemicals and solvent/vehicle*

89

90 Test chemicals are dissolved or suspended in an appropriate solvent or vehicle and diluted with the
91 solvent/vehicle to each individual concentration before being added to cell cultures or culture media so that
92 all chemical treatment media contain an equal concentration of the solvent/vehicle. Distilled water, DMSO,
93 acetone, and ethanol can be used to dissolve chemicals, and the final solvent/vehicle concentrations in the
94 medium should not exceed 5%, 0.5%, 0.5% and 0.1 %, respectively. Although the concentration of DMSO
95 can be as high as 0.5%, it is recommended that it does not exceed 0.1%. The absence of cytotoxicity should
96 dictate the highest allowable concentration of solvent/vehicle.

97

98 Test-chemical solutions are prepared before use as a general rule. To reduce labor, stock and/or working
99 solutions may be preserved in aliquots at -20 °C for less than 10 days and thawed before use, but must not

100 be re-frozen. The suspensions of test chemicals must be prepared before use and cannot be stored for the
101 future use.

102 103 *2.5.2 Test chemical concentrations*

104
105 The highest concentration of test chemical to be tested is 5 mg/mL or 10 mM, whichever is the lowest
106 [9,10]. In the case of chemicals difficult to dissolve, the highest concentration(s) may be one or two
107 precipitating dose levels which exceed the highest soluble concentration.

108
109 Five to nine concentrations should be tested and these are determined according to the results of cell
110 growth assay.

111 112 *2.5.3 Negative and Positive controls*

113
114 The solvent/vehicle for a test chemical is used as the negative control. A known tumor-initiator, 3-
115 methylcholanthrene (MCA, final concentration of 1 µg/mL), and a known tumor-promoter, 12-*O*-
116 tetradecanoylphorbol-13- acetate (TPA, final concentration of 50 ng/mL), are used for the positive
117 controls in the initiation assay and promotion assay, respectively. MCA and TPA are dissolved in
118 DMSO. When the solvent/vehicle for the test chemical is not DMSO, DMSO is still necessary as
119 the negative control for MCA or TPA.

120
121 The stock solutions of MCA and TPA in DMSO can be stored in aliquots at -20 °C for at least two
122 years, provided they are not thawed.

123 124 125 **2.6 Stock cells**

126
127 The cells should be obtained from a reliable source. Health Science Research Resources Bank (HSRRB,
128 Osaka, Japan) [http://www.jhsf.or.jp/index_b.html]. supplies frozen Bhas 42 cells. The cells are expanded
129 by growing them up to 50-70% confluence in culture plates with M10F so as to avoid the increase of
130 transformed variants. It is important to maintain a sub-confluent state in all the area of the cultures since
131 the transformed cells, which lose the characteristic of cell-to-cell contact inhibition, preferentially multiply
132 at confluence. The cells collected from cell populations that do not exceed 50-70 % confluence are washed
133 once with 0.02% EDTA-PBS(-), trypsinized using 0.25% trypsin and subcultured. After two passages, the
134 expanded cells are suspended at a cell density of 5×10^5 cells/mL in cold fresh M10F containing 5%
135 DMSO and cryo-preserved in 0.5 mL aliquots to make a master stock in liquid nitrogen. Using one tube of
136 the master stock, working cell stock for transformation assays are prepared using the same procedures and
137 stored in liquid nitrogen until required for experiments. Generally, large numbers of tubes of the master
138 and working stock cells are prepared so as not to be depleted prematurely.

139
140 The transformation assay must be started using the cryo-preserved stock cells. Conventionally, the cells at
141 higher passages can be used for the cell growth assay for dose setting, but should not be used for the
142 transformation assays themselves in order to avoid excessive spontaneous transformation.

143 144 145 **2.7 Checking of the Bhas 42 cells**

146
147 In order to determine the utility of the cryo-preserved cells, every batch of working stock cells must be
148 confirmed for (a) adequate plating efficiency ($\geq 50\%$), (b) low background of spontaneous transformation
149 and (c) their ability to be transformed by archetypical chemical carcinogens. To be considered suitable for

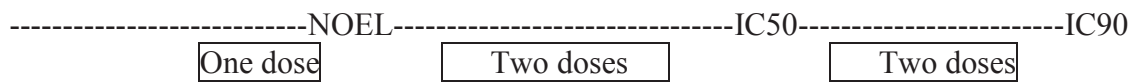
201
202
203

$$\text{relative cell growth} = [(At - Ab)/(Ac - Ab)] \times 100$$

204 where At is the absorbance of CV extract from a well with the chemically-treated cells, Ac is the
205 absorbance of CV extract from a well with the solvent/vehicle-treated cells and Ab is the absorbance
206 of CV extract from a well with the medium only.

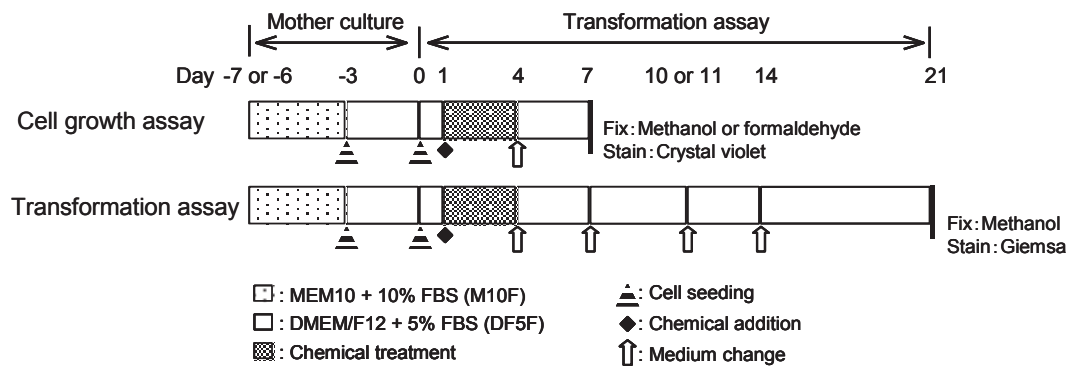
207
208 *3.1.2 Dose setting for the transformation assay*

209
210 Five to nine concentrations are set up based on the results of cell growth assays. These concentrations
211 cover a range from the highest cytotoxicity (less than 20% survival compared to the negative control) to
212 little or no toxicity. Ideally, one concentration below the no observed effect level (NOEL), two
213 concentrations between the NOEL and the 50% inhibitory concentration (IC50) and two concentrations
214 between the IC50 and the IC90 are selected. The ratio between neighboring concentrations should be
215 less than square root 10 (Fig. 3).



218
219
220 Fig. 3. Dose setting for the transformation assay in the initiation assay

221
222
223 *3.1.3 Transformation assay*



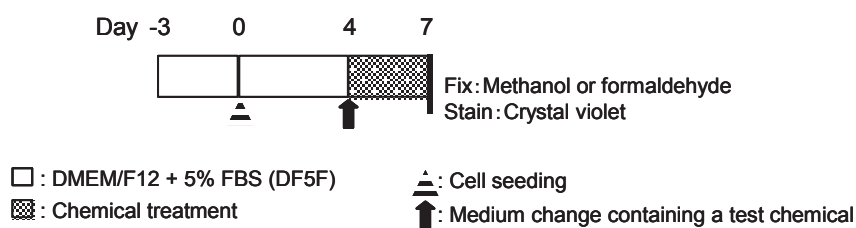
238
239
240 Fig. 4. Schematic protocol of transformation assay in the initiation assay

241 The frozen stock cells are rapidly thawed, suspended in M10F and cultured in Φ 100-mm culture plates at
242 a volume of 10 mL medium. When the cells reached about 70% confluence, they are trypsinized,
243 suspended in DF5F at an appropriate density (7,000 to 10,000 cells /mL is suggested) and cultured in Φ
244 100-mm culture plates (Day -3). The cells at about 70% confluence are trypsinized, and suspended in
245 DF5F at 2,000 cells/mL. The cell suspension is seeded into each well at a volume of 2 mL (4,000
246 cells/well) for the transformation assay and the concurrent cell growth assay (Day 0). Nine wells are
247 prepared for each dose (one plate of 6 wells for the transformation assay and 3 wells for the concurrent cell
248 growth assay). 20-24 hours after seeding, a chemical solution is added to each well without replacement of
249 medium. The cells are exposed to the chemical for three days (Day 1-4). The medium is changed with fresh
250 DF5F on Day 4, 7, 10 (or 11) and 14. On Day 21 the cells are fixed with methanol, and stained with 5%

251 Giemsa solution (Fig. 4). The positive control (1 $\mu\text{g}/\text{mL}$ MCA) and the negative control are included in
 252 the transformation assay for each chemical (when the solvent/vehicle of test chemical is not DMSO,
 253 DMSO is still necessary as the negative control for MCA). The cell growth assay is carried out in parallel
 254 with the transformation assay.
 255
 256

257 3.2 Promotion assay

258 3.2.1 Cell growth assay



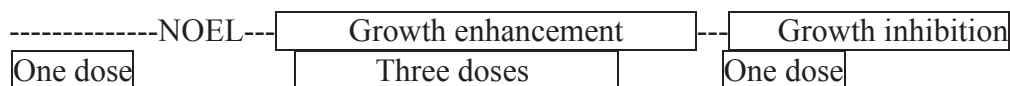
272 Fig. 5. Schematic protocol of cell growth assay in the promotion assay

273 The cells at $\leq 70\%$ confluence in DF5F are trypsinized, and 14000 cells are seeded into a well in 2 mL of
 274 medium (Day 0). Three wells are prepared for each treatment condition. Wells containing medium alone
 275 are also prepared for the blank control in photometry (The blank control can be shared with the cell growth
 276 assay in the initiation assay which is performed simultaneously). On Day 4, chemical treatment is started
 277 by exchanging existing medium with fresh medium containing a test chemical solution or solvent/vehicle
 278 alone. On Day 7, the cultures are fixed with methanol or 3.7% formaldehyde for 30 min, washed and dried.
 279 The cells are stained with 1.5 mL of crystal violet solution for 15 min, rinsed well with water and dried
 280 (Fig. 5).

281 The following procedures are the same as those in the cell growth assay in the initiation assay (section
 282 3.1.1).

283 3.2.2 Dose setting for the transformation assay

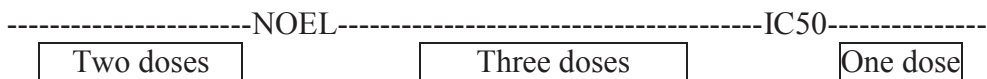
284
 285 For the chemicals that exhibit marked growth enhancement, concentrations are selected to cover the range
 286 from little effect to enhancement on cell growth. Ideally, one concentration below the NOEL, three
 287 concentrations in the range of growth enhancement, and one concentration in the range of weak growth
 288 inhibition are assessed (Fig. 6).
 289
 290
 291
 292



300 Fig. 6. Dose setting for transformation assay in the promotion assay of chemicals
 301 that exhibit marked growth enhancement.

302 For the chemicals that inhibit growth, concentrations are selected to cover the range from the NOEL to the
 303 level below 50% inhibition on cell growth. Ideally, two concentrations below the NOEL, three

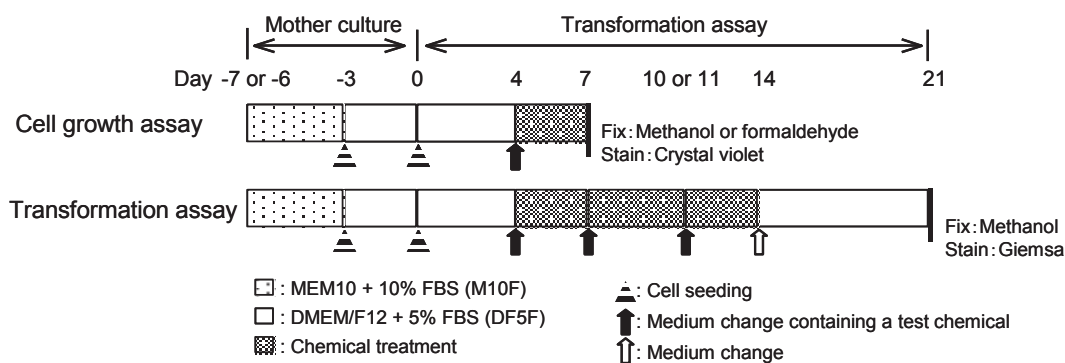
302 concentrations between the NOEL and the IC50 and one concentration above the IC50 are evaluated (Fig.
 303 7).



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 308
 309 Fig. 7. Dose setting for transformation assay in the promotion assay of chemicals that inhibit growth

310
 311
 312 Whether a chemical shows growth enhancement or inhibition, other experimental conditions, such as the
 313 number of concentrations set up and the ratio between neighboring concentrations, are the same as
 314 described in the initiation assay.

315 3.2.3 Transformation assay



327
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 330
 331 Fig. 8. Schematic protocol of transformation assay in the promotion assay.

332
 333
 334 The transformation assay is carried out in the same manner as in the initiation assay except for the
 335 following (Fig. 8):

- 336
- 337 ▪ The cells are suspended in medium at 7,000 cells/mL, and 14,000 cells are plated into each well (Day 0)
- 338
- 339 ▪ The cells are exposed to a test chemical for 10 days from Day 4 to Day 14.
- 340
- 341 ▪ The chemical treatment is carried out by exchanging existing medium with fresh medium containing a chemical solution or solvent/vehicle alone on Day 4, 7 and 10 (or 11).
- 342
- 343 ▪ On day 14 the medium is changed with the normal medium containing neither solvent/vehicle nor test chemical..
- 344
- 345 ▪ TPA (50 ng/mL) is used for the positive control instead of MCA.

346 347 348 349 4. Evaluation of the results in the 6-well method

350 351 352 4.1 Record of transformation frequency

353
354 Transformed foci are scored using a stereomicroscope. Transformed foci are characterized by the following
355 morphological properties: (a) more than 100 cells, (b) spindle-shaped cells differing in appearance from the
356 contact-inhibited monolayer cells, (c) deep basophilic staining, (d) random orientation of cells at the edge
357 of foci (criss-cross), (e) dense multilayering of cells (piling up) and (f) invasive growth into the
358 surrounding monolayer of contact-inhibited cells. It should be noted that there are transformed foci that do
359 not exhibit some of these characteristics.

360
361 The number of transformed foci in each well are recorded for every concentration.
362

363 364 **4.2 Statistical analysis** 365

366 For the increases in the number of the transformed foci produced by a test chemical, a statistical analysis is
367 performed by multiple comparison using the one-sided Dunnett test ($p < 0.05$). For the positive controls, the
368 statistical significance is evaluated by one-sided t-test or Aspin-Welch test ($p < 0.05$) depending on the
369 result of F-test for homoscedasticity (homogeneity of variance).
370

371 372 **4.3 Assay acceptance criteria** 373

374 The following criteria must be fulfilled for a given assay to be considered valid. The initiation or
375 promotion assay is repeated independently, as needed, to satisfy the assay acceptance criteria.
376

377 *4.3.1 Concurrent cell growth assay* 378

- 379 • When contamination or technical problems are observed, two undamaged wells are necessary at the
380 minimum for each concentration for cell growth assessment.

381 382 *4.3.2 Transformation assay* 383

384 *4.3.2.1 Initiation assay and promotion assay*

- 385 • If a given chemical concentration results in excessive cell death and/or cells fail to reach confluence at
386 the end of transformation assay because of chemical toxicity, the concentration is not valid for
387 transformation assessment and is excluded from focus-counting, statistical analysis and judgment, and
388 “toxicity” is recorded in the data sheet.
- 389 • When contamination or technical problems are observed, if, for a given concentration, the number of
390 damaged wells is two or more, the concentration is not considered valid for transformation assessment
391 and is excluded from focus-counting, statistical analysis and judgment. In such cases,
392 “contamination”, “accident”, “technical error”, etc. are recorded in the data sheet.
- 393 • In the positive control, there must be a statistically significant increase in the number of transformed
394 foci per well compared to the corresponding negative control (one-sided t-test or Aspin-Welch test,
395 $p < 0.05$).

396 397 *4.3.2.2 Initiation assay*

- 398 • In the negative control, the number of transformed foci must be ten or less per well.
- 399 • If there is no statistically significant increase in the number of transformed foci at any dose (in case of
400 negative results), four valid test chemical concentrations are necessary, at a minimum, to
401 accept the transformation assay for evaluating a chemical. Those concentrations should
402 include at least one concentration near the NOEL and three concentrations in the range
403 between the NOEL and the IC_{90} in the concurrent cell growth assay.

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4.3.2.3 Promotion assay

- In the negative control, the number of transformed foci must be twelve or less per well.
- If there is no statistically significant increase in the number of transformed foci at any dose (in case of negative results), four valid test chemical concentrations are necessary, at a minimum, to accept the transformation assay for evaluating a chemical. Those concentrations should include at least one concentration near the NOEL and two concentrations in the range of growth enhancement when the chemical enhances cell growth (increases cell density in the cell growth assay). When the chemical does not induce growth enhancement but induces cytotoxicity, the cytotoxicity observed in the concurrent cell growth assay may not be similar to that observed in the transformation assay, since the durations of chemical exposure to the cells are different between the cell growth assay (3 days) and the transformation assay (10 days). Consequently, chemical toxicity is sometimes accumulated over the 10 day duration of the transformation promotion assay and the valid plates may be lost because of chemical toxicity. In such cases, four valid plates are also necessary in the concentration range where cells are not killed and are confluent at the end of the transformation assay. If excessive toxicity is encountered, it may be necessary to repeat the assay in a lower concentration range.

4.4 Assessment criteria

The assay results are judged as follows:

- The results in the initiation and promotion assays are judged positive if there exist two or more sequential doses that induce statistically significant increases in the number of transformed foci per well relative to the corresponding vehicle control (multiple comparison using one-sided Dunnett test, $p < 0.05$).
- The results in the initiation and promotion assays are judged negative if there is no dose showing a statistically significant increase in the number of transformed foci per well.
- If the statistically significant increase is at only one or non-sequential doses, the assay result is regarded as equivocal, and then the initiation or promotion assay is repeated together with the concomitant cell growth assay, including the dose that caused the statistically significant increase of transformation frequency in the first assay. The chemical is judged to be positive if a statistically significant increase in the number of transformed foci per well is induced at one or more concentrations in the second assay; otherwise, the chemical is judged to be negative.
- A test chemical is positive in the Bhas 42 CTA, if it is positive in either the initiation or promotion assay.

5. Procedures of 96-well method

5.1 Medium change

The final volume of medium is 0.1 mL per well. The medium is removed by inverting the plates and shaking out the medium over paper towels or disposable diapers three times to drain off the medium

454 completely. The medium is replaced by gently dispensing it down the walls of the wells over the cells
455 using micro-tips attached to a multichannel micropipette.

456

457

458 **5.2 Initiation assay**

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460 *5.2.1 Cell growth assay*

461

462 The cell growth assay is carried out in the same manner as in the initiation assay of the 6-well method
463 except for the following (see section 3.1.1):

464

- 465 ▪ Into each well, 200 cells are seeded with 0.05 mL of DF5F (Day 0).

- 466 ▪ For each treatment condition, 8 wells are prepared.

- 467 ▪ The cultures are treated by 0.05 mL of medium containing a test chemical solution or
468 solvent/vehicle alone at two times the final concentration added to each well so that the final
469 volume of medium is 0.1 mL.

- 470 ▪ The volumes of crystal violet solution and dye extraction solution are 0.1 mL/well.

471

472 *5.2.2 Dose setting for the transformation assay*

473

474 The concentrations to be tested in the transformation assay are determined in the same way that they are in
475 the initiation assay of 6-well method (see section 3.1.2).

476

477 *5.2.3 Transformation assay*

478

479 The transformation assay is carried out in the same manner as in the initiation assay of the 6-well method
480 except for the following:

481

- 482 ▪ Into each well, 200 cells are seeded with 0.05 mL of DF5F (Day 0).

- 483 ▪ For each treatment condition, one 96-well plate is prepared for the transformation assay and 8
484 wells are prepared for the cell growth assay, respectively.

- 485 ▪ The cultures are treated by the addition of 0.05 mL of medium containing a chemical solution or
486 solvent/vehicle alone at two times the final concentration added to each well so that the final
487 volume of medium is 0.1 mL.

488

489

490 **5.3. Promotion assay**

491

492 *5.3.1 Cell growth assay*

493

494 A cell growth assay is carried out in the same manner as in the promotion assay of the 6-well method
495 except for the following (see section 3.2.1):

496

- 497 ▪ Into each well, 400 cells are seeded with 0.1 mL of DF5F (Day 0).

- 498 ▪ For each treatment condition, 8 wells are prepared.

- 499 ▪ The volumes of crystal violet solution and dye extraction solution are 0.1 mL/well.

500

501 *5.3.2 Dose setting for the transformation assay*

502

503 The concentrations to be tested in the transformation assay are determined in the same was that they are in
504 the promotion assay of 6-well method (See section 3.2.2).

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5.3.3 Transformation assay

The transformation assay is carried out in the same manner as in the promotion assay of the 6-well method except for the following (see section 3.2.3):

- Into each well, 400 cells are seeded with 0.1 mL of DF5F (Day 0).
- For each treatment condition, one 96-well plate is prepared for the transformation assay and 8 wells are prepared for the cell growth assay, respectively.

6. Evaluation of the results in the 96-well method

6.1 Record of transformation frequency

Transformed foci are judged using the same criteria as in the 6-well method.

The number of wells having transformed foci versus the number of wells observed is recorded for every concentration: a well having one focus is counted as one and a well having two or more foci is also counted as one.

6.2 Statistical analysis

For the increases in the proportion of wells with transformed foci in the plate treated with a test chemical, a statistical analysis is performed by chi-square test with Bonferroni adjustment (p -value <0.05 , upper-sided) [11]. For multiplicity, the number of concentrations that satisfy the assay acceptance criteria is considered by this methodology. For the positive controls, the statistical significance is evaluated by one-sided chi-square test ($p<0.05$, upper-sided).

6.3 Assay acceptance criteria

The following criteria must be fulfilled for a given assay to be considered valid. The initiation or promotion assay is repeated independently, as needed, to satisfy the assay acceptance criteria.

6.3.1 Concurrent cell growth assay

- When contamination or technical problems are observed, four undamaged wells are necessary at the minimum for each concentration for cell growth assessment.

6.3.2 Transformation assay

6.3.2.1 Initiation assay and promotion assay

- If a given chemical concentration results in excessive cell death and/or cells fail to reach confluence at the end of transformation assay because of chemical toxicity, the concentration is not valid for transformation assessment and excluded from focus-counting, statistical analysis and judgment, and “toxicity” is recorded in the data sheet.

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- When contamination or technical problems are observed, if, for a given concentration, the number of damaged wells is ≥ 7 , the concentration is not considered valid for transformation assessment and is excluded from focus-counting, statistical analysis and judgment. In such cases, “contamination”, “accident”, “technical error”, etc. are recorded in the data sheet.
 - In the positive control, there must be a statistically significant increase in the proportion of wells having transformed foci compared to the corresponding negative control (one-sided chi-square test, $p < 0.05$, upper-sided).

564 **6.3.2.2 Initiation assay**

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- In the negative control, the number of wells having transformed foci must be 15 wells/plate or less (if there exist damaged wells, $\leq 15.625\%$ of undamaged wells).
 - If there is no statistically significant increase in the proportion of wells having transformed foci at any dose (in case of negative results), four valid test chemical concentrations are necessary, at a minimum, to accept the transformation assay for evaluating a chemical. Those concentrations should include at least one concentration near the NOEL and three concentrations in the range between the NOEL and the IC_{90} in the concurrent cell growth assay.

573 **6.3.2.3 Promotion assay**

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- In the negative control, the number of wells having transformed foci must be 20 wells/plate or less (if there exist damaged wells, $\leq 20.833\%$ of undamaged wells).
 - If there is no statistically significant increase in the proportion of wells having transformed foci at any dose (in case of negative results), four valid test chemical concentrations are necessary, at a minimum, to accept the transformation assay for evaluating a chemical. Those concentrations should include at least one concentration near the NOEL and two concentrations in the range of growth enhancement when the chemical enhances cell growth (increases cell density in the cell growth assay). When the chemical does not induce growth enhancement but induces cytotoxicity, the cytotoxicity observed in the concurrent cell growth assay may not be similar to that observed in the transformation assay, since the durations of chemical exposure to the cells are different between the cell growth assay (3 days) and the transformation assay (10 days). Consequently, chemical toxicity is sometimes accumulated over the 10 day duration of the transformation promotion assay and the valid plates may be lost because of chemical toxicity. In such cases, four valid plates are also necessary in the concentration range where cells are not killed and are confluent at the end of the transformation assay. If excessive toxicity is encountered, it may be necessary to repeat the assay in a lower concentration range.

591

592 **6.4 Assessment criteria**

593

594 The assessment criteria are the same as those of 6-well method except that a statistical analysis is

595 performed for the proportion of wells having transformed foci (number of wells having transformed

596 foci/number of wells observed) using chi-square test with Bonferroni adjustment, $p < 0.05$, upper-sided.

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601 **7. References**


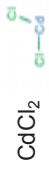
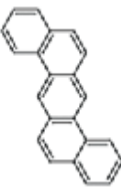
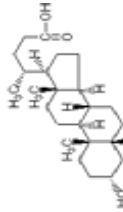
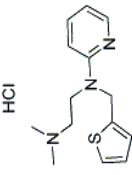


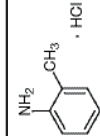
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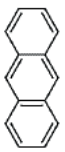
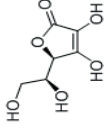
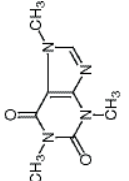
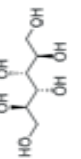


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Annex 14

Table 58.

Chemical properties and classes of 12 coded test chemicals and positive controls for the 6-well method validation study^a

#	Chemical Name	M.W.	Chemical structure	CAS	IARC [as of May 2010]	Physical appearance	Water solubility ^b	Solubility (Other solvents)	Vapor pressure (Volatility)	Chemical class
1	2-Acetylaminofluorene	223		53-96-3	Not listed (C ^c)	Solid	144 mg/L at 25 °C (est) <very poor>	Sol in ether, acetic acid	(Possibly Non-volatile)	Aromatic amine; requires metabolic activation
2	Cadmium chloride	183		10108-64-2	1 (as Cadmium compounds) (C)	Solid	140 g/100 mL at 20°C <very good>	Soluble in acetone	10 mm Hg at 656 °C (Non-volatile)	Inorganic cadmium compound
3	Dibenz[<i>a,h</i>]anthracene	278		53-70-3	2A (C)	Solid	0.0005 mg/L at 27°C <very poor (almost insoluble)>	Slightly soluble in ethanol, soluble in acetone and benzene.	1.1X10 ⁻¹⁰ mm Hg (est) (Possibly Non-volatile)	Polycyclic aromatic hydrocarbon; requires metabolic activation
4	Lithocholic acid	377		434-13-9	Not listed (TP ^d)	Solid	0.38 mg/L at 20 °C <very poor (almost insoluble)>	Freely soluble in hot alcohol	(Unknown, Possibly Non-volatile)	Steroid derivative (Bile acid)
5	Methapyrilene hydrochloride	298		135-23-9	Not listed (TP)	Solid	1 gm dissolves in 0.5 mL water <very good>	1 gm dissolves in 5 mL alcohol, in 3 mL chloroform	(Unknown, Possibly Non-volatile)	Heterocyclic amine; Anti-histamine drug
6	Mezerein	655		34807-41-5	Not listed (TP)	Solid	Not soluble <very poor (almost insoluble)>	Soluble in DMSO or ethanol	(Unknown, Possibly Non-volatile)	Diterpene compound Protein kinase activator (Non-phorbol type)
7	Sodium arsenite	130		7784-46-5	1 (as Arsenic compounds) (C)	Solid	Freely soluble <very good>	Slightly soluble in alcohol	0% volatiles by volume at 21 °C (Possibly Non-volatile)	Inorganic arsenite compound
8	<i>o</i> -Toluidine hydrochloride	144		636-21-5	1 (as <i>o</i> -Toluidine) (C)	Solid	Very soluble <very good>	Soluble in alcohol; insoluble in ether, benzene	as VOC (Possibly Volatile)	Aromatic amine

#	Chemical Name	M.W.	Chemical structure	CAS	IARC [as of May 2010]	Physical appearance	Water solubility ^b	Solubility (Other solvents)	Vapor pressure (Volatility)	Chemical class
9	Anthracene	178		120-12-7	3 (NC ^e)	Solid	0.04-1.3 mg/L at 25°C <very poor>	1 gm dissolves in 67 mL absolute alcohol, 70 mL methanol, 62 mL benzene, 85 mL chloroform	2.67X10 ⁻⁶ mm Hg at 25°C (eat) (Non-volatile)	Polycyclic aromatic hydrocarbon; requires metabolic activation
10	L-Ascorbic acid	176		50-81-7	Not listed (NC)	Solid	0.33 g/mL <good>	0.02 mg/mL in ethanol; insoluble in ether, chloroform, benzene	7.9179 Pa @ 465.15 deg K= 0.06 mm Hg at 192 °C (Non-volatile)	Lactone structure; Antioxidant
11	Caffeine	194		58-08-2	3 (NC)	Solid	2.16X10 ⁻⁴ mg/L at 25°C <moderate>	1 gm dissolves in 66 mL alcohol, 50 mL acetone, 5.5 mL chloroform, 530 mL ether, 100 mL benzene	7.3X10 ⁻⁹ mm Hg at 25°C (est) (Non-volatile)	Purine alkaloid
12	D-Mannitol	182		69-65-8	Not listed (NC)	Solid	2.16X10 ⁻⁵ mg/L at 25°C <good>	Insoluble in ether	(Non-volatile)	Sugar alcohol
	3-Methylcholanthrene	268		56-49-5	Not listed (C)	Solid	2.9X10 ⁻³ mg/L at 25°C <very poor>	Soluble in xylene, toluene, and benzene	4.3X10 ⁻⁸ mm Hg at 25°C (Non-volatile)	Polycyclic aromatic hydrocarbon; requires metabolic activation
	12-O-tetradecanoylphorbol-13-acetate (TPA)	617		16561-29-8	Not listed (TP)	Solid	No data available	Soluble to 100 mM in DMSO	(Possibly Non-volatile)	Diterpene compound Protein kinase activator (phorbol type)

^a Chemical property data from Hazardous Substances Data Bank (HSDB) in Toxicology Data Network (TOXNET: <http://toxnet.nlm.nih.gov/index.htm>) except for methapyrilene HCL and mezerein.

^b Very good >1000 g/L; good 100-1000 g/L; moderate 10-100 g/L; poor 1-10 g/L; very poor <1 g/L

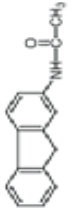


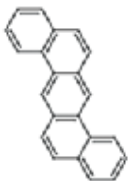
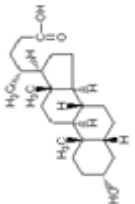
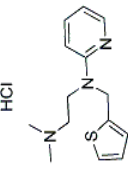
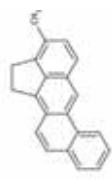
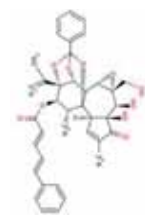
^c Carcinogen

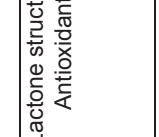
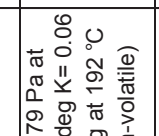
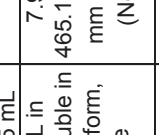

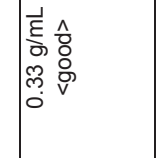
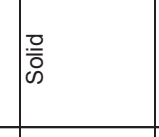
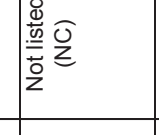

^d Tumor-promoter

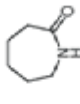
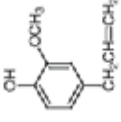
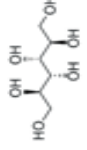
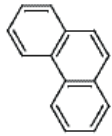
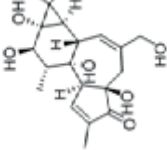

^e Non-carcinogen

Annex 15

Chemical properties and classes of test chemicals for the pre-validation phase, phase I and phase II of 96-well method validation study^a

#	Chemical Name	M.W.	Chemical structure	CAS	IARC [as of May 2010]	Physical appearance	Water solubility ^b	Solubility (Other solvents)	Vapor pressure (Volatility)	Chemical class
1	2-Acetylaminofluorene	223		53-96-3	Not listed (C ⁵)	Solid	144 mg/L at 25 °C (est) <very poor>	Soluble in ether, acetic acid	Possibly non-volatile	Aromatic amine; requires metabolic activation
2	Benzo[a]pyrene	252		50-32-8	1 (C)	Solid	1.60X10 ⁻³ mg/L at 25°C <very poor (almost insoluble)>	Soluble in benzene, toluene, xylene, ether	5.49X10 ⁻⁹ mm Hg at 25 °C (extrapolated) (Non-volatile)	Polycyclic aromatic hydrocarbon; Metabolic activation
3	Cadmium chloride	183		10108-64-2	1 (as Cadmium compounds) (C)	Solid	140 g/100 mL at 20°C <very good>	Soluble in acetone	10 mm Hg at 656 °C (Non-volatile)	Inorganic cadmium compound
4	Dibenz[a,h]anthracene	278		53-70-3	2A (C)	Solid	0.0005 mg/L at 27 °C <very poor (almost insoluble)>	Slightly soluble in ethanol, soluble in acetone and benzene.	1.1X10 ⁻¹⁰ mm Hg (est) (Possibly non-volatile)	Polycyclic aromatic hydrocarbon; requires metabolic activation
5	Lithocholic acid	377		434-13-9	Not listed (TP ⁵)	Solid	0.38 mg/L at 20 °C <very poor (almost insoluble)>	Freely soluble in hot alcohol	(Unknown, Possibly non-volatile)	Steroid derivative (Bile acid)
6	Methapyrilene hydrochloride	298		135-23-9	Not listed (TP)	Solid	1 gm dissolves in 0.5 mL water <very good>	1 gm dissolves in 5 mL alcohol, in 3 mL chloroform	(Unknown, Possibly non-volatile)	Heterocyclic amine; Anti-histamine drug
7	3-Methylcholanthrene	268		56-49-5	Not listed (C)	Solid	2.9X10 ⁻³ mg/L at 25°C <very poor>	Soluble in xylene, toluene, and benzene	4.3X10 ⁻⁸ mm Hg at 25 °C (Non-volatile)	Polycyclic aromatic hydrocarbon; requires metabolic activation
8	Mezerein	655		34807-41-5	Not listed (TP)	Solid	Not soluble <very poor (almost insoluble)>	Soluble in DMSO or ethanol	(Unknown, Possibly non-volatile)	Diterpene compound Protein kinase activator (Non-phorbol type)

#	Chemical Name	M.W.	Chemical structure	CAS	IARC [as of May 2010]	Physical appearance	Water solubility ^b	Solubility (Other solvents)	Vapor pressure (Volatility)	Chemical class
9	<i>N</i> -Methyl- <i>N'</i> -nitro- <i>N</i> -nitrosoguanidine (MNNG)	147		70-25-7	1 (C)	Solid	Slightly soluble (< 0.5%) <poor>	Soluble in DMSO	No data available (Possibly non-volatile)	Nitrosoguanidine; Alkylating agent
10	Sodium arsenite	130		7784-46-5	1 (as Arsenic compounds) (C)	Solid	Freely soluble <very good>	Slightly soluble in alcohol	0% volatile by volume at 21 °C (Possibly non-volatile)	Inorganic arsenite compound
11	<i>o</i> -Toluidine	107		95-53-4	1 (C)	Liquid	Slightly soluble; 1.66X10+4 mg/L at 25°C <moderate (close to poor)>	Soluble in alcohol, ether;	2.60X10-1 mm Hg at 25 °C; 0.32 torr; Volatile with steam; Volatile organic compound	Aromatic amine
12	12- <i>O</i> -tetradecanoylphorbol-13-acetate (TPA)	617		16561-29-8	Not listed (TP)	Solid	No data available	Soluble to 100 mM in DMSO	(Possibly non-volatile)	Diterpene compound; Protein kinase activator (phorbol type)
13	Ampicillin sodium salt	371		69-52-3	3 (as Ampicillin, 69-53-4) (NC ^e)	Solid	50 mg/mL <moderate>	No data available	(Unknown, Possibly non-volatile)	Beta-lactam antibiotics
14	Anthracene	178		120-12-7	3 (NC)	Solid	0.04-1.3 mg/L at 25°C <very poor>	1 gm dissolves in 67 mL absolute alcohol, 70 mL methanol, 62 mL benzene, 85 mL	2.67X10-6 mm Hg at 25 °C (eat???) (Non-volatile)	Polycyclic aromatic hydrocarbon
15	L-Ascorbic acid	176		50-81-7	Not listed (NC)	Solid	0.33 g/mL <good>	0.02 mg/mL in ethanol; insoluble in ether, chloroform, benzene	7.9179 Pa at 465.15 deg K= 0.06 mm Hg at 192 °C (Non-volatile)	Lactone structure; Antioxidant
16	Caffeine	194		58-08-2	3 (NC)	Solid	2.16X10+4 mg/L at 25°C <moderate>	1 gm dissolves in 66 mL alcohol, 50 mL acetone, 5.5 mL chloroform, 530 mL ether, 100 mL benzene	7.3X10-9 mm Hg at 25 °C (est) (Non-volatile)	Purine alkaloid

#	Chemical Name	M.W.	Chemical structure	CAS	IARC [as of May 2010]	Physical appearance	Water solubility ^b	Solubility (Other solvents)	Vapor pressure (Volatility)	Chemical class
17	Caprolactam	113		105-60-2	4 (NC)	Solid	5.25X10+6 mg/L at 25 °C <very good>	Freely soluble in methanol, ethanol; Soluble in benzene, ethanol, chloroform	1.9X10-3 mm Hg at 25 °C (Non-volatile)	Amid; Lactam
18	Eugenol	164		97-53-0	3 (NC)	Liquid	2.463X10+3 mg/L at 25 °C <poor>	Miscible with alcohol, chloroform, ether	2.26X10-2 mm Hg at 25 °C (Possibly volatile)	Phenylpropanoid compound
19	D-Mannitol	182		69-65-8	Not listed (NC)	Solid	2.16X10+5 mg/L at 25 °C <good>	Insoluble in ether	(Non-volatile)	Sugar alcohol
20	Phenanthrene	178		85-01-8	3 (NC)	Solid	0.977 - 1.18 mg/L at 25 °C <very poor>	Soluble in ethanol, diethyl ether, acetone, benzene, and carbon disulfide	1.21X10-4 mm Hg at 25 °C (Non-volatile)	Polycyclic aromatic hydrocarbon
21	Phorbol	364		17673-25-5	Not listed (NC)	Solid	Quite soluble <good/very good>	Highly soluble in polar solvents	(Possibly non-volatile)	Diterpene compound
22	Pyrene	202		129-00-0	3 (NC)	Solid	0.135 mg/L at 25 °C <very poor (almost insoluble)>	Soluble in alcohol, ether, benzene	8.92X10-5 mm Hg at 25 °C (Non-volatile)	Polycyclic aromatic hydrocarbon; requires metabolic activation

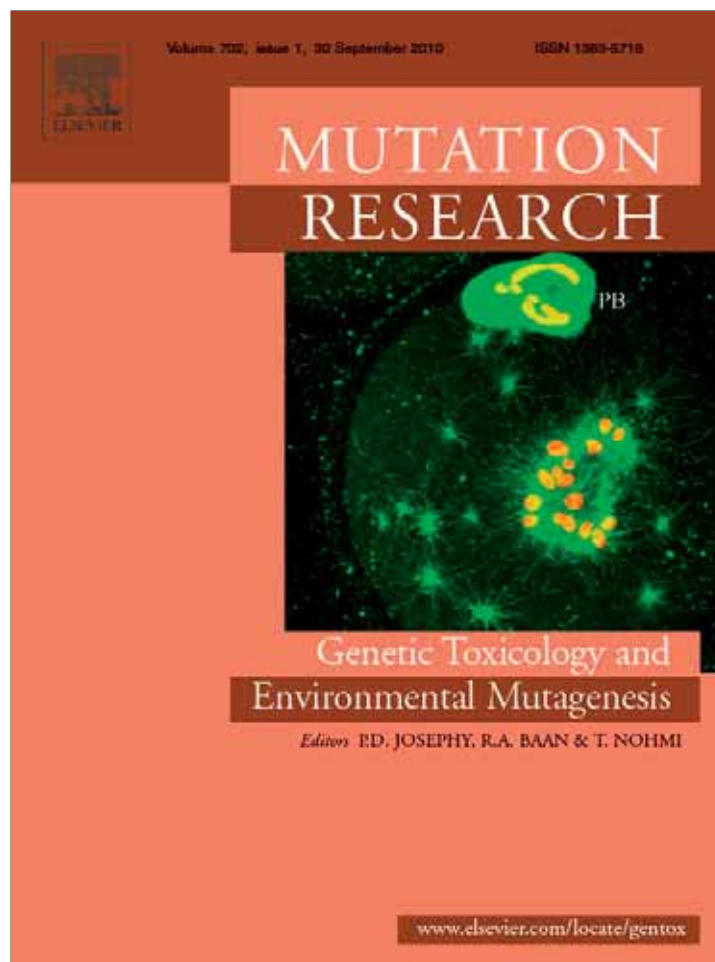
^a Chemical property data from Hazardous Substances Data Bank (HSDB) in Toxicology Data Network (TOXNET: <http://toxnet.nlm.nih.gov/index.html>) except for methapyrene HCL and mezer

^b very good > 1000 g/L; good 100-1000 g/L; moderate 10-100 g/L; poor 1-10 g/L; very poor <1 g/L

^c Carcinogen

^d Tumor-promoter

^e Non-carcinogen



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A Bhas 42 cell transformation assay on 98 chemicals: The characteristics and performance for the prediction of chemical carcinogenicity

Ayako Sakai*, Kiyoshi Sasaki, Dai Muramatsu, Shoko Arai, Nobuko Endou, Sachiko Kuroda, Kumiko Hayashi, Yeon-mi Lim, Shojiro Yamazaki, Makoto Umeda, Noriho Tanaka

Laboratory of Cell Carcinogenesis, Hatano Research Institute, Food and Drug Safety Center, Hadano, Kanagawa 257-8525, Japan

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ABSTRACT

The Bhas 42 cell transformation assay is a short-term system using a clone of the BALB/c 3T3 cells transfected with an oncogenic murine *ras* gene (*v-Ha-ras*). The assay has previously been reported to be capable of detecting the tumor-initiating and tumor-promoting activities of chemical carcinogens according to the different protocols, an initiation assay and a promotion assay, respectively. We applied this short-term assay to 98 chemicals to characterize the assay and evaluate its performance for the detection of chemical carcinogenicity. When the assay results were compared with the existing genotoxicity data, the Bhas 42 cell transformation assay could detect a considerable number of Ames-negative and Ames-discordant carcinogens; and the promotion assay detected most of those Ames-negative and -discordant carcinogens. This fact suggested that the Bhas 42 cells behaved as initiated cells in the transformation assay. The performance indices were calculated from the assay results of 52 carcinogens and 37 non-carcinogens. The concordance was 78%, sensitivity 73%, specificity 84%, positive predictivity 86%, negative predictivity 69%, false negative 27% and false positive 16%. Of these values, the concordance, specificity, negative predictivity and false positive were superior and the other performance indices were equivalent to those of conventional genotoxicity tests. From overall results, we concluded that the accuracy of prediction of chemical carcinogenicity would be improved by introducing the Bhas 42 cell transformation assay into the battery of *in vitro* assays.

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1. Introduction

The carcinogenicity of chemicals has been predicted with genotoxicity assays. *In vitro* cell transformation assays can detect chemical carcinogens but are classified into a category different from genotoxicity assays [1–4]. The cell transformation assays attract attention as an additional *in vitro* method to predict the carcinogenicity of chemicals, since there are concerns about the carcinogens to which conventional genotoxicity assays are insensitive [5]. An analysis estimated non-genotoxic carcinogens to be present in 12% of carcinogens classified into Group 1, 2A and 2B by the International Agency for Research on Cancer (IARC) [6].

BALB/c 3T3 cells and C3H 10T1/2 cells have been used for the cell transformation assays [7]. The assays using these cell lines require longer period compared with the conventional genotoxicity assays, but achieve production of visible transformed foci. Also, in the assay there are initiation and promotion stages that mimic the two stages in the carcinogenicity tests in experimental animals, and most of

the non-genotoxic carcinogens can be detected as promoters of cell transformation in the two-stage cell transformation assay, where the target cells are initiated with a known tumor initiator prior to treatment with a test chemical [8–10].

Recently a short-term cell transformation assay has been developed [11], using the Bhas 42 cells which were established from the BALB/c 3T3 cells through the transfection with a plasmid pBR322 containing Ha-MuSV-DNA, clone H1 (*v-Ha-ras*) [12–14]. The Bhas 42 cells are transformed by known tumor promoters, including 12-*O*-tetradecanoylphorbol-13-acetate (TPA), okadaic acid and lithocholic acid, without initiating treatment with a known tumor initiator such as 3-methylcholanthrene (MCA) [11], and are presumed to be initiated toward transformation by the introduced viral *ras* sequence [15]. Thus the Bhas 42 cell transformation assay does not require pretreatment to detect tumor-promoting activity, and the assay period is 3 weeks instead of 6 weeks in the BALB/c 3T3 cell transformation assay. Because of the increased sensitivity of cells to transformation, the assay scales such as the dish size, the dish number, the medium volume, the serum concentration in the cultures and the number of medium exchange have been diminished, so that the cost performance is improved. The Bhas 42 cell transformation assay has been proved by Asada et al. to

* Corresponding author. Tel.: +81 463 82 4751; fax: +81 463 82 9627.
E-mail address: sakai.a@fdsc.or.jp (A. Sakai).

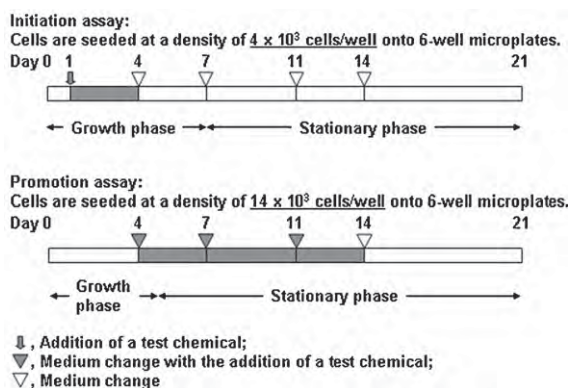


Fig. 1. Illustration of the Bhas 42 cell transformation assay.

be capable of detecting tumor-initiating activity as well as tumor-promoting activity of chemicals depending on a change between the two protocols, the initiation assay and the promotion assay [16]. The protocols are illustrated in Fig. 1. In the initiation assay, the cells are inoculated at a low density and treated with a test chemical in the beginning of the assay period so that the target cells can undergo cell division several times before contact inhibition of growth at confluence, and thus DNA damage is fixed as mutations in the genes. In the promotion assay, the cells are seeded more densely than in the initiation assay, and the treatment with a test chemical is started just before confluence (at sub-confluence) and continued longer than in the initiation assay.

Employing arsenic compounds, Muramatsu et al. verified that the sensitivity of Bhas 42 cell transformation assay for the detection of initiating activity is similar to that of the conventional BALB/c 3T3 cell transformation assay, and the sensitivity for the detection of promoting activity is equivalent to that of the two-stage BALB/c 3T3 cell transformation assay where the target cells are initiated with a sub-threshold dose of MCA [17]. Tanaka et al. conducted an interlaboratory collaborative study on the Bhas 42 cell transformation assay by 6 laboratories, and demonstrated that the assay on 9 chemicals is reproducible and applicable to the detection of both initiating activity and promoting activity of carcinogens [18].

We applied the Bhas 42 cell transformation assay to 98 chemicals including known carcinogens and non-carcinogens to evaluate its performance for predicting the carcinogenicity of chemical substances under a project financed by New Energy and Industrial Technology Development Organization (NEDO) in Japan. In this paper, we present the assay results of individual chemicals and the performance indices calculated from the results. We also compared the assay results and performances with existing genotoxicity data to characterize this assay.

2. Materials and methods

2.1. Cell culture

Bhas 42 cells were expanded and cryopreserved in a large batch prior to transformation assays. The cells were proliferated in Eagle's minimum essential medium (Nissui Pharmaceutical, Tokyo, Japan) supplemented with 10% fetal bovine serum (FBS; Moregate Biotech, Bulimba, Australia, and GIBCO™, Invitrogen, Carlsbad, CA, USA) (M10F) in a humidified 5% CO₂ incubator at 37 °C, and subcultured using 0.25% trypsin (Invitrogen) when the cultures reached about 70% confluence. The multiplied cells were suspended at 5×10^5 cells/mL in fresh M10F containing 5% dimethyl sulfoxide (DMSO; Wako Pure Chemical Industries, Osaka, Japan), frozen in 0.5 mL volumes at -80 °C and stored in liquid nitrogen. The transformation assays were always started from this frozen stock. Dulbecco's modified Eagle's medium/Ham's F12 (Invitrogen) supplemented with 5% FBS (DF5F) was used for the transformation assays. The two batches of FBS, which were prescreened to produce a minimal number of transformed foci in the Bhas 42 cells treated with DMSO alone and an appropriate number of foci in the cells treated with 1 µg/mL of MCA or 0.05 µg/mL of TPA in the transformation assay, were used [2,7].

2.2. Chemicals

Benzo[*a*]anthracene [56-55-3], 5-azacytidine [320-67-2], barium chromate [10294-40-3], benzo[*a*]pyrene [50-32-8], cyclophosphamide [50-18-0], cyclosporin A [59865-13-3], 2,4-diaminotoluene [95-80-7], melphalan [148-82-3], MCA [56-49-5], mitomycin C [50-07-7], sterigmatocystin [10048-13-2], thio-TEPA [52-24-4], chenodeoxycholic acid [474-25-9], 4-chloro-*o*-toluidine hydrochloride [3165-93-3], cholic acid [81-25-4], deoxycholic acid [83-44-3], dichlorvos [62-73-7], epichlorohydrin [106-89-8], *D*-limonene [5989-27-5], methapyrilene hydrochloride [135-23-9], mezerein [34807-41-5], phorbol 12,13-didecanoate [24928-17-4], sodium arsenate, [10048-95-0], styrene oxide [96-09-3], TPA [16561-29-8], *o*-toluidine [95-53-4], zinc chloride [7646-85-7], *o*-anisidine [90-04-0], benzene [71-43-2], cobalt sulfate heptahydrate [10026-24-1], diethylstilbestrol [56-53-1], 1,4-dioxane [123-91-1], urethane (ethyl carbamate) [51-79-6], methyl carbamate [598-55-0], nickel monoxide [1313-99-1], 4-acetylaminofluorene [28322-02-3], chromotrope FB (acid red 14) [3567-69-9], ampicillin sodium salt [69-52-3], anthracene [120-12-7], L-ascorbic acid [50-81-7], benzoin [119-53-9], caffeine [58-08-2], caprolactam [105-60-2], 2-chloroethanol [107-07-3], chromium(III) chloride, anhydrous [10025-73-7], diazepam [439-14-5], *N,N*-dimethylformamide [68-12-2], eugenol [97-53-0], HC blue no. 2 [33229-34-4], hydrocortisone [50-23-7], phenanthrene [85-01-8], phenol [108-95-2], *p*-phenylenediamine dihydrochloride [624-18-0], rotenone [83-79-4], sodium chloride [7647-14-5], sodium nitrite [7632-00-0], sunset yellow FCF [2783-94-0], thiabendazole [148-79-8], triphenyltin hydroxide [76-87-9], barium chloride dihydrate [10326-27-9], *tert*-butylhydroquinone [1948-33-0], 8-hydroxyquinoline [148-24-3], propyl gallate [121-79-9], tetracycline hydrochloride [64-75-5], sodium valproate [1069-66-5] and sodium orthovanadate [13721-39-6] were purchased from Sigma-Aldrich (St. Louis, MO, USA). 2-Acetylaminofluorene [53-96-3], 2-amino-3-methylimidazo[4,5-*f*]quinoline (IQ) [76180-96-6], dibenz[*a,h*]anthracene [53-70-3], cadmium chloride [10108-64-2], lithocholic acid [434-13-9], methylarsonic acid [124-58-3], dimethylarsinic acid [75-60-5], formaldehyde [50-00-0], nickel (II) chloride [7718-54-9], *p*-toluidine [106-49-0], aspartame [22839-47-0], 2,6-diaminotoluene [823-40-5], *D*-mannitol [69-65-8], methotrexate [59-05-2], phthalic anhydride [85-44-9], *m*-toluidine [108-44-1], sodium fluoride [7681-49-4], valproic acid [99-66-1], 2,3-diaminotoluene [2687-25-4], 2,5-diaminotoluene dihydrochloride [615-45-2], 3,4-diaminotoluene [496-72-0], capsaicin [404-86-4] and salicylic acid [69-72-7] were obtained from Wako Pure Chemical Industries. Quercetin [117-39-5], sodium saccharin [128-44-9], phenobarbital sodium salt [57-30-7] and 1-naphthylamine [134-32-7] were from Tokyo Chemical Industry (Tokyo, Japan). *N*-Methyl-*N*-nitro-*N*-nitrosoguanidine (MNNG) [70-25-7] and ethidium bromide [1239-45-8] were from Kanto Chemical (Tokyo, Japan). Sodium arsenite [7784-46-5] solution (0.05 M) was from Merck KGaA (Darmstadt, Germany). 2-Naphthylamine [91-59-8] solution (1001 µg/mL) was from Ultra Scientific (North Kingstown, RI, USA). Furfurylamide [3688-53-7] was from Ueno Fine Chemicals Industry (Tokyo, Japan).

Test chemicals were dissolved in sterile distilled water, DMSO, methanol (Wako) or acetone (Wako) as indicated in the parenthesis following "0" of the concentration columns in Table 1. MCA and TPA were in DMSO. The final concentration of distilled water was 5% in the culture medium, that of methanol was 0.5%, that of acetone was 0.5% and that of DMSO was 0.1 or 0.5% depending on the solubility of test chemicals in DMSO.

2.3. Transformation assays

The transformation assays were conducted according to the procedures reported by Asada et al. with a slight modification [16].

2.3.1. Initiation assay to examine initiating activity of chemicals

One tube of the frozen Bhas 42 cells was rapidly thawed and proliferated once in M10F up to about 70% confluence and then subcultured in DF5F to reach about 70% confluence again. Thereafter cells were cultured in DF5F. The cells were trypsinized and suspended at a density of 2000 cells/mL and seeded onto each well of 6-well microplates in 2 mL volumes (4000 cells/well, day 0). Six wells were prepared for each treatment group. At 24 h after seeding, the cells were treated by the addition of a test chemical solution or the vehicle alone to the cultures, and the treatment in the initiation phase was continued for 72 h. Following the exposure period, all treatment media were removed and the cells were refed with medium without the test chemical (day 4) and subsequently cultured in the normal medium until day 21, receiving medium exchanges on day 7, day 11 and day 14 (Fig. 1). The cells were fixed with methanol and stained with Giemsa's solution. Each assay was performed including MCA (1 µg/mL) as the positive control and 0.1% DMSO as its negative control.

2.3.2. Promotion assay to examine promoting activity of chemicals

The promotion assay was carried out in the same manner as the initiation assay except for the following procedures. The cells were seeded at a density of 7000 cells/mL (14,000 cells/well, day 0) and cultured for 4 days without a medium exchange. On day 4, day 7, and day 11, the culture medium was replaced with a fresh medium containing a test chemical or vehicle alone and the treatment in the promotion phase was continued until day 14 (for a total of 10 days). The cells were then subsequently cultured in the normal medium without the test chemical for 1

Table 1
Results of Bhas 42 cell transformation assay of 92 chemicals.

Chemical ^a	Initiation assay				Promotion assay						
	Concentration (µg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (µg/mL)	CG	Foci/well	Judgment			
<i>Carcinogens</i>											
2-Acetylaminofluorene (53-96-3)	0 ^e (0.1% DMSO)	100	3.3 ± 1.5	+	0 (0.1% DMSO)	100	7.0 ± 2.4	+			
	10	99	6.2 ± 3.3		1	103	8.7 ± 2.3				
	20	62	9.0 ± 3.8 [†]		5	100	13.0 ± 1.9 [†]				
	25	19	9.5 ± 2.6 [†]		10	82	21.7 ± 6.3 [†]				
	30	13	7.7 ± 1.9		15	71	16.0 ± 4.6 [†]				
	40	9	6.7 ± 2.6		20	62	8.8 ± 2.5				
	50	9	7.5 ± 3.3		25	61	4.0 ± 2.2				
	MCA ^f	1	58		37.2 ± 5.8 [#]	TPA ^g	0.05		146	20.8 ± 3.5 [#]	
Benz[<i>a</i>]anthracene (56-55-3)	0 (0.1% DMSO)	100	1.0 ± 1.1	+	0 (0.1% DMSO)	100	3.0 ± 1.3	+			
	0.25	84	2.7 ± 1.6		0.25	79	4.7 ± 2.1				
	0.5	73	3.7 ± 2.0		0.5	70	6.3 ± 2.7				
	1	71	8.5 ± 2.3 [†]		1	68	9.3 ± 2.7 [†]				
	2.5	67	8.0 ± 3.2 [†]		2.5	64	12.2 ± 2.7 [†]				
	5	63	11.2 ± 2.6 [†]		5	65	8.5 ± 3.2 [†]				
	10	60	13.2 ± 2.8 [†]		10	69	9.7 ± 3.7 [†]				
	MCA	1	30		15.0 ± 3.8 [#]	TPA	0.05		151	16.5 ± 4.8 [#]	
2-Amino-3-methylimidazo[4,5- <i>f</i>]quinoline (IQ) (76180-96-6)	0 (0.5% DMSO)	100	4.2 ± 0.8	+	0 (0.5% DMSO)	100	2.2 ± 1.9	-			
	20	73	7.2 ± 2.3		3.2	95	2.7 ± 0.5				
	40	63	11.3 ± 3.2 [†]		5.6	72	3.2 ± 1.8				
	60	56	13.2 ± 4.4 [†]		10	92	1.0 ± 0.9				
	70	55	15.8 ± 2.3 [†]		18	66	0.8 ± 1.2				
	80	48	10.8 ± 1.2 [†]		32	65	1.0 ± 0.9				
	90	42	7.5 ± 2.8		56	51	0.5 ± 0.5				
	100	37	8.0 ± 3.0		TPA	0.05	131		17.5 ± 5.9 [#]		
	MCA	1	56		14.2 ± 1.6 [#]						
	5-Azacytidine (320-67-2)	0 (5% Water)	100		2.0 ± 0.9	+	0 (5% Water)		100	5.7 ± 1.2	-
0.1		40	2.7 ± 1.8	0.01	104		6.0 ± 2.6				
0.13		35	4.2 ± 1.0	0.018	103		8.2 ± 4.0				
0.18		26	8.3 ± 1.5 [†]	0.032	101		6.5 ± 1.9				
0.25		18	7.0 ± 4.5 [†]	0.056	104		6.2 ± 1.2				
0.3		12	7.8 ± 3.5 [†]	0.1	105		6.8 ± 1.3				
0.5		2	Toxic	0.18	100		6.3 ± 2.1				
0 ^h (0.1% DMSO)		100	1.5 ± 1.0	0.32	99		4.2 ± 0.8				
1		47	18.7 ± 3.2 [#]	0 ⁱ (0.1% DMSO)	100		7.7 ± 2.1				
MCA				TPA	0.05		149	33.7 ± 5.0 [#]			
Barium chromate (10294-40-3)		0 (5% Water)	100	1.0 ± 0.6	+		0 (5% Water)	100	2.7 ± 1.4	-	
		1	101	0.7 ± 0.8			0.5	95	2.7 ± 1.6		
	2	95	2.7 ± 0.8	1		105	1.7 ± 1.2				
	3	77	2.0 ± 0.0	2		100	2.3 ± 1.4				
	4	58	3.7 ± 1.5 [†]	3.5		106	2.3 ± 1.9				
	5	66	4.3 ± 1.2 [†]	5		105	2.0 ± 0.9				
	7	24	4.5 ± 2.2 [†]	7		109	1.8 ± 1.2				
	10	15	4.5 ± 2.4 [†]	10		108	1.8 ± 1.5				
	15	10	3.5 ± 1.8 [†]	15		99	2.3 ± 2.6				
	0 (0.1% DMSO)	100	2.2 ± 1.5	0 (0.1% DMSO)		100	2.5 ± 1.6				
	1	47	27.5 ± 3.4 [#]	TPA		0.05	176	20.5 ± 1.6 [#]			
	Benzo[<i>a</i>]pyrene (50-32-8)	0 (0.1% DMSO)	100	1.8 ± 1.2		+	0 (0.1% DMSO)	100	2.5 ± 0.8		-
		0.01	75	4.3 ± 2.0 [†]			0.01	85	1.0 ± 0.9		
		0.025	35	7.5 ± 2.7 [†]			0.025	79	1.8 ± 1.3		
0.05		26	4.3 ± 1.5 [†]	0.05	70		1.3 ± 1.5				
0.1		12	4.5 ± 2.1 [†]	0.1	57		1.3 ± 1.5				
0.25		4	Toxic	0.25	43		0.5 ± 0.8				
1		27	14.0 ± 3.2 [#]	TPA	0.05		152	14.5 ± 3.9 [#]			
Cyclophosphamide (50-18-0)		0 (5% Water)	100	3.3 ± 2.2	+		0 (5% Water)	100	4.5 ± 1.6	-	
	100	103	5.2 ± 1.2	160		100	3.7 ± 1.9				
	500	101	10.8 ± 1.0 [†]	250		105	2.8 ± 1.6				
	1100	55	14.8 ± 3.6 [†]	390		108	4.2 ± 2.5				
	1300	37	9.7 ± 4.3 [†]	620		97	3.5 ± 1.0				
	1500	26	9.8 ± 3.3 [†]	1000		95	1.8 ± 1.7				

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay			
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment
MCA	2000	16	1.2 ± 0.8		1600	87	0.0 ± 0.0	
	2200	13	Toxic		2500	81	0.0 ± 0.0	
	2400	10	Toxic		0 (0.1% DMSO)	100	3.7 ± 1.9	
	0 (0.1% DMSO)	100	6.5 ± 3.1		TPA 0.05	154	18.5 ± 4.1 [#]	
	1	63	44.5 ± 2.1 [#]					
Cyclosporin A (59865-13-3)				+				-
First experiment				±				-
	0 (0.1% DMSO)	100	1.7 ± 0.5		0 (0.1% DMSO)	100	4.5 ± 1.8	
	2	95	4.0 ± 1.7		0.0625	100	7.5 ± 3.3	
	2.5	89	4.7 ± 2.6 [*]		0.125	98	6.0 ± 4.1	
	3	89	2.5 ± 2.2		0.25	99	6.8 ± 2.1	
	3.5	86	2.7 ± 1.2		0.5	93	6.3 ± 3.4	
	4	89	1.0 ± 1.1		1	90	3.5 ± 3.2	
	4.5	79	1.0 ± 1.1		2	83	1.8 ± 1.8	
	5	78	1.3 ± 1.0		4	79	2.3 ± 1.8	
MCA	1	45	21.7 ± 2.3 [#]		6	74	1.2 ± 1.0	
					TPA 0.05	140	37.8 ± 6.6 [#]	
Second experiment				+				
	0 (0.1% DMSO)	100	2.8 ± 0.8					
	0.5	104	4.3 ± 1.5					
	1	107	4.7 ± 1.0					
	1.6	106	5.5 ± 2.4					
	2.5	108	7.0 ± 3.2 [*]					
	4	99	4.8 ± 2.4					
	6	86	5.8 ± 1.0					
	10	40	6.2 ± 1.9 [*]					
	15	31	4.0 ± 1.3					
MCA	1	35	33.0 ± 3.0 [#]					
2,4-Diaminotoluene (95-80-7)				+				-
	0 (0.1% DMSO)	100	5.2 ± 1.0		0 (0.1% DMSO)	100	6.7 ± 1.4	
	10	90	8.0 ± 2.2		1	92	7.3 ± 1.8	
	15	98	7.5 ± 2.3		3	92	8.5 ± 2.2	
	20	92	10.2 ± 4.2 [*]		5	89	7.2 ± 1.5	
	25	90	10.5 ± 3.8 [*]		7.5	78	7.5 ± 3.9	
	30	80	9.3 ± 2.9		10	75	5.2 ± 1.9	
	35	67	12.3 ± 4.4 [*]		15	75	3.0 ± 1.1	
	40	52	12.2 ± 3.1 [*]		20	67	0.7 ± 1.2	
	45	45	12.7 ± 2.1 [*]		TPA 0.05	138	38.0 ± 5.9 [#]	
	50	33	8.0 ± 2.4					
MCA	1	42	45.8 ± 3.8 [#]					
Dibenz[<i>a,h</i>]anthracene (53-70-3)				+				-
	0 (0.1% DMSO)	100	1.0 ± 1.3		0 (0.1% DMSO)	100	3.0 ± 1.1	
	0.003	109	2.8 ± 1.2		0.0003	85	1.0 ± 1.1	
	0.01	86	2.2 ± 1.2		0.001	78	3.0 ± 1.8	
	0.03	65	6.5 ± 1.9 [*]		0.003	79	2.8 ± 0.8	
	0.1	36	13.5 ± 3.1 [*]		0.01	79	4.2 ± 2.6	
	0.3	22	13.8 ± 2.5 [*]		0.03	70	2.0 ± 2.2	
	1	24	12.0 ± 2.2 [*]		0.1	62	2.3 ± 1.2	
	3	20	10.5 ± 3.6 [*]		0.3	56	2.7 ± 1.9	
	10	26	Toxic		1	63	1.0 ± 1.1	
MCA	1	43	11.7 ± 5.3 [#]		TPA 0.05	227	10.8 ± 2.6 [#]	
Melphalan (148-82-3)				+				-
	0 (5% Water)	100	3.2 ± 1.0		0 (5% Water)	100	8.2 ± 4.0	
	0.1	99	2.3 ± 1.8		0.05	93	5.0 ± 2.8	
	0.25	107	5.5 ± 3.3		0.1	98	8.2 ± 4.3	
	0.5	105	6.2 ± 1.6		0.25	93	4.0 ± 1.5	
	0.75	109	8.7 ± 2.3 [*]		0.5	94	0.7 ± 0.8	
	1	80	9.5 ± 2.9 [*]		0.75	87	Toxic	
	1.5	48	15.3 ± 4.1 [*]		1	90	Toxic	
	2	20	8.5 ± 3.4 [*]		0 (0.1% DMSO)	100	4.8 ± 1.5	
	0 (0.1% DMSO)	100	2.2 ± 1.0		TPA 0.05	131	33.3 ± 3.1 [#]	
MCA	1	61	29.7 ± 3.4 [#]					
3-Methylcholanthrene (MCA) (56-49-5)				+				-
	0 (0.1% DMSO)	100	2.2 ± 1.5		0 (0.1% DMSO)	100	2.3 ± 1.0	
	0.001	96	1.0 ± 1.5		0.001	95	0.8 ± 1.0	
	0.008	97	1.0 ± 0.9		0.008	93	0.5 ± 0.8	
	0.016	92	2.5 ± 1.0		0.016	90	0.5 ± 0.5	
	0.031	93	4.0 ± 1.8		0.031	87	1.0 ± 0.9	
	0.063	81	4.8 ± 1.6		0.063	83	1.5 ± 1.5	
	0.125	77	7.7 ± 2.2 [*]		0.125	81	2.7 ± 1.6	

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay						
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment			
MCA	0.25	66	12.7 ± 2.7 ⁺		0.25	73	1.3 ± 1.2				
	0.5	53	18.8 ± 3.9 ⁺		0.5	70	0.3 ± 0.8				
	1	42	24.8 ± 3.4 ⁺		1	68	0.5 ± 0.5				
	2	35	28.7 ± 5.1 ⁺		2	69	0.5 ± 0.5				
	1	45	21.2 ± 3.6 [#]		TPA 0.05	148	10.2 ± 3.4 [#]				
Mitomycin C (50-07-7)	0 (0.1% DMSO)	100	2.0 ± 1.8	+	0 (0.1% DMSO)	100	3.2 ± 2.1	-			
	0.006	84	1.2 ± 1.2		0.004	94	2.8 ± 1.8				
	0.01	64	3.7 ± 1.6		0.006	98	3.0 ± 1.7				
	0.016	37	5.3 ± 1.0 ⁺		0.01	94	1.0 ± 0.6				
	0.025	8	5.2 ± 2.8 ⁺		0.016	95	0.5 ± 0.5				
	0.039	3	Toxic		0.025	90	0.2 ± 0.4				
	0.062	2	Toxic		0.039	86	0.0 ± 0.0				
	0.1	2	Toxic		0.062	88	0.0 ± 0.0				
	1	36	14.7 ± 3.0 [#]		0.1	84	0.0 ± 0.0				
					TPA 0.05	123	19.5 ± 3.6 [#]				
N-Methyl-N'-nitro-N-nitrosoguanidine (MNNG) (70-25-7)	0 (0.1% DMSO)	100	1.7 ± 1.2	+	0 (0.1% DMSO)	100	2.2 ± 1.3	-			
	0.01	92	0.5 ± 0.5		0.01	92	1.0 ± 1.3				
	0.05	94	0.5 ± 0.5		0.05	97	2.2 ± 1.6				
	0.1	99	1.7 ± 0.8		0.1	101	2.3 ± 1.2				
	0.15	94	2.2 ± 1.5		0.15	108	1.8 ± 1.0				
	0.22	93	3.0 ± 1.3		0.22	103	1.0 ± 0.6				
	0.32	80	5.0 ± 1.8		0.32	117	1.7 ± 1.2				
	0.47	47	6.0 ± 1.8 ⁺		0.47	103	2.5 ± 1.5				
	0.7	15	9.7 ± 4.8 ⁺		0.7	116	2.0 ± 1.5				
	1	6	3.8 ± 3.3		1	112	0.7 ± 0.8				
	2	2	Toxic		2	109	0.0 ± 0.0				
	1	36	27.7 ± 3.8 [#]		TPA 0.05	143	13.8 ± 5.9 [#]				
	Sterigmatocystin (10048-13-2)	0 (0.1% DMSO)	100		1.2 ± 1.5	+	0 (0.1% DMSO)		100	7.8 ± 3.0	-
		0.005	118		5.8 ± 3.3		0.00003		100	7.3 ± 2.4	
		0.01	112		9.5 ± 2.5 ⁺		0.0001		100	7.2 ± 3.5	
		0.02	82		13.8 ± 5.5 ⁺		0.0003		95	7.2 ± 1.9	
		0.03	56		18.0 ± 4.1 ⁺		0.001		95	8.5 ± 3.4	
0.04		37	23.5 ± 7.3 ⁺	0.003	102		5.2 ± 1.6				
0.05		36	19.8 ± 4.4 ⁺	0.01	99		1.7 ± 0.5				
0.1		11	Toxic	0.03	89		0.0 ± 0.0				
1		62	30.0 ± 5.8 [#]	TPA 0.05	142		35.8 ± 6.0 [#]				
Thio-TEPA (52-24-4)		0 (5% Water)	100	1.0 ± 1.1	+		0 (5% Water)	100	1.2 ± 0.8	-	
	0.06	98	0.7 ± 0.8	0.02		96	1.2 ± 1.0				
	0.1	82	1.7 ± 1.9	0.04		95	0.7 ± 0.8				
	0.16	79	1.7 ± 1.0	0.06		93	1.5 ± 1.0				
	0.25	67	5.2 ± 2.3 ⁺	0.08		88	1.5 ± 1.5				
	0.4	43	8.8 ± 2.6 ⁺	0.1		84	0.3 ± 0.5				
	0.6	16	3.5 ± 2.8	0 (0.1% DMSO)		100	2.7 ± 1.4				
	1	3	Toxic	TPA 0.05		114	12.0 ± 5.0 [#]				
	0 (0.1% DMSO)	100	1.0 ± 1.5								
	1	42	15.2 ± 5.3 [#]								
Cadmium chloride (10108-64-2)	0 (5% Water)	100	2.3 ± 1.8	-	0 (5% Water)	100	2.8 ± 1.8	+			
	0.5	86	3.2 ± 2.3		0.05	106	1.7 ± 1.0				
	0.63	87	2.8 ± 2.1		0.07	103	2.0 ± 1.9				
	0.8	76	1.8 ± 2.6		0.1	106	3.7 ± 2.1				
	1	76	2.8 ± 1.7		0.2	106	7.0 ± 2.0 ⁺				
	1.3	40	1.2 ± 1.9		0.3	109	7.3 ± 2.2 ⁺				
	1.6	14	0.7 ± 0.8		0.4	113	12.5 ± 3.1 ⁺				
	2	0	Toxic		0.5	119	19.2 ± 4.0 ⁺				
	0 (0.1% DMSO)	100	3.3 ± 2.3		0 (0.1% DMSO)	100	3.8 ± 1.2				
	1	36	16.5 ± 6.4 [#]		TPA 0.05	119	21.5 ± 4.8 [#]				
Chenodeoxycholic acid (474-25-9)	0 (0.1% DMSO)	100	0.5 ± 0.5	-	0 (0.1% DMSO)	100	2.7 ± 0.8	+			
	10	94	1.2 ± 1.2		10	74	3.0 ± 0.9				
	12	90	1.2 ± 1.0		12	87	6.8 ± 3.8 ⁺				
	14	100	2.0 ± 0.6		14	99	5.2 ± 2.0				
	16	105	1.2 ± 0.4		16	100	5.3 ± 1.4				
	19	103	1.0 ± 1.3		19	101	6.7 ± 2.6				
	23	103	1.5 ± 1.5		23	101	10.7 ± 4.0 ⁺				

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay				
	Concentration (µg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (µg/mL)	CG	Foci/well	Judgment	
MCA	27	109	1.0 ± 1.5		27	112	14.3 ± 2.7*		
	32	122	1.0 ± 0.9		32	115	18.3 ± 3.9*		
	37	72	2.2 ± 1.2		37	139	0.2 ± 0.4		
	1	48	27.8 ± 6.7 [#]		TPA 0.05	126	22.2 ± 4.2 [#]		
4-Chloro- <i>o</i> -toluidine hydrochloride (3165-93-3)				–				+	
	0 (5% Water)	100	1.0 ± 1.1		0 (5% Water)	100	1.7 ± 0.8		
	3	72	0.7 ± 0.8		1.8	87	5.2 ± 1.0		
	6	76	0.0 ± 0.0		3	80	4.8 ± 2.1		
	10	90	1.7 ± 1.6		6	78	4.8 ± 2.9		
	18	84	1.0 ± 1.3		10	72	6.7 ± 3.1*		
	30	71	1.5 ± 1.8		18	61	6.7 ± 1.9*		
	60	57	1.7 ± 1.2		30	62	6.0 ± 1.5*		
	100	42	0.3 ± 0.5		60	43	8.3 ± 3.4*		
	180	50	0.2 ± 0.4		100	39	9.8 ± 1.9*		
	0 (0.1% DMSO)	100	1.0 ± 1.5		0 (0.1% DMSO)	100	2.2 ± 1.6		
	1	42	15.2 ± 5.3 [#]		TPA 0.05	227	11.8 ± 2.8 [#]		
	Cholic acid (81-25-4)				–				+
0 (0.1% DMSO)		100	2.0 ± 0.9		0 (0.1% DMSO)	100	4.3 ± 1.2		
10		64	0.3 ± 0.5		18	74	5.5 ± 2.4		
18		80	0.7 ± 0.8		24	87	5.0 ± 1.7		
32		90	0.7 ± 0.8		32	99	7.7 ± 2.4		
56		96	1.2 ± 0.8		42	100	10.2 ± 2.8*		
100		100	1.2 ± 1.2		56	101	10.2 ± 3.9*		
180		109	2.0 ± 1.1		75	101	10.2 ± 2.8*		
320		23	2.0 ± 1.4		100	112	12.7 ± 3.6*		
1		48	33.3 ± 3.8 [#]		130	115	14.5 ± 4.8*		
					180	139	20.5 ± 2.1*		
					TPA 0.05	152	24.5 ± 3.4 [#]		
Deoxycholic acid (83-44-3)					–				+
	0 (0.1% DMSO)	100	2.3 ± 1.4		0 (0.1% DMSO)	100	2.7 ± 0.8		
	6	100	0.7 ± 0.8		19	94	3.8 ± 2.0		
	10	97	1.2 ± 1.5		23	94	6.8 ± 2.8		
	15	109	1.3 ± 1.4		27	112	14.0 ± 6.2*		
	22	82	2.0 ± 1.3		32	125	18.0 ± 4.2*		
	32	46	2.0 ± 1.7		37	141	14.0 ± 4.6*		
	47	24	2.2 ± 1.5		44	100	0.0 ± 0.0		
	1	48	25.2 ± 8.0 [#]		52	41	0.0 ± 0.0		
					61	9	Toxic		
					TPA 0.05	152	27.2 ± 6.7 [#]		
	Dichlorvos (62-73-7) First experiment				–				±
		0 (0.1% DMSO)	100	1.7 ± 1.0	–	0 (0.1% DMSO)	100	4.5 ± 1.4	
5.5		92	1.2 ± 1.2		5.5	100	3.7 ± 1.0		
7		88	2.0 ± 1.3		7	100	5.5 ± 2.7		
10		32	1.8 ± 1.2		10	104	4.8 ± 2.6		
13		16	0.8 ± 0.8		13	105	6.3 ± 2.7		
16		5	0.3 ± 0.8		16	105	6.3 ± 1.8		
20		2	0.2 ± 0.4		20	116	8.2 ± 3.3		
24		1	0.5 ± 1.2		24	123	8.5 ± 2.0*		
30		0	Toxic		30	123	4.2 ± 2.7		
1		36	14.5 ± 3.7 [#]		TPA 0.05	116	22.8 ± 4.5 [#]		
MCA Second experiment									±
		0 (0.1% DMSO)	100			0 (0.1% DMSO)	100	4.7 ± 1.0	
	4	93			4	93	3.3 ± 3.3		
	7	95			7	95	4.2 ± 2.2		
	10	102			10	102	4.3 ± 2.5		
	13	106			13	106	6.8 ± 1.0		
	16	101			16	101	5.2 ± 2.9		
	20	109			20	109	4.5 ± 2.3		
	24	124			24	124	5.5 ± 2.3		
	30	126			30	126	8.7 ± 2.4*		
	1	147			TPA 0.05	147	21.8 ± 3.2 [#]		
	Epichlorohydrin (106-89-8)				–				+
		0 (0.1% DMSO)	100	1.8 ± 1.3		0 (0.1% DMSO)	100	4.7 ± 1.4	
20		121	2.7 ± 0.5		10	108	7.0 ± 4.0		
40		103	2.0 ± 1.3		20	99	7.7 ± 1.5		
60		116	1.5 ± 1.5		30	116	15.0 ± 5.1*		
80		42	2.8 ± 1.8		40	113	19.2 ± 7.5*		
100		3	Toxic		50	115	Toxic		
1		56	21.5 ± 1.5 [#]		60	145	Toxic		
				70	117	Toxic			

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay				
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment	
D-Limonene (5989-27-5)				–	TPA 0.05	163	35.0 ± 5.0 [#]	+	
	0 (0.5% DMSO)	100	6.2 ± 2.0		0 (0.5% DMSO)	100	4.5 ± 2.0		
	10	104	9.2 ± 3.1		10	98	3.8 ± 2.1		
	20.5	48	6.2 ± 2.8		25	92	6.3 ± 3.5		
	21.5	35	6.3 ± 2.3		26	99	3.5 ± 2.2		
	22.5	12	2.2 ± 1.2		27	95	5.0 ± 3.9		
	23.5	18	2.2 ± 1.3		28	102	10.7 ± 2.3 ⁺		
	24.5	6	1.3 ± 1.0		29	81	14.2 ± 8.3 ⁺		
	25	11	1.2 ± 1.9		30	83	9.8 ± 3.7		
	26	3	0.7 ± 1.0		32	57	2.2 ± 2.4		
	MCA 1	66	35.3 ± 7.3 [#]		34	22	Toxic		
					TPA 0.05	132	17.8 ± 4.2 [#]		
	Lithocholic acid (434-13-9)				–				+
0 (0.1% DMSO)		100	1.0 ± 0.6		0 (0.1% DMSO)	100	1.8 ± 1.5		
12		109	1.0 ± 0.9		12	94	15.8 ± 3.3 ⁺		
14		108	0.7 ± 0.8		14	88	17.7 ± 3.6 ⁺		
16		119	0.8 ± 0.8		16	89	18.5 ± 3.0 ⁺		
19		120	0.2 ± 0.4		19	92	24.5 ± 4.5 ⁺		
23		129	0.8 ± 0.8		23	139	9.0 ± 1.7 ⁺		
27		63	1.5 ± 1.4		27	34	0.0 ± 0.0		
MCA 1		48	23.7 ± 3.6 [#]		TPA 0.05	152	22.8 ± 3.7 [#]		
Methapyrilene hydrochloride (135-23-9)					–				+
		0 (5% Water)	100	2.3 ± 2.5		0 (5% Water)	100	2.5 ± 2.0	
		100	105	2.3 ± 1.8		10	114	7.0 ± 3.0	
		150	88	2.2 ± 1.5		25	127	8.5 ± 1.9 ⁺	
	200	67	2.7 ± 1.0		50	155	18.5 ± 6.3 ⁺		
	250	30	0.8 ± 0.8		75	165	9.5 ± 3.9 ⁺		
	300	5	0.5 ± 0.8		100	162	7.8 ± 1.8 ⁺		
	350	1	1.0 ± 1.3		150	143	2.0 ± 0.9		
	400	1	Toxic		200	125	0.5 ± 0.5		
	0 (0.1% DMSO)	100	2.5 ± 2.0		250	109	1.0 ± 0.6		
	MCA 1	29	17.2 ± 3.2 [#]		0 (0.1% DMSO)	100	2.7 ± 1.5		
					TPA 0.05	150	17.3 ± 3.4 [#]		
	Methylarsonic acid (124-58-3)				–				+
0 (5% Water)		100	2.5 ± 2.3		0 (5% Water)	100	5.8 ± 2.6		
700		103	1.7 ± 1.2		1	101	5.3 ± 3.2		
900		100	2.7 ± 2.3		3	94	8.3 ± 2.2		
1000		96	1.8 ± 0.8		7	100	14.3 ± 4.8 ⁺		
1100		80	2.8 ± 1.9		10	97	19.2 ± 1.7 ⁺		
1200		60	2.3 ± 2.2		30	107	43.3 ± 3.6 ⁺		
1300		41	0.5 ± 0.8		70	107	55.0 ± 3.3 ⁺		
1400 (10.0 mM)		23	1.7 ± 1.4		100	115	60.8 ± 3.3 ⁺		
0 (0.1% DMSO)		100	3.2 ± 1.6		140	119	56.3 ± 3.5 ⁺		
MCA 1		54	24.5 ± 2.4 [#]		280	124	36.3 ± 3.0 ⁺		
					400	123	Toxic		
					500	111	Toxic		
				0 (0.1% DMSO)	100	8.2 ± 1.0			
				TPA 0.05	140	30.0 ± 3.7 [#]			
Mezerein (34807-41-5)				–				+	
	0 (0.1% DMSO)	100	2.2 ± 1.3		0 (0.1% DMSO)	100	4.3 ± 2.0		
	0.00001	86	3.0 ± 1.4		0.00001	98	4.2 ± 2.1		
	0.00003	89	0.3 ± 0.5		0.00003	96	5.2 ± 3.2		
	0.00005	87	2.3 ± 0.5		0.00005	99	7.0 ± 2.2		
	0.0001	86	0.7 ± 1.2		0.0001	99	9.3 ± 2.9 ⁺		
	0.0003	84	1.7 ± 1.6		0.0003	123	19.5 ± 4.2 ⁺		
	0.0005	81	1.5 ± 0.5		0.0005	152	33.7 ± 3.3 ⁺		
	0.001	83	1.5 ± 1.4		0.001	191	40.2 ± 3.1 ⁺		
	0.003	114	3.0 ± 1.9		0.003	163	Toxic		
	MCA 1	31	17.2 ± 3.5 [#]		TPA 0.05	150	20.0 ± 3.3 [#]		
	2-Naphthylamine (91-59-8)				±				+
		0 (0.5% Methanol)	100	2.2 ± 1.5		0 (0.5% Methanol)	100	0.7 ± 1.2	
0.01		111	1.8 ± 1.8		0.01	71	1.2 ± 0.8		
0.03		113	1.8 ± 1.8		0.03	74	0.8 ± 0.4		
0.1		116	4.2 ± 2.4		0.1	72	0.5 ± 0.8		
0.3		106	2.2 ± 1.0		0.3	81	0.8 ± 0.8		
1		99	2.2 ± 0.8		1	84	2.0 ± 1.3		
3		76	4.2 ± 1.0		3	81	6.0 ± 2.6 ⁺		
5		72	5.0 ± 2.4 ⁺		5	72	10.2 ± 2.5 ⁺		

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay			
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment
MCA	0 (0.1% DMSO)	100	2.3 ± 2.1		TPA	0 (0.1% DMSO)	100	2.0 ± 1.9
	1	36	12.7 ± 4.1 [#]			0.05	104	4.7 ± 1.8 [#]
Phorbol 12,13-didecanoate (24928-17-4)				+				+
	0 (0.1% DMSO)	100	3.2 ± 1.8		0 (0.1% DMSO)	100	5.5 ± 2.3	
	0.01	124	1.8 ± 1.2		0.01	156	63.2 ± 10.7 ⁺	
	0.03	99	3.0 ± 1.4		0.03	178	65.0 ± 10.3 ⁺	
	0.1	97	7.5 ± 1.9		0.1	180	53.2 ± 5.4 ⁺	
	0.18	104	8.5 ± 2.3 ⁺		0.18	189	51.3 ± 2.9 ⁺	
	0.3	110	12.7 ± 3.5 ⁺		0.3	176	52.3 ± 1.6 ⁺	
	1	133	29.8 ± 7.2 ⁺		1	184	45.5 ± 4.3 ⁺	
MCA	1	59	32.7 ± 4.8 [#]		TPA	0.05	125	22.7 ± 3.1 [#]
Quercetin (117-39-5)				–				+
	0 (0.1% DMSO)	100	3.5 ± 2.2		0 (0.1% DMSO)	100	7.3 ± 1.0	
	2	98	4.5 ± 2.0		0.5	98	8.5 ± 2.1	
	3	98	3.3 ± 2.3		1	93	9.5 ± 3.7	
	4	77	1.8 ± 1.2		2	84	13.5 ± 3.1 ⁺	
	5	27	3.3 ± 2.7		3	70	16.8 ± 4.2 ⁺	
	6	11	2.0 ± 1.3		5	32	40.8 ± 6.9 ⁺	
	7	3	1.8 ± 0.8		7	25	Toxic	
MCA	1	42	27.8 ± 2.6 [#]		TPA	0.05	120	35.2 ± 5.6 [#]
Sodium arsenate (10048-95-0)				–				+
	0 (5% Water)	100	1.5 ± 1.4		0 (5% Water)	100	4.5 ± 2.0	
	0.5	102	1.5 ± 1.0		0.5	103	6.0 ± 2.2	
	0.7	97	1.0 ± 1.3		0.7	110	5.8 ± 1.9	
	1	86	1.5 ± 0.5		1	106	9.7 ± 3.7 ⁺	
	1.5	52	2.3 ± 2.0		1.5	103	17.0 ± 4.4 ⁺	
	2.2	31	1.7 ± 0.8		1.7	111	13.2 ± 3.1 ⁺	
	3.2	12	1.0 ± 0.9		2.2	102	11.0 ± 1.8 ⁺	
	5	6	0.8 ± 1.6		3.2	108	8.3 ± 2.5	
	0 (0.1% DMSO)	100	1.2 ± 1.5		5	102	1.7 ± 1.2	
MCA	1	49	27.7 ± 1.9 [#]		TPA	0.05	117	26.7 ± 2.9 [#]
Sodium arsenite (7784-46-5)				±				+
	0 (5% Water)	100	0.5 ± 0.5		0 (5% Water)	100	4.7 ± 1.9	
	0.056	117	0.5 ± 0.5		0.01	92	3.3 ± 1.4	
	0.1	109	1.2 ± 0.8		0.056	100	6.3 ± 1.0	
	0.18	76	2.2 ± 1.2		0.1	94	10.2 ± 4.5 ⁺	
	0.32	33	2.8 ± 2.0 ⁺		0.16	100	17.0 ± 5.0 ⁺	
	0.56	10	1.3 ± 1.2		0.18	93	23.3 ± 5.7 ⁺	
	1	0	Toxic		0.25	93	12.0 ± 3.3 ⁺	
	0 (0.1% DMSO)	100	1.3 ± 1.4		0.32	97	5.3 ± 2.3	
	MCA	1	48	21.0 ± 4.0 [#]		TPA	0.05	121
Sodium saccharin (128-44-9)				–				+
	0 (5% Water)	100	3.7 ± 1.9		0 (5% Water)	100	4.7 ± 1.4	
	410	100	3.2 ± 1.2		410	95	4.5 ± 0.8	
	821	100	3.3 ± 1.2		821	85	4.0 ± 1.4	
	1231	107	3.2 ± 1.8		1231	75	6.0 ± 2.8	
	1641	108	2.8 ± 1.3		1641	71	11.3 ± 2.7 ⁺	
	1847	109	2.3 ± 1.2		1847	70	15.2 ± 4.6 ⁺	
	2052 (10 mM)	108	3.8 ± 2.0		2052 (10 mM)	70	18.3 ± 6.6 ⁺	
	0 (0.1% DMSO)	100	3.2 ± 2.5		0 (0.1% DMSO)	100	6.0 ± 2.8	
	MCA	1	57	49.2 ± 5.5 [#]		TPA	0.05	146
Styrene oxide (96-09-3)				–				+
	0 (0.1% DMSO)	100	1.3 ± 0.8		0 (0.1% DMSO)	100	4.5 ± 2.6	
	10	107	1.3 ± 1.5		10	94	5.8 ± 3.3	
	15	111	0.8 ± 1.2		20	97	5.8 ± 1.0	
	20	111	3.2 ± 2.3		30	95	12.7 ± 4.2 ⁺	
	25	104	3.2 ± 2.5		40	91	13.7 ± 5.6 ⁺	
	30	102	3.5 ± 1.9		50	88	11.8 ± 7.3 ⁺	
	40	104	4.0 ± 3.6		70	85	Toxic	
	50	96	3.7 ± 3.1		TPA	0.05	121	19.5 ± 3.9 [#]

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay				
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment	
MCA	1	49	32.5 ± 5.0 [#]						
12-O-Tetradecanoylphorbol-13-acetate (TPA) (16561-29-8)				±				+	
	0 (0.1% DMSO)	100	0.8 ± 1.2		0 (0.1% DMSO)	100	1.3 ± 1.4		
	0.0003	97	1.8 ± 1.2		0.0003	102	1.8 ± 0.8		
	0.001	81	1.3 ± 1.5		0.001	112	2.7 ± 1.0		
	0.003	83	0.7 ± 0.5		0.003	117	3.7 ± 2.9		
	0.01	84	0.2 ± 0.4		0.01	139	8.2 ± 3.7 [†]		
	0.03	86	0.5 ± 0.5		0.03	143	13.0 ± 2.5 [†]		
	0.05	76	0.3 ± 0.8		0.05	164	13.8 ± 2.3 [†]		
	0.1	81	0.5 ± 0.8		0.1	194	12.7 ± 4.9 [†]		
	0.3	75	0.5 ± 0.5		0.3	212	20.8 ± 5.5 [†]		
	0.5	85	1.3 ± 1.2		0.5	220	24.3 ± 2.3 [†]		
	1	88	1.5 ± 1.0		1	231	34.8 ± 5.5 [†]		
	3	88	2.8 ± 2.2 [†]		3	211	47.5 ± 5.2 [†]		
MCA	1	36	19.8 ± 2.5 [#]		TPA 0.05	158	10.3 ± 2.7 [#]		
o-Toluidine (95-53-4)				–				+	
	0 (0.5% DMSO)	100	1.8 ± 1.7		0 (0.5% DMSO)	100	4.8 ± 1.5		
	100	99	1.7 ± 1.6		100	82	5.3 ± 1.6		
	200	92	3.5 ± 2.3		200	69	2.5 ± 2.9		
	300	87	1.5 ± 1.2		300	63	8.3 ± 1.9 [†]		
	400	76	4.5 ± 2.0		400	54	8.0 ± 1.3 [†]		
	600	50	3.3 ± 2.6		600	39	1.8 ± 1.5		
	800	36	2.7 ± 2.0		800	40	0.7 ± 0.8		
	0 (0.1% DMSO)	100	4.2 ± 1.3		0 (0.1% DMSO)	100	7.0 ± 2.6		
	MCA	1	59	17.5 ± 2.3 [#]		TPA 0.05	142	15.3 ± 4.4 [#]	
Zinc chloride (7646-85-7)				–				+	
	0 (5% Water)	100	0.8 ± 0.8		0 (5% Water)	100	1.3 ± 1.0		
	7	94	0.7 ± 0.5		1	112	1.3 ± 1.2		
	10	104	0.0 ± 0.0		3	126	2.2 ± 1.5		
	11	98	0.7 ± 0.8		7	131	4.0 ± 2.0		
	12	73	0.2 ± 0.4		10	162	14.0 ± 3.7 [†]		
	13	21	0.5 ± 0.8		11	162	15.3 ± 2.5 [†]		
	14	3	0.0 ± 0.0		13	175	19.8 ± 4.4 [†]		
	15	1	Toxic		15	100	8.5 ± 3.2 [†]		
	0 (0.1% DMSO)	100	1.3 ± 1.5		0 (0.1% DMSO)	100	2.5 ± 1.5		
	MCA	1	47	25.0 ± 6.2 [#]		TPA 0.05	47	11.2 ± 2.9 [#]	
	o-Anisidine (90-04-0)				–				–
		0 (0.1% DMSO)	100	2.0 ± 0.9		0 (0.1% DMSO)	100	3.5 ± 2.2	
		60	100	2.0 ± 1.8		10	91	2.7 ± 0.5	
		80	97	3.8 ± 2.8		30	89	1.3 ± 1.9	
100		79	2.7 ± 0.8		70	88	1.0 ± 1.1		
130		58	3.0 ± 2.0		130	60	1.7 ± 1.2		
170		32	4.2 ± 1.8		170	67	4.3 ± 3.1		
220		10	4.0 ± 1.3		220	53	4.3 ± 3.1		
260		8	4.3 ± 2.2		TPA 0.05	169	25.7 ± 2.3 [#]		
MCA		1	49	29.5 ± 4.2 [#]					
Benzene (71-43-2)				–				–	
	0 (0.5% DMSO)	100	2.3 ± 2.5		0 (0.5% DMSO)	100	2.7 ± 1.4		
	31.25	94	3.7 ± 3.3		31.25	95	2.3 ± 2.1		
	62.5	97	1.0 ± 1.3		62.5	104	3.8 ± 2.5		
	125	106	0.8 ± 1.6		125	111	3.8 ± 3.1		
	250	100	1.5 ± 1.4		250	107	5.0 ± 3.2		
	500	104	1.8 ± 1.5		500	89	3.3 ± 2.0		
	750	98	2.3 ± 2.0		750	104	1.7 ± 1.2		
	1000 (12.8 mM)	109	2.2 ± 0.8		1000 (12.8 mM)	100	2.3 ± 1.2		
	MCA	1	34	19.7 ± 6.4 [#]		TPA 0.05	106	15.8 ± 4.5 [#]	
Cobalt sulfate heptahydrate (10026-24-1)				–				–	
	0 (5% Water)	100	4.8 ± 1.8		0 (5% Water)	100	7.5 ± 3.1		
	1	101	4.3 ± 2.2		1	105	6.0 ± 3.0		
	3	113	2.8 ± 1.7		3	101	6.8 ± 1.5		
	10	125	2.3 ± 1.9		10	99	6.0 ± 1.1		
	15	133	3.0 ± 1.3		15	104	8.3 ± 3.6		
	20	126	3.0 ± 1.4		20	98	6.3 ± 0.8		
	30	118	3.5 ± 1.5		30	96	4.7 ± 1.9		
	40	84	3.7 ± 1.4		40	95	0.7 ± 0.8		
	0 (0.1% DMSO)	100	3.8 ± 1.3		0 (0.1% DMSO)	100	5.2 ± 1.2		
	MCA	1	60	39.8 ± 4.8 [#]		TPA 0.05	131	34.2 ± 6.5 [#]	
	Diethylstilbestrol (56-53-1)				–				–

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay			
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment
MCA	0 (0.1% DMSO)	100	2.3 ± 2.6		0 (0.1% DMSO)	100	7.2 ± 3.5	
	0.7	94	2.8 ± 1.6		0.01	94	4.5 ± 1.9	
	1	94	1.8 ± 1.9		0.022	95	3.8 ± 1.3	
	1.3	67	4.8 ± 3.3		0.046	94	5.0 ± 1.7	
	1.5	39	4.2 ± 2.6		0.1	95	3.3 ± 1.4	
	1.7	16	3.3 ± 1.9		0.22	90	3.8 ± 2.9	
	2	11	3.0 ± 1.9		0.46	84	2.3 ± 2.2	
	3	20	3.2 ± 2.4		1	68	0.0 ± 0.0	
	1	48	42.8 ± 5.2 [#]		TPA 0.05	136	20.0 ± 2.5 [#]	
	Dimethylarsinic acid (75-60-5)	0 (5% Water)	100	3.3 ± 1.6	–	0 (5% Water)	100	2.8 ± 1.2
140		119	2.3 ± 2.1		0.3	98	1.8 ± 1.2	
200		88	2.3 ± 1.2		0.7	94	2.7 ± 2.0	
280		60	2.0 ± 1.3		1	93	2.7 ± 2.3	
400		35	1.8 ± 1.0		3	94	0.8 ± 1.2	
630		21	2.0 ± 1.3		7	92	0.8 ± 0.4	
1000		13	1.3 ± 1.6		10	94	0.8 ± 0.4	
1380		9	0.5 ± 0.5		30	94	0.0 ± 0.0	
0 (0.1% DMSO)		100	3.3 ± 2.0		0 (0.1% DMSO)	100	1.8 ± 1.8	
1		60	23.3 ± 4.2 [#]		TPA 0.05	118	17.0 ± 3.2 [#]	
1,4-Dioxane (123-91-1)	0 (5% Water)	100	1.2 ± 0.4	–	0 (5% Water)	100	3.7 ± 1.5	–
	150	102	1.7 ± 1.6		150	100	2.7 ± 1.5	
	300	91	1.7 ± 1.4		300	101	2.7 ± 2.0	
	625	97	1.7 ± 1.9		625	99	3.2 ± 1.0	
	1250 (14.2 mM)	95	1.8 ± 1.5		1250 (14.2 mM)	94	3.0 ± 1.4	
	0 (0.1% DMSO)	100	1.8 ± 0.8		0 (0.1% DMSO)	100	2.8 ± 1.3	
	1	36	12.0 ± 3.9 [#]		TPA 0.05	119	23.8 ± 3.6 [#]	
Ethyl carbamate (Urethane) (51-79-6)	0 (5% Water)	100	1.3 ± 1.9	–	0 (5% Water)	100	1.2 ± 1.2	–
	89	104	1.0 ± 1.3		89	100	1.8 ± 0.8	
	151	102	0.7 ± 0.5		151	99	1.7 ± 1.0	
	294	97	0.7 ± 0.8		294	95	1.3 ± 0.8	
	445	96	0.7 ± 0.8		445	94	1.5 ± 1.4	
	597	90	1.5 ± 1.5		597	90	1.2 ± 1.0	
	739	95	0.7 ± 0.8		739	93	0.8 ± 0.8	
	891 (10 mM)	92	1.3 ± 0.8		891 (10 mM)	89	1.5 ± 1.6	
	0 (0.1% DMSO)	100	1.5 ± 1.5		0 (0.1% DMSO)	100	3.0 ± 1.5	
	1	40	23.8 ± 7.5 [#]		TPA 0.05	114	13.7 ± 3.4 [#]	
Formaldehyde (50-00-0)	0 (5% Water)	100	1.3 ± 0.5	–	0 (5% Water)	100	2.3 ± 1.9	–
	0.5	102	1.7 ± 0.8		0.1	98	1.0 ± 0.6	
	0.7	103	0.3 ± 0.5		0.15	97	0.7 ± 0.8	
	0.8	104	1.0 ± 1.3		0.2	98	2.0 ± 1.4	
	1	90	1.7 ± 0.8		0.3	105	1.8 ± 1.2	
	1.2	70	1.3 ± 1.0		0.5	90	0.8 ± 0.8	
	1.4	25	1.3 ± 1.0		0.7	95	1.7 ± 0.8	
	1.6	11	1.5 ± 1.0		1	99	0.7 ± 0.8	
	1.8	6	1.0 ± 1.5		1.4	97	1.2 ± 0.8	
	2	6	0.8 ± 1.2		2	96	1.0 ± 1.3	
	0 (0.1% DMSO)	100	1.0 ± 0.9		3	99	1.0 ± 0.9	
	1	53	19.2 ± 1.3 [#]		5	96	0.0 ± 0.0	
					7	47	Toxic	
					10	13	Toxic	
					0 (0.1% DMSO)	100	2.3 ± 2.0	
					TPA 0.05	134	12.2 ± 4.1 [#]	
	Furylfuramide (AF-2) (3688-53-7)	0 (0.1% DMSO)	100	2.3 ± 2.3	–	0 (0.1% DMSO)	100	6.5 ± 1.9
1		88	2.8 ± 1.7		0.6	95	7.7 ± 3.7	
1.25		90	2.8 ± 1.5		1.25	94	3.2 ± 1.6	
1.5		75	3.7 ± 2.9		2	92	2.7 ± 1.6	
1.75		61	3.8 ± 1.2		3	88	1.2 ± 1.0	
2		52	3.0 ± 0.9		5	100	0.5 ± 0.5	
2.25		24	3.2 ± 1.9		7.5	104	0.2 ± 0.4	
2.5		12	1.5 ± 0.5		10	94	Toxic	
1		43	28.7 ± 3.1 [#]		20	21	Toxic	
					TPA 0.05	151	33.0 ± 6.2 [#]	
Methyl carbamate (Methyl urethane) (598-55-0)	0 (5% Water)	100	1.2 ± 0.8	–	0 (5% Water)	100	3.2 ± 2.0	–

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay				
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment	
MCA	100	79	1.3 ± 0.8		100	101	3.2 ± 2.1		
	200	74	3.0 ± 1.8		200	94	3.0 ± 2.8		
	500	81	1.2 ± 1.5		500	104	3.0 ± 2.1		
	1000 (13.3 mM)	78	2.2 ± 1.3		1000 (13.3 mM)	99	2.2 ± 1.7		
	0 (0.1% DMSO)	100	1.3 ± 0.5		0 (0.1% DMSO)	100	3.2 ± 2.2		
	1	48	37.3 ± 6.2 [#]		TPA 0.05	152	28.5 ± 3.1 [#]		
Nickel (II) chloride (7718-54-9)	0 (5% Water)	100	1.5 ± 1.2	–	0 (5% Water)	100	0.8 ± 1.2	–	
	0.69	114	2.3 ± 1.5		0.4	114	1.8 ± 1.3		
	1	102	0.5 ± 0.5		0.6	116	1.5 ± 1.0		
	1.5	114	1.0 ± 1.3		1	121	1.0 ± 2.3		
	2.2	109	1.2 ± 1.0		1.5	119	1.2 ± 1.2		
	3.2	104	1.2 ± 1.2		2.5	126	0.8 ± 0.8		
	4.7	87	1.8 ± 1.5		4	126	Toxic		
	6.9	53	1.5 ± 1.9		6	126	Toxic		
	10	14	0.7 ± 0.8		0 (0.1% DMSO)	100	1.2 ± 0.8		
	16	0	Toxic		TPA 0.05	138	10.2 ± 1.5 [#]		
	25	0	Toxic						
	0 (0.1% DMSO)	100	1.2 ± 1.5						
	1	14	17.8 ± 5.6 [#]						
	Nickel monooxide (1313-99-1)	0 (5% Water)	100	3.0 ± 0.9	–	0 (5% Water)	100	3.5 ± 2.1	–
1.875		104	2.5 ± 1.6		1.875	104	2.0 ± 1.7		
3.75		105	1.3 ± 1.4		3.75	108	2.5 ± 1.9		
7.5		115	3.0 ± 1.9		7.5	100	1.2 ± 1.5		
10		107	2.5 ± 1.9		10	98	0.8 ± 0.8		
15		117	1.3 ± 1.4		15	87	0.8 ± 0.8		
20		105	2.3 ± 1.9		20	93	Toxic		
30		63	1.0 ± 1.1		30	87	Toxic		
60		34	0.5 ± 0.8		0 (0.1% DMSO)	100	3.8 ± 1.0		
0 (0.1% DMSO)		100	4.3 ± 2.3		TPA 0.05	112	28.5 ± 4.1 [#]		
1		60	40.3 ± 4.8 [#]						
Phenobarbital sodium salt (57-30-7)		0 (5% Water)	100	3.3 ± 2.7	–	0 (5% Water)	100	2.8 ± 1.5	–
		100	106	3.8 ± 1.2		100	94	1.0 ± 1.1	
		300	109	3.0 ± 1.3		300	85	1.3 ± 0.8	
	500	104	2.7 ± 1.0		500	81	0.7 ± 0.8		
	700	79	1.7 ± 1.9		700	77	1.0 ± 1.3		
	1000	49	1.5 ± 1.0		1000	77	0.2 ± 0.4		
	1500	15	0.8 ± 1.2		1500	60	0.8 ± 0.8		
	2000	3	0.0 ± 0.0		2000	31	Toxic		
	0 (0.1% DMSO)	100	3.2 ± 1.6		0 (0.1% DMSO)	100	2.3 ± 2.0		
	1	62	29.2 ± 3.7 [#]		TPA 0.05	131	14.8 ± 3.2 [#]		
	<i>p</i> -Toluidine (106-49-0)	0 (0.1% DMSO)	100	2.7 ± 0.8	–	0 (0.1% DMSO)	100	2.8 ± 2.6	–
		10	101	1.8 ± 2.2		5	89	3.0 ± 0.6	
		25	92	3.0 ± 0.9		10	97	2.5 ± 1.8	
		50	88	3.0 ± 0.9		25	87	2.5 ± 2.3	
75		84	2.7 ± 2.3		50	84	1.8 ± 1.3		
100		81	3.0 ± 2.3		75	82	1.7 ± 0.5		
150		65	2.3 ± 1.6		150	80	1.0 ± 0.9		
200		32	4.5 ± 2.3		300	62	3.5 ± 1.8		
300		13	5.2 ± 3.5		TPA 0.05	117	17.8 ± 2.2 [#]		
1		42	42.0 ± 8.2 [#]						
Non-carcinogens 4-Acetylaminofluorene (28322-02-3)		0 (0.1% DMSO)	100	0.8 ± 1.2	–	0 (0.1% DMSO)	100	1.5 ± 0.8	–
		50	115	1.3 ± 1.0		5	85	1.3 ± 1.2	
		75	106	1.7 ± 1.0		10	78	1.0 ± 0.9	
		100	92	1.7 ± 1.2		25	70	1.3 ± 1.4	
	120	87	2.0 ± 1.7		50	59	0.2 ± 0.4		
	135	73	2.2 ± 2.0		100	54	0.0 ± 0.0		
	150	76	2.5 ± 1.4		150	60	0.0 ± 0.0		
	1	44	19.5 ± 4.1 [#]		TPA 0.05	129	13.8 ± 2.6 [#]		
	Acid red 14 (Chromotrope FB) (3567-69-9)	0 (5% Water)	100	4.2 ± 3.8	–	0 (5% Water)	100	4.8 ± 2.1	–
		30	94	5.8 ± 3.7		30	105	3.7 ± 1.8	
50		98	4.5 ± 3.2		70	110	2.7 ± 1.4		

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay			
	Concentration (µg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (µg/mL)	CG	Foci/well	Judgment
MCA	100	105	4.2 ± 1.7		100	100	2.0 ± 2.1	
	300	150	4.5 ± 2.4		300	97	4.7 ± 3.1	
	500	104	7.3 ± 2.7		700	97	4.3 ± 1.0	
	700	82	6.5 ± 1.4		1000	106	5.2 ± 3.4	
	1000	24	2.2 ± 1.6		1500	110	5.3 ± 1.9	
	0 (0.1% DMSO)	100	2.3 ± 1.5		0 (0.1% DMSO)	100	5.3 ± 2.7	
	1	62	35.7 ± 2.1 [#]		TPA 0.05	122	27.2 ± 4.1 [#]	
Ampicillin sodium salt (69-52-3)				–				–
	0 (5% Water)	100	1.5 ± 0.8		0 (5% Water)	100	5.0 ± 1.9	
	300	102	2.2 ± 1.6		700	102	7.2 ± 1.3	
	700	99	1.5 ± 1.8		1000	95	3.5 ± 2.3	
	1000	67	1.7 ± 1.2		1500	98	4.8 ± 2.3	
	1200	29	1.0 ± 0.9		3000	90	4.2 ± 3.1	
	1250	27	1.3 ± 1.2		5000	87	Toxic	
	1300	17	2.3 ± 1.0		0 (0.1% DMSO)	100	8.8 ± 4.2	
	1500	6	3.5 ± 1.8		TPA 0.05	120	28.5 ± 5.0 [#]	
	1750	2	1.8 ± 1.0					
MCA	0 (0.1% DMSO)	100	1.7 ± 1.9					
	1	42	32.7 ± 8.4 [#]					
Anthracene (120-12-7)				–				–
	0 (0.5% DMSO)	100	5.3 ± 1.5		0 (0.5% DMSO)	100	6.3 ± 1.0	
	2.5	96	7.7 ± 2.7		2.5	116	6.7 ± 2.2	
	5	89	7.3 ± 1.5		5	106	6.3 ± 2.1	
	10	108	7.3 ± 3.8		10	111	6.8 ± 2.4	
	25	99	6.8 ± 1.5		25	121	6.0 ± 1.8	
	50	105	8.2 ± 1.5		50	115	7.2 ± 3.3	
	100	100	7.3 ± 1.8		100	114	6.2 ± 2.1	
MCA	0 (0.1% DMSO)	100	5.7 ± 2.0		0 (0.1% DMSO)	100	8.8 ± 3.5	
	1	37	23.8 ± 3.9 [#]		TPA 0.05	141	27.2 ± 2.1 [#]	
L-Ascorbic acid (50-81-7)				–				–
	0 (5% Water)	100	3.0 ± 3.2		0 (5% Water)	100	3.0 ± 1.3	
	80	94	2.5 ± 1.0		100	104	1.0 ± 0.9	
	100	88	2.0 ± 1.3		140	107	0.5 ± 0.5	
	130	79	1.5 ± 1.0		190	106	0.5 ± 0.5	
	170	64	3.8 ± 1.7		260	103	0.2 ± 0.4	
	220	33	3.0 ± 1.3		370	101	0.8 ± 1.0	
	280	20	4.0 ± 2.4		520	97	Toxic	
	370	4	3.2 ± 1.5		720	45	Toxic	
MCA	0 (0.1% DMSO)	100	1.2 ± 1.2		0 (0.1% DMSO)	100	3.5 ± 1.6	
	1	49	26.5 ± 7.3 [#]		TPA 0.05	119	22.5 ± 2.4 [#]	
Aspartame (22839-47-0)				–				–
	0 (5% Water)	100	1.0 ± 0.6		0 (5% Water)	100	2.3 ± 1.2	
	80	97	1.3 ± 0.8		80	107	2.3 ± 1.2	
	100	107	1.7 ± 2.0		100	120	1.8 ± 1.8	
	200	100	0.3 ± 0.5		200	106	2.8 ± 1.8	
	500	109	1.3 ± 1.2		500	108	2.3 ± 1.0	
	1000	103	1.3 ± 1.0		1000	101	1.5 ± 1.0	
	2000	111	1.8 ± 1.2		2000	94	0.0 ± 0.0	
	3000 (10.2 mM)	99	1.3 ± 0.8		3000 (10.2 mM)	79	0.2 ± 0.4	
MCA	0 (0.1% DMSO)	100	0.8 ± 0.8		0 (0.1% DMSO)	100	1.3 ± 0.8	
	1	49	27.7 ± 4.6 [#]		TPA 0.05	150	21.7 ± 2.9 [#]	
Benzoin (119-53-9)				–				–
	0 (0.1% DMSO)	100	8.8 ± 1.0		0 (0.1% DMSO)	100	3.8 ± 1.9	
	1	94	9.8 ± 4.4		12.5	92	4.3 ± 2.0	
	10	98	9.3 ± 2.5		25	86	4.5 ± 1.9	
	30	102	11.2 ± 4.0		50	85	4.2 ± 2.0	
	50	109	7.7 ± 2.7		100	83	1.5 ± 0.8	
	70	106	10.0 ± 1.8		200	84	0.0 ± 0.0	
	100	105	8.3 ± 4.6		400	87	0.2 ± 0.4	
	200	67	10.2 ± 2.3		800	82	0.8 ± 1.0	
MCA	1	66	42.2 ± 3.5 [#]		TPA 0.05	158	14.5 ± 2.9 [#]	
Caffeine (58-08-2)				–				–
	0 (5% Water)	100	0.5 ± 0.5		0 (5% Water)	100	3.5 ± 2.2	
	50	108	1.7 ± 2.1		30	105	2.7 ± 0.8	
	75	89	1.5 ± 0.8		50	95	1.5 ± 1.4	
	100	95	1.2 ± 0.8		75	94	1.5 ± 1.4	
	150	79	1.2 ± 1.5		100	83	1.2 ± 1.5	
	200	68	0.7 ± 1.2		150	77	1.0 ± 1.1	
	300	48	1.0 ± 0.6		200	70	0.0 ± 0.0	
	400	13	1.5 ± 1.4		300	60	0.0 ± 0.0	
	500	3	Toxic		400	52	0.0 ± 0.0	

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay				
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment	
MCA	30	90	4.3 ± 1.4		75	91	7.8 ± 3.9		
	70	92	3.2 ± 1.6		150	88	7.8 ± 2.1		
	100	87	2.7 ± 1.8		300	84	4.7 ± 1.9		
	200	84	4.0 ± 2.1		500	77	2.3 ± 1.5		
	400	81	4.3 ± 1.8		750	73	0.2 ± 0.4		
	700	87	3.8 ± 1.5		1000	78	Toxic		
	1000	44	Toxic		1250	78	Toxic		
	0 (0.1% DMSO)	100	3.7 ± 2.6		1500	76	Toxic		
	1	49	32.2 ± 4.4 [#]		0 (0.1% DMSO)	100	9.3 ± 2.3		
					TPA 0.05	143	39.0 ± 5.8 [#]		
Hydrocortisone (50-23-7)	0 (0.5% DMSO)	100	6.5 ± 1.6	–	0 (0.5% DMSO)	100	0.8 ± 0.4	–	
	1	84	6.8 ± 2.3		3	81	2.8 ± 1.8		
	10	75	7.0 ± 2.8		10	85	2.5 ± 2.1		
	50	79	7.2 ± 1.6		25	79	2.7 ± 2.3		
	100	75	6.5 ± 2.4		50	109	1.7 ± 1.4		
	150	68	4.3 ± 2.2		100	90	1.2 ± 1.5		
	300	25	4.2 ± 1.6		250	96	1.0 ± 0.9		
	500	7	4.2 ± 1.9		500	45	Toxic		
	0 (0.1% DMSO)	100	3.5 ± 2.1		0 (0.1% DMSO)	100	1.0 ± 0.9		
	1	74	30.0 ± 7.5 [#]		TPA 0.05	152	10.8 ± 2.8 [#]		
D-Mannitol (69-65-8)	0 (5% Water)	100	0.5 ± 0.5	–	0 (5% Water)	100	2.5 ± 1.2	–	
	100	105	0.5 ± 0.8		100	112	2.8 ± 1.2		
	200	101	1.8 ± 1.2		200	107	1.7 ± 1.4		
	500	101	0.3 ± 0.5		500	108	3.5 ± 1.4		
	1000	102	0.7 ± 0.5		1000	102	4.0 ± 2.7		
	2000 (11.0 mM)	103	0.8 ± 0.8		2000 (11.0 mM)	116	2.7 ± 1.2		
	0 (0.1% DMSO)	100	1.5 ± 0.5		0 (0.1% DMSO)	100	2.8 ± 1.8		
	1	42	13.2 ± 3.3 [#]		TPA 0.05	227	17.3 ± 3.4 [#]		
	Methotrexate (59-05-2)	0 (5% Water)	100	1.5 ± 2.1	–	0 (5% Water)	100	2.3 ± 2.3	–
		0.01	88	1.5 ± 1.2		0.0003	96	1.5 ± 1.0	
0.03		105	1.5 ± 1.4		0.001	98	0.8 ± 0.8		
0.1		94	0.8 ± 0.8		0.003	91	2.5 ± 1.9		
0.2		102	0.8 ± 0.8		0.01	102	1.2 ± 1.2		
0.5		105	0.7 ± 0.8		0.03	94	1.0 ± 1.5		
1		40	0.2 ± 0.4		0.1	84	0.0 ± 0.0		
3		44	Toxic		0.3	77	0.0 ± 0.0		
0 (0.1% DMSO)		100	1.0 ± 0.6		0 (0.1% DMSO)	100	2.5 ± 1.0		
1		50	17.2 ± 2.9 [#]		TPA 0.05	122	12.2 ± 3.3 [#]		
1-Naphthylamine (134-32-7)	0 (0.1% DMSO)	100	5.5 ± 2.4	–	0 (0.1% DMSO)	100	5.0 ± 4.1	–	
	2.5	100	6.5 ± 2.4		2.5	90	6.0 ± 3.1		
	5	96	5.7 ± 2.4		5	94	6.3 ± 1.5		
	10	93	3.8 ± 1.5		10	83	5.2 ± 2.3		
	20	79	6.8 ± 3.3		20	80	3.5 ± 1.9		
	30	64	8.5 ± 1.5		30	77	1.0 ± 0.9		
	50	34	4.5 ± 2.1		50	60	2.2 ± 2.0		
	70	15	3.0 ± 2.1		100	43	Toxic		
	100	17	2.8 ± 2.5		TPA 0.05	118	21.8 ± 6.2 [#]		
	1	56	34.7 ± 3.5 [#]						
Phenanthrene (85-01-8)	0 (0.1% DMSO)	100	4.0 ± 3.0	–	0 (0.1% DMSO)	100	6.2 ± 1.8	–	
	10	97	4.5 ± 2.7		2.5	99	5.7 ± 2.8		
	20	87	4.8 ± 1.9		5	101	7.2 ± 1.9		
	40	70	4.3 ± 2.0		10	90	8.7 ± 1.9		
	60	77	4.5 ± 2.9		20	84	8.8 ± 2.9		
	100	103	5.8 ± 2.4		40	83	7.8 ± 2.7		
	150	98	4.7 ± 2.4		60	85	2.7 ± 1.5		
	1	44	27.8 ± 5.0 [#]		TPA 0.05	91	29.7 ± 4.2 [#]		
	Phenol (108-95-2)	0 (5% Water)	100	1.8 ± 1.8	–	0 (5% Water)	100	3.5 ± 1.4	–
		50	89	2.2 ± 1.6		5	98	2.3 ± 1.2	
75		88	1.3 ± 2.0		10	99	1.8 ± 1.5		
100		71	1.0 ± 1.5		25	93	1.0 ± 0.6		
125		61	3.2 ± 1.3		50	84	0.8 ± 0.8		
150		48	1.5 ± 1.0		100	75	1.3 ± 1.0		
175		18	1.8 ± 1.0		200	51	0.2 ± 0.4		
200		20	1.3 ± 1.2		300	45	Toxic		
300		4	2.8 ± 1.2		500	38	Toxic		

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay				
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment	
MCA	19	83	2.2 ± 1.9		5	101	5.5 ± 2.6		
	27	82	2.8 ± 2.9		10	98	2.8 ± 1.5		
	37	82	1.3 ± 2.0		50	83	0.0 ± 0.0		
	50	76	3.8 ± 2.6		100	72	Toxic		
	75	36	3.7 ± 2.3		500	73	Toxic		
	1	68	18.7 ± 3.9 [#]		TPA 0.05	139	19.5 ± 6.0 [#]		
<i>m</i> -Toluidine (108-44-1)	0 (0.1% DMSO)	100	2.0 ± 1.3	–	0 (0.1% DMSO)	100	7.0 ± 2.2	–	
	100	104	2.7 ± 1.4		100	74	5.3 ± 2.3		
	200	96	2.3 ± 2.0		200	67	3.0 ± 2.8		
	300	87	2.8 ± 1.6		300	54	5.0 ± 2.1		
	400	54	2.2 ± 1.5		400	60	5.2 ± 2.1		
	500	63	2.8 ± 1.2		500	44	2.8 ± 0.4		
	600	40	2.7 ± 1.6		600	60	5.0 ± 1.7		
	800	33	2.0 ± 1.4		700	63	2.2 ± 2.0		
	1	45	25.3 ± 4.0 [#]		TPA 0.05	107	22.5 ± 4.7 [#]		
Triphenyltin hydroxide (76-87-9)	0 (0.1% DMSO)	100	4.2 ± 1.9	–	0 (0.1% DMSO)	100	5.7 ± 2.4	–	
	0.01	88	2.2 ± 1.6		0.001	138	6.2 ± 1.9		
	0.015	88	2.0 ± 1.8		0.003	131	3.5 ± 1.6		
	0.017	82	3.0 ± 1.3		0.005	137	4.5 ± 1.9		
	0.02	72	1.7 ± 1.5		0.01	131	4.3 ± 0.8		
	0.023	49	1.2 ± 1.2		0.03	119	5.3 ± 2.2		
	0.025	10	1.7 ± 1.9		0.07	117	Toxic		
	0.03	5	0.8 ± 1.2		0.08	64	Toxic		
	0.04	5	Toxic		TPA 0.05	153	31.8 ± 3.3 [#]		
	1	43	32.8 ± 6.5 [#]						
	Barium chloride dihydrate (10326-27-9)	0 (5% Water)	100	5.3 ± 1.9	–	0 (5% Water)	100	3.5 ± 2.2	+
		500	45	5.7 ± 3.6		100	114	4.7 ± 2.1	
		600	37	6.5 ± 2.1		300	132	9.8 ± 0.8 [*]	
700		42	2.8 ± 2.7		500	144	10.0 ± 2.4 [*]		
1000		27	4.3 ± 1.0		1000	120	6.3 ± 2.7		
1500		25	3.2 ± 1.9		2000	83	11.7 ± 3.7 [*]		
3000 (12.3 mM)		12	5.8 ± 3.7		2500 (10.2 mM)	67	11.8 ± 2.9 [*]		
0 (0.1% DMSO)		100	5.3 ± 3.2		0 (0.1% DMSO)	100	3.3 ± 1.8		
1		72	32.0 ± 2.4 [#]		TPA 0.05	144	32.7 ± 4.8 [#]		
<i>tert</i> -Butylhydroquinone (1948-33-0)		0 (0.1% DMSO)	100	1.2 ± 1.0	–	0 (0.1% DMSO)	100	8.3 ± 2.4	+
		0.1	96	0.8 ± 0.8		1	102	12.0 ± 2.8	
	0.3	92	1.5 ± 0.8		2	95	24.5 ± 5.0 [*]		
	0.7	91	1.8 ± 1.2		3	104	38.7 ± 2.7 [*]		
	1	95	0.5 ± 0.5		4	130	49.7 ± 5.2 [*]		
	1.3	91	1.0 ± 1.3		5	138	33.0 ± 4.1 [*]		
	1.7	66	0.5 ± 0.5		6	130	17.8 ± 3.9 [*]		
	2	26	0.5 ± 0.5		8	79	Toxic		
	1	46	17.8 ± 4.4 [#]		TPA 0.05	140	27.3 ± 4.0 [#]		
	8-Hydroxyquinoline (148-24-3)	0 (0.1% DMSO)	100	6.3 ± 3.3	–	0 (0.1% DMSO)	100	7.2 ± 1.8	+
		0.03	100	6.5 ± 1.5		0.3	98	5.3 ± 2.7	
0.06		101	6.8 ± 2.6		1	105	10.2 ± 2.5		
0.09		103	7.8 ± 3.9		2	106	17.2 ± 2.3 [*]		
0.12		104	5.3 ± 3.0		3	123	20.7 ± 3.4 [*]		
0.15		102	4.5 ± 2.0		5	131	21.3 ± 3.6 [*]		
0.18		112	4.3 ± 1.2		7	67	Toxic		
0.21		113	4.3 ± 1.6		9	85	Toxic		
0.24		49	2.8 ± 1.2		11	55	Toxic		
0.28		72	8.3 ± 3.6		13	35	Toxic		
1		76	41.5 ± 2.8 [#]		TPA 0.05	136	23.0 ± 3.4 [#]		
Propyl gallate (121-79-9)		0 (0.1% DMSO)	100	0.8 ± 1.3	+	0 (0.1% DMSO)	100	6.2 ± 2.2	+
		1	71	3.2 ± 1.5 [*]		0.5	98	7.5 ± 1.9	
	1.25	32	3.3 ± 1.4 [*]		0.8	98	9.0 ± 3.6		
	1.5	17	3.5 ± 1.4 [*]		1.3	91	9.3 ± 2.7		
	1.75	7	2.3 ± 1.2		2	83	13.8 ± 6.1 [*]		
	2	5	3.0 ± 1.7		3	80	13.8 ± 2.4 [*]		
	2.25	3	1.5 ± 1.8		5	64	19.0 ± 7.0 [*]		
	2.5	2	1.2 ± 1.0		7.5	26	Toxic		
	1	50	11.0 ± 0.6 [#]		10	16	Toxic		

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay						
	Concentration (μg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (μg/mL)	CG	Foci/well	Judgment			
Sodium fluoride (7681-49-4)				–	TPA	0.05	120	25.0 ± 3.6 [#]	+		
	0 (5% Water)	100	1.8 ± 2.8		0 (5% Water)	100	5.0 ± 2.2				
	4	107	1.8 ± 1.6		18	120	7.5 ± 1.2				
	6.2	105	0.7 ± 0.5		24	149	10.8 ± 2.6 [†]				
	10	112	1.0 ± 0.9		32	221	13.3 ± 1.5 [†]				
	16	98	1.3 ± 1.2		42	223	8.5 ± 2.3 [†]				
	25	92	2.0 ± 1.4		56	128	0.8 ± 0.8				
	40	13	1.2 ± 1.6		75	76	Toxic				
	62	0	0.5 ± 0.8		100	47	Toxic				
	100	0	0.0 ± 0.0		0 (0.1% DMSO)	100	4.0 ± 1.7				
MCA	0 (0.1% DMSO)	100	3.2 ± 1.5		TPA	0.05	143	14.7 ± 1.9 [#]			
	1	42	25.5 ± 2.6 [#]								
Tetracycline hydrochloride (64-75-5)				–					+		
	0 (5% Water)	100	1.5 ± 2.3		0 (5% Water)	100	3.7 ± 2.1				
	5	107	1.3 ± 1.4		5	101	8.2 ± 2.0 [†]				
	10	104	2.5 ± 1.9		10	117	11.3 ± 4.5 [†]				
	15	113	1.3 ± 1.0		15	121	5.7 ± 1.2				
	20	110	2.3 ± 1.2		20	130	3.3 ± 1.5				
	25	101	3.3 ± 2.6		25	129	8.3 ± 2.1 [†]				
	40	29	2.3 ± 2.1		40	107	25.3 ± 4.0 [†]				
	50	7	3.0 ± 1.5		50	61	1.0 ± 1.3				
	60	3	3.2 ± 1.2		60	36	0.0 ± 0.0				
	0 (0.1% DMSO)	100	1.5 ± 1.4		0 (0.1% DMSO)	100	5.7 ± 1.8				
	MCA	1	49	26.3 ± 3.3 [#]	TPA	0.05	145	32.2 ± 3.1 [#]			
	<i>Chemicals of unknown carcinogenicity</i>										
Sodium valproate (1069-66-5)				+					+		
	0 (5% Water)	100	2.2 ± 0.8		0 (5% Water)	100	2.8 ± 2.5				
	50	123	2.5 ± 1.4		6	141	12.8 ± 5.5 [†]				
	100	102	5.0 ± 2.5		10	161	23.0 ± 4.6 [†]				
	150	62	6.2 ± 1.7 [†]		25	183	29.5 ± 2.0 [†]				
	200	37	5.5 ± 2.4 [†]		50	228	9.7 ± 2.1 [†]				
	250	22	5.3 ± 1.2 [†]		100	206	0.8 ± 0.8				
	300	12	5.2 ± 2.4 [†]		200	164	0.0 ± 0.0				
	400	7	3.8 ± 1.6		500	83	Toxic				
	0 (0.1% DMSO)	100	2.5 ± 2.4		0 (0.1% DMSO)	100	5.7 ± 1.6				
	MCA	1	55	18.8 ± 2.6 [#]	TPA	0.05	138	20.3 ± 5.5 [#]			
	Valproic acid (99-66-1)				+						+
		0 (0.1% DMSO)	100	2.3 ± 1.9		0 (0.1% DMSO)	100	4.8 ± 1.5			
25		118	3.7 ± 0.5		3.125	107	10.3 ± 4.9 [†]				
50		119	5.0 ± 2.3		6.25	119	15.7 ± 2.7 [†]				
75		104	6.8 ± 1.5 [†]		12.5	138	28.7 ± 1.6 [†]				
100		80	6.0 ± 1.7 [†]		25	169	24.8 ± 4.1 [†]				
140		47	5.7 ± 2.9 [†]		50	193	9.5 ± 2.1 [†]				
200		21	5.5 ± 2.7		100	161	0.2 ± 0.4				
300		11	3.3 ± 2.2		200	125	0.0 ± 0.0				
MCA		1	43	20.3 ± 3.1 [#]	TPA	0.05	146	22.7 ± 2.3 [#]			
2,3-Diaminotoluene (2687-25-4)				–					+		
	0 (0.1% DMSO)	100	1.8 ± 1.3		0 (0.1% DMSO)	100	3.8 ± 2.7				
	3	83	3.3 ± 1.8		5	90	14.0 ± 2.6 [†]				
	5	78	3.5 ± 2.5		7	98	12.2 ± 1.9 [†]				
	7	75	4.7 ± 1.0		15	133	11.5 ± 4.8 [†]				
	15	58	4.2 ± 2.4		20	133	9.5 ± 2.9 [†]				
	20	47	3.8 ± 1.8		30	134	2.5 ± 3.0				
	30	34	4.2 ± 2.1		50	118	Toxic				
	50	4	0.8 ± 0.8		100	64	Toxic				
	MCA	1	47	11.2 ± 1.9 [#]	TPA	0.05	153	12.7 ± 3.6 [#]			
2,5-Diaminotoluene dihydrochloride (615-45-2)				–					+		
	0 (5% Water)	100	2.7 ± 1.0		0 (5% Water)	100	1.3 ± 1.0				
	1	98	3.5 ± 1.6		1	92	3.7 ± 2.2				
	2.5	96	2.7 ± 2.0		5	79	4.8 ± 2.8				
	4	100	3.7 ± 0.8		10	77	10.2 ± 2.1 [†]				
	5.5	90	3.8 ± 4.3		15	76	15.0 ± 0.9 [†]				
	7	95	2.7 ± 2.3		20	77	16.7 ± 3.5 [†]				
	8.5	86	4.2 ± 0.8		25	81	16.2 ± 4.5 [†]				
	10	79	3.2 ± 2.0		30	80	17.2 ± 2.8 [†]				
	11.5	84	4.0 ± 1.1		35	87	17.3 ± 4.4 [†]				
	13	76	3.7 ± 1.4		40	87	15.2 ± 3.5 [†]				
	0 (0.1% DMSO)	100	3.2 ± 1.8		0 (0.1% DMSO)	100	4.5 ± 1.9				

Table 1 (Continued)

Chemical ^a	Initiation assay				Promotion assay			
	Concentration (µg/mL)	CG ^b	Foci/well ^c	Judgment ^d	Concentration (µg/mL)	CG	Foci/well	Judgment
MCA	1	50	25.5 ± 3.8 [#]		TPA 0.05	152	10.3 ± 2.4 [#]	
3,4-Diaminotoluene (496-72-0)				–				+
	0 (5% Water)	100	0.7 ± 0.5		0 (5% Water)	100	4.0 ± 2.0	
	1	88	1.2 ± 1.3		0.3	91	3.5 ± 2.9	
	2.5	88	0.8 ± 0.8		1	80	5.3 ± 2.7	
	7.5	65	1.7 ± 1.2		2.5	73	11.7 ± 4.8 [*]	
	10	46	1.8 ± 1.2		5	61	15.0 ± 4.4 [*]	
	13.5	30	0.5 ± 0.8		10	49	19.7 ± 1.4 [*]	
	15	25	1.5 ± 0.5		20	38	1.3 ± 1.4	
	17.5	17	1.7 ± 1.6		30	32	Toxic	
	20	14	0.8 ± 1.2		0 (0.1% DMSO)	100	3.7 ± 2.0	
	0 (0.1% DMSO)	100	1.5 ± 1.6		TPA 0.05	120	25.7 ± 5.0 [#]	
MCA	1	47	21.7 ± 4.5 [#]					
Sodium orthovanadate (13721-39-6)				–				+
	0 (5% Water)	100	1.8 ± 2.8		0 (5% Water)	100	1.5 ± 0.8	
	0.01	95	1.0 ± 0.9		0.022	95	2.0 ± 2.3	
	0.022	95	1.3 ± 0.5		0.046	96	2.5 ± 1.6	
	0.046	92	0.8 ± 1.2		0.1	106	2.0 ± 1.3	
	0.1	92	0.5 ± 0.5		0.22	138	3.3 ± 0.8	
	0.22	110	0.8 ± 0.8		0.46	226	12.0 ± 4.7 [*]	
	0.46	107	0.5 ± 0.5		1	403	15.5 ± 4.3 [*]	
	1	40	0.2 ± 0.4		2.2	473	7.8 ± 2.9 [*]	
	2.2	0	0.2 ± 0.4		4.6	249	Toxic	
	0 (0.1% DMSO)	100	3.2 ± 1.5		0 (0.1% DMSO)	100	2.3 ± 1.8	
MCA	1	42	25.5 ± 2.6 [#]		TPA 0.05	227	9.2 ± 3.3 [#]	
Capsaicin (404-86-4)				–				–
	0 (0.1% DMSO)	100	0.5 ± 0.5		0 (0.1% DMSO)	100	2.0 ± 1.7	
	10	108	0.3 ± 0.8		1	82	1.8 ± 0.8	
	30	103	1.2 ± 1.2		5	95	2.7 ± 1.0	
	50	76	0.7 ± 0.8		10	109	2.2 ± 1.2	
	53	68	0.7 ± 1.0		20	100	2.0 ± 1.1	
	57	67	0.8 ± 0.8		25	95	2.0 ± 1.7	
	60	38	0.7 ± 0.8		30	90	1.2 ± 1.2	
MCA	1	45	13.8 ± 3.5 [#]		TPA 0.05	131	10.0 ± 2.2 [#]	
Ethidium bromide (1239-45-8)				–				–
	0 (5% Water)	100	2.8 ± 1.5		0 (5% Water)	100	3.5 ± 1.8	
	0.01	97	1.0 ± 1.1		0.0001	92	3.2 ± 2.0	
	0.03	103	1.2 ± 1.2		0.0003	101	4.0 ± 3.0	
	0.1	112	1.2 ± 1.0		0.001	101	3.7 ± 1.2	
	0.3	96	2.3 ± 1.2		0.003	106	1.8 ± 0.8	
	1	83	1.0 ± 0.6		0.01	100	0.0 ± 0.0	
	3	73	Toxic		0.03	113	0.0 ± 0.0	
	10	36	Toxic		0.1	104	Toxic	
	15	13	Toxic		0 (0.1% DMSO)	100	3.2 ± 1.8	
	30	1	Toxic		TPA 0.05	176	26.5 ± 3.9 [#]	
	0 (0.1% DMSO)	100	2.2 ± 1.3					
MCA	1	60	27.5 ± 2.4 [#]					
Salicylic acid (69-72-7)				–				–
	0 (0.1% DMSO)	100	3.5 ± 1.4		0 (0.1% DMSO)	100	6.7 ± 1.5	
	100	117	3.2 ± 2.6		50	96	5.3 ± 2.3	
	200	119	1.0 ± 1.3		100	94	7.7 ± 2.3	
	250	120	1.8 ± 1.3		200	86	7.5 ± 2.1	
	300	128	0.5 ± 0.5		300	85	6.0 ± 2.4	
	400	103	2.2 ± 0.8		400	75	4.5 ± 1.5	
	600	67	1.3 ± 1.4		600	83	6.8 ± 2.1	
	800	27	0.3 ± 0.5		800	80	3.3 ± 2.2	
MCA	1	63	30.5 ± 7.6 [#]		TPA 0.05	211	28.7 ± 6.0 [#]	

^a CAS registry number in parentheses.

^b % of cell growth compared to that of solvent control.

^c Average number of transformed foci/well ± SD.

^d Positive or negative call for the test chemical based on the assay data presented.

^e Solvent control: final solvent concentration of the working culture media in parentheses.

^f Positive control in the initiation assay.

^g Positive control in the promotion assay.

^h Solvent control for the positive control (MCA in the initiation assay or TPA in the promotion assay).

^{*} $p < 0.05$; Dunnett test, vs solvent control.

[#] $p < 0.05$; t -test, vs DMSO (the solvent of MCA and TPA).

Table 2
The performance of Bhas 42 cell transformation assay for the prediction of chemical carcinogenicity.

		In vivo carcinogenicity		Total		
		Carcinogen	Non-carcinogen			
(a) Contingency table of the results in the Bhas 42 cell transformation assay					(b) Performance	
Bhas 42 cell transformation assay	+	38	6	44	Concordance	78%
	–	14	31	45	Sensitivity	73
Total		52	37	89	Specificity	84
					Positive predictivity	86
					Negative predictivity	69
					False negative	27
					False positive	16

week till day 21 (Fig. 1). Each assay was conducted including TPA (0.05 µg/mL) as the positive control and 0.1% DMSO as its negative control.

2.3.3. Counting of transformed foci and statistical analysis

The transformed foci were judged on the basis of the morphological characteristics: (a) more than 100 cells, (b) spindle-shaped cells different from the contact-inhibited monolayer cells, (c) deep basophilic staining, (d) random orientation of cells at the edge of foci, (e) dense multilayering of cells and (f) invasive growth into the monolayer of surrounding contact-inhibited cells. The number of transformed foci in every well was recorded and a statistical analysis for the increase of the transformed foci was performed by multiple comparison using the Dunnett method ($p < 0.05$).

2.4. Cell growth assay

The cell growth assays, using a crystal violet (CV) staining method, were performed prior to the transformation assays to determine the doses applicable to the Bhas 42 cell transformation, and also concurrently with every transformation assay (every initiation and promotion assay) to estimate the effect of each treatment on the cell growth and survival. The cells were seeded and treated with a chemical in the same manner as in the initiation assay or in the promotion assay. Three wells were prepared for each test concentration. The cultures were fixed with 10% formalin on day 7 and stained with a 0.1% CV solution. CV was extracted from the stained cells with 50% ethanol containing 0.03 M sodium citrate and 0.02 M hydrochloric acid. The optical density of extract was measured at 540 nm. The survival rate of cells treated with a chemical was expressed as a percentage of the control cells.

2.5. Dose setting for transformation assays

Five or more concentrations were set up based on the results of cell growth assays.

In the initiation assay, the concentrations were determined to cover a range from little or no toxicity to the highest toxicity (less than 20% survival compared to the control cultures). In practice, one dose below NOEL, two doses between NOEL and IC_{50} and two doses between IC_{50} and IC_{90} were arranged, at least.

In the promotion assay, the test concentrations were selected to cover a range from little effect on cell growth to growth enhancement, for the chemicals that exhibited marked growth enhancement. In practice, one dose below NOEL, three doses in the range of growth enhancement and one dose in the range of weak growth inhibition were arranged, at least. For the chemicals that did not induce marked growth enhancement, the test concentrations were selected to range from a dose two or three levels lower than the no-effect concentration to that showing a survival less than 50%. In practice, two doses below NOEL, two doses between NOEL and IC_{50} and one dose above IC_{50} were arranged, at least.

For a chemical which caused a sharp decline of cell growth within a narrow concentration range, one or two more doses outside the predicted concentration range were set up as a precaution against the fluctuation of cell response among experiments. For compounds with little cytotoxicity, the highest concentration was 5 mg/mL or 10 mM, whichever was the lowest.

3. Results

The individual results of 98 chemicals in the Bhas 42 cell transformation assay are presented in Table 1. The tested chemicals are listed being classified into carcinogens, non-carcinogens and chemicals of unknown carcinogenicity according to the existing data of rodent *in vivo* carcinogenicity and human epidemiology. Tumor promoters are assorted into carcinogens. The bases of classification are shown in the footnote of Table 3. The results in the Bhas 42 cell transformation assays were judged positive when there existed

two or more doses that induced statistically significant increases of transformed foci, and negative when there was no dose showing statistically significant increase of foci. When the statistically significant increase was at only one dose, the assay result was regarded as equivocal, and then the initiation or promotion assay together with the concomitant cell growth assay was repeated including the positive dose in the first assay. The chemical was judged to be positive, when the chemical again statistically significantly increased the number of transformed foci at one or more concentrations in the second assay (Cyclosporin A and Dichlorvos). As a rule, the assays were carried out at the concentrations less than 5 mg/mL or 10 mM, whichever was the lowest, when the test chemical did not cause cell growth inhibition or enhancement. In Table 1 the molar concentrations are indicated in the parentheses for the concentrations that are equal to 10 mM or exceptionally exceed 10 mM.

Among the 52 tested carcinogens, 2-acetylaminofluorene, benz[a]anthracene and phorbol 12,13-didecanoate were judged positive both in the initiation assay and in the promotion assay. IQ, 5-azacytidine, barium chromate, benzo[a]pyrene, cyclophosphamide, cyclosporin A, 2,4-diaminotoluene, dibenz[a,h]anthracene, melphalan, MCA, mitomycin C, MNNG, sterigmatocystin and thio-TEPA were called positive in the initiation assay. Cadmium chloride, chenodeoxycholic acid, 4-chloro-*o*-toluidine hydrochloride, cholic acid, deoxycholic acid, dichlorvos, epichlorohydrin, *D*-limonene, lithocholic acid, methapyrilene hydrochloride, methylarsonic acid, mezerein, 2-naphthylamine, quercetin, sodium arsenate, sodium arsenite, sodium saccharin, styrene oxide, TPA, *o*-toluidine and zinc chloride were positive in the promotion assay. However, *o*-anisidine, benzene, cobalt sulfate heptahydrate, diethylstilbestrol, dimethylarsinic acid, 1,4-dioxane, ethyl carbamate, formaldehyde, furylfuramide, methyl carbamate, nickel (II) chloride, nickel monoxide, phenobarbital sodium salt and *p*-toluidine were negative in either assay.

Out of the 37 non-carcinogens, 4-acetylaminofluorene, acid red 14, ampicillin sodium salt, anthracene, L-ascorbic acid, aspartame, benzoic acid, caffeine, caprolactam, 2-chloroethanol, chromium (III) chloride, 2,6-diaminotoluene, diazepam, *N,N*-dimethylformamide, eugenol, HC blue no. 2, hydrocortisone, *D*-mannitol, methotrexate, 1-naphthylamine, phenanthrene, phenol, *p*-phenylenediamine dihydrochloride, phthalic anhydride, rotenone, sodium chloride, sodium nitrite, sunset yellow FCF, thiabendazole, *m*-toluidine and triphenyltin hydroxide were judged negative both in the initiation assay and in the promotion assay. However, barium chloride dihydrate, *tert*-butylhydroquinone, 8-hydroxyquinoline, sodium fluoride and tetracycline hydrochloride were positive in the promotion assay, and propyl gallate was positive both in the initiation and in the promotion assay.

The number of chemicals of unknown carcinogenicity was 9. Valproic acid and sodium valproate were positive in both assays. 2,3-Diaminotoluene, 2,5-diaminotoluene dihydrochloride,

Table 3
Comparison of results in the Bhas 42 cell transformation assay with those in genotoxicity assays.

Chemical	Bases of classification ^a	Bhas 42 CTA ^b		Ames ^{c,d}	MLC ^e	Chrom. Ab. ^{c,f}		MN ^{c,g}
		Initiation	Promotion			<i>In vitro</i>	<i>In vivo</i>	
<i>Carcinogens</i>								
2-Acetylaminofluorene	TA#31 ^h	+	+	+	+,+,+	+	+	+
Benz[<i>a</i>]anthracene	TA#31	+	+	+,+,+	+,+,+,+/-	+/-	+/-	+
IQ	TA#31	+	-	+				-(CCRIS ⁱ)
5-Azacytidine	TA#31	+	-	+,+/-	+,+,+,+	+,+/-		+
Barium chromate	TA#31	+	-	-				
Benzo[<i>a</i>]pyrene	TA#31	+	-	+,+	+,+,+	+	+	+,+
Cyclophosphamide	TA#31	+	-	+,+/-,+/-	+,+,+,+	+,+	+,+/-	+,+,+
Cyclosporin A	TA#31	+	-	-				-
2,4-Diaminotoluene	TA#31	+	-	+,+	+	+		-
Dibenz[<i>a,h</i>]anthracene	TA#31	+	-	+,+/-,+	+,+	+		
Melphalan	TA#31	+	-	+,+,+,+/-	+	+,+	+	+
MCA	TA#31	+	-	+,+	+	-,-		
Mitomycin C	TA#31	+	-	+(CCRIS)	+,+	+,+	+,+,+	+,+,+
MNNG	TA#31	+	-	+,+	+,+	+/-		+
Sterigmatocystin	IARC ^j	+	-	+(CCRIS)				+(CCRIS)
Thio-TEPA	TA#31	+	-	+		+	+	+
Cadmium chloride	TA#31	-	+	-,-	+	+,+		-
Chenodeoxycholic acid	Orig ^k , TP ^{l,m}	-	+	-/+ (CCRIS)				
4-Chloro- <i>o</i> -toluidine HCl	TA#31	-	+	-	+	+w ⁿ		
Cholic acid	TA#31, TP	-	+	+/-				
Deoxycholic acid	CCRIS, TP	-	+	-/+ (CCRIS)				
Dichlorvos	TA#31	-	+w	+,+	+,+,+	+	-,-	-
Epichlorohydrin	TA#31	-	+	+,+,+/-	+,+	+,+/-	+,+/-	-
D-Limonene	NTP(347) ^o	-	+	-,-	-,+	-		
Lithocholic acid	Orig., TP ^p	-	+	-,-	+/-,+/-	-		
Methapyrilene HCl	TA#31	-	+	-	-,+,+/-	+		
Methylarsonic acid	Review, TP ^q	-	+	-(Orig. ^r)	+(CCRIS)	+(Orig. ^r)		+(CCRIS)
Mezerein	TA#31, TP	-	+	-,-				
2-Naphthylamine	TA#31	±	+	+,+,+/-,+,+/-	+,+,+	+,+		-,+,-,+
Phorbol 12,13-didecanoate	CCRIS, TP	+	+					
Quercetin	TA#31	-	+	+,+,+/-,+/-	+	+,+	+	+,+
Sodium arsenate	TA#31	-	+	-(CCRIS)	-(CCRIS)			-
Sodium arsenite	TA#31	±	+	-	+,+			+
Sodium saccharin	TA#31	-	+	-,-				
Styrene oxide	TA#31	-	+	+,+,+/-,+/-	+,+,+	+	+/-	-
TPA	TA#31, TP	±	+	-				
<i>o</i> -Toluidine	TA#31	-	+	+,+	+,+/-,+/-	+,+/-		+/-,-
Zinc chloride	TA#31, TP	-	+	+				
<i>o</i> -Anisidine	TA#31	-	-	+	+,+,+	+		
Benzene	TA#31	-	-	-	-,+,+/-,+/-	-,+,+	+,+	+,+
Cobalt sulfate	TA#31	-	-	+w				
Diethylstilbestrol	TA#31	-	-	-,-,-,-	+,+,+/-,+/-	+,+/-	+	+/-,+,-
Dimethylarsinic acid	CCRIS	-	-	-(CCRIS)	+(CCRIS)	+(Orig. ^r)	+(Review ^d)	+/- (CCRIS)
1,4-Dioxane	TA#31	-	-	-,-	-,-			-
Ethyl carbamate	TA#31	-	-	+/-,-,-	-	-,+		+,+
Formaldehyde	TA#31	-	-	+,+/-,+/-	+	+,+	+/-	+/-
Furylfuramide	TA#31	-	-	+,+,+	+,+	+		
Methyl carbamate	TA#31	-	-	-,-,+	-,-	-		-
Nickel (II) chloride	TA#31	-	-	-	+	+	+	-
Nickel monooxide	TA#31	-	-	-				-
Phenobarbital sodium	TA#31	-	-	-,+/-				
<i>p</i> -Toluidine	Orig. ^s	-	-	-(CCRIS)				
<i>Non-carcinogens</i>								
4-Acetylaminofluorene	TA#31	-	-	+	+/-,+,+/-	-	-	-
Acid red 14	TA#31	-	-	-,-	-,-	-		-
Ampicillin	TA#31	-	-	-,-	-,-	-		-
Anthracene	TA#31	-	-	+w, -,+/-	+,+,+,+/-	-		
L-Ascorbic acid	TA#31	-	-	+w/-,-			+	+
Aspartame	CCRIS	-	-	-(NTP) ^t				-(NTP) ^t
Benzoin	TA#31	-	-	+w/-,+/-	+,+,-,+/-	-		-
Caffeine	TA#31	-	-	-,-,-		+		+
Caprolactam	TA#31	-	-	-,-	-,-,-,-	-	-	-,-
2-Chloroethanol	TA#31	-	-	+,+,+/-	+,+	+	-	-
Chromium(III) Chloride	TA#31	-	-	-		+		
2,6-Diaminotoluene	TA#31	-	-	+				
Diazepam	TA#31	-	-	-,-				
<i>N,N</i> -Dimethylformamide	TA#31	-	-	-,-	+/-,-,-,+/-	-		
Eugenol	TA#31	-	-	-	+,+,+	+		-,+
HC Blue 2	TA#31	-	-	+	+,+	-	-	
Hydrocortisone	TA#31	-	-	-		+		
D-Mannitol	TA#31	-	-	-,-	-,-	-	-	-
Methotrexate	TA#31	-	-	-	+,+	+	+	+

tive or discordant in the Ames test. They are barium chromate, cyclosporin A, cadmium chloride, chenodeoxycholic acid, 4-chloro-*o*-toluidine hydrochloride, cholic acid, deoxycholic acid, α -limonene, lithocholic acid, methapyrilene hydrochloride, methylarsonic acid, mezerein, sodium arsenate, sodium arsenite, sodium saccharin, TPA and *o*-toluidine. The Ames-negative and Ames-discordant carcinogens except barium chromate and cyclosporin A were judged positive in the promotion assay. They included TPA and mezerein which are typical tumor promoters in mouse skin [19,20], bile acids which are regarded as endogenous tumor promoters [21–23] and sodium saccharin which has been demonstrated to be a promoter of bladder carcinogenesis in rats [24]. Phorbol 12,13-didecanoate that was positive in the promotion assay is also a tumor promoter in mouse skin [25,26]. However, we did not underline it in Table 3 because of the absence of published data in the Ames test and the other genotoxicity tests. In the previous papers, other tumor promoters such as okadaic acid [11,16,27], *o,p'*-dichlorodiphenyltrichloroethane (*o,p'*-DDT) [11], *p,p'*-dichlorodiphenyl trichloroethane (*p,p'*-DDT) [11], fumonisin B1 [28] and T-2 toxin [28] were also reported to be positive in the promotion assay of Bhas 42 cell transformation. These facts indicated that the promotion assay of Bhas 42 cell transformation detected known tumor promoters and suggested that the Bhas 42 cells behaved as initiated cells in the transformation assay. In this paper, arsenate and arsenite were judged positive in the promotion assay. Inorganic arsenics have epidemiologically been defined to be carcinogenic in the bladder, lung and skin of human and found to increase cancer incidence in various experimental animal models [29]. However, their mutagenicity is negative in bacteria and weekly positive in mammalian cells *in vitro*, though they have been reported to induce chromosomal aberrations, aneuploidy and micronuclei formation [29–31]. Thus, the Bhas 42 cell transformation assay can detect a considerable number of the carcinogens that are negative or difficult to be detected in the conventional genotoxicity assays. The carcinogens escaping scrutiny from *in vitro* screening would be diminished in number if the Bhas 42 cell transformation assay is performed in addition to genotoxicity assays to predict chemical carcinogenicity. Incidentally, we have demonstrated that the Bhas 42 cells retain the transfected *v-Ha-ras* gene and express its mRNA at the same level as *c-Ha-ras* mRNA (in preparation for publication). Our presumption is that the expression of transfected active oncogene is involved in the high sensitivity of Bhas 42 cells to the non-genotoxic carcinogens in transformation.

According to a review paper [5], the performance indices of the Ames, mouse lymphoma, *in vitro* and *in vivo* chromosomal aberration and micronucleus assays to predict rodent carcinogenicity of chemicals are as follows: concordance, 52–72%; sensitivity, 39.5–86%; specificity, 26–79%; positive predictivity, 76.5–83%; negative predictivity, 27–45%; false negative, 14–60.5%; false positive, 21–74%. As compared to those values, the concordance, specificity, negative predictivity and false positive of Bhas 42 cell transformation assay were superior and the other performances were equivalent to those of genotoxicity assays. It must be mentioned, however, that the chemical sets used for the calculation of performances are different between the Bhas 42 cell transformation assay in the present paper and the genotoxicity assays in the review paper.

The inorganic compounds tested in this study were 13 in all. This number may not be enough to evaluate the performance of Bhas 42 cell transformation assay for the prediction of carcinogenicity of inorganic compounds. When we dared to calculate the performances for inorganic chemicals, the concordance, sensitivity, specificity, positive predictivity, negative predictivity, false negative and false positive were 62%, 63%, 60%, 71%, 50%, 38% and 40%, respectively (the contingency and performance tables are not shown). The organic carcinogens and non-carcinogens tested were

76 in total. For the organic chemicals, the concordance, sensitivity, specificity, positive predictivity, negative predictivity, false negative and false positive were 80%, 75%, 88%, 89%, 72%, 25% and 13%, respectively (the contingency and performance tables are not shown). The Bhas 42 cell transformation assay may predict the carcinogenicity of inorganic chemicals with a little lower probability than that of organic chemicals.

The Bhas 42 cell transformation assay judged diethylstilbestrol negative in the present study, and also called estradiol and zearalenone as negative in the previous reports [11,27,28]. This assay may be insensitive to carcinogenicity of female sex hormones. Progesterone, a corpus luteum hormone, has been reported to be positive in the promotion assay [11].

The Bhas 42 cell transformation could detect a considerable number of Ames-negative or Ames-discordant carcinogens. Its performances to predict the carcinogenicity of chemicals are superior or equivalent to those of genotoxicity assays. As shown in Table 3, any of the genotoxicity assays is not perfect to predict the carcinogenicity of chemicals. We consider that the carcinogens escaping from *in vitro* screening would be reduced in number and the accuracy of prediction for chemical carcinogenicity would be improved by introducing the Bhas 42 cell transformation into the battery of *in vitro* assays.

Conflict of interest statement

The authors declare that there are no conflicts of interest.

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Table 3 (Continued)

Chemical	Bases of classification ^a	Bhas 42 CTA ^b		Ames ^{c,d}	ML ^{c,e}	Chrom. Ab. ^{c,f}		MN ^{c,g}
		Initiation	Promotion			<i>In vitro</i>	<i>In vivo</i>	
1-Naphthylamine	TA#31	–	–	+,+,+/-		+,+/-		+,+
Phenanthrene	TA#31	–	–	+w,-,+/-		–		
Phenol	TA#31	–	–	–	+,+,+	+,+	+	+
<i>p</i> -Phenylenediamine 2HCl	TA#31	–	–	+	+,eq. ^u ,+	+		
Phthalic anhydride	TA#31	–	–	–,-	+	–		
Rotenone	TA#31	–	–	–	+	+		+
Sodium chloride	TA#31	–	–	–	+/-			
Sodium nitrite	Orig. ^v ,NTP(495)	–	–	+,+	+	+	+	–,+,+
Sunset yellow FCF	TA#31	–	–	–	+,-	–	–	–
Thiabendazole	TA#31	–	–	+				+
<i>m</i> -Toluidine	Orig. ^s	–	–	–(CCRIS)				
Triphenyltin hydroxide	TA#31	–	–	–,-	+	–		
Barium chloride	TA#31	–	+	–,-	+	–		
<i>tert</i> -Butylhydroquinone	TA#31	–	+	–,-		+	–	–
8-Hydroxyquinoline	TA#31	–	+	+,+	+,+	+w	–	–
Propyl gallate	TA#31	+	+	–,-	+,+	+	+	+/-
Sodium fluoride	TA#31	–	+	–,-	+,+,+	+/-	–	–,+
Tetracycline HCl	TA#31	–	+	–,-	–,+/-	–		
<i>Chemicals of unknown carcinogenicity</i>								
Sodium valproate		+	+					
Valproic acid		+	+				+/- (human) Orig. ^w	
2,3-Diaminotoluene		–	+	+(CCRIS)				
2,5-Diaminotoluene 2HCl		–	+					
3,4-Diaminotoluene		–	+	+(CCRIS)				
Sodium orthovanadate		–	+					–(CCRIS)
Capsaicin		–	–	+/- (CCRIS)	+(CCRIS)	–(CCRIS)		–(CCRIS)
Ethidium bromide		–	–	+(CCRIS)	+/- (CCRIS)			–(CCRIS)
Salicylic acid		–	–	+/- (CCRIS)				

The underline shows the carcinogens that were positive in the Bhas 42 cell transformation assay but are negative or discordant in the Ames test.

^a The databases, or review and original papers referred to for the *in vivo* carcinogenic potency of chemicals.

^b The assay results obtained in the present study.

^c Assay results obtained from existing databases, or review and original papers. Most results are taken from a review paper, TA#31^h. In TA#31, several databases are consulted for each chemical. The results listed in TA#31 are arranged in this table so that “+” means “positive in one database”, “+,+” means “positive in two different databases”, “+,+,+” means “positive in three different databases”, “+,-,-” means “positive in a database and negative in two other databases”, “+/-” means that there are diverging results inside a database and so on. The assay results which do not exist in TA#31 are based on CCRISⁱ, or review and original papers, and the sources of those data are indicated in the parentheses.

^d Ames test.

^e Mouse lymphoma test.

^f Chromosomal aberration test.

^g Micronucleus test.

^h “OECD Environment, Health and Safety Publications, Series on Testing and Assessment No. 31: Detailed Review Paper on Cell transformation Assays for Detection of Chemical Carcinogens” [5].

ⁱ The Chemical Carcinogenesis Research Information System.

^j IARC Monographs.

^k Original paper.

^l *In vivo* tumor promoter.

^m Ref. [22].

ⁿ Weakly positive.

^o The United State National Toxicology Program. The NTP technical report number was indicated in the parenthesis.

^p Ref. [21].

^q Ref. [30].

^r Ref. [32].

^s Ref. [33].

^t Studies in Genetically Modified Models, GMM-01.

^u Equivocal.

^v Ref. [34].

^w Ref. [35].

3,4-diaminotoluene and sodium orthovanadate were positive in the promotion assay. Capsaicin and ethidium bromide were negative in either assay.

The performance indices of Bhas 42 cell transformation assay for predicting the carcinogenicity of chemicals were calculated from the assay results of known carcinogens and non-carcinogens. As shown in Table 2, we applied the Bhas 42 cell transformation assay to 52 *in vivo* carcinogens and obtained positive judgments from 38 carcinogens and negative judgments from 14. We tested 37 *in vivo* non-carcinogens and 6 were positive and 31 were negative. Consequently, concordance was 78%, sensitivity 73%, specificity 84%, positive predictivity 86%, negative predictivity 69%, false negative 27% and false positive 16%.

4. Discussion

The results of 98 chemicals tested in the present study were summarized and compared with existing data for genotoxicity in Table 3. Among genotoxicity assays, the Ames test is the most popular and has been applied to the largest number of chemicals including carcinogens and non-carcinogens. Of the 52 carcinogens tested in this assay, about half are negative or discordant in the Ames test from our consulting of databases or review of original papers. In the Bhas 42 cell transformation assay, 17 of the Ames-negative or Ames-discordant carcinogens were judged positive. In Table 3, we underline the carcinogens that were positive in the Bhas 42 cell transformation assay but are nega-

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